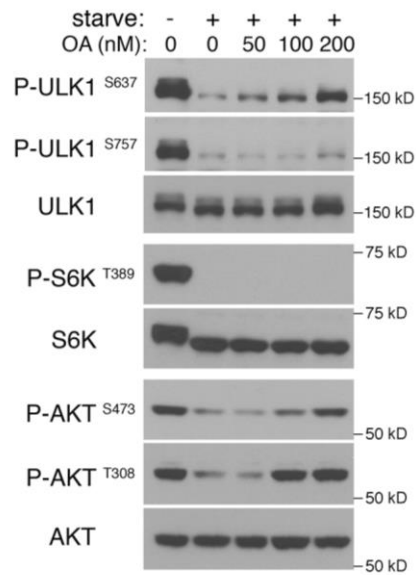
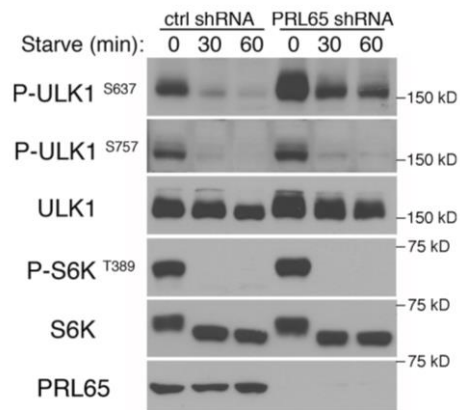


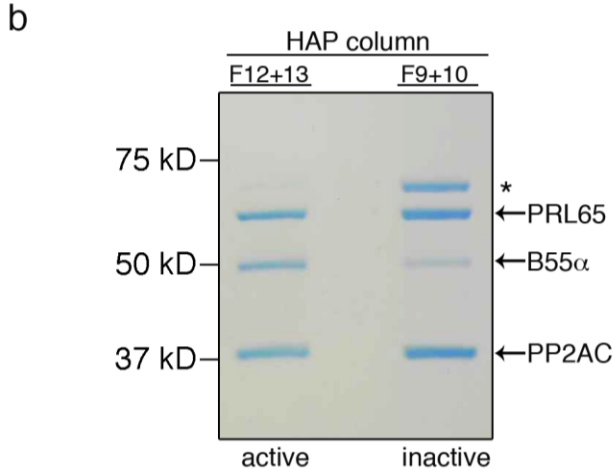
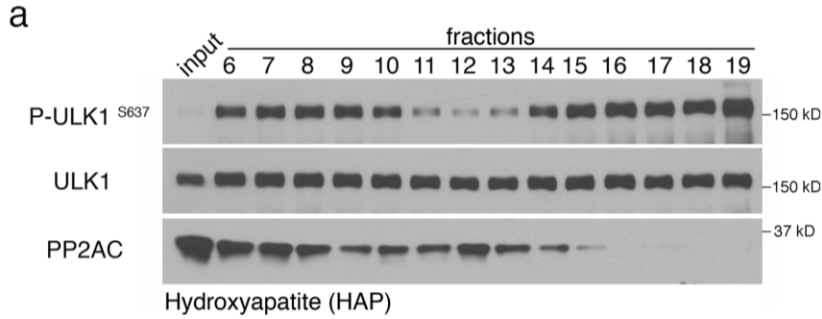
Supplementary Figure 1 Phosphatase activity is required for starvation induced autophagy. **(a, b)** ULK1 S637 and S757 phospho antibodies are specific. In **(a)**, the antibodies cannot recognize ULK1 when the respective sites of ULK1 (S637, S757) are mutated to alanine. In **(b)**, the antibodies recognized ectopic ULK1 (K46I mutant) expressed in ULK1/2DKO MEFs, and the signal was reduced upon starvation, correspondent with dephosphorylation of ULK1. **(c)** Starvation increases phosphatase activity in MEF cells. In vitro phosphatase assay was carried out using Flag-S-ULK1 as a substrate and increasing amounts of total cell lysate from MEFs that were kept in full media or starvation media. Reactions were incubated at 30°C for 30 min. **(d)** Starvation increases phosphatase activity in HT1080 cells. In vitro phosphatase assay was carried out using 10 μ g of total cell lysate from HT1080 cells that were kept in full media or starvation media. Reactions were terminated at the indicated time.



