Direct Asymmetric Amination of α-Branched Cyclic Ketones Catalyzed by a Chiral Phosphoric Acid

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General Information:

Unless otherwise noted, all commercial reagents were used without further purification. 5Å MS were dried in oven at 200 °C overnight and used when after cooling to rt. Dichloromethane, toluene, ether, THF and triethylamine were purified by passage through an activated alumina column under argon. Thin-layer chromatography (TLC) analysis of reaction mixtures was performed using Merck silica gel 60 F254 TLC plates, and visualized under UV or by staining with ceric ammonium molybdate or potassium permanganate. Flash column chromatography was carried out on Merck Silica Gel 60 Å, 230 X 400 mesh. Nuclear magnetic resonance (NMR) spectra were recorded using Bruker AV-600, DRX-500, AV-500, AVQ-400, AVB-400 and AV-300 spectrometers. ¹H and ¹³C chemical shifts are reported in ppm downfield of tetramethylsilane and referenced to residual solvent peak (CHCl₃; $\delta H = 7.26$ and $\delta C = 77.16$, CH₃OH; $\delta H = 3.31$ and $\delta C = 49.00$). Multiplicities are reported using the following abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad resonance. Mass spectral data were obtained from the Micro-Mass/Analytical Facility operated by the College of Chemistry, University of California, Berkeley. Enantiomeric excesses were determined on a Shimadzu VP Series Chiral HPLC using IA, IB, IC, OD, ASH columns. The synthesis of phosphoric acids (S)-**TRIP**¹, (*R*)-**TCYP**² has been previously described. Racemic products were synthesized by carrying out the amination reactions using (\pm) -**TRIP** as catalyst.

Synthesis of the chiral phosphoric acids:

(R)-6,6'-dioctyl-3,3'-bis(2,4,6-tricyclohexylphenyl)-[1,1'-binaphthalene]-2,2'-diol (S2)



S1³ and (2,4,6-tricyclohexylphenyl)magnesium bromide² were prepared according previous reports. Into a flame dried 250 mL flask, equipped with a stir bar was added (*R*)-S1 (2.0g, 2.35 mmol) and NiCl₂(PPh₃)₂ (154 mg, 0.24 mmol) and THF 20 mL. The heterogeneous mixture was purged with N₂ and the above Grignard solution added slowly at 0 °C. The resulting mixture was warmed to rt and stirred overnight, after which the mixture was quenched with satd. NH₄Cl solution. After extraction with Et₂O three times, the combined organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give the residue, which was purified by flash chromatography (20:1 Hexane/DCM) to afford a syrup 2.3 g.

To the solution of the above syrup in dioxane (40 mL) was added HCl (conc., 4 mL) at rt. The mixture was heated to 70 °C and stirred overnight. The mixture was then cooled to rt and concentrated *in vacuo* to afford a residue, which was diluted with DCM and washed with satd. NaHCO₃ solution and brine. The combined organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography (40:1 – 20:1, Hexane/DCM) to afford **S2** (2.0 g, 74% yield) as a white foam. ¹H NMR (500 MHz, CDCl₃) δ 7.74 (s, 2H), 7.71 (s, 2H), 7.32 (d, *J* = 8.6 Hz, 2H), 7.26 (d, *J* = 8.5 Hz, 2H), 7.21 (s, 2H), 7.17 (s, 2H), 4.88 (s, 2H), 2.91 – 2.78 (m, 4H), 2.71 – 2.54 (m, 4H), 2.44 – 2.29 (m, 2H), 2.13 – 2.03 (m, 4H), 2.01 – 1.91 (m, 6H), 1.91 – 0.87 (m, 80H). ¹³C NMR (125 MHz, CDCl₃) δ 150.14, 147.92, 146.97, 146.71, 138.25, 131.72, 131.27, 130.06, 129.35, 129.20, 128.16, 126.88, 124.28, 122.39, 122.36, 112.96, 44.95, 41.96, 41.83, 36.15, 34.89, 34.68, 34.64, 34.61, 34.37, 34.05, 32.04, 31.62, 29.80, 29.67, 29.44, 27.27, 27.18, 27.10, 27.00, 26.91, 26.42, 26.21, 22.83, 14.27. m/z HRMS (ESI) found [M+H]⁺ 1153.8767, C₈₄H₁₁₃O₂⁺ requires 1153.746.

(R)-C₈-TCYP



To a solution of S2 (2.0 g, 1.73 mmol) in anhydrous pyridine (7 mL) was added POCl₃ (473 uL, 5.20 mmol) at rt. The mixture was warmed to 90 °C and stirred overnight, after which the mixture was cooled to rt and added to H₂O (7 mL) slowly. The mixture was then warmed to 105 °C for 4 h, after which it was cooled to rt and acidified with 3N HCl solution. The mixture was extracted with DCM three times, and the combined organic layer was dried over Na_2SO_4 , filtered and concentrated in vacuo to give a residue, which was purified by flash chromatography (20:1 - 2:1, Hexane/ Ethyl Acetate) to afford a white foam, which was diluted in Et₂O, washed with 3N HCl solution and concentrated in vacuo to give (R)-Cs-TCYP (1.5 g, 71% yield) as white a foam. ¹H NMR (400 MHz, CDCl₃) δ 7.59 (d, J = 15.9 Hz, 4H), 7.20 – 7.07 (m, 4H), 6.89 (d, J = 8.3 Hz, 4H), 2.75 (td, J = 7.1, 3.9 Hz, 4H), 2.50 - 2.36 (m, 2H), 2.23 - 2.03 (m, 4H), 1.96 (m,-1.82 (m, 10H), 1.80 - 0.49 (m, 80H). ¹³C NMR (125 MHz, CDCl₃) δ 146.88, 146.67, 146.28, 145.84, 145.77, 140.03, 132.05, 132.00, 131.20, 130.65, 127.71, 126.75, 126.45, 122.38, 121.74, 121.56, 44.85, 42.19, 41.86, 37.16, 36.05, 35.15, 34.79, 34.25, 33.28, 32.70, 32.06, 31.42, 29.72, 29.57, 29.46, 27.44, 27.29, 27.26, 27.06, 26.84, 26.50, 26.41, 26.03, 22.84, 14.27. ³¹P NMR (162 MHz, CDCl₃) δ 1.35. m/z HRMS (ESI) found [M-H]⁻ 1215.8308, C₈₄H₁₁₂O₄P⁻ requires 1215.8304.

(R)-H₈-TCYP



(*R*)-H₈-TCYP was prepared from (*R*)-3,3'-diiodo-2,2'-bis(methoxymethoxy)-4a,5,5',6,6',7,7',8,8a,8'-decahydro-1,1'-binaphthalene⁴ as the above procedures. ¹H NMR (500 MHz, CDCl₃) δ 6.89 – 6.81 (m, 6H), 2.89 – 2.78 (m, 2H), 2.72 (tt, J = 13.2, 6.1 Hz, 4H), 2.50 – 2.38 (m, 2H), 2.31 (dd, J = 16.8, 6.1 Hz, 2H), 2.16 (t, J = 12.1 Hz, 4H), 2.00 – 1.68 (m, 20H), 1.69 – 1.55 (m, 12H), 1.51 – 1.36 (m, 12H), 1.33 – 1.07 (m, 16H), 1.07 – 0.83 (m, 8H). ¹³C NMR (125 MHz, CDCl₃) δ 146.52, 146.46, 145.81, 144.91, 144.84, 136.10, 133.27, 132.31, 129.45, 129.42, 126.75, 122.37, 121.44, 44.80, 41.94, 41.80, 37.21, 34.96, 34.85, 34.18, 33.67, 32.77, 29.13, 27.98, 27.53, 27.30, 27.26, 27.01, 26.83, 26.58, 26.52, 23.10, 22.97. ³¹P NMR (162 MHz, CDCl₃) δ -1.58. m/z HRMS (ESI) found [M-H]⁻ 999.6391, C₆₈H₈₈O₄P⁻ requires 999.6426.

