

Supplementary Figure 1. Targeted gene disruption strategy and confirmation. (A) Positions

of flanking sequences relative to the genes of interest in the *Z. tritici* genome, sizes of

flanking sequences and sizes of regions to be replaced by the hygromycin resistance

cassette under a *trpC* promoter (*trpC+hph*). Primers used for confirmation of successful

gene disruption are marked with arrows and amplicon size is indicated above. (B)

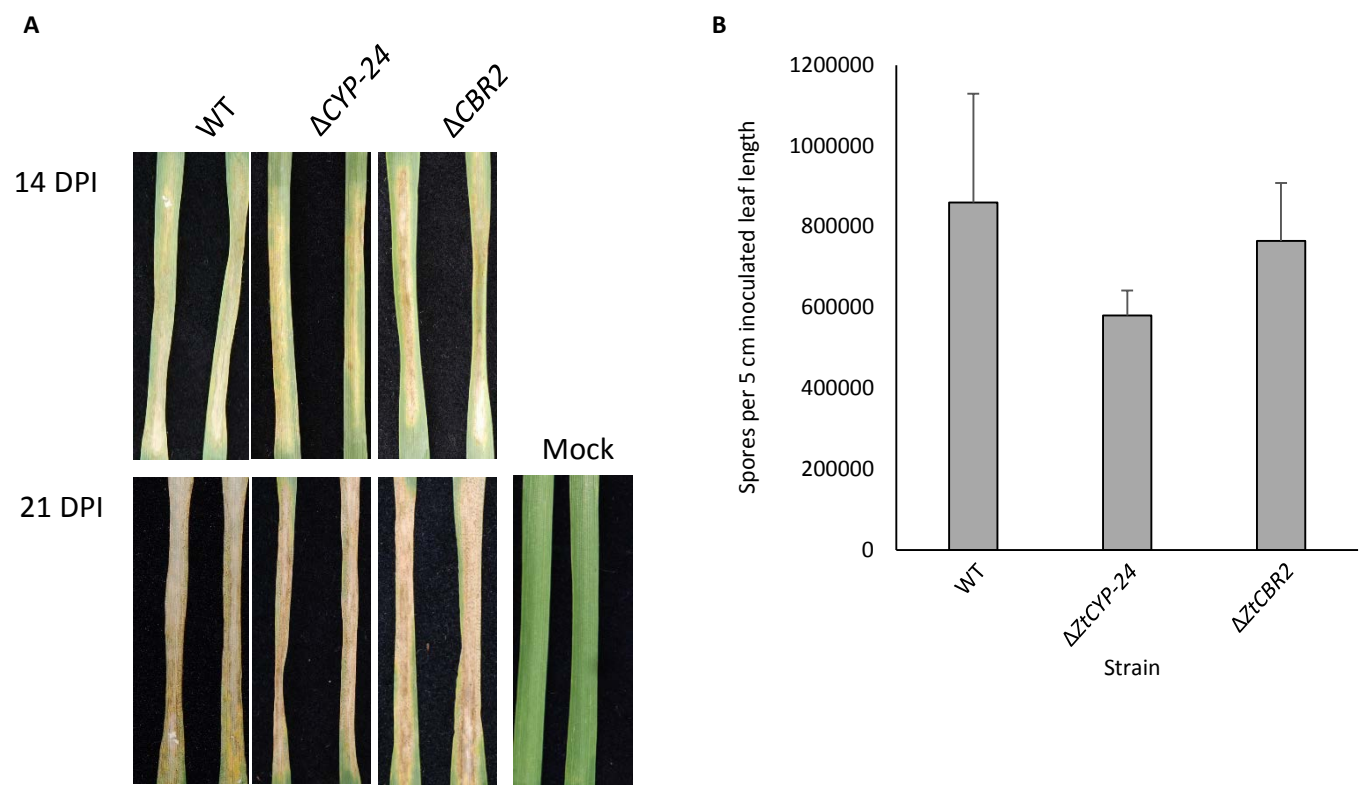
Confirmation of integration of *trpC+hph* at the correct location in the correct orientation

using a primer from within *hph* and primer from within 200 bp of genomic DNA adjacent to

one of the flanking sequences used to guide insertion. A single band of the correct amplicon

size is indicative of successful gene targeting. WT gDNA was used as a control. Strains taken

forward for further studies and their names are indicated above the relevant PCR lanes.



Supplementary Figure 2. Wheat leaf infection assays with Δ ZtCYP-24 and Δ ZtCBR2 strains

(A) Leaves infected with WT, Δ ZtCYP-24 and Δ ZtCBR2 after 14 and 21 DPI; mock-inoculated control leaves at 21 DPI are shown to the right. A total of 12 leaves per strain/mock were inoculated and two representative leaves are shown. (B) Mean number of spores (recovered by washing) per 5 cm length of inoculated leaf for WT, Δ ZtCYP-24 and Δ ZtCBR2.

