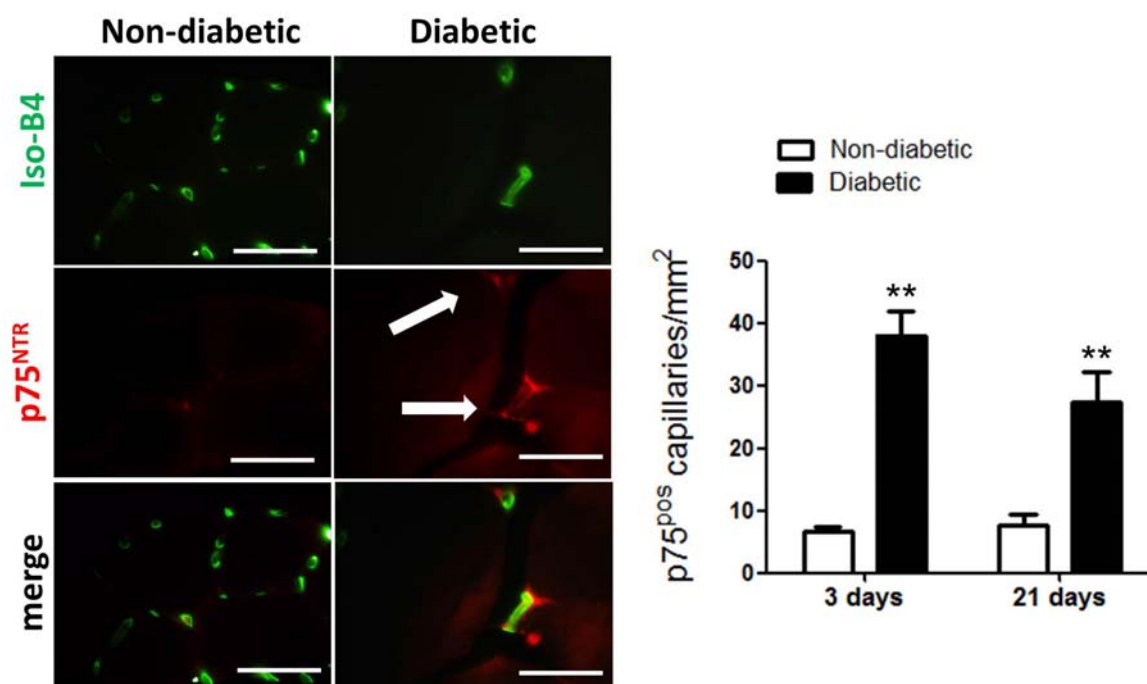
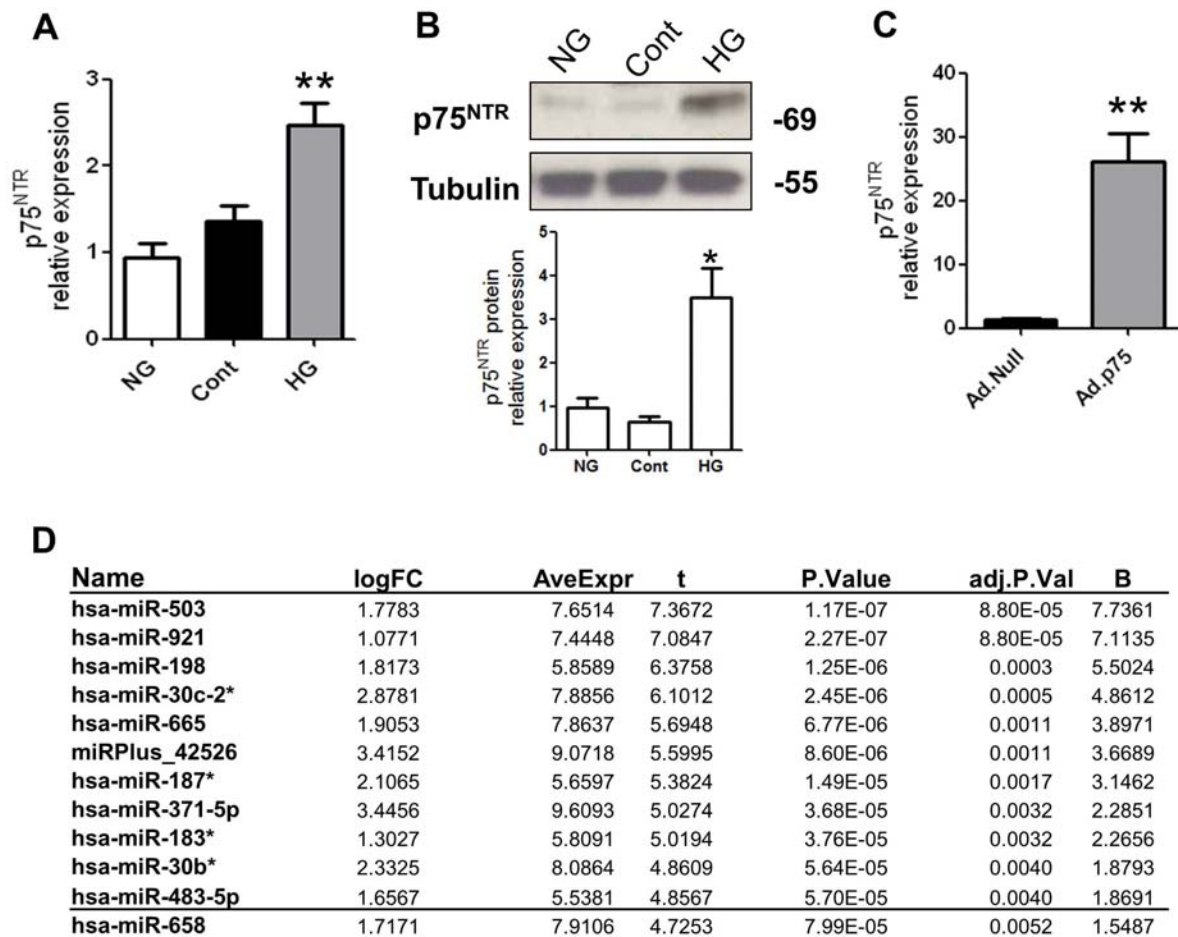