Synthesis of the substrates:



Substrates 1a, 1s were commercial available. Synthesis of substrate $1b^5$, $1c^5$, $1e^5$, $1d^6$, $1f^5$, $1l^5$ and $1m^5$ was reported previous.

Substrate $1g^7$, $1i^8$, $1k^8$, $1x^9$ was synthesized in **method A**, whose ¹H NMR data matched the literature.

General procedure for **method A**: To a solution of ArBr or ArI (10 mmol) in THF was added *n*BuLi (4 mL, 2.5 N in heaxane, 10 mmol) at -78 °C. After stirring for 1h at this temperature, cyclohexene oxide (5 mmol) was added followed by adding BF₃.Et₂O (7.5 mmol). The reaction was quenched with satd. NaHCO₃ solution after stirring for another 3h at -78 °C. The mixture was diluted with ethyl acetate and washed with 2N NaOH solution and brine. The combined organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography to afford the alcohol.

To the solution of alcohol (1.0 equiv.) in DCM was added DMP (1.2 equiv). After TLC monitoring the full conversion of the SM, the mixture was quenched with satd. NaHCO₃ solution and Na₂SO₃ solution. The mixture was extracted with DCM for three times, the combined organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography to afford the desired ketone.

1h, $1j^{10}$ was synthesized in **method B**.

General procedure for **method B**: To a flame dried flask was added Cs_2CO_3 (6.5 g, 20 mmol), $Pd_2(dba)_3$ (184 mg, 0.2 mmol) and Xantphos (139 mmol, 0.24 mmol). After purging with N_2 , toluene (10 mL), ketone (15 mmol) and phenyl iodide (1.11 mL, 10 mmol) were added. After stirring overnight at 80 °C, the mixture was filtered and concentrated *in vacuo* to afford a residue, which was purified by flash chromatography to provide the desired ketone.



Substrate 10^{11} , $1p^{12}$ was synthesized in method C^{11} .

General procedure for **method C**: To a solution of CuI (190 mg, 1.0 mmol) in THF (10 mL) was added alkenyl magnesium bromide (13 mL, 1.0 N in THF, 13 mmol) at -78°C. After stirring for 30 min, cyclohexene oxide (1.0 mL, 10 mmol) was added and the mixture was warmed to -20°C. After stirring for another 2 h at this temperature, the mixture was quenched with satd. NH₄Cl solution and extracted with Et₂O for 3 times. The combined organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography to afford the alcohol.

To the solution of $(ClCO)_2$ (889 µL, 10.5 mmol) in DCM (20 mL) was added DMSO (1.5 mL, 21.0 mmol) at -78 °C. After stirring for 0.5 h, a solution of the above alcohol (7.0 mmol) was added slowly into the mixture. After stirring for another 2h at -78 °C, Et₃N (5.8 mL, 42.0 mmol) was added and the mixture was slowly warmed to rt. The mixture was quenched with satd. NH₄Cl solution after stirring for another 1h stirring. The mixture was extracted with DCM three times, and the combined organic layer was washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography to afford desired ketone.

Substrates $1n^{13}$, 1t, 1u, $1v^{14}$, $1w^{14}$ was synthesized in method D^{13-14} .

General procedure for **method D**: To a solution of cyclic ketone (10 mmol) in DMSO (20 mL) was added phenylacetylene (1.1 mL, 10 mmol) and *t*BuOK (1.1 g, 10 mmol) at 80 °C or 100 °C.

After stirring of 45 mins after this temperature, the mixture was poured into a mixture of ice and satd. NH₄Cl solution. Upon extraction with Et₂O three times, the combined organic layer was washed with H₂O, brine, dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography to afford desired ketone.

Data for the new compounds was given below:

tert-butyl 4-(2-oxocyclohexyl)benzoate (1h)



¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, *J* = 8.2 Hz, 2H), 7.18 (d, *J* = 8.3 Hz, 2H), 3.66 (dd, *J* = 12.2, 5.4 Hz, 1H), 2.63 – 2.40 (m, 2H), 2.27 (ddd, *J* = 12.2, 6.0, 3.6 Hz, 1H), 2.22 – 2.12 (m, 1H), 2.10 – 1.96 (m, 2H), 1.90 – 1.78 (m, 2H), 1.58 (s, 9H). ¹³C NMR (125 MHz, CDCl₃) δ 209.76, 165.78, 143.57, 130.77, 129.59, 128.59, 80.92, 57.51, 42.34, 35.14, 28.32, 27.88, 25.43. m/z HRMS (ESI) found [M+Na]⁺ 297.1465, C₁₇H₂₂O₃Na⁺ requires 297.1461.

(*Z*)-2-styrylcyclohexanone (**1q**)



To a solution of **S3** (500 mg, 2.5 mmol) in MeOH (8 mL) was added Lindlar's catalyst (200 mg) and quinolone (1.2 uL). After purging with H₂, the mixture was stirred at rt for 2h. The mixture was filtered and concentrated in vacuo to afford a residue, which was purified by flash chromatography (4: 1, DCM/ Hexane) to afford the alkene-substituted alcohol (302 mg, 60% yield). The Swern Oxidation was carried out following the procedure in **Method C** to afford the desired ketone **1q**. ¹H NMR (400 MHz, CDCl₃) δ 7.33 (dd, *J* = 8.2, 6.7 Hz, 2H), 7.25 (d, *J* = 5.5 Hz, 1H), 7.19 (d, *J* = 7.1 Hz, 2H), 6.64 (d, *J* = 11.7 Hz, 1H), 5.78 (dd, *J* = 11.7, 9.7 Hz, 1H), 3.50 (td, *J* = 10.1, 5.4 Hz, 1H), 2.52 – 2.42 (m, 1H), 2.31 (td, *J* = 12.7, 6.1 Hz, 1H), 2.19 – 2.03 (m, 2H), 1.93 – 1.86 (m, 1H), 1.80 – 1.62 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 211.30, 136.95,

130.83, 129.37, 128.31, 128.26, 126.93, 50.16, 41.86, 34.98, 27.68, 24.33. m/z HRMS (EI) found $[M]^+$ 200.11991, $C_{14}H_{16}O^+$ requires 200.1201.

(E)-7-styryl-1,4-dioxaspiro[4.5]decan-8-one (1s)



¹H NMR (500 MHz, CDCl₃) δ 7.43 – 7.33 (m, 2H), 7.30 (dd, J = 8.7, 6.6 Hz, 2H), 7.24 – 7.19 (m, 1H), 6.47 – 6.40 (m, 1H), 6.37 (d, J = 16.1 Hz, 1H), 4.14 – 4.01 (m, 4H), 3.53 (dt, J = 12.8, 6.3 Hz, 1H), 2.72 (dddd, J = 14.3, 13.0, 6.5, 1.2 Hz, 1H), 2.47 (ddd, J = 14.4, 4.9, 3.4 Hz, 1H), 2.21 (ddd, J = 13.3, 6.0, 3.2 Hz, 1H), 2.14 – 1.98 (m, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 209.64, 136.96, 131.59, 128.50, 127.47, 126.46, 126.36, 107.15, 64.87, 64.71, 50.02, 40.84, 38.16, 34.52. m/z HRMS (EI) found [M]⁺258.1254, C₁₆H₁₈O₃⁺ requires 258.1256.

