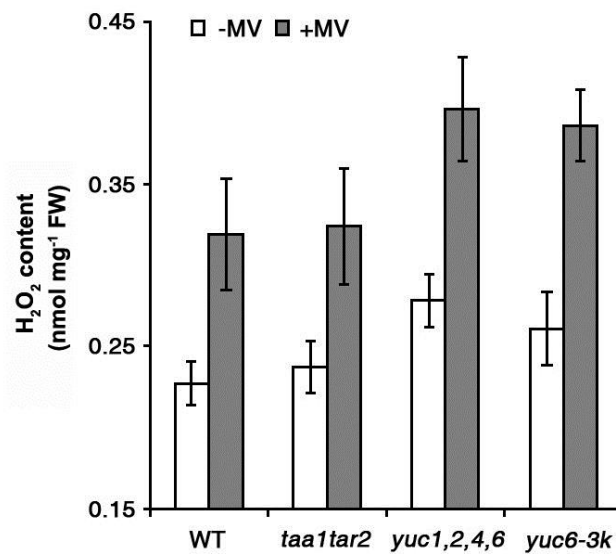


Supplementary Figure 1 | Overexpression of *YUC6* increases drought tolerance.

(a and b) Comparison of drought tolerance symptoms. (a) Shown are wild-type (WT, *Col-gl*) and *yuc6-1D* plants grown under well-watered conditions for 3 weeks (Control) and thereafter without watering for 14 days (Drought). (b) Shown are soil grown plants grown under well-watered conditions for 3 weeks and then allowed to grow without irrigation. Background ecotype of *yuc6-1D* and *yuc6-2D* is *Col-gl*, and that of *yuc6-3k* and *35S::YUC6* is *Col-0*. The *YUC6* overexpression (*yuc6-1D*, *yuc6-2D* and *35S::YUC6*) and their control WT plants were photographed after 12 days of drought. The loss-of function *yuc6-3k* mutant and its WT control were photographed after 10 days of drought. Isogenic WT lines were used for the comparisons. All plant materials have been described⁵.



Supplementary Figure 2 | YUC proteins, but not auxin, are involved in ROS control under oxidative stress.

Three-week-old auxin deficient mutants, *taa1 tar2*, quadruple *yuc1,2,4,6*, and loss-of function *yuc6-3k* plants were untreated (-MV) or exposed to 10 μM methyl viologen (+MV) for 4 h. Quantification of H₂O₂ was done as described in Methods. Data represent means±SE, n=3.

Supplementary Figure 3

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Q8VZ59|YUC6_ARATH MDFCWKREMEGKLAHDH-----R-----GMTSPRRICVV 29
Q39242|NTRA_ARATH --MCWISMSQSRFIIKSLFSTAGGFLLGSALSNPPLATAFSSSSSSSSAAAAMDVMEHK 58
A9LN30|NTR2_HORVD -----MEGSAAAPLR 10
P0A9P4|TRXB_ECOLI -----MGTTKH 6
Q8KCB2|FENR_CHLTE -----MLDIHNPATDHHDM 14

          FAD
Q8VZ59|YUC6_ARATH TGPVIVGAGPSGLATAACLKERGITSVLLERSNCIASLWQLKTYDRLLHLLPKQFCLEFI 89
Q39242|NTRA_ARATH TKVCIVGSGPAAHTAAIYASRAELKPLLFEGW-----MAND-IAPGGQLTTT---TDV 107
A9LN30|NTR2_HORVD TRVCIIGSGPAAHTAAIYAARAELKPVLFEGW-----MAND-IAAGGQLTTT---TDV 59
P0A9P4|TRXB_ECOLI SKLLIIGSGPAGYTAAVYAARANLQPVLITG-----MEKGGQLTTT---TEV 50
Q8KCB2|FENR_CHLTE RDLTIIIGGPTGIFAAPQCGMNNISCRIIESM-----PQLGGQLAALYPEKHI 62
          *:*.***. : * : : : * :