## SUPPLEMENTARY FIGURES



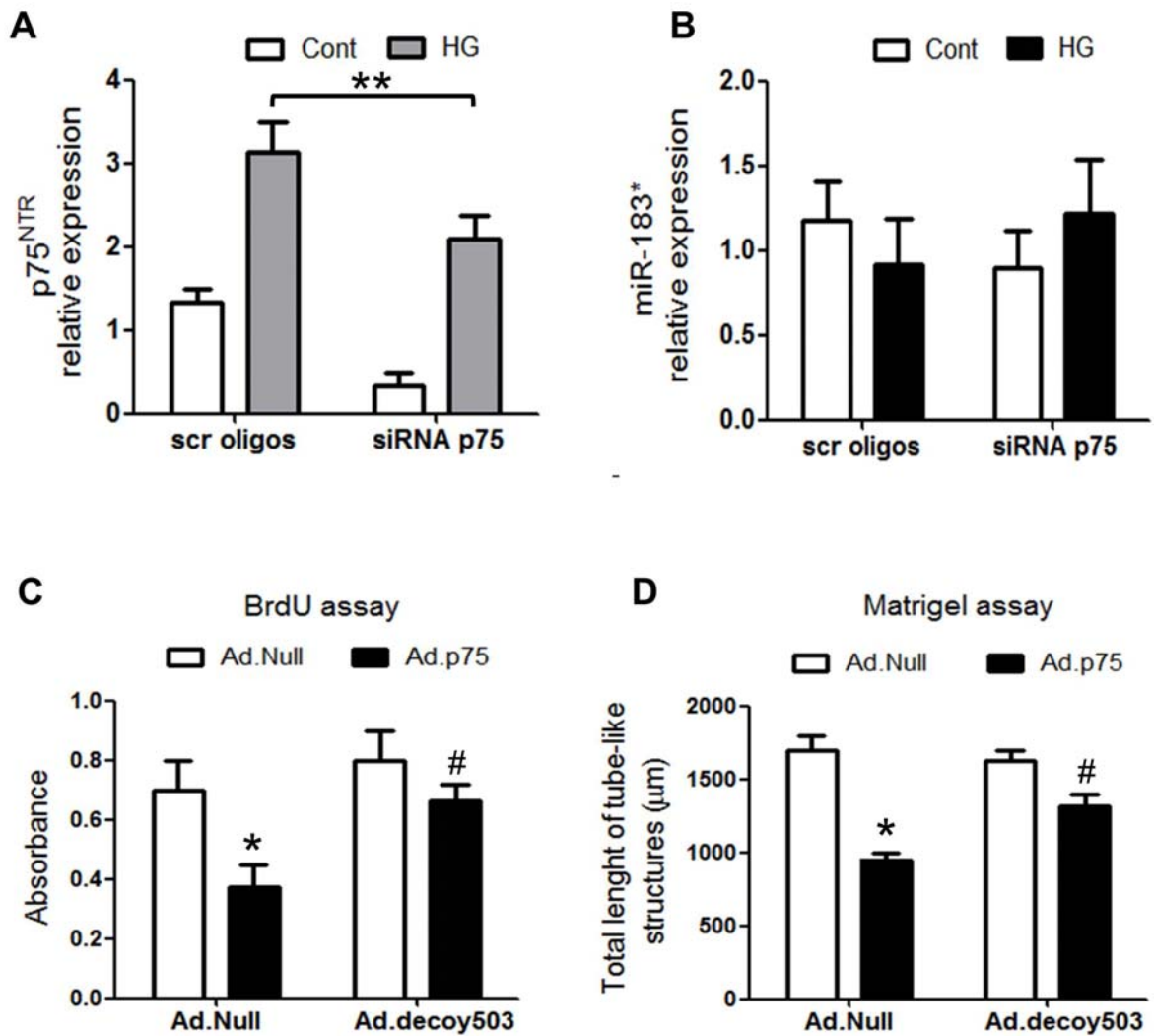
### Supplementary Figure 1: Expression of p75<sup>NTR</sup> in microvascular ECs of limb muscle

**A**, Representative pictures of ischemic muscular sections from diabetic and non-diabetic mice (n=6/group) stained with an antibody for mouse p75<sup>NTR</sup> (red fluorescence) and with the Isolectin-B4 (Iso-B4; green fluorescence) at 21 days after ischemia. Capillary EC which are positive for p75<sup>NTR</sup> express yellow fluorescence (from merging of the red and green fluorescence) and are pointed by arrows. Scale bar: 50 $\mu$ m **B**, Bar graph quantifies the density of capillaries expressing p75<sup>NTR</sup> in ischemic limb muscles of normoglycemic and diabetic mice. Values are means $\pm$ SEM (n=6/group). \*\*P<0.01 vs. non-diabetic mice. Unpaired two-tailed Student's t-test was applied when comparing two groups. All values are mean $\pm$ SEM of three independent experiments.