(*E*)-4,4-dimethyl-2-styrylcyclohexanone (1t)



¹H NMR (300 MHz, CDCl₃) δ 7.42 – 7.35 (m, 2H), 7.29 (ddd, J = 7.7, 6.7, 1.4 Hz, 2H), 7.24 – 7.16 (m, 1H), 6.43 (dd, J = 16.1, 6.5 Hz, 1H), 6.34 (d, J = 16.2 Hz, 1H), 3.38 – 3.22 (m, 1H), 2.54 (dddd, J = 14.3, 13.2, 6.6, 1.0 Hz, 1H), 2.33 (ddd, J = 14.4, 4.6, 3.0 Hz, 1H), 1.87 (ddd, J = 13.2, 5.9, 3.1 Hz, 1H), 1.81 – 1.63 (m, 3H), 1.28 (s, 3H), 1.06 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 211.69, 137.21, 131.19, 128.50, 127.68, 127.33, 126.32, 49.70, 47.12, 39.76, 38.26, 31.43, 30.79, 24.58. m/z HRMS (EI) found [M]⁺228.1514, C₁₆H₂₀O⁺ requires 228.1514.

Synthesis of Products

General procedure for asymmetric amination:

To the substrate **1** (0.30 mmol) in a 1 dram (15 x 45 mm) vial equipped with an 8 mm magnetic stirrer bar was added DCM (0.3 ml). Subsequently, *di*-tert-butyl azodicarboxylates (90 mg, 0.39

mmol), 5Å MS (50 mg), and (*R*)-C₈-TCYP (36 mg, 0.03 mmol or 18 mg, 0.015 mmol) were added. After all the reagents were dissolved, the mixture was warmed to 45 °C with the cap open. After about 2h, the DCM was evaporated, leaving the mixture as syrup. After heated at 45 °C for another 40h or 60h, the mixture was cooled to rt and directly purified by flash column chromatography to afford the desired product.

The relevant racemic products were synthesized in the same procedure expect (\pm)-**TRIP** (10 mol%) was used as catalyst.

(S)-di-*tert*-butyl 1-(2-oxo-1-phenylcyclohexyl)hydrazine-1,2-dicarboxylate (2a)



The reaction was carried out according the general procedure using **1a** (52 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60 h, the mixture was purified by flash column chromatography (7: 1, Hexane/ethyl acetate) to afford the title product **2a** (117 mg, 97% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.08 (m, 5H), 6.63 – 5.66 (m, 1H), 3.06 – 2.90 (m, 1H), 2.85 – 2.73 (m, 1H), 2.49 – 2.28 (m, 2H), 2.22 – 2.09 (m, 1H), 2.09 – 1.97 (m, 1H), 1.86 – 1.69 (m, 1H), 1.66 – 1.24 (m, 19H). ¹³C NMR (125 MHz, CDCl₃) δ 205.95, 155.55, 155.06, 151.03, 137.07, 128.44, 127.95, 127.78, 127.62, 83.63, 81.60, 80.98, 40.53, 28.13, 28.06, 27.93, 21.54. m/z HRMS (ESI) found [M+Na]⁺ 427.2205, C₂₂H₃₂N₂O₅Na⁺ requires 427.2203. [α]_D²⁰ = +5.1 (c 1.0, CHCl₃). HPLC (Chiralpak OD column, 98:2 hexanes/isopropanol, 0.5 ml/min; tr= 17.0 min (minor), 18.2 min (major); 99% ee.

(S)-di-tert-butyl 1-(2-oxo-1-(p-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2b)



The reaction was carried out according the general procedure using **1b** (56 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45°C for 60 h, the mixture was purified by flash column chromatography (7: 1, Hexane/ EA) to afford the title product **2b** (122 mg, 97%)

yield). ¹H NMR (400 MHz, CDCl₃) δ 7.18 – 7.04 (m, 4H), 6.59 – 5.63 (m, 1H), 3.08 – 2.66 (m, 2H), 2.46 – 2.32 (m, 2H), 2.31 (d, J = 4.8 Hz, 3H), 2.22 – 1.94 (m, 2H), 1.86 – 1.67 (m, 1H), 1.63 – 1.52 (m, 1H), 1.55 – 1.28 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 206.11, 155.62, 155.10, 137.75, 134.06, 129.34, 129.23, 128.58, 127.58, 81.55, 80.97, 40.58, 40.26, 28.18, 28.12, 27.98, 21.63, 21.03. m/z HRMS (ESI) found [M+Na]⁺ 441.2368, C₂₃H₃₄N₂O₅Na⁺ requires 441.2360. [α]_D²⁰ = +6.5 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 92:8 hexanes/ isopropanol, 1 ml/min; tr= 11.9 min (minor), 13.1 min (major); 99% ee.

(S)-di-*tert*-butyl 1-(1-(naphthalen-2-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2c)



The reaction was carried out according the general procedure using **1c** (67 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (8: 1, Hexane/ EA) to afford the title product **2c** (122 mg, 90% yield). ¹H NMR (600 MHz, CDCl₃) δ 7.86 – 7.74 (m, 3H), 7.71 – 7.60 (m, 1H), 7.53 – 7.42 (m, 3H), 6.63 – 5.71 (m, 1H), 3.13 – 2.70 (m, 2H), 2.66 – 2.36 (m, 2H), 2.28 – 2.15 (m, 1H), 2.12 – 2.05 (m, 1H), 1.91 – 1.72 (m, 1H), 1.71 – 1.60 (m, 1H), 1.51 – 1.35 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 206.12, 155.61, 155.17, 134.88, 133.03, 132.84, 128.35, 128.11, 127.41, 126.54, 126.16, 125.98, 81.07, 40.73, 28.19, 28.11, 27.96, 21.69. m/z HRMS (ESI) found [M+Na]⁺ 477.2362, C₂₆H₃₄N₂O₅Na⁺ requires 477.2360. HPLC (Chiralpak OD column, 98:2 hexanes/ isopropanol, 0.5 ml/min; tr= 17.9 min (major), 20.3 min (minor); 97% ee.

(S)-di-*tert*-butyl 1-(1-(benzo[d][1,3]dioxol-5-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2d)



The reaction was carried out according the general procedure using **1d** (65 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 40h, the mixture was purified by

flash column chromatography (5: 1, Hexane/ EA) to afford the title product **2d** (108 mg, 80% yield). ¹H NMR (500 MHz, CDCl₃) δ 6.85 – 6.76 (m, 1H), 6.76 – 6.65 (m, 2H), 6.00 – 5.63 (m, 3H), 3.05 – 2.69 (m, 2H), 2.43 – 2.22 (m, 2H), 2.17 – 1.92 (m, 2H), 1.79 – 1.66 (m, 1H), 1.66 – 1.52 (m, 1H), 1.54 – 1.32 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 205.95, 155.47, 155.13, 148.00, 147.34, 130.92, 130.26, 121.05, 108.69, 107.94, 101.30, 101.02, 81.07, 40.63, 28.20, 28.14, 27.85, 21.70. m/z HRMS (ESI) found [M+Na]⁺ 471.2100, C₂₃H₃₂N₂O₇Na⁺ requires 471.2102. [α]_D²⁰ = +12.5 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 85:15 hexanes/ isopropanol, 1 ml/min; tr = 10.9 min (minor), 17.2 min (major); 98% ee.