Supplementary Figure 2 Okadaic acid (OA) inhibits the dephosphorylation of ULK1 at S637 in a dose dependent manner. MEF cells were incubated in starvation media with increasing amounts of OA for 1 hr. Phosphorylation on ULK1 and S6K were monitored using site specific phospho-antibodies. AKT, previously reported as OA sensitive, was included as a positive control for OA treatment.



Supplementary Figure 3 PRL65 is required for ULK1 S637 dephosphorylation. MEF cells were transduced with lentivirus containing control shRNA or shRNA targeting PRL65, the scaffolding subunit of PP2A. 48-72 hrs post transduction, cells were incubated in starvation media for the indicated time and analysed by immunoblotting.



c Serine/threonine-protein phosphatase 2A 56 kDa regulatory subunit gamma isoform (PPP2R5C)

```

1  MLTCNKAGSG  MVVDAASSNG  PFQPVALLHI  RDVPPADQEK  LFIQKLRQCC
51  VLFDFVSDPL  SDLKWKEVKR  AALSEMVEYI  THNRNVITEP  IYPEAVHMFA
101 VNMFRTLPPS  SNPTGAEFDP  EEDEPTLEAA  WPHLQLVYEF  FLRFLESPDF
151 QPNIAKKYID  QKFVLQLEL  FDESDPRERD  FLKTTLHRIY  GKFLGLRAYI
201 RKQINNIFYR  FIYETEHNHG  IAELLEILGS  IINGFALPLK  EEHKIFLLKV
251 LLPLHKVKSL  SVYHPQLAYC  VVQFLEKDST  LTEPVVMALL  KYWPKTHSPK
301 EVMFLNELEE  ILDVIEPSEF  VKIMEPLFRQ  LAKCVSSPHF  QVAERALYYW
351 NNEYIMSLIS  DNAAKILPIM  FPSLYRNSKT  HWNKTIHGLI  YNALKLFMEM
401 NQKLFDDCTQ  QFKAEKLKEK  LMKKEREEAW  VKIENLAKAN  PQAQKELKKKD
451 RPLVRRKSEL  PQDPHTEKAL  EAHCRASELL  SQDGR

```

sequence coverage: 24%

Supplementary Figure 4 Only a subset of PP2A complexes in the cell can dephosphorylate ULK1. (a) Not all fractions containing PP2AC have activity against ULK1. Fractions from Hydroxyapatite column were assessed for activity against ULK1 and presence of PP2AC. Active fractions (11-13) were combined as input for the next purification step. (b, c) Regulatory subunit purified from inactive fraction of HAP column. (b) shows a Coomassie blue gel of PP2A purified from active and inactive fractions of the HAP column. * indicates unknown band that was excised and analyzed by mass spectrometry, which was identified as the B γ regulatory subunit. (c) shows results of mass spectrometry analysis. Peptide fragments matched to the B γ regulatory subunit of PP2A with a sequence coverage of 24%. Identified proteins: At $p < 0.01$, average false discovery rate, FDR: 0%.

- a** Serine/threonine-protein phosphatase 2A catalytic subunit beta isoform (PPP2CB)
- ```

1 MDDKAFKEL DQWVEQLNEC KQLNENQVRT LCEKAKEILT KESNVQEVRC
51 PVTVCGDVHG QFHDLMELEFR IGGKSPDTNY LFMGDYVDRG YYSVETVLL
101 VALKVRYPER ITILRGNHES RQITQVYGFY DECLRKYGNA NVWKYFTDLF
151 DYLLPLTALVD GQIFCLHGGL SPSIDTLDHI RALDRLQEVV HEGPMC DLLW
201 SDPDRGGWG ISPRGAGYTF QDISSETFNH ANGLTLVSRV HQLVMEGYNW
251 CHDRNVVTIF SAPNYCYRCG NQAAIMELDD TLKYSFLQFD PAPERGEPHV
301 TRRTPDYFL