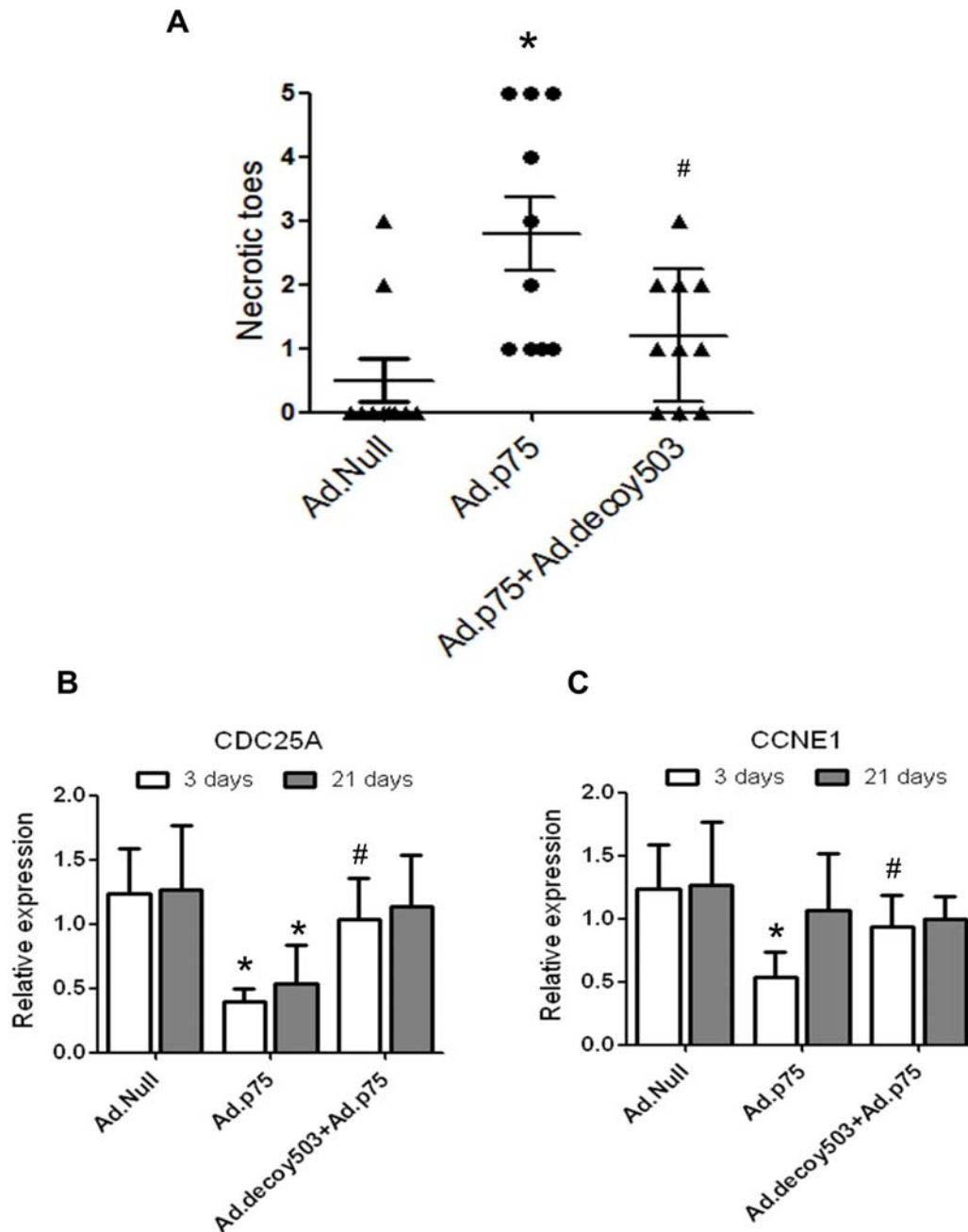
### Supplementary Figure 2: miR-503 expression in microvascular ECs and miRNA array analysis

**A**, HMVECs were exposed to high-glucose (HG), cultured in normal glucose (NG) or osmotic control (Cont; L-Glucose) conditions for 24hrs and expression of p75<sup>NTR</sup> was analyzed by qPCR. \*\*p<0.01 vs NG or Cont (n=3), unpaired two-tailed Student's t-test; **B**, Representative Western blot for p75<sup>NTR</sup> and tubulin and protein quantification \*p<0.05 vs NG or Cont (n=3), unpaired two-tailed Student's t-test **C**, Expression of p75<sup>NTR</sup> in HUVECs after transduction with *Ad.p75*. Gene expression was normalized to 18S expression. \*\*p<0.01 vs. *Ad.Null* (n=3), unpaired two-tailed Student's t-test; All values are mean±SEM of three independent experiments. **D**, Ranked list of miRNA differentially expressed in HUVECs transduced with *Ad.p75* or *Ad.Null* (as control). miRNAs were ranked by differential expression and statistical significance. The default table of Limma is ranked by B-statistics (B), closely related to the adjusted p-value. The B-statistic used for this analysis is the log-odds that that gene is differentially expressed and it is automatically adjusted for multiple testing by assuming that 1% of the genes are expected to be differentially expressed.