(S)-di-tert-butyl 1-(1-(3-methoxyphenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2e)



The reaction was carried out according the general procedure using **1e** (61 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (6: 1, Hexane/ EA) to afford the title product **2e** (114 mg, 88% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.26 – 7.19 (m, 1H), 6.88 – 6.75 (m, 3H), 6.42 – 5.63 (m, 1H), 3.75 (s, 3H), 3.09 – 2.70 (m, 2H), 2.49 – 2.27 (m, 2H), 2.25 – 1.95 (m, 2H), 1.82 – 1.69 (m, 1H), 1.62 – 1.53 (m, 1H), 1.48 – 1.35 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 205.91, 159.69, 159.14, 155.64, 155.10, 138.90, 129.50, 129.39, 120.72, 119.94, 114.07, 113.84, 113.34, 81.06, 75.25, 55.26, 40.70, 36.69, 28.18, 28.12, 21.61. m/z HRMS (ESI) found [M+Na]⁺ 457.2310, C₂₃H₃₄N₂O₆Na⁺ requires 457.2309. [α]_D²⁰ = +5.3 (c 1.0, CHCl₃). HPLC (Chiralpak IA column, 90:10 hexanes/ isopropanol, 1 ml/min; tr= 10.1 min (minor), 13.1 min (major); 99% ee.

(S)-di-tert-butyl 1-(1-(4-bromophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2f)



The reaction was carried out according the general procedure using **1f** (76 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by

flash column chromatography (8: 1, Hexane/ethyl acetate) to afford the title product **2f** (136 mg, 80% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, J = 8.6 Hz, 2H), 7.18 – 7.09 (m, 2H), 6.66 – 5.75 (m, 1H), 3.10 – 2.93 (m, 1H), 2.89 – 2.58 (m, 1H), 2.48 – 2.33 (m, 1H), 2.32 – 1.94 (m, 3H), 1.86 – 1.68 (m, 1H), 1.57 – 1.26 (m, 19H). ¹³C NMR (125 MHz, CDCl₃) δ 205.70, 155.56, 155.16, 136.63, 131.54, 130.81, 129.51, 125.30, 122.27, 81.36, 40.48, 28.19, 28.09, 27.99, 21.38. m/z HRMS (ESI) found [M+Na]⁺ 505.1313, C₂₂H₃₁N₂O₅BrNa⁺ requires 505.1309. [α]_D²⁰ = +0.4 (c 1.0, CHCl₃).HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr = 9.0 min (minor), 16.0 min (major); 98% ee.

(S)-di-*tert*-butyl 1-(2-oxo-1-(4-(trifluoromethyl)phenyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2g)



The reaction was carried out according the general procedure using **1g** (72 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (8: 1, Hexane/ EA) to afford the title product **2g** (101 mg, 71% yield). ¹H NMR (600 MHz, CDCl₃) δ 7.72 – 7.51 (m, 2H), 7.44 – 7.30 (m, 2H), 6.64 – 5.69 (m, 1H), 3.17 – 2.95 (m, 1H), 2.92 – 2.67 (m, 1H), 2.61 – 2.36 (m, 1H), 2.32 – 2.15 (m, 1H), 2.15 – 2.05 (m, 1H), 1.87 – 1.71 (m, 1H), 1.64 – 1.55 (m, 1H), 1.52 – 1.34 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 205.71, 155.70, 155.21, 141.90, 130.07 (q, *J* = 32.6 Hz), 129.09, 128.11, 125.32 (q, *J* = 3.7 Hz), 125.14, 124.62 (q, *J* = 2.9 Hz), 122.98, 81.57, 40.48, 28.65, 28.20, 28.09, 28.00, 21.22. m/z HRMS (ESI) found [M+Na]⁺ 495.2078, C₂₃H₃₁F₃N₂O₅Na⁺ requires 495.2077. [α]_D²⁰ = -3.4 (c 0.5, CHCl₃). HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr = 6.7 min (minor), 10.6 min (major); 98% ee.

(*S*)-di-*tert*-butyl 1-(1-(4-(tert-butoxycarbonyl)phenyl)-2-oxocyclohexyl)hydrazine-1,2dicarboxylate (**2h**)



The reaction was carried out according the general procedure using **1h** (82 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (7: 1, Hexane/ EA) to afford the title product **2h** (115 mg, 76% yield). ¹H NMR (600 MHz, CDCl₃) δ 8.01 – 7.84 (m, 2H), 7.57 – 7.20 (m, 2H), 6.80 – 5.00 (m, 1H), 3.13 – 2.51 (m, 2H), 2.45 – 1.95 (m, 4H), 1.86 – 1.67 (m, 1H), 1.65 – 1.09 (m, 28H). ¹³C NMR (125 MHz, CDCl₃) δ 205.58, 165.65, 165.39, 155.58, 155.20, 142.11, 131.52, 129.43, 128.84, 127.52, 81.94, 81.35, 81.16, 40.49, 36.68, 28.22, 28.17, 28.07, 27.97, 21.34. m/z HRMS (ESI) found [M+Na]⁺ 527.2728, C₂₇H₄₀N₂O₇Na⁺ requires 527.2728. [α]_D²⁰ = +4.7 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 85:15 hexanes/ isopropanol, 1 ml/min; tr= 8.0 min (minor), 16.8 min (major); 97% ee.

(S)-di-tert-butyl 1-(2-oxo-1-(o-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2i)



The reaction was carried out according the general procedure using **1i** (56 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (9: 1, Hexane/ethyl acetate) to afford the title product **2i** (117 mg, 73% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.20 – 7.11 (m, 3H), 7.01 (d, *J* = 7.5 Hz, 1H), 6.45 – 5.96 (m, 1H), 2.99 – 2.57 (m, 3H), 2.44 – 2.32 (m, 1H), 2.57 – 2.22 (m, 3H), 2.11 – 1.58 (m, 4H), 1.53 – 1.31 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 204.87, 155.40, 155.01, 138.46, 138.27, 137.65, 137.37, 135.36, 135.18, 133.41, 132.79, 129.45, 128.02, 127.85, 127.20, 125.95, 125.88, 125.19, 82.40, 81.49, 81.28, 80.88, 78.18, 41.52, 40.84, 40.53, 35.97, 35.90, 35.74, 28.29, 28.13, 27.76, 26.83, 23.05, 22.18, 21.68. m/z HRMS (ESI) found [M+Na]⁺ 441.2358, C₂₃H₃₄N₂O₅Na⁺ requires 441.2360. [α]_D²⁰ = +57.3 (c 1.0, CHCl₃). HPLC (Chiralpak OD column, 98:2 hexanes/ isopropanol, 1 ml/min; tr = 7.6 min (major), 8.6 min (minor); 99% ee.