```
- sequence coverage: 38.5%
- b** Serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A alpha isoform (PPP2R1A)
- ```

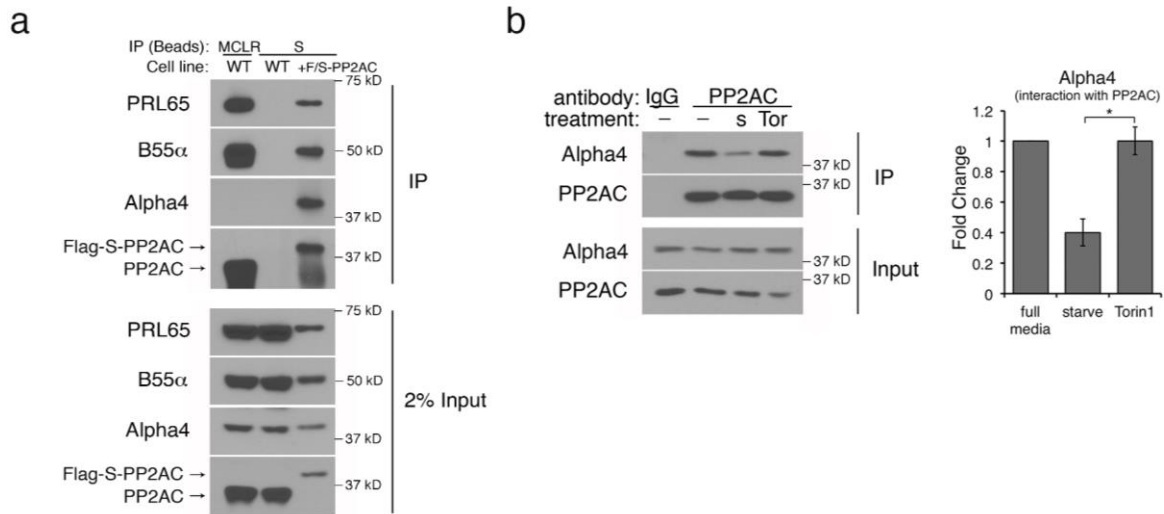
1 MAAADGDDSL YPIAVLIDEL RNEDVQLRLN SIKKLSIAL ALGVERTSRSE
51 LLPFLTDTIY DEDEVLLALA EQLGFTTTLV GGPEYVHCLL PPLESLATVE
101 ETVVRDKAVE SLRAISHEHS PSDLEAHFVP LVKRLAGGDW FTSRTSACGL
151 FSVCYPRVSS AVKAELRQYF RNLCSDDTPM VRRRAASKLG EFAKVLLELDN
201 VKSEIIPMFS NLASDEQDSV RLLAVEACVN IAQLLPQEDL EALVMPTRLRQ
251 AAEDKSWRVR YMVADKFTEL QKAVGPEITK TDLVPAFQNL MKDCEAEVRA
301 AASHKVKEFC ENLSADCREN VIMTQILPCI KELVSDANQH VKSALASVIM
351 GLSPILGKDN TIEHLLPLFL AQLKDECEPV RLNIISNLDC VNEVIGIRQL
401 SQSLLPAIVE LAEDAKWRVR LAIEYMPLL AGQLGVEFFD EKLNLSLMAW
451 LVDHVYAIRES AATSNLKKLV EKFGKEWAHA TIIPKVLAMS GDPNYLHRMT
501 TLF CINVLSE VCGQDITTKH MLPTVLRMAG DPVANVRFNV AKSLQKIGPI
551 LDNSTLQSEV KPILEKLTQD QDVDVKYFAQ EALTVLSLA