**Supplementary Figure 3: The p75<sup>NTR</sup> receptor regulates EC function**

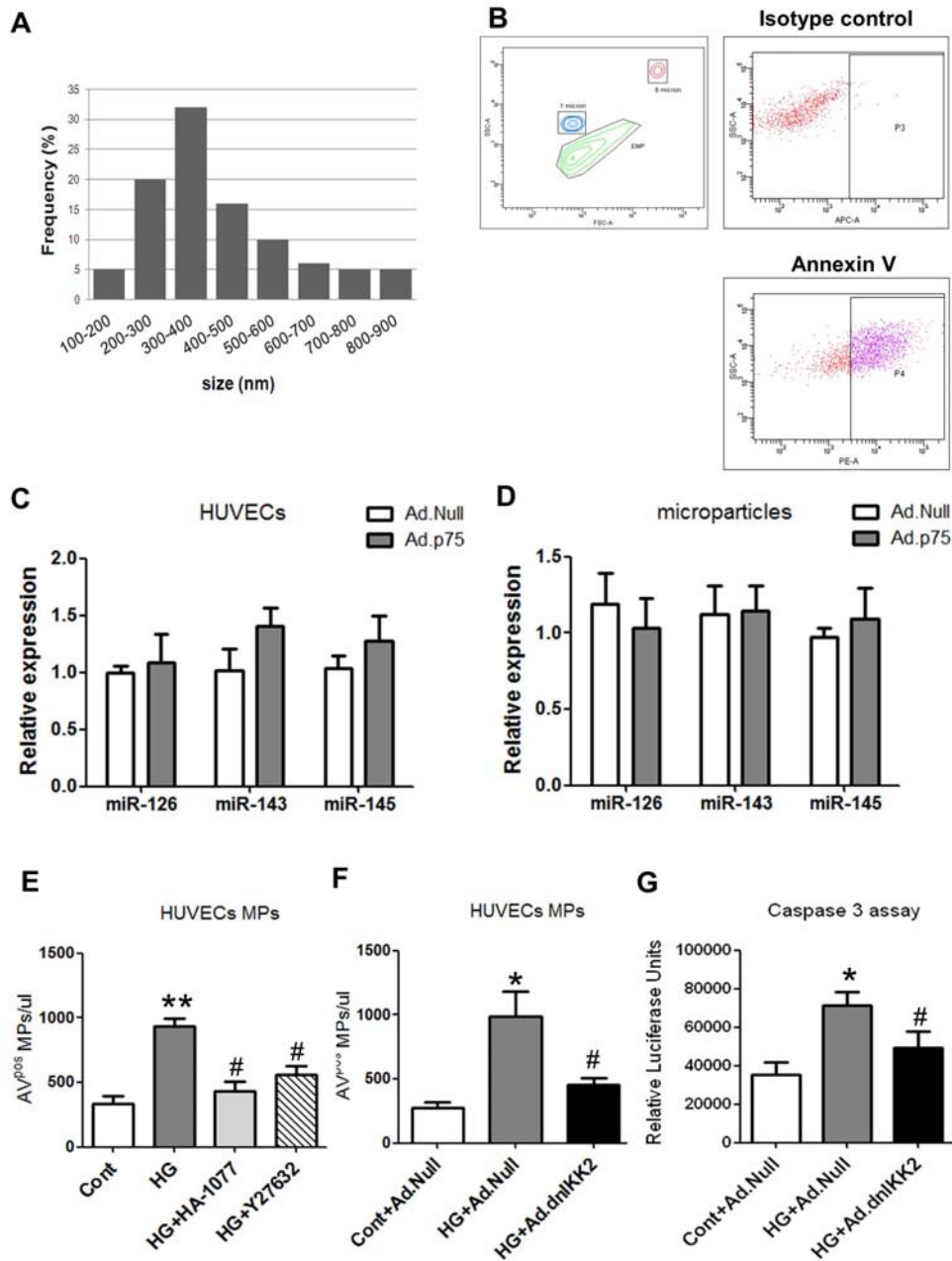
**A**, p75<sup>NTR</sup> expression in HUVECs transfected with siRNA for p75<sup>NTR</sup> or scramble oligos (scr oligos) and treated with HG or L-Glucose (Cont) for 24hrs; \*\*p<0.01; two-way ANOVA (n=5); **B**, miR-183\* expression in the conditions described above; **E**, Proliferation assay in HUVECs transduced with *Ad.Null*, *Ad.p75* or *Ad.dec503* adenovirus; **F**, Matrigel assay in the same conditions described above. Results are described as total tube length. For **E** and **F**: \*p<0.05 vs. *Ad.Null*, #p<0.05 vs. *Ad.p75* (n=3), unpaired two-tailed Student's t-test. All values are mean±SEM of three independent experiments.



**Supplementary Figure 4: Clinical outcomes and target gene expression after p75<sup>NTR</sup> gene transfer in limb muscles of ischemic mice**

**A**, Analysis of necrotic toes of non-diabetic mice injected with *Ad.p75*, *Ad.Null* or *Ad.p75* and *Ad.decoy503* together in ischemic limb muscles (n=12/group). \*p<0.05 vs. *Ad.Null*, #p<0.05 vs. *Ad.p75*, necrotic toes endpoint was analyzed using Cochran-Armitage trend test  
**B**, Relative expression of *CDC25A* and *CCNE1* in ischemic adductors of non-diabetic mice given *Ad.p75*, *Ad.Null* or *Ad.p75* and *Ad.decoy503* together (n=6/group). \*p<0.05 vs. *Ad.Null*, #p<0.05 vs. *Ad.p75*, Mann-Whitney nonparametric test was applied. All values are mean±SEM of three independent experiments.