(S)-di-tert-butyl 1-(1-(2-chlorophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2j)



The reaction was carried out according the general procedure using **1j** (62 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (9: 1, Hexane/ethyl acetate) to afford the title product **2j** (98 mg, 78% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.44 – 7.34 (m, 1H), 7.33 – 7.11 (m, 3H), 6.67 – 6.24 (m, 1H), 3.40 – 2.74 (m, 2H), 2.71 – 2.58 (m, 1H), 2.58 – 2.44 (m, 1H), 2.11 – 1.83 (m, 3H), 1.66 – 1.53 (m, 1H), 1.62 – 1.38 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 204.20, 155.16, 154.92, 151.07, 135.75, 134.44, 132.34, 131.63, 129.11, 128.91, 128.73, 126.81, 100.04, 83.72, 81.60, 81.10, 77.67, 77.55, 40.84, 40.41, 40.15, 36.59, 36.40, 28.37, 28.25, 28.16, 28.06, 27.97, 27.93, 26.89, 21.55. m/z HRMS (ESI) found [M+Na]⁺ 461.1813, C₂₂H₃₁ClN₂O₅Na⁺ requires 461.1814. [α]_D²⁰ = +33.2 (c 1.0, CHCl₃). HPLC (Chiralpak OD column, 98:2 hexanes/ isopropanol, 1 ml/min; tr= 9.8 min (minor), 10.8 min (major); 99% ee.

(S)-di-tert-butyl 1-(2-oxo-1-phenylcyclopentyl)hydrazine-1,2-dicarboxylate (2k)



The reaction was carried out according the general procedure using **1k** (48 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 40h, the mixture was purified by flash column chromatography (8: 1, Hexane/ EA) to afford the title product **2k** (108 mg, 78% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.58 – 7.46 (m, 2H), 7.38 – 7.31 (m, 2H), 7.31 – 7.27 (m, 1H), 6.12 – 5.18 (m, 1H), 3.00 – 2.72 (m, 2H), 2.70 – 2.53 (m, 1H), 2.17 (dd, *J* = 19.0, 8.5 Hz, 1H), 2.11 – 2.02 (m, 1H), 1.72 – 1.57 (m, 1H), 1.48 – 1.28 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 211.82, 211.29, 155.00, 154.77, 154.57, 154.23, 154.06, 133.92, 133.14, 128.94, 128.71, 128.48, 128.40, 128.33, 128.29, 83.76, 82.13, 81.68, 80.91, 73.80, 73.55, 35.12, 34.87, 31.48, 31.18, 28.31, 28.22, 28.17, 28.13, 28.00, 18.19. m/z HRMS (ESI) found [M+Na]⁺

413.2050, $C_{21}H_{30}N_2O_5Na^+$ requires 417.2047. HPLC (Chiralpak OD column, 98.5:1.5 hexanes/ isopropanol, 0.5 ml/min; tr= 19.3 min (major), 20.7 min (minor); >95% ee.

(R)-di-tert-butyl 1-(4-oxo-3-phenyltetrahydro-2H-pyran-3-yl)hydrazine-1,2-dicarboxylate (2I)



The reaction was carried out according the general procedure using **11** (53 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 40h, the mixture was purified by flash column chromatography (20: 1, DCM/ Et₂O) to afford the title product **21** (118 mg, 97% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.68 – 7.36 (m, 2H), 7.34 (t, *J* = 7.4 Hz, 2H), 7.30 (d, *J* = 7.2 Hz, 1H), 6.93 – 5.75 (m, 1H), 4.72 (d, *J* = 12.2 Hz, 1H), 4.60 – 4.36 (m, 1H), 4.29 – 3.84 (m, 2H), 2.91 – 2.57 (m, 1H), 2.52 – 2.35 (m, 1H), 1.48 – 1.26 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 200.96, 155.05, 134.58, 128.84, 128.69, 128.56, 128.42, 128.21, 128.04, 100.06, 82.25, 81.91, 81.14, 73.25, 71.45, 67.61, 67.06, 40.26, 28.20, 28.11, 28.00. m/z HRMS (ESI) found [M+Na]⁺ 429.1997, C₂₁H₃₀N₂O₆Na⁺ requires 429.1996. [α]_D²⁰ = +83.0 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 85:15 hexanes/ isopropanol, 1 ml/min; tr = 5.6 min (major), 8.3 min (minor); 99% ee.

(*R*)-di-*tert*-butyl 1-(1-(tert-butoxycarbonyl)-4-oxo-3-phenylpiperidin-3-yl)hydrazine-1,2dicarboxylate (**2m**)



The reaction was carried out according the general procedure using **1m** (82 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (10: 1, DCM/ Et₂O) to afford the title product **2m** (120 mg, 79% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.56 – 7.38 (m, 2H), 7.33 – 7.25 (m, 3H) , 6.51 – 5.50 (m, 1H) , 5.20 – 3.92 (m, 2H) , 3.81 – 3.25 (m, 1H) , 2.91 – 2.43 (m, 1H) , 2.48 – 1.98 (m, 1H) , 1.60 – 1.06 (m, 28H). ¹³C NMR (125 MHz, CDCl₃) δ 202.85, 155.84, 155.08, 154.00, 133.73, 129.03,

128.92, 128.80, 128.64, 128.56, 82.19, 81.70, 81.33, 80.88, 80.78, 80.41, 73.60, 48.07, 41.03, 39.20, 38.86, 28.63, 28.36, 28.21, 28.16, 28.04, 27.99, 27.77. m/z HRMS (ESI) found $[M+Na]^+$ 528.2681, $C_{26}H_{39}N_3O_7Na^+$ requires 528.2680. $[\alpha]_D{}^{20} = +38.0$ (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 85:15 hexanes/ isopropanol, 1 ml/min; tr = 6.6 min (major), 8.8 min (minor); 96% ee.

(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2n**)



The reaction was carried out according the general procedure using **1n** (60 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (10: 1, Hexane/ethyl acetate) to afford the title product **2n** (128 mg, 99% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.34 (m, 2H), 7.30 (t, *J* = 7.7 Hz, 2H), 7.26 – 7.19 (m, 1H), 6.51 – 6.25 (m, 2H), 6.24 – 6.05 (m, 1H), 3.14 – 2.80 (m, 1H), 2.64 (dt, *J* = 14.4, 4.6 Hz, 1H), 2.37 (dt, *J* = 12.5, 5.0 Hz, 1H), 2.20 – 2.00 (m, 2H), 1.99 – 1.84 (m, 1H), 1.82 – 1.67 (m, 1H), 1.65 – 1.34 (m, 19H). ¹³C NMR (100 MHz, CDCl₃) δ 207.94, 155.49, 136.26, 130.93, 128.64, 128.43, 128.10, 127.80, 126.73, 81.35, 72.60, 39.47, 28.24, 28.11, 21.80, 20.89. m/z HRMS (ESI) found [M+Na]⁺ 453.2347, C₂₄H₃₄N₂O₅Na⁺ requires 453.2360. [α]_D²⁰ = -18.0 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 94:6 hexanes/ isopropanol, 1 ml/min; tr= 11.1 min (minor), 11.9 min (major); 98% ee.