Q8VZ59|YUC6_ARATH IPFPGDFPTYPTK---QQFIEYLEDYARRFDIKPEFNQTVESA-AFDENLGMWRVTSVGE 145
Q39242|NTRA_ARATH E----NFPFGPEGILGIDIVEKFRKQSERFGTTIFTET-VNKV-DFS--SKPFKL---FT 156
A9LN30|NTR2_HORVD E----NFPFGPPTGIMGIDLMDNCRAQSVRFGTNILSET-VTEV-DFS--ARPPRV---TS 108
P0A9P4|TRXB_ECOLI E----NWPFGDPNDLTGPLLMERMHEHATKFETEIIFDH-INKV-DLQ---NRPFR---NG 99
Q8KCB2|FENR_CHLTE Y----DVAGFPE-VPAIDLVESLWQAERYNPVVLNETVTKYTKLD--DGTFFET---RT 112
          : * : : : : : : : : : : :

Q8VZ59|YUC6_ARATH EGTTEYVCRWLVAATGENAEPV--VPRF-EGMDKFAAAGVVKHTCHYKTKGGDFAGKRVLV 202
Q39242|NTRA_ARATH DSRT-VLADSVIISTGAVAKRL-SFTGSGEGNGGFWNRGISAFAVGDGAAPIFRNKPLVV 214
A9LN30|NTR2_HORVD DSST-VLADTVVAVATGAVARRL-YFSGS----DTYWNRGISAFAVGDGAAPIFRNKPIAV 162
P0A9P4|TRXB_ECOLI DNGE-YTCDALIIATGASARYL-GLPSE----EAFKGRGVSAAATGDF--FYRNQKVAV 151
Q8KCB2|FENR_CHLTE NTGNVYRSRAVLI AAGLGAFEPKRLPQLGNI-DHL--TGSS-VYYAVKSVEDFKGRVVI 168
          : : : : * * : : : :

          NADP
Q8VZ59|YUC6_ARATH VCGGNSGMEVCLDLNFGAQPSSLVVRDAVHVLPREM----LGTSTFGLSMFLLKWLPIRL 258
Q39242|NTRA_ARATH IGGGDSAMEEANFLTKYGSKVYIIHRRDTFRASKIMQQRALSNPKI-----EVIWNS-AV 268
A9LN30|NTR2_HORVD IGGGDSAMEEGNFLT KYGSQVYIIHRRNTFRASKIMQQRALSNPKI-----QVVWDS-EV 216
P0A9P4|TRXB_ECOLI IGGGNTAVEEALYLSNIASEVHLIHRRDGFRAEKILIKRLMDKVENGI--ILHTNR-TL 208
Q8KCB2|FENR_CHLTE VGGGDSALDWTVGLIKNAASVTLVHRGHEFQGHGKTAHEVERARANGTI--DVYLET-EV 225
          :* * : : : : * : : : : :

Q8VZ59|YUC6_ARATH VDRFLLVVSRLFILGDTTLLGLNRPRLGPLELKNISGKTPVLDVGTLAKIKTGDIVKCSGI 318
Q39242|NTRA_ARATH -----VEAYGDENG-----RVLGGLKVK-N----- 287
A9LN30|NTR2_HORVD -----VEAYGGAGG-----GPLAGVKVK-N----- 235
P0A9P4|TRXB_ECOLI -----EEVTGDQMG-----VTGVRLRDT----- 226
Q8KCB2|FENR_CHLTE -----ASI--EESN-----GVLTRVHLRSS----- 243
          : : : :

Q8VZ59|YUC6_ARATH RRLKRHEVEFDNGKTERFDAILLATGYKSNVPSWLKENKMFSSKKGDFPIQEF----PEGW 374
Q39242|NTRA_ARATH -----VVTGDSVSLKVSGLFFAIGHEPATKF-LDQG-LELDEDGYVVTKP----GTK 334
A9LN30|NTR2_HORVD -----LVTGEVSDLVQVSLFFAIGHEPATKF-LNGQ-LELHADGYVATKP----GSTH 282
P0A9P4|TRXB_ECOLI -----QNSDNIESLDVAGLFVAIGHSPNTAI-FEQQ-LELEN-GYIKVQSGIHGNATQ 276
Q8KCB2|FENR_CHLTE -----DGSKWTV EADRLLLILIGFKSNLGLPLARWD-LELYENALVV-----DSHMK 287
          . : : * . : : .