```
- sequence coverage: 37.2%
- c** Serine/threonine-protein phosphatase 2A 55 kDa regulatory subunit B alpha isoform (PPP2R2A)
- ```

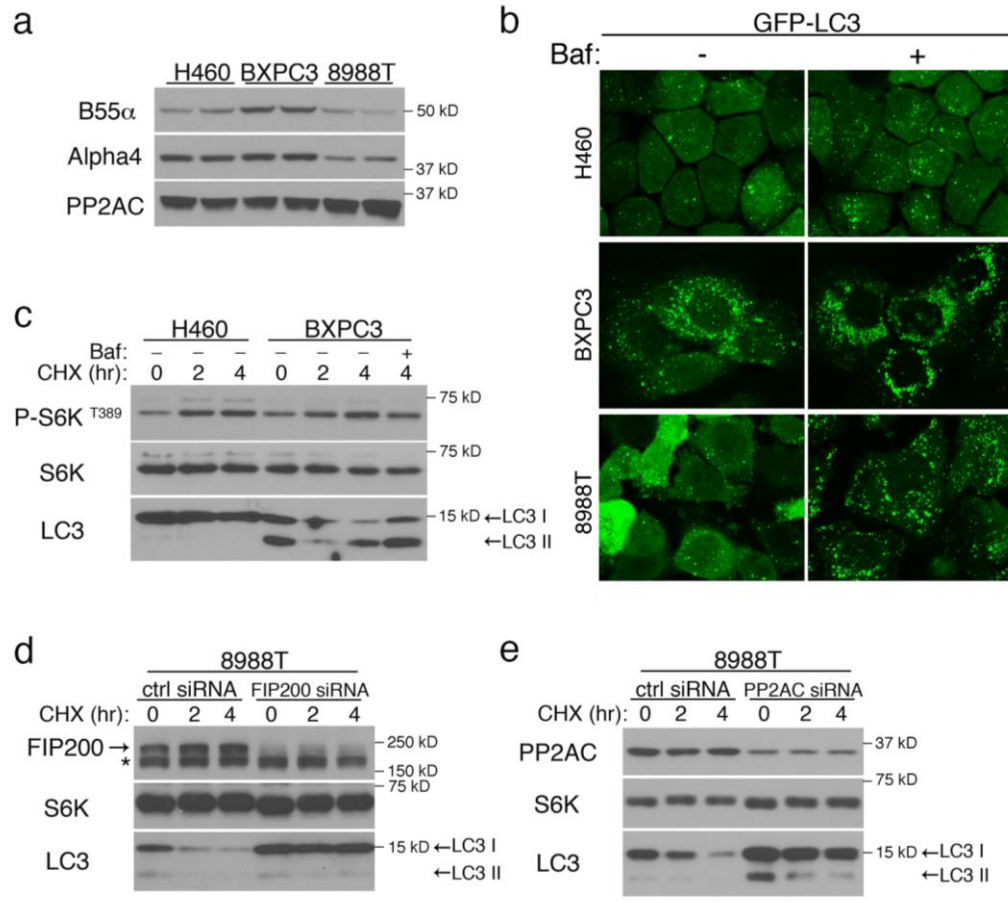
1 MAGAGGGNDI QWCF SQVKG VDDVAEADI ISTVEFNHSG ELLATGDKGG
51 RVVIFQQEQE NKIQSHSRGE YNVYTFQSH EPEFDYKSL EIEEKINKIR
101 WLPQKNAAQF LLSTNDKTIK LWKISERDKR PEGYNLKEED GRYRDP TTVT
151 TLRVPVFRPM DLMVEASPRR IFANAHTYHI NSISINS DYE TYLSADDLRI
201 NLWHLEITDR SFNIVDIKPA NMEELTEVIT AAEFHPNSCN T FVYSSSKGT
251 IRLCDMRASA LCDRHSKLF EPE DPSNRSF FSEI ISSIS VKF SHSGRYM
301 MTRDYL SVKI WDLNMENRPV ETYQVHE YLR SKLCSLYEND CIFDKFECCW
351 NGSDSVMTG SYNFFR MFD RNTKRDITL E ASRENNKPR T VLKPRKVCAS
401 GKRKDEISV DSLDFNKKIL HTAWHPKENI IAVATTNNLY IFQDKVN