(S)-di-tert-butyl 1-(2-oxo-1-vinylcyclohexyl)hydrazine-1,2-dicarboxylate (20)



The reaction was carried out according the general procedure using **10** (37 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 40h, the mixture was purified by flash column chromatography (7: 1, Hexane/ethyl acetate) to afford the title product **20** (95 mg, 89% yield). ¹H NMR (500 MHz, CDCl₃) δ 6.34 – 6.02 (m, 1H), 5.95 (dd, *J* = 17.7, 10.8 Hz, 1H),

5.23 (d, J = 10.9 Hz, 1H), 4.96 (d, J = 17.7 Hz, 1H), 3.20 – 2.71 (m, 1H), 2.54 – 2.40 (m, 1H), 2.31 (dd, J = 10.9, 5.9 Hz, 1H), 2.16 – 1.92 (m, 2H), 1.82 – 1.57 (m, 2H), 1.58 – 1.25 (m, 19H). ¹³C NMR (125 MHz, CDCl₃) & 208.08, 155.53, 136.37, 115.65, 81.37, 72.64, 39.33, 38.97, 29.04, 28.25, 28.13, 28.05, 27.99, 21.60, 20.58. m/z HRMS (ESI) found [M+Na]⁺ 377.2049, C₁₈H₃₀N₂O₅Na⁺ requires 377.2047. [α]_D²⁰ = -54.5 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr= 10.1 min (minor), 25.0 min (major); 96% ee.

(*S*,*Z*)-di-*tert*-butyl 1-(2-oxo-1-(prop-1-en-1-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2p**)



The reaction was carried out according the general procedure using **1p** (41 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (8: 1, Hexane/ EA) to afford the title product **2p** (108 mg, 98% yield). ¹H NMR (500 MHz, CDCl₃) δ 6.61 – 5.86 (m, 1H), 5.80 – 5.30 (m, 2H), 2.86 – 2.27 (m, 3H), 2.22 – 1.97 (m, 1H), 2.00 – 1.70 (m, 4H), 1.59 (dd, *J* = 7.5, 1.9 Hz, 3H), 1.41 (d, *J* = 16.1 Hz, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 207.26, 205.82, 155.59, 155.43, 155.25, 154.90, 130.95, 128.31, 128.08, 127.69, 81.61, 81.04, 80.78, 73.94, 73.11, 39.79, 39.15, 38.34, 37.11, 28.26, 28.16, 28.10, 27.97, 27.24, 21.93, 21.66, 14.56, 14.46. m/z HRMS (ESI) found [M+Na]⁺ 391.2205, C₁₉H₃₂N₂O₅Na⁺ requires 391.2203. [α]_D²⁰ = +2.3 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr = 11.6 min (minor), 15.7 min (major); 99% ee.

(*S*,*Z*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2q**)



The reaction was carried out according the general procedure using **1q** (60 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (9: 1, Hexane/ethyl acetate) to afford the title product **2q** (128 mg, 99% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.44 – 7.32 (m, 2H), 7.30 – 7.25 (m, 1H), 7.19 – 6.99

(m, 2H), 6.83 - 6.59 (m, 1H), 6.53 - 6.28 (m, 1H), 4.75 - 2.86 (m, 1H), 2.58 - 2.22 (m, 1H), 2.17 (d, J = 11.6 Hz, 1H), 2.07 - 1.88 (m, 1H), 1.84 - 1.54 (m, 3H), 1.53 - 1.45 (m, 1H), 1.45 - 1.31 (m, 19H). ¹³C NMR (125 MHz, CDCl₃) δ 208.98, 155.51, 155.11, 154.23, 138.16, 137.83, 132.06, 131.60, 129.65, 129.34, 128.86, 128.71, 128.45, 128.12, 127.76, 127.62, 127.10, 82.05, 80.70, 71.41, 39.74, 39.15, 37.51, 36.93, 36.18, 29.75, 28.24, 28.08, 28.05, 27.94, 21.66, 20.19. m/z HRMS (ESI) found [M+Na]⁺ 453.2367, C₂₄H₃₄N₂O₅Na⁺ requires 453.2360. HPLC (Chiralpak IC column, 92:8 hexanes/ isopropanol, 1 ml/min; tr= 10.7 min (minor), 14.5 min (major); 99% ee.

(S)-di-*tert*-butyl 1-(2-oxo-[1,1'-bi(cyclohexan)]-1'-en-1-yl)hydrazine-1,2-dicarboxylate (2r)



The reaction was carried out according the general procedure using **1r** (53 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (10: 1, Hexane/ethyl acetate) to afford the title product **2r** (118 mg, 96% yield). ¹H NMR (500 MHz, CDCl₃) δ 6.24 – 5.47 (m, 1H), 5.41 (t, *J* = 3.8 Hz, 1H), 2.66 (dq, *J* = 12.0, 6.2 Hz, 1H), 2.52 (ddd, *J* = 13.5, 9.2, 3.5 Hz, 1H), 2.32 (ddd, *J* = 14.3, 8.9, 5.5 Hz, 1H), 2.21 (ddd, *J* = 14.1, 7.5, 3.4 Hz, 1H), 2.14 – 2.01 (m, 2H), 1.98 – 1.89 (m, 2H), 1.88 – 1.78 (m, 2H), 1.76 – 1.69 (m, 1H), 1.66 – 1.58 (m, 1H), 1.58 – 1.47 (m, 4H), 1.47 – 1.35 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 207.12, 206.43, 155.74, 155.25, 134.25, 133.78, 127.64, 127.45, 82.15, 81.21, 80.71, 40.87, 40.78, 40.73, 36.69, 33.98, 28.24, 28.20, 28.14, 27.96, 27.02, 26.06, 26.00, 25.87, 25.78, 22.85, 22.73, 22.06, 21.94. m/z HRMS (ESI) found [M+Na]⁺ 431.2523, C₂₂H₃₆N₂O₅Na⁺ requires 431.2516. HPLC (Chiralpak IC column, 96:4 hexanes/ isopropanol, 1 ml/min; tr= 16.4 min (minor), 37.6 min (major); 95% ee.

(*S*,*E*)-di-*tert*-butyl 1-(8-oxo-7-styryl-1,4-dioxaspiro[4.5]decan-7-yl)hydrazine-1,2-dicarboxylate (2s)



The reaction was carried out according the general procedure using **1s** (77 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (3: 1, Hexane/ethyl acetate) to afford the title product **2s** (120 mg, 82% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.36 (dd, *J* = 15.3, 7.5 Hz, 2H), 7.33 – 7.13 (m, 3H), 6.93 – 5.87 (m, 3H), 4.22 – 3.85 (m, 4H), 3.00 – 2.71 (m, 1H), 2.74 – 2.47 (m, 1H), 2.45 – 2.30 (m, 1H), 2.23 (dd, *J* = 24.3, 14.5 Hz, 1H), 2.11 (m, 1H), 2.00 – 1.82 (m, 1H), 1.53 – 1.07 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 205.18, 204.93, 204.08, 155.34, 154.71, 154.17, 153.76, 137.21, 136.57, 136.51, 134.30, 130.73, 130.01, 128.61, 128.45, 128.28, 128.21, 127.80, 127.43, 127.34, 127.27, 126.88, 126.71, 107.69, 107.19, 107.07, 82.34, 81.60, 81.41, 80.92, 80.76, 68.98, 65.37, 65.14, 64.68, 64.17, 64.11, 64.02, 43.25, 42.07, 41.11, 36.78, 36.31, 36.20, 35.98, 35.09, 34.93, 32.13, 28.22, 28.16, 28.13, 27.66, 14.23. m/z HRMS (ESI) found [M+Na]⁺ 511.2415, C₂₆H₃₆N₂O₇Na⁺ requires 511.2415. [α]_D²⁰ = -9.3 (c 1.0, CHCl₃). HPLC (Chiralpak ASH column, 80:20 hexanes/ isopropanol, 1 ml/min; tr= 24.1 min (minor), 28.3 min (major); 97% ee.