Q8VZ59|YUC6_ARATH RGECGLYAV-----GFTKRGISGASMDAKRIAEDIHKCWKQDEQVKKI----- 417
Q39242|NTRA_ARATH TSVVGVFAAGDVQDKKYRQAITAAGT--GCMAALDAEHYL----QEIGS-----Q 378
A9LN30|NTR2_HORVD TSVEGVFAAGDVQDKKYRQAITAAGS--GCMAALDAEHYL----QEVGA-----Q 326
P0A9P4|TRXB_ECOLI TSI PGVFAAGDVMDHIYRQAITSAIT--GCMAALDAERYL----DGLAD-----A 320
Q8KCB2|FENR_CHLTE TSV DGLYAAGDIAYYPGKLLKIQTGLSEATMAVRHSLSYI----KPGEKIRNVFSSVKMA 343
          . * : * . : * . : : .

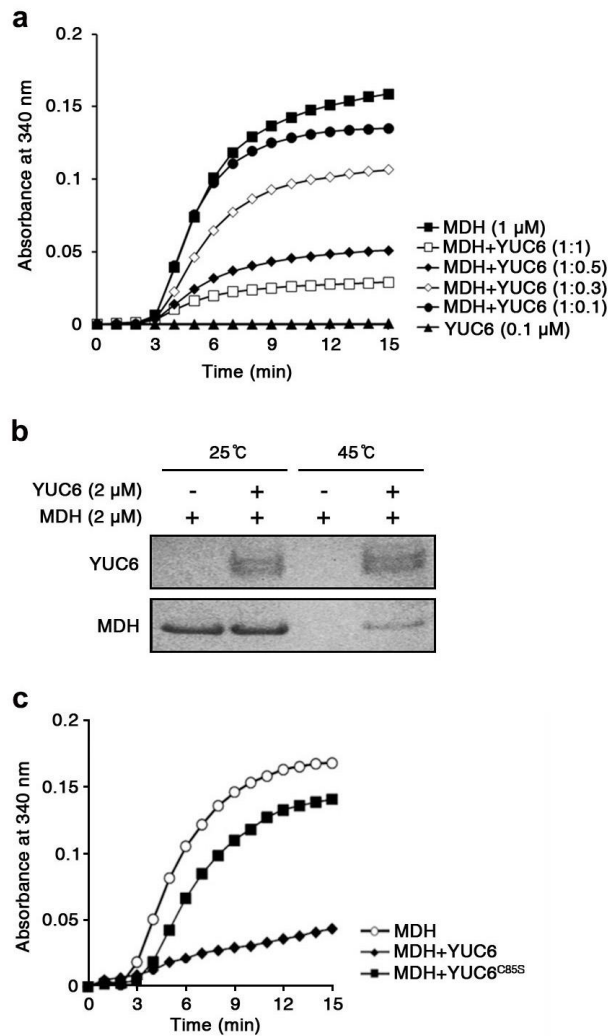
Q8VZ59|YUC6_ARATH ----- 383
Q39242|NTRA_ARATH EGKSD----- 383
A9LN30|NTR2_HORVD VGKSD----- 331
P0A9P4|TRXB_ECOLI K----- 321
Q8KCB2|FENR_CHLTE KEKKAEEAGNATENKAE 360

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Supplementary Figure 3 | Conservation of FAD and NADP binding sites are evolutionarily conserved in thiol-reductases and ferredoxin-reductases.

