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Supplemental Figure S3



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Supplemental Figure S5



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NMDA reduced top motifs				NMDA induced top motifs				
	Motif	Name	P-value		Motif	Name	P-value	
	<u><u>S</u>POTTOCCATOO</u>	RFX	1e-75	ET	GGCSSZ	NF1	1e-18	
	GTIGCCATGGCAA	Rfx2	1e-74	귵 T G	CAAS	NF1- halfsite	1e-6	
		Hoxc9	1e-71	<u>FTG</u>	<u>CASSCTCC</u>	Tlx	1e-5	
	ECATERATCA	Pdx1	1e-53	<u>E</u>AA(CAGCTGI	Myf5	1e-4	
	SSESATEAATCE	HOXA9	1e-53	AACA	GCTG	MyoG	1e-4	
	<u>FGTTFCCATGGEA</u>	X-box	1e-51	SGT	<mark>F⊊CCATGG</mark> ⊊	X-box	1e-4	
	STTAATTAS	Lhx3	1e-45	ZEA		BMYB	1e-4	
	물론TAAT	Nkx6.1	1e-43	<u>Set</u>	GAGTCAESE	Jun- AP1	1e-4	
	<u><u>FTAAL</u>GE</u>	Isl1	1e-38	<u>zęr(</u>	<mark>GAAT</mark> ବୁୁ	TEAD	1e-4	
	<u>SGTISCCATGGSA</u>	Rfx1	1e-33	울 <mark>GT</mark>	I CCATCC S	Rfx1	1e-3	
	TAATTAGE	Lhx2	1e-32	<u>ê</u>CA	CTGEEE	Tef12	1e-3	
	ETGATGEAAE	Atf4	1e-31	êê T(ASTCATS	Fosl2	1e-3	
	ATGRATAATICA	Pit1	1e-27	들<u>C</u>Ç/	TTGTIS	Sox2	1e-3	
	ŞECAGÇTG	SCL	1e-26		<mark>GAAT</mark> ହ୍ମ	TEAD4	1e-3	
	TGATTCATSSS	Hoxb4	1e-25	GGGG	GGGG	Maz	1e-3	
	EGTGIIGCAA	CEBP:AP1	1e-24		IGTTTACES	FOXP1	1e-3	
	SCACCTCFESS	MyoD	1e-22	<u> </u>	CAGCTGZ	Ap4	1e-3	
	êCAGCTG울롯	Tcf12	1e-22		TCC ² CCAA	Stat3	1e-3	
	STTSCATCA	Chop	1e-21	STCI	TTAC	Foxo1	1e-3	
	STAATESSATTA	Phox2a	1e-17	<u>çç</u>	TGTᡓᢓᡓ	Sox6	1e-3	
	SSCCCAGGGG	EBF1	1e-17	GTI	CCATGGÇA	Rfx2	1e-3	
	GGGGGGGG	Maz	1e-17	GGA	ATTCCC	NFkB	1e-2	
	GGGGCTGTCCAT	REST- NRSF	1e-17	FCT	TGTICS	Sox4	1e-2	
	ESTCCCESCOR	EBF	1e-17	<mark>Şê</mark> C	ATGESS	p53	1e-2	
	<u>ATTAACACCT</u>	Eomes	1e-17	\$ \$C4	<u>ĢÇTĢ</u>	SCL	1e-2	

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Gene count

Supplemental Figure S8







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Control Super Enhancers

Eef2k Junb 9430041J12Rik MIx Pipox Prrg2 Sept9 Psmg4 St8sia2 Med25 Myl6 Ndufa6 Dnajb5 Ppp2r4 Gap43 Hoxb7 Gm1943 Nnat Dclk2 Cyb561d2 Ripply3 2900041M22Rik Atp6v1g2 Tssc1 Ccdc106 Celf4 Wbscr17 Slitrk5 4933400F21Rik Shank2 Ppp3ca Gtpbp6 Ppp2r2b Zfhx3 Gse1 Rhof Suv420h1 Cyth1 Epha4 Naa15 Kif26b Pcdh9 Nrg3 Syn3 Tet3 Dscam Unc80 Frmd4a Rbfox3 Tcf20 Trim9 Lrfn5 Wdr6 Acsf3 Tmem130 Dpysl3 Mab21I2 Mapre3 Mapt Dbn1 Crmp1 Psd2 Sema6d Nrp2 Tmem55b Ncam1 Pcdh1 Tom1I2 Add2 Gria2 Pax2 Lhx5 Trim3 Ctnnd2 Magi1 Kif5a Nrxn1 Kcnc1 Prickle2 Slc12a5 Atp1a3 Kiḟ21b Cadm2 Fam168b