```
- sequence coverage: 43.2%

**Supplementary Figure 5** Mass spectrometry analysis of p36, p65 and p50 bands from ULK1 phosphatase purification. The peptides detected during mass spectrometry analysis are shown in red. **(a)** The p36 band produced peptide fragments that matched to the catalytic subunit of PP2A with a sequence coverage of 38.5%. **(b)** Peptide fragments from the p65 band matched to the scaffolding subunit of PP2A with a sequence coverage of 37.2%. **(c)** Peptide fragments from the p50 band matched to the B55 $\alpha$  regulatory subunit of PP2A with a sequence coverage of 43.2%. Identified proteins: At  $p < 0.01$ , average false discovery rate, FDR: 0%.

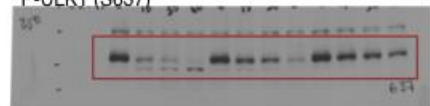


**Supplementary Figure 6** Starvation induces Alpha4 dissociation from PP2AC. **(a)** Endogenous PP2AC-Alpha4 complex is catalytically inactive. WT MEFs (ctrl) were lysed and incubated with microcystin-LR (MCLR) beads which bind to the catalytic pocket of PP2AC. As a comparison, MEFs expressing Flag-S-PP2AC were lysed and incubated with S-beads to pull down total PP2A complexes in the cell. **(b)** WT MEFs were incubated in starvation media or media containing 1  $\mu$ M Torin1 for 1 hour, lysed and incubated with control antibody or antibody against PP2AC. The amount of Alpha4 interacting with PP2AC under each condition was monitored by Immunoblotting and quantitated on the right (fold change relative to full media  $\pm$  s.d., n=5. two-tail student's t-Test, \* p<0.05).

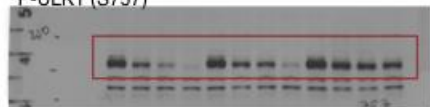


**Supplementary Figure 7** High basal autophagy in Pancreatic Ductal Adenocarcinoma cell lines is ULK1 complex dependent. **(a)** Immunoblot for endogenous levels of PP2A regulatory proteins B55 $\alpha$  and Alpha4 in control cell line H460 and PDAC cell lines BXPC3 and 8988T. **(b)** Cell lines stably expressing GFP-LC3 were kept in complete media in the presence or absence of 20 nM bafilomycin (Baf) for 90 min. Representative images from two independent experiments shown. **(c)** BXPC3 has high basal autophagy. BXPC3 and H460 were kept in complete media with 20  $\mu\text{g ml}^{-1}$  cycloheximide (CHX) for the indicated amount of time. Where indicated, 10 nM bafilomycin was added at the start of CHX treatment. Cells were lysed and immunoblotted for endogenous LC3. **(d)** FIP200 is required for basal autophagy in 8988T cells. 8988T was transduced with control siRNA or siRNA targeting FIP200, another member of the ULK1 complex. Cells were then treated as in (c). \* indicates non-specific band. **(e)** PP2AC is required for basal autophagy in 8988T cells. 8988T was transduced with control siRNA or siRNA targeting the catalytic subunit of PP2A. Cells were then treated as in (c).

Figure 1c  
P-ULK1 (S637)



P-ULK1 (S757)



P-S6K (T389)



ULK1



S6K

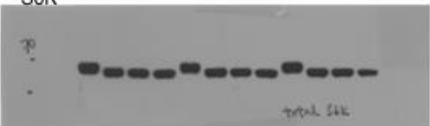


Figure 1e  
P-ULK1 (S637)



P-ULK1 (S757)



S-tag



P-S6K (T389)