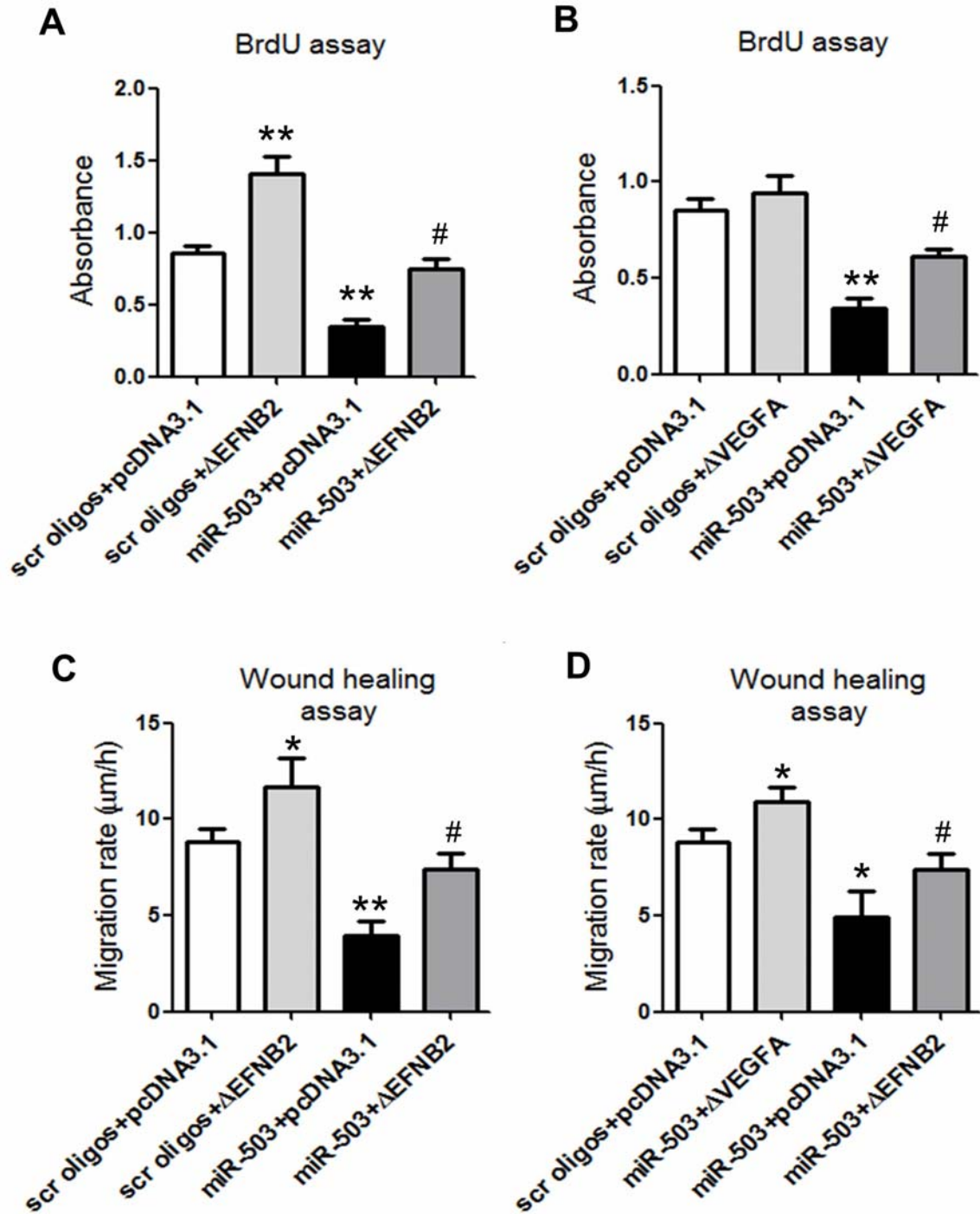


### Supplementary Figure 6: Endothelial microparticles analysis

**A**, Size distribution of isolated MPs from p75<sup>NTR</sup>-transduced HUVECs (n=500 cells). **B**, Flow cytometric analysis of MPs from p75<sup>NTR</sup>-transduced HUVECs. p75<sup>NTR</sup> increased the release of HUVEC-derived MPs expressing AnnexinV (AnnV<sup>Pos</sup>) on the MP-surface; **C**, Expression of miR-126, miR-143 and miR-145 in HUVECs and in **D**, isolated MPs from medium of p75<sup>NTR</sup>-transduced HUVECs. **E**, HUVECs were cultured in HG and osmotic control medium (Cont) and treated with Rho kinase inhibitors Y27632 (10μM) or HA-1077 (10μM) and MPs were collected from the medium by centrifugation and analyzed for Annexin V by flow cytometry. \*\*p<0.01 vs Cont; #p<0.05 vs HG (n=3), unpaired two-tailed Student's t-test. **F**, HUVECs were cultured in same conditions described above and transduced with the *Ad.Null* or *Ad.dnIKK2*. **G**, Caspase-3 activity in HUVECs treated as described above, For **F** and **E**: \*p<0.05 vs Cont/*Ad.Null*; #p<0.05 vs HG/*Ad.Null* (n=3), unpaired two-tailed Student's t-test. All values are mean±SEM of three independent experiments.