(*S*,*E*)-di-*tert*-butyl 1-(5,5-dimethyl-2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (2t)



The reaction was carried out according the general procedure using **1t** (68 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (9: 1, Hexane/ethyl acetate) to afford the title product **2t** (113 mg, 82% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.43 – 7.23 (m, 5H), 6.55 – 6.21 (m, 2H), 6.04 (d, *J* = 16.5 Hz, 1H), 2.73 (d, *J* = 14.1 Hz, 1H), 2.63 – 2.39 (m, 2H), 2.28 – 2.01 (m, 1H), 2.01 – 1.73 (m, 1H), 1.60 – 1.52 (m, 1H), 1.49 – 1.33 (m, 18H), 1.07 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 207.01, 205.97, 155.55, 154.28, 136.30, 133.79, 129.05, 128.73, 128.37, 126.77, 81.98, 80.87, 71.55, 48.84, 40.46, 36.79, 36.56, 32.68, 31.10, 28.32, 28.17, 27.97, 26.58, 22.72, 22.70, 14.20,

14.17. m/z HRMS (ESI) found $[M+Na]^+ 481.2670$, $C_{26}H_{38}N_2O_5Na^+$ requires 481.2673. $[\alpha]_D^{20} = +66.0$ (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 92:8 hexanes/ isopropanol, 1 ml/min; tr= 6.1 min (minor), 8.2 min (major); 98% ee.

(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcycloheptyl)hydrazine-1,2-dicarboxylate (**2u**)



The reaction was carried out according the general procedure using **1u** (64 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 60h, the mixture was purified by flash column chromatography (9: 1, Hexane/ EA) to afford the title product **2u** (80 mg, 60% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.46 – 7.35 (m, 2H), 7.30 (t, *J* = 7.6 Hz, 2H), 7.23 (t, *J* = 7.4 Hz, 1H), 6.94 – 5.86 (m, 3H), 3.55 – 2.57 (m, 1H), 2.52 – 2.34 (m, 1H), 2.33 – 2.18 (m, 1H), 2.12 – 1.89 (m, 1H), 1.91 – 1.53 (m, 6H), 1.53 – 1.38 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 208.25, 155.90, 155.14, 137.38, 136.50, 135.73, 128.63, 128.51, 128.36, 127.89, 126.72, 126.46, 82.06, 81.44, 29.40, 28.28, 28.20, 28.12, 28.01, 24.18, 22.74. m/z HRMS (ESI) found [M+Na]⁺ 467.2519, C₂₅H₃₆N₂O₅Na⁺ requires 467.2516. [α]_D²⁰ = -9.3 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr= 7.0 min (major), 7.9 min (minor); 72% ee.

(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclopentyl)hydrazine-1,2-dicarboxylate (**2v**)



The reaction was carried out according the general procedure using **1v** (56 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45 °C for 40h, the mixture was purified by flash column chromatography (9: 1, Hexane/ethyl acetate) to afford the title product **2v** (123 mg, 99% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.33 (m, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 7.28 – 7.24 (m, 1H), 6.59 (d, *J* = 16.3 Hz, 1H), 6.48 – 6.14 (m, 1H), 5.90 (dd, *J* = 16.3, 4.3 Hz, 1H), 2.87 – 2.36 (m, 3H), 2.27 (dd, *J* = 18.8, 8.3 Hz, 1H), 2.13 – 1.98 (m, 1H), 1.86 – 1.66 (m, 1H),

1.53 – 1.32 (m, 18H). ¹³C NMR (125 MHz, CDCl₃) δ 212.56, 212.01, 155.42, 154.12, 153.62, 136.25, 135.97, 133.47, 133.42, 128.71, 128.48, 128.35, 126.82, 124.93, 124.58, 82.17, 81.77, 81.19, 72.67, 35.04, 34.81, 31.93, 28.28, 28.21, 28.17, 27.97, 18.30. m/z HRMS (ESI) found [M+Na]⁺ 439.2208, C₂₃H₃₂N₂O₅Na⁺ requires 439.2203. [α]_D²⁰ = +115.5 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 93:7 hexanes/ isopropanol, 1 ml/min; tr= 6.6 min (minor), 7.4 min (major); 99% ee.

(*S*)-di-tert-butyl 1-(2-oxo-1-(phenylethynyl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2w**)



The reaction was carried out according the general procedure using **1w** (59 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45°C for 40 h, the mixture was purified by flash column chromatography (10: 1, hexanes/ ethyl acetate) to afford the title product **2w** (42 mg, 33% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.36 (m, 2H), 7.34 – 7.26 (m, 3H), 6.63 – 6.02 (m, 1H), 3.07 – 2.13 (m, 4H), 2.13 – 1.90 (m, 2H), 1.90 – 1.65 (m, 2H), 1.46 (d, *J* = 6.7 Hz, 18H). ¹³C NMR (101 MHz, CDCl₃) δ 155.27, 153.50, 152.30, 149.75, 131.97, 131.75, 129.05, 128.79, 128.72, 128.33, 122.27, 115.47, 85.08, 83.63, 81.67, 81.32, 79.67, 38.44, 36.73, 28.29, 28.23, 28.16, 28.02, 27.61, 27.55, 22.51, 21.97, 21.20. m/z HRMS (EI) found [M]⁺ 428.2307, C₂₄H₃₂N₂O₅⁺ requires 428.2311. [α]_D²⁰ = +44.5 (c 1.0, CHCl₃). HPLC (Chiralpak OD column, 98:2 hexanes/isopropanol, 0.5 ml/min; tr= 17.3 min (minor), 24.2 min (major); 93% ee.

(*R*)-di-tert-butyl 1-(1-methyl-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2**x**)



The reaction was carried out according the general procedure using 1x (33 mg, 0.3 mmol) and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring at 45°C for 40 h, the mixture was purified by flash column chromatography (5: 1, hexanes/ ethyl acetate) to afford the title product 2x (96 mg, 94% yield). ¹H NMR (400 MHz, CDCl₃) δ 6.85 – 6.19 (m, 1H), 3.03 – 2.48 (m, 1H), 2.49 – 2.12

(m, 2H), 2.05 - 1.80 (m, 1H), 1.73 - 0.99 (m, 25H). ¹³C NMR (100 MHz, CDCl₃) δ 209.46, 208.91, 155.65, 155.04, 154.81, 82.14, 81.26, 81.08, 69.96, 68.86, 39.89, 38.76, 38.31, 29.42, 28.15, 28.02, 21.77, 20.99, 20.23. m/z HRMS (EI) found [M]⁺ 342.2158, C₁₇H₃₀N₂O₅⁺ requires 342.2155. [α]_D²⁰ = -18.9 (c 1.0, CHCl₃).

(R)-N-(1-methyl-2-oxocyclohexyl)benzamide (S4)



To a solution of 2x (78 mg, 0.23 mmol) in DCM (3 mL) was added TFA (1.5 ml) at rt. After stirring for 3h, acetone (1.5 mL) was added and the mixture was stirred for another 10 min before concentration *in vacuo*.

To the solution of the above residue in HOAc (3 ml) was added active zinc powder (300 mg). After purging with N₂, the mixture was stirred overnight. After filtration, the filtrate was concentrated *in vacuo* to afford a residue, which was diluted with DCM and basified with satd. Na₂CO₃ solution. Extraction with DCM five times to afford the organic layer, which was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue.