S6K

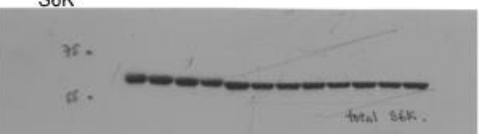


Figure 2a

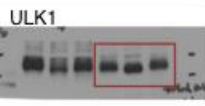
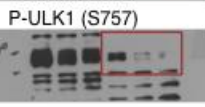
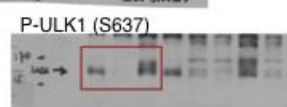
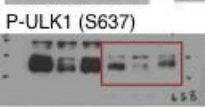
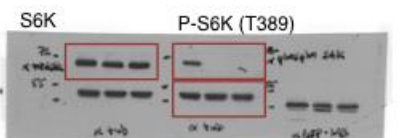
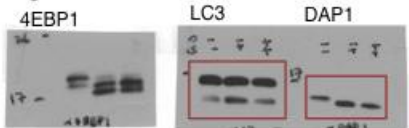


Figure 2b



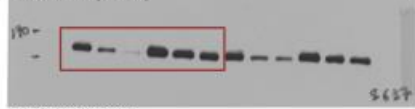
Tubulin



Figure 3a  
P-ULK1 (S757)



P-ULK1 (S637)



P-S6K (T389)



S6K



PP2AC

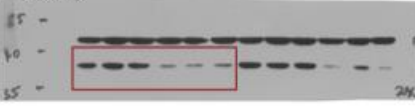
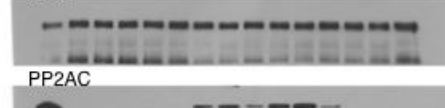


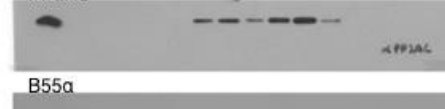
Figure 4e  
P-ULK1 (S637)



ULK1



PP2AC



B55a

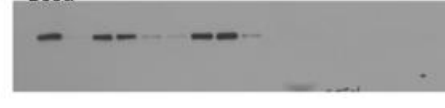
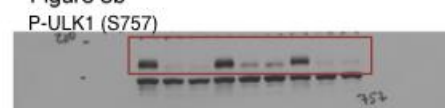
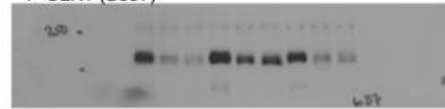


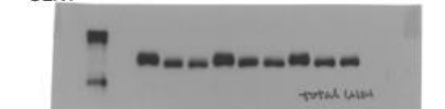
Figure 5b  
P-ULK1 (S757)



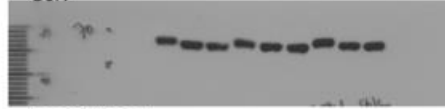
P-ULK1 (S637)



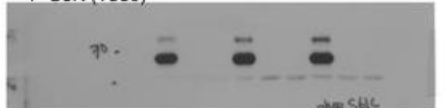
ULK1



S6K



P-S6K (T389)



B55a

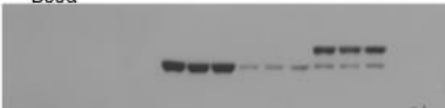


Figure 3c  
P-ULK1 (S637)



P-ULK1 (S757)



PP1C



PP2AC



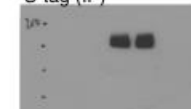
Figure 5d  
GFP (input)



S-tag (input)



S-tag (IP)



GFP (IP)



