Supplemental Data

Supplemental Figures



Supplement Fig. 1. Labeling, binding affinity and biological activity of modified glycolipid. (A) Amine-modified α GalCer and 7DW8-5 were synthesized by Aptuit (Greenwich, CT). Succinimide ester of either AF680 or BTR (Molecular Probes) was used to label the modified (B) Labeled glycolipid products were confirmed by mass spectrometry. (C) glycolipids. Affinities between glycolipid analogs and mCD1d molecule were measured by a competitive ELISA. Glycolipid analogs were serially diluted in the presence of 1 µg/ml 18:1 Biotinyl PE lipid, as the competitor. CD1d/biotinyl PE complex was detected with HRP-labeled avidin. 18:1 Caproylamine PE lipid was used as a positive control. Inhibition curves were generated using GraphPad Prism 4.0 software. Mean ± SD of triplicate assays is shown. (D, E) Biological activity of modified glycolipids was compared with that of unlabeled glycolipids. In (D), 2×10^4 human *I*NKT cells were co-cultured with 2 x 10⁴ Hela cells transfected with a human CD1d gene, or in (E) 2 x 10⁴ mouse *I*NKT hybridoma cells were co-cultured with 2 x 10⁴ A20-mCD1d cells, in the presence of an indicated concentration of each glycolipid. After 24-hr incubation, the culture supernatants were collected and the concentrations of human IFN- γ and mouse IL-2 in the supernatants were determined by ELISA. Mean ± SD of four triplicate assays is shown.



Supplemental Fig. 2. Use of L363 antibody, which equally binds α GalCer/mCD1d and 7DW8-5/mCD1d complexes, in order to determine the glycolipid/CD1d complex expressed by various APCs *in vivo*. (A) After incubating 2 µg of rat anti-mouse IgG₁ antibody with anti-rat IgG_K chain antibody coated microbeads for 2 hr, the microbeads were incubated with 1 µg of mCD1d:mIgG1 dimer for 2 hr. After washing, microbeads were loaded with 2 µg of 7DW8-5 and α GalCer at 37°C overnight. Then, microbeads were stained with serial diluted AF647-labeled L363 antibody for 30 min, and then analyzed with BD LSRII flow cytometer. (B) The binding kinetics of L363 antibody staining was fit using GraphPad Prism 4.0 software. Mean ± SD of triplicate assays is shown. (C) Lymphocytes, isolated from PLNs and spleens of mice 8 hr after i.m. or i.v. injection with glycolipids, were stained with L363 antibody, anti-mouse MHC-II, antimouse CD11c, anti-mouse F4/80, anti-mouse CD11b and anti-B220 antibodies. Macrophages were gated as CD11c⁻CD11b⁺F4/80⁻, B cells were gated as B220⁺CD11c⁻F4/80⁻, and DCs were gated as CD11c⁺MHC-II⁺F4/80⁻. Representative histograms from one mouse receiving glycolipid by (D) i.m. or (E) i.v. are shown.



Supplemental Fig. 3. DCs are required for early *i*NKT cell activation. CD11c-DTR mice were administered 100 ng diphtheria toxin by i.p. injection, and 36 hr later, mice were injected i.m. or i.v. with glycolipid. (A) PLNs and spleens were isolated 2 hr after injection, stained with CD3 antibody and α GalCer-loaded mCD1d dimer, and *i*NKT cells were gated by being a double positive. *i*NKT cell activation was assessed by monitoring CD69 expression and the results were quantified by flow cytometry. Representative histograms from one mouse receiving glycolipid by (B) i.m. or (C) i.v. are shown.



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Supplemental Fig. 4. Kinetics and molecular modeling of 7DW8-5 and CD1d binding. (A) CD1d-expressing cells capture 7DW8-5 faster than α GalCer. A20-mCD1d cells (2 x 10⁵) were loaded with 2 µg α GalCer or 7DW8-5 at 37°C. After 0, 15, 30, 60, 120, 240, 480 and 960 min, binding was stopped by fixing the cells with ice-cold paraformaldehyde, and L363 antibody was used to quantify CD1d-bound glycolipid. The results were expressed as mean ± SD of triplicates at each time point. (B) Proposed binding model of 7DW8-5 (cyan), a fluorinated analog of α GalCer, to mCD1d. To generate the model of 7DW8-5 binding CD1d, the coordinates of α GalCer (PDB entry 3TO4) were loaded into the program. MacPyMOL and the Build/Fragment and Sculpting Wizards were used to modify the compound and rearrange positioning to avoid collisions with mCD1d. The yellow surface (PDB entry 3TO4 using protein atoms only) describes the pocket of mCD1d in the presence of α GalCer (blue violet). The assumption is that the crystallization detergent molecule will be displaced by the 7DW8-5, due to lack of space to accommodate the longer aliphatic chain and the substitution of a hydrogen atom for a fluorine atom.