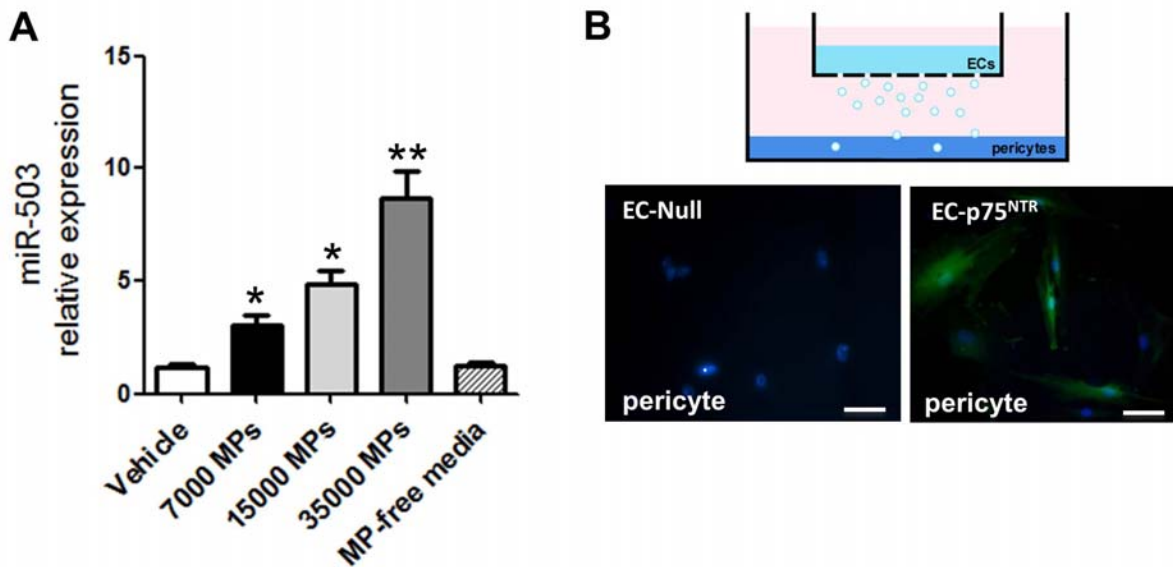




**Supplementary Figure 8: Overexpression of 3'UTR-mutated target gene restores proliferation and migration in pericytes overexpressing miR-503**

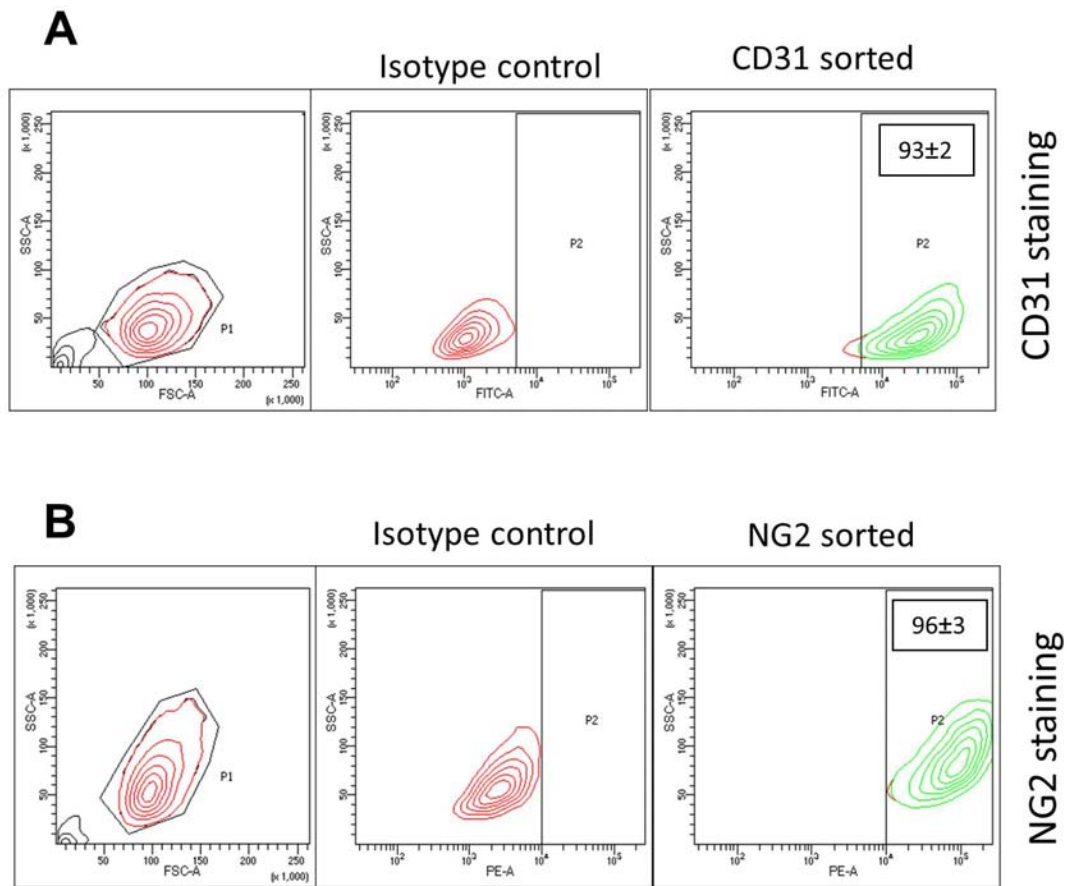
HUVECs expressing either  $\Delta$ EFNB2 or  $\Delta$ VEGFA or vector alone (pcDNA3.1) were transfected with mature miR-503 or a scramble sequence and further cultivated for 24 hrs. Then, proliferation (A and B) or migration (C and D) were assayed.  $\Delta$ EFNB2 or  $\Delta$ VEGFA partially restored the proliferation (A and B, respectively) and migration (C and D, respectively) in HUVEC transfected with pre-miR-503. \*\*p<0.01 or \*p<0.05 vs. scr oligos+pcDNA3.1; #p<0.05 vs. miR-503+ pcDNA3.1 (n=5), unpaired two-tailed Student's t-test. All values are mean $\pm$ SEM of three independent experiments.