Figure 6B

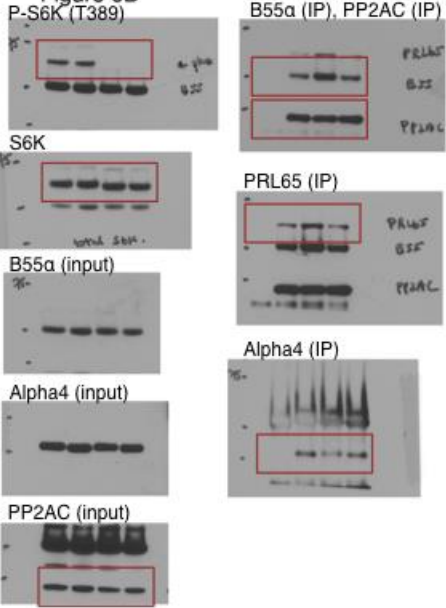


Figure 6C

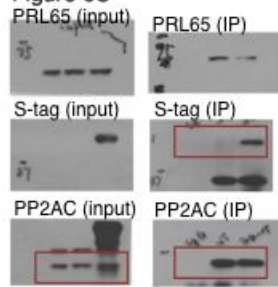


Figure 6D

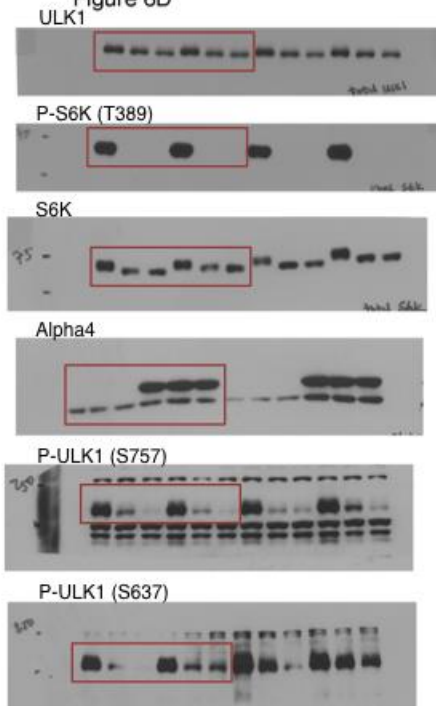
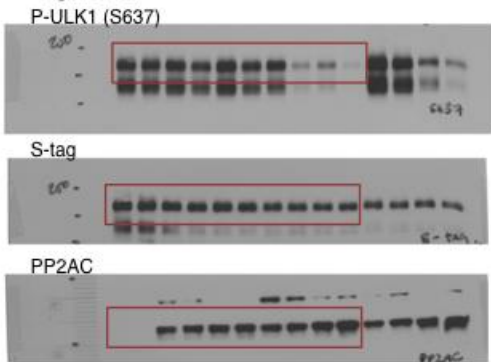
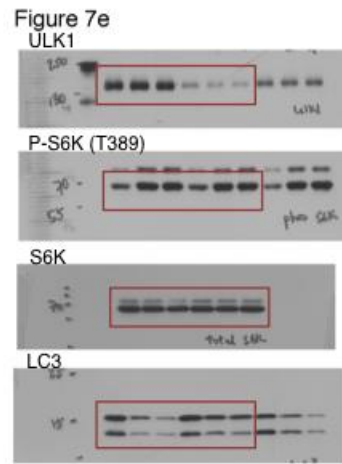
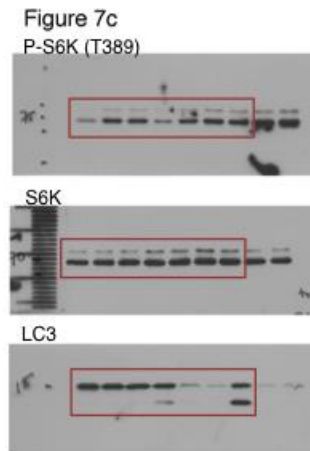
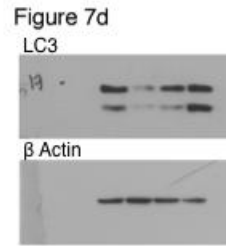
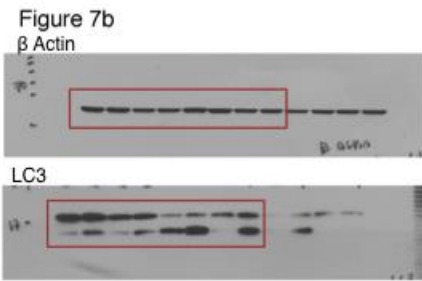


Figure 7a





**Supplementary Figure 8** Full scans of western blots related to respective figures as indicated. The protein of interest being detected is labeled on the top left hand corner of each blot.