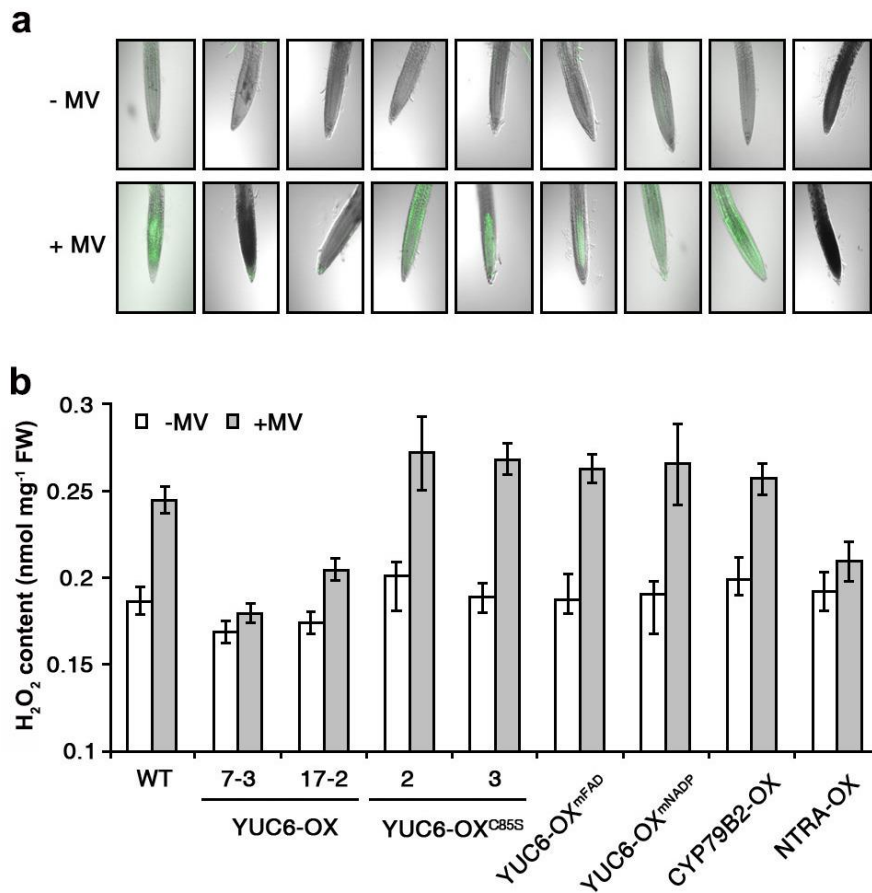
Sequences of thioredoxin-reductases from *A. thaliana* (NTRA), barley (NTR2) and *E. coli* (TRXB) were aligned with YUC6. In addition, the ferredoxin-NADP reductase of the green alga *Chlorobium tepidum* (FENR) was aligned using the Clustal-O program tool at the UniProt web site. UniProt protein codes are given for each peptide. Essential Gly residues in FAD and NADP binding sites are marked in *yellow*.

Cys residues at the active sites (CxxC) of Arabidopsis NTRA, barley NTR2 and *E. coli* TrxB thioredoxin reductases are marked in *green*. Cys-63 in the conserved motif CIASLW, and Cys-85 in the conserved motif C(E/Q)LP of YUC6 are marked in *blue*.



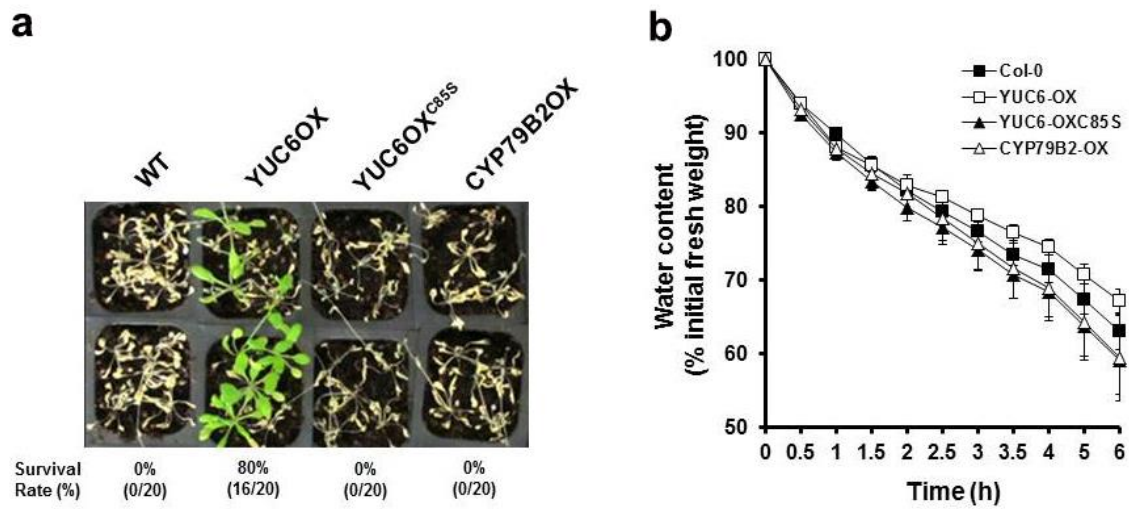
Supplementary Figure 4 | YUC6 exhibits holdase chaperone activity in a Cys-85 dependently.