Primer name	Sequence	Use	Restriction site
NB5.5-1F	aaaagatctTTGTCATTGCTGAGCCTGTGT	Amplification of flanking sequence NB5.5-1 for targeting <i>Ztcb1</i>	BglII
NB5.5-1R	aaaggcgcaccactagTACTGCTGGAGA GACTCTGCG	Amplification of flanking sequence NB5.5-1 for targeting <i>Ztcb1</i>	SpeI
NB5.3-1F	aaaggcgcaccactCTGTGGTCTCCTCTATGGTT	Amplification of flanking sequence NB5.3-1 for targeting <i>Ztcb1</i>	NarI
NB5.3-1R	aaaggcgcaccCAGGCCAAAAAGGCGCTATGA	Amplification of flanking sequence NB5.3-1 for targeting <i>Ztcb1</i>	SgsI
HYG-N1F	aaaactagTATATTGAAGGAGCATTTTTTGG	Amplification of hygromycin resistance cassette to clone into pNOV2114	SpeI
HYG-N1R	aaaggcgcctACTACTATTTCCTTTGCCCTCG	Amplification of hygromycin resistance cassette to clone into pNOV2114	NarI
CBR2 F1 F	aagagctcACGATGAACGATCGGTTT CATCACGATG CTACGGTTTTG	Amplification of flanking sequence CBR2F1 for targeting <i>Ztcb2</i>	SacI
CBR2 F1 R	aaggtaccAAGTGAATTGCAGCCAC GCA	Amplification of flanking sequence CBR2F1 for targeting <i>Ztcb2</i>	KpnI
CBR2 F2 F	aaaagcttGCCGACATTCCGG GTC CCAA	Amplification of flanking sequence CBR2F2 for targeting <i>Ztcb2</i>	HindIII
CBR2 F2 R	aaactagtCGAGTACCGCGCTTTGGGGG	Amplification of flanking sequence CBR2F2 for targeting <i>Ztcb2</i>	SpeI
CYP-24 F1 F	aaaagcttATGTGAGAGATCGTGAGTTGTTTGCA	Amplification of flanking sequence CYP-24F1 for targeting <i>Ztcbp-24</i>	HindIII
CYP-24 F1 R	aatcttagaGATCAACTTTGCACTGCTAGAAGT GAAGAT	Amplification of flanking sequence CYP-24F1 for targeting <i>Ztcbp-24</i>	XbaI
CYP-24 F2 F	aaggtaccGATGACTGCTGTGATCCTCTCGAA	Amplification of flanking sequence CYP-24F2 for targeting <i>Ztcbp-24</i>	KpnI
CYP-24 F2 R	aagagctctTGTCTGAACACCTGGCTTTGCGGT	Amplification of flanking sequence CYP-24F2 for targeting <i>Ztcbp-24</i>	SacI
CBR1 + Hyg F	GCTGATCTGACCCAGTTGCCT	CBR1 KO confirmation	
CBR1 + Hyg R	CAATCGTGAGGGGTTCACT	CBR1 KO confirmation	
CBR2 + Hyg F	AGTCGTCCAGCGGGTGA GCA	CBR2 KO confirmation	
CBR2 + Hyg R	GGCGAAGCTCTCCGGCCATC	CBR2 KO confirmation	
CYP-24 + Hyg F	AGTCGTCCAGCGGGTGA GCA	CYP-24 KO confirmation	
CYP-24 + Hyg R	GCCTCCCTGAACTCCCA	CYP-24 KO confirmation	

Supplementary Table 1 All primers used in this study with added adenines followed by restriction sites in lower case at the 5' end.

Fungal genome	URL
<i>Zyloseptoria tritici</i>	http://genome.jgi-psf.org/Mycgr3/Mycgr3.home.html
<i>Ustilago maydis</i>	http://genome.jgi.doe.gov/Ustma1/Ustma1.home.html
<i>Stagonospora nodorum</i>	http://genome.jgi.doe.gov/Stano2/Stano2.home.html
<i>Puccinia graminis</i>	http://genome.jgi-psf.org/Pucgr1/Pucgr1.home.html
<i>Piriformospora indica</i>	http://genome.jgi-psf.org/Pirin1/Pirin1.home.html
<i>Neurospora tetrasperma</i>	http://genome.jgi-psf.org/Neute_matA2/Neute_matA2.home.html
<i>Neurospora crassa</i>	http://genome.jgi-psf.org/Neucr2/Neucr2.home.html
<i>Mycosphaerella fijiensis</i>	http://genome.jgi.doe.gov/Mycfi2/Mycfi2.home.html
<i>Fusarium graminearum</i>	http://genome.jgi.doe.gov/Fusgr1/Fusgr1.home.html
<i>Dothistroma septosporum</i>	http://genome.jgi-psf.org/Dotse1/Dotse1.home.html
<i>Colletotrichum graminicola</i>	http://genome.jgi-psf.org/Colgr1/Colgr1.home.html
<i>Cochliobolus sativus</i>	http://genome.jgi.doe.gov/Cocsa1/Cocsa1.home.html
<i>Cochliobolus heterostrophus</i>	http://genomeportal.jgi-psf.org/CocheC5_3/CocheC5_3.home.html
<i>Blumeria graminis</i> f. sp. <i>hordei</i>	http://genome.jgi.doe.gov/Blugr1/Blugr1.home.html
<i>Aspergillus terreus</i>	http://genome.jgi.doe.gov/Aspte1/Aspte1.home.html
<i>Aspergillus nidulans</i>	http://genome.jgi.doe.gov/Aspnid1/Aspnid1.home.html
<i>Aspergillus fumigatus</i>	http://genome.jgi.doe.gov/Aspfu1/Aspfu1.home.html

Supplementary Table 2 URLs of all genome sequences incorporated into genome-wide analysis of *CBR* genes.