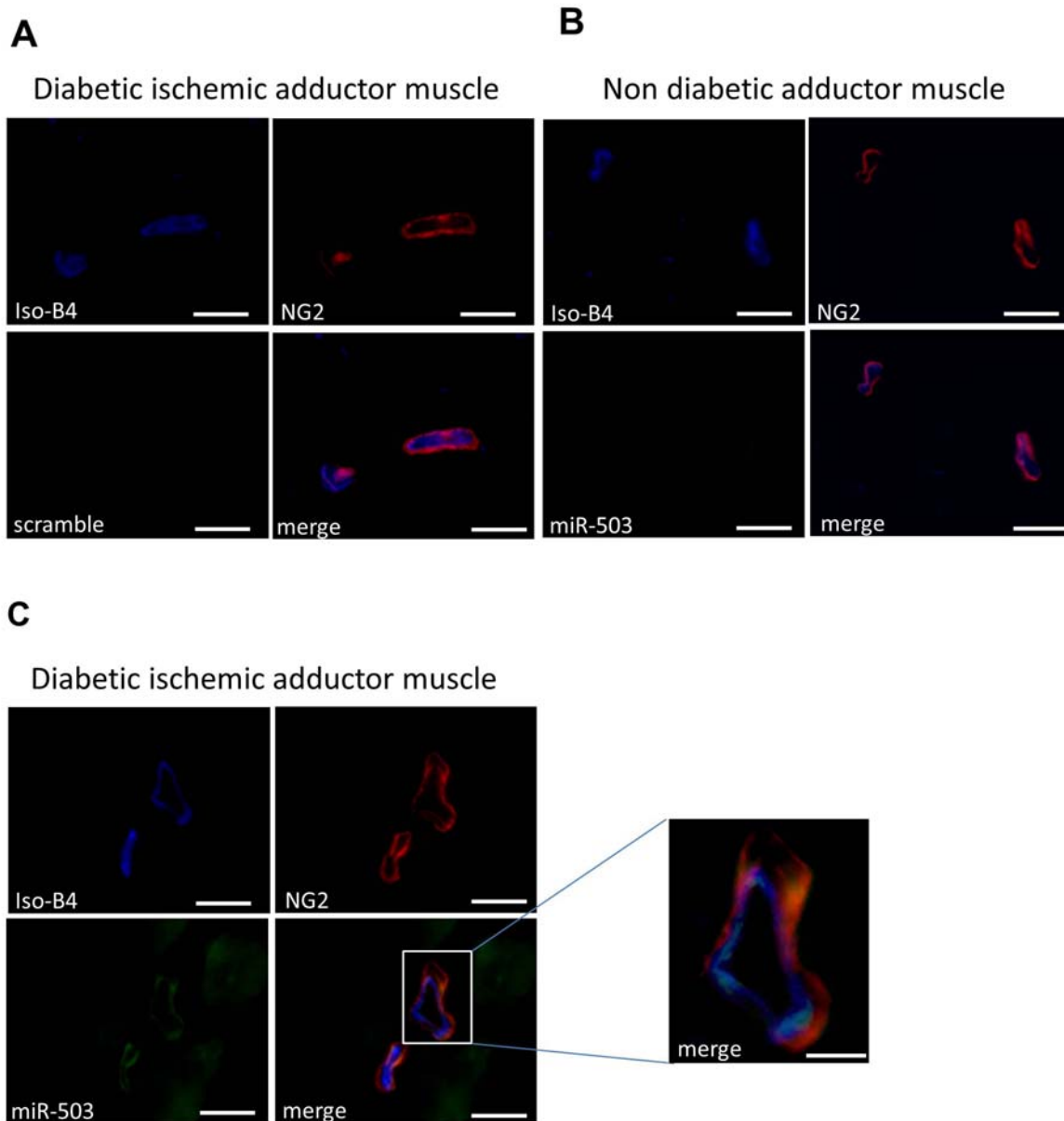
**Supplementary Figure 9: The uptake of MPs by pericytes**

**A**, pericytes incubated for 24h with MPs-miR-503 increased significantly the intracellular levels of miR-503 in dose-dependent manner. \* $p < 0.05$ , \*\* $p < 0.01$  vs. vehicle` (n=5), unpaired two-tailed Student's t-test. All values are mean $\pm$ SEM of three independent experiments. **B**, An *in vitro* co-culture system of HUVECs (top compartment) with pericytes (bottom compartment) in which the cells are separated by a membrane to prevent direct cell contact has been setup HUVECs were labeled with DiO (green fluorescence) and transduced with *p75<sup>NTR</sup>*-overexpressing adenovirus or *Null*-control adenovirus and fluorescence was analyzed in pericytes after 48hrs of co-culture by fluorescent microscopy; Scale bar 50  $\mu$ m.