Holdase chaperone activity of YUC6 was determined spectrophotometrically (**a** and **c**) and by SDS-PAGE migration (**b**). (**a**) A solution containing purified Arabidopsis malate dehydrogenase (MDH, 1 μ M) and purified recombinant MBP-tagged YUC6 protein in the indicated molar ratios was heated at 45 °C for the indicated time period. Thermal aggregation of MDH was monitored as increase in Abs_{340nm}. (**b**) MDH (2 μ M) was incubated in the absence or presence of YUC6 (2 μ M) for 15 min at 25 or 45 °C. After centrifugation, soluble fractions were rescued and separated on SDS-PAGE. (**c**) Holdase chaperone activity was measured as in Supplementary Figure 4a using a 1:1 molar ratio of MDH and indicated recombinant MBP-tagged YUC6 and YUC6^{C85S} proteins.



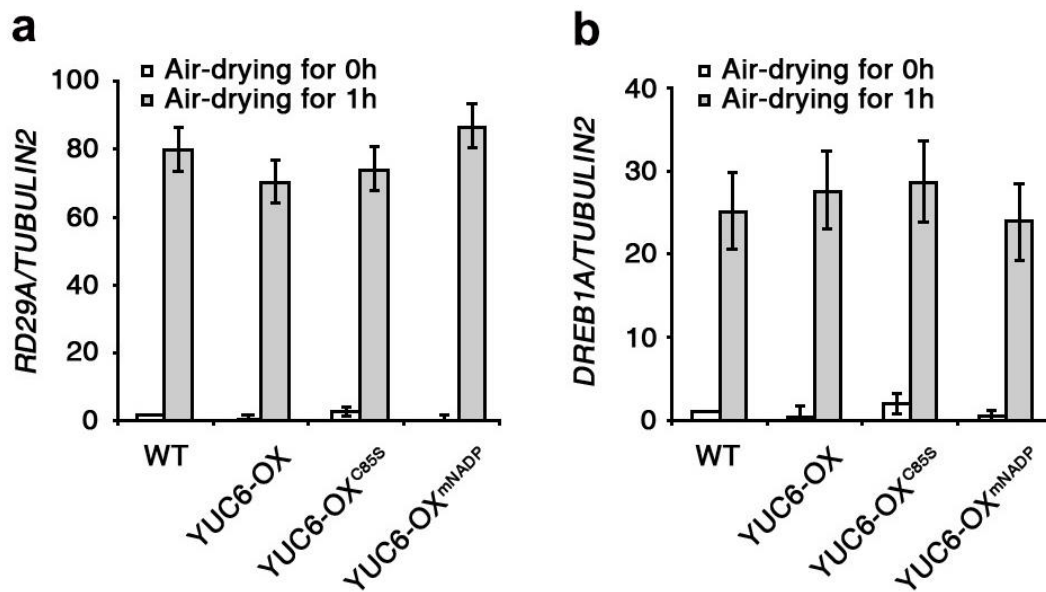
Supplementary Figure 5 | Cys-85 is essential for reduced ROS accumulation under oxidative stress in *YUC6* overexpression plants.

(a) Shown are roots of 5-day-old plants grown on MS agar, treated without (-MV) or with 10 μ M MV (+MV) for 4 h, and stained with H₂DCF-DA for H₂O₂ detection. (b) Three-week-old aerial parts of plants were treated with MV as in (a) and H₂O₂ contents were quantitatively analyzed. Data represent means \pm SE, n=3.



Supplementary Figure 6 | Auxin-overproduction driven by the overexpression of *CYP79B2* is not involved for the drought tolerance.