NMDA_R2 NMDA_R1 Control_R2 Control_R1





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LEGENDS FOR SUPPLEMENTAL FIGURES

Supplemental Figure S1.

(A) Heat map showing correlation of FAIRE-seq replicates. (B-D) Scatter plot showing replicate correlation and enrichment distribution of all identified peaks for NP (B), TND1 (C) and TND10 (D). (E) Bar plot showing mapping statistics of reads obtained for each FAIRE-seq sample. X-axis represents percentage of alignment. (F) A profile plot showing normalized FAIRE enrichment around TSS of all genes in different stages of neurogenesis. X-axis shows distance from TSS, while y-axis represents library size normalized enrichment. (G) Bar plot showing distribution of FAIRE peaks in different genomic locations in NP, TND1 and TND10.
(H) Bar plot showing feature size normalized distribution of FAIRE peaks in different genomic locations in NP, TND1 and TND10.

Supplemental Figure S2.

(A) Heat map showing the correlation of H3K27ac replicates. (B-D) Scatter plot showing the replicate correlation and enrichment distribution of all identified H3K27ac peaks for NP (B), TND1 (C) and TND10 (D). (E) Bar plot showing the mapping statistics of H3K27ac ChIP-seq reads obtained for each sample. The x-axis represents the percentage of alignment. (F) Bar plot showing the total number of enriched peaks identified in NP, TND1 and TND10. (G) Bar plot showing the number of unique enriched peaks identified in NP, TND1 and TND10. (H) Box plot showing the distribution of peak width in NP, TND1 and TND10. P-values are calculated using Wilcoxon test. (I) A profile plot showing normalized H3K27ac enrichment around the TSS of all genes. The x-axis shows the distance from the TSS; the y-axis represents the library size-normalized enrichment at different stages of neurogenesis. (J) Bar plot showing the overlap of all FAIRE peaks with H3K27ac peaks. A peak was considered to be overlapping if the coordinates of FAIRE peaks showed at least 20% overlap with H3K27ac peaks using the same approach as in J. (L) Bar plot showing the overlap of all H3K27ac peaks with FAIRE peaks using 20% overlapping criteria. (M) Bar plot showing the overlap of unique

H3K27ac peaks with FAIRE peaks using same criteria as in (L). (N) Bar plot showing the number of all H3K27ac peaks that lie within 1 kb from FAIRE peaks. (O) Bar plot showing the number of unique H3K27ac peaks that lie within 1 kb from FAIRE peaks. (P) Bar plot showing feature size normalized distribution of FAIRE and H3K27ac overlapping peaks in different genomic locations in NP, TND1 and TND10. (Q) Box plot showing the expression of adjacent stages for the genes closest to NP, TND1 and TND10 unique H3K27ac sites. Genes were divided into overlapping bins based on the distance of the distal regulatory elements from the TSS. Expression is reported as reads per kilobase of transcript per million mapped reads (RPKM). (R) Stacked bar plot showing the overlap of all uniquely identified H3K27ac sites in TND10 with H3K27ac sites from other tissues.

Supplemental Figure S3.