**Supplementary Figure 10: Flow cytometry analysis of CD31 and NG2 sorted cells from limb muscle**

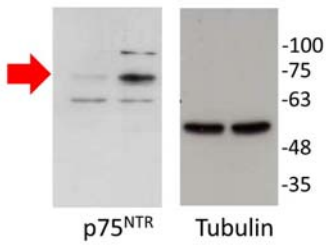
Cells were isolated using magnetic beads conjugated with CD31 and NG2 antibodies. Sorted cells were stained using **A**, CD31-FITC or **B**, NG2-PE conjugated antibody. Boxed: percentage of CD31 and NG2 in sorted cells (n=3); All values are mean±SEM of three independent experiments.



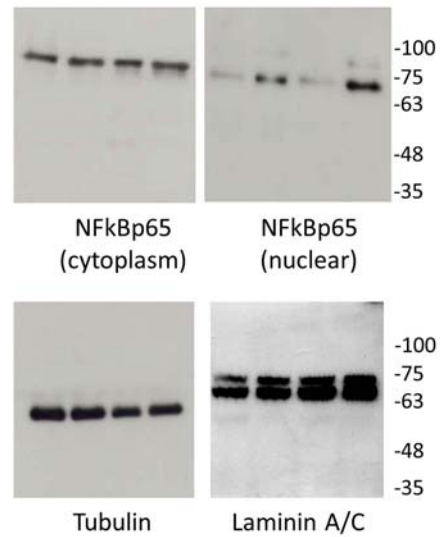
**Supplementary Figure 11: miR-503 *in situ* hybridization**

Localization of miR-503 (green fluorescence) by *in situ* hybridization in endothelial cells (Isolectin-B4 blue fluorescence) and pericytes (NG2, red fluorescence) in the ischemic limb muscles of diabetic and non-diabetic mice. **A**, Ischemic adductor muscle of diabetic mice hybridized with scrambled sequence probes as negative control (n=6). **B**, Non diabetic adductor muscle hybridized with miR-503 probe (n=6). **C**, Ischemic adductor muscle of diabetic mice hybridized with miR-503 probe (n=6). Scale bars 50  $\mu$ m. Insert: Higher magnification image of small vessel in ischemic adductor muscles of diabetic mice. Scale bars 10 $\mu$ m.

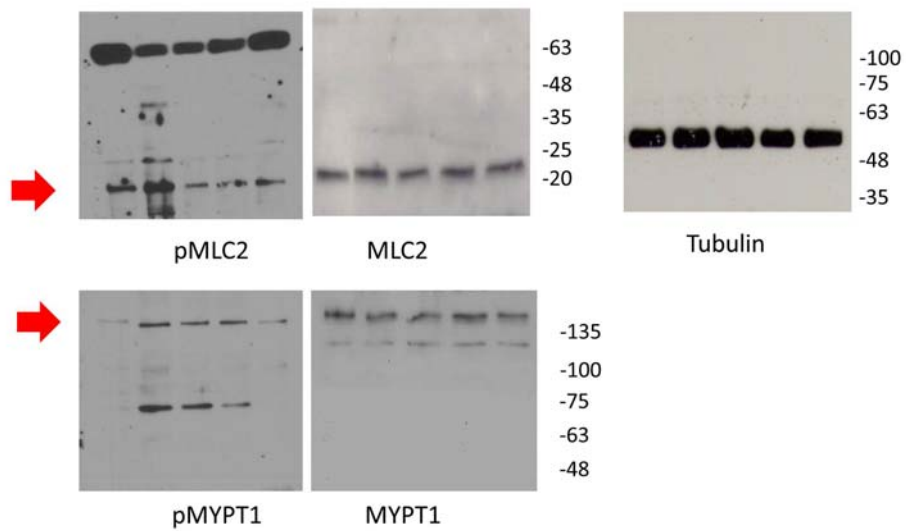
Full unedited gel for Figure 1



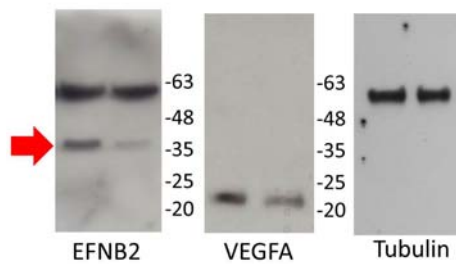
Full unedited gel for Figure 3



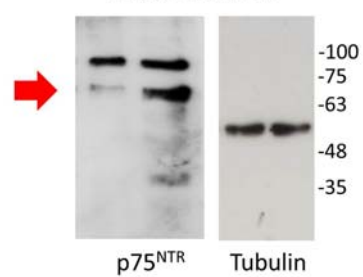
Full unedited gel for Figure 5



Full unedited gel for Figure 6



Full unedited gel for Suppl. Figure 1



Supplementary Figure 12: Original data for immunoblot related to main Figures

## SUPPLEMENTARY TABLES

**Table 1 - List of primers used in the study**

	<b>Forward</b>	<b>Reverse</b>
mut 3'UTR EFNB2	5'ATGATTTTTTAAGCAGACTCAATTTAA TATACTTATCA-3'	5'TGAGTCTGCTTAAAAAATCATC CAAAGCAGA-3'
mut 3' VEGFA	5'ATTCGCCATTTTATTTTTCTATTTAATT AATCACCGAG-3'	5'AGAAAATAAAATGGCGAATCCA ATTCCAA-3'
<b>ChIP primers</b>		
NF-kBp65	5'-CTTCCTCCTCCCTGTCTCC-3'	5'-GTTTTTCTGCCTGCCGTAAC-3'
IkB $\alpha$	Millipore primer set 17-10060	