Col-0 plants (WT) were compared to YUC6-OX, YUC6-OX^{C85S}, and CYP79B2-OX transgenics in the same background. **(a)** Drought survival assay. Three-week-old well-watered plants were left for 7 days without irrigation. At 4 days after resuming irrigation, the plants were photographed, and the survival rate (percent) in each sample was quantified and is given at the bottom of the picture. **(b)** Water loss assay. The assay was performed as described in Figs. 1b and 5b.



Supplementary Figure 7 | Expression of the drought-responsive genes *RD29A* and *DREB1A* is not affected by overexpression of *YUC6*.

RD29A (a) and *DREB1A* (b) transcript levels were measured by qRT-PCR in total RNA from aerial parts of 3-week-old soil grown WT (Col-0), YUC6-OX, YUC6-OX^{C85S} and YUC6-OX^{mNADP} plants that were allowed to air-dry naturally for the indicated time periods. The expression levels were normalized compared to those of *TUB*. Bars represent means±SE, n=3.

Supplementary Table 1 Expression of ROS homeostasis genes is elevated in *yuc6-1D*.

AGI ID	Protein and GO Function	Mean log ²	Fold
At5g38000	Zinc-binding dehydrogenase, Oxidoreductase	1.181	2.27
At5g37940	Zinc-binding dehydrogenase, Oxidoreductase	1.138	2.20
At5g17000	Zinc-binding dehydrogenase, Oxidoreductase	0.568	1.48
At4g36090	2-oxoglutarate and Fe(II)-dependent oxygenase, Oxidoreductase	0.295	1.23
At5g59540	2-oxoglutarate and Fe(II)-dependent oxygenase, Oxidoreductase	0.757	1.69
At3g46500	2-oxoglutarate and Fe(II)-dependent oxygenase, Oxidoreductase	0.844	1.80
At3g46490	2-oxoglutarate and Fe(II)-dependent oxygenase, Oxidoreductase	0.890	1.85
At3g28480	2-oxoglutarate and Fe(II)-dependent oxygenase, Oxidoreductase	0.269	1.20
At2g18150	Peroxidase, Oxidation reduction, heme and metal binding	0.423	1.34
At4g32580	Thioredoxin, Redox homeostasis	0.259	1.20

Microarray hybridization was carried in a previous study⁵. Fold induction of ROS homeostasis genes in *yuc6-1D* versus the wild-type are listed.

Supplementary Table 2 Primer sequences used in this study.

Name	Primer sequence (5' to 3')	Purpose
<i>YUC6</i>	F: AAAAAGCAGGCTTCATGGATTTCTGTTGGAAGAG R: AGAAAGCTGGGTTCGATTTTTTTTACTTGCTCGTC	Gateway cloning
<i>YUC6^{C85S}</i>	F: CTTCTAAACAATTCTCTGAACTTCCGATTATA R: TATAATCGGAAGTTCAGAGAATTGTTTAGGAAG	Mutagenesis
<i>YUC6^{mFAD}</i>	F: CCGGTGATCGTAGGCCGCCGACCGTCCGTACTAG R: CATGCTGCCGTGGCTAGTACCGACGGTCCG	Mutagenesis
<i>NTRA</i>	F: GCGGATCCATGAGCCAGTCAAGATTC R: GCCTCGAGTCAATCACTCTTACCCTC	Cloning
<i>IAA1</i>	F: ATGGAAGTCACCAATGGGGCTTAACCT R: TCATAAGGCAGTAGGAGCTTCGGATC	qRT-PCR
<i>RD29A</i>	F: ATCACTTGGCTCCACTGTTGTTC R: ACAAAACACACATAAACATCCAAAGT	qRT-PCR
<i>DREB1A</i>	F: TGC GTTGGCGTTTCAGGATG R: CAAACTCGGCATCTCAAACATCG	qRT-PCR
<i>TUB</i>	F: AGCAAATGTGGGACTCCAAG R: CACCTTCTTCATCCGCAGTT	qRT-PCR
<i>CYP79B2</i>	F: AAAAAGCAGGCTTCATGAACACTTTTACCTCAA R: AGAAAGCTGGGTTCCTTCACCGTCCGGTAGAG	Gateway cloning