(A) Cell viability assay showing total number of viable cells in control and NMDA treated neurons. (B) Representative immunoblots (n=2) showing caspase3 in NMDA induced neuron. β-actin is shown as a loading control. (C) Immunofluorescence microscopy for β III Tubulin staining in control and NMDA treated neurons confirming no symptoms of neuronal degeneration. Images were taken at 63X magnification (D) mRNA levels for classical activity modulated gene Npas4 in neuron treated with NMDA or NMDA in presence of NMDAR antagonist DAPV or L-glutamate were measured by gRT-PCRs relative to Actb plotted on the y-axis. Error bars represent SEM from independent biological replicates. (E-F) mRNA expression kinetics of neuronal activity modulated genes upon various concentrations of NMDA were measured by qRT-PCRs relative to Actb and plotted on the y-axis for ESderived neurons (E) and cortical neurons (F). (G-H) Bar plot showing the enrichment of known mouse phenotypes in genes down-regulated (G) and upregulated (H) upon NMDA treatment. The bars reflect the number of genes in each category; the lines represent the multiple tested corrected p-value of the corresponding GO term. (I) Volcano plot showing upand down-regulated lincRNAs upon NDMA treatment in TND10 neurons. The x-axis represents the fold change between control and NDMA in log2; the y-axis shows the adjusted p-value in -log10. (J) Heat map showing the expression of the genes nearest to differentially expressed lincRNAs. **(K)** Density plot showing distance (in kb, log2) of lincRNA from nearest gene TSS. **(L)** Scatter plot displaying changes in expression of lincRNA and corresponding nearest genes. **(M-N)** Bar plots showing enrichment of biological processes in down-(M) and up-regulated (N) genes near down- or up-regulated lincRNAs upon NDMA treatment respectivelly. The bars reflect the number of genes in each category; the lines represent the multiple tested corrected p-value of the corresponding GO term.

Supplemental Figure S4.

(A) mRNA levels for knockdown efficiency of target gene in epithelial cells pre-depleted for two days with specific siRNA against pro-survival genes were measured by qRT-PCR relative to *Actb* and plotted on the y-axis. Error bars represent SEM from independent biological replicates. (B) Representative images of colony forming assay performed on epithelial cell after 7 days of siRNA-mediated depletion of pro-survival factors compared to non-targeting control (NTC). (C) Representative Bright field images of the cells described in (B).

Supplemental Figure S5.

(A) Representative browser tracks of few progenitor genes showing increase in expression upon NMDA. (B) Same as in (A), but showing down-regulation of selected neuronal genes.

Supplemental Figure S6.

(A) Comparison of NMDA modulated H3K27ac peaks with H3K27ac peaks changing between NP and TND10. Peaks that are either losing or gaining two fold enrichment upon NMDA treatment were compared with two fold changing H3K27ac peaks between NP and TND10. A peak was considered to be overlapping, if it was found to be within 1kb distance of peaks modulated during neurogenesis (NP to TND10). (B) Venn diagram showing overlap of significantly down-regulated genes upon NMDA with significantly down or up-regulated genes between NP and TND10. (C) Same as in (B) but for genes significantly up-regulated upon NMDA. (D-E) Stacked bar plot showing the overlap of NMDA-induced (D) or -reduced

(E) peaks with H3K27ac peaks from several embryonic and adult stages taken from E11.5, E14.5, E.17.5, P0, P7, P21, P56 cortex (Nord et al. 2013). **(F-G)** List of top 25 transcription factors motifs identified in NMDA-reduced (F) and NMDA-induced (G) peaks.

Supplemental Figure S7.

(A-B) Heat map showing the expression of genes, binned based on distance (10kb, 20kb, 30kb, 50kb and 75kb) from NMDA-reduced (A) or NMDA-induced distal peaks (B) in controland NMDA-treated day 10 neurons. (C-D) Bar plots showing the enrichment of biological processes in genes that are near to down- and up-regulated distal regulatory regions and their expression reproducibly changed by at least 1.5-fold. The bars reflect the number of genes in each category; the lines represent the multiple tested corrected p-value of the corresponding GO term. (E) mRNA levels for genes shown in Fig. 7H upon NMDA mediated activation in mouse cortical neurons isolated from E16.5 mouse brain measured by qRT-PCR relative to *Actb* and plotted on the y-axis. Error bars represent SEM from independent biological replicates.

Supplemental Figure S8.

(A) Enrichment of biological process in genes near to non-promoter control only superenhancers of day 10 neurons. Bars reflect count of gene in each category while line represent multiple tested corrected p-value, displayed as alternate x-axis, of corresponding GO term. (B) Same as in (A) but for NMDA unique super-enhancers.

Supplemental Figure S9.

(A-B) Heat maps showing expression of genes near to control (A) and NDMA (B) nonpromoter super-enhancers in control and NDMA-treated neurons.