To the solution of the above residue in DCM (3 mL) was added Et₃N (95 uL, 0.68 mmol) and BzCl (53 uL, 0.46 mmol) at rt. After stirring for 3 h, satd. NaHCO₃ solution was added into the mixture, which was then extracted with DCM for 3 times. The combined organic layer was washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography (3: 1, Hexane/ ethyl acetate) to afford **S4** (11 mg, 21% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.80 (d, *J* = 7.6 Hz, 1H), 7.66 (s, 1H), 7.53 – 7.46 (m, 1H), 7.43 (dd, *J* = 8.2, 6.7 Hz, 1H), 3.07 (ddd, *J* = 11.0, 4.2, 2.2 Hz, 1H), 2.66 (ddd, *J* = 13.8, 12.1, 6.3 Hz, 1H), 2.56 – 2.45 (m, 1H), 2.09 (ddd, *J* = 9.4, 5.9, 3.2 Hz, 1H), 1.88 – 1.75 (m, 3H), 1.72 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 210.94, 166.21, 135.17, 131.54, 128.66, 127.05, 62.80, 38.92, 37.55, 27.98, 22.04, 21.36. [α]_D²⁰ = +9.2 (c 0.5, CHCl₃). HPLC (Chiralpak IC column, 80:20 hexanes/isopropanol, 1.0 ml/min; tr= 12.6 min (minor), 15.7 min (major); 86% ee.

(S)-di-tert-butyl 1-(1-benzyl-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2y)



The reaction was carried out according the general procedure using **1y** (57 mg, 0.3 mmol) and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring at 45°C for 40 h, the mixture was purified by flash column chromatography (10: 1, Hexane/ EA) to afford the title product **2y** (111 mg, 89% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.26 (m, 3H), 7.11 – 6.99 (m, 2H), 5.18 – 4.43 (m, 1H), 3.58 – 2.87 (m, 2H), 2.83 – 2.63 (m, 1H), 2.57 – 2.38 (m, 1H), 2.35 – 2.19 (m, 1H), 2.15 – 2.00 (m, 1H), 1.88 – 1.57 (m, 2H), 1.53 – 1.33 (m, 20H). ¹³C NMR (101 MHz, CDCl₃) δ 210.42, 210.05, 155.71, 155.41, 155.12, 154.59, 137.65, 137.23, 130.34, 130.24, 128.99, 128.79, 127.32, 126.96, 82.62, 82.47, 82.02, 81.23, 72.16, 71.48, 71.40, 40.48, 39.20, 38.96, 38.52, 38.16, 37.98, 36.74, 30.44, 30.11, 29.95, 28.18, 20.07. [α]_D²⁰ = -92.5 (c 1.0, CHCl₃). HPLC (Chiralpak IA column, 97:3 hexanes/isopropanol, 1 ml/min; tr= 11.6 min (minor), 12.8 min (major); 99% ee.

Kinetic resolution procedure:

(*R*)-di-*tert*-butyl 1-(2-oxo-1-(thiophen-2-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (2z)



To a solution of **1z** (54 mg, 0.3 mmol) in DCM (0.6 mL) was added *di*-tertbutyl azodicarboxylates (90 mg, 0.39 mmol), 5Å MS (50 mg), and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring for 60 at rt, the mixture was purified by flash column chromatography (10: 1 to 6: 1, Hexane/ethyl acetate) to afford the recovered starting material **1z** (24 mg, 45% yield, 96% ee) and the title product **2z** (68 mg, 55% yield, 93% ee). HPLC data for recovered **1z**: HPLC (Chiralpak IC column, 95:5 hexanes/ isopropanol, 1 ml/min; tr = 17.8 min (major), 20.2 min (minor); 96% ee. Data for **2z**: ¹H NMR (500 MHz, CDCl₃) δ 7.29 (d, *J* = 5.1 Hz, 1H), 6.95 (t, *J* = 4.4 Hz, 1H), 6.93 – 6.87 (m, 1H, 6.06 – 5.55 (m, 1H), 3.17 – 2.80 (m, 2H), 2.50 – 2.36 (m, 1H), 2.29 – 1.98 (m, 3H), 1.80 – 1.66 (m, 1H), 1.63 – 1.55 (m, 1H), 1.52 – 1.38 (m, 18H). ¹³C

NMR (125 MHz, CDCl₃) δ 155.24, 141.54, 129.32, 126.47, 126.42, 120.92, 114.48, 112.17, 83.71, 81.73, 81.28, 72.40, 39.65, 28.20, 28.08, 27.97, 21.24. m/z HRMS (ESI) found [M+Na]⁺ 433.1767, C₂₀H₃₀N₂O₅SNa⁺ requires 433.1768. [α]_D²⁰ = -53.8 (c 1.0, CHCl₃). HPLC (Chiralpak IC column, 92:8 hexanes/ isopropanol, 1 ml/min; tr= 12.4 min (minor), 17.8 min (major); 93% ee.

Kinetic resolution conditions for 1n and 1t:

To a solution of **1n or 1t** (0.3 mmol) in DCM (3 mL) was added *di*-tert-butyl azodicarboxylates (90 mg, 0.36 mmol), 5 Å MS (50 mg), and (*R*)-C₈-TCYP (18 mg, 0.015 mmol). After stirring for 40h at rt, the mixture was purified by flash column chromatography (15: 1 to 10: 1, hexane/ ethyl acetate) to afford the recovered starting material and the amination product.

For **1n**: Starting ketone **1n** (25 mg, 41% yield, 99% ee) was recovered with product **2a** (76 mg, 59% yield, 99% ee) obtained. HPLC data for recovered **1n**: HPLC (Chiralpak IC column, 95:5 hexanes/ isopropanol, 1 ml/min; tr = 15.8 min (major), 20.5 min (minor); 99% ee.

For **1t**: Starting ketone **1t** (20 mg, 30% yield, 97% ee) was recovered with product **2t** (96 mg, 70% yield, 97% ee) obtained. HPLC data for recovered **1a**: HPLC (Chiralpak IC column, 95:5 hexanes/ isopropanol, 1 ml/min; tr = 14.6 min (major), 21.1 min (minor); 97% ee.

Determine of the absolute configuration of recovered 1a:



To a solution of **1a** (14 mg, 0.07 mmol) in ethyl acetate (2 mL) was added 10% Pd/C (40 mg) at rt. After purging with H₂, the mixture was stirred at rt for 3h. Filtration and concentration in vacuo gave a residue, which was purified by flash column chromatography (20 :1, hexane/ ethyl acetate) to give the title product **S4** (6.7 mg, 47% yield). The absolute configuration of the stereocenter was determined to be (*S*) by comparison of the optical rotation of **S4** with literature¹⁵.

Large-scale synthesis example:

To the substrate **1a** (348 mg, 2.0 mmol) in a 1 dram (15 x 45 mm) vial equipped with an 8 mm magnetic stirrer bar was added DCM (1.0 ml). Subsequently, *di*-tert-butyl azodicarboxylates (598 mg, 2.6 mmol), 5Å MS (300 mg), and (*R*)-C₈-TCYP (240 mg, 0.2 mmol) were added. After all the reagents were dissolved, the mixture was warmed to 45 °C with the cap open. After about 2h, the DCM was evaporated, leaving the mixture as syrup. After heated at 45 °C for another 48 h, the mixture was cooled to rt and directly purified by flash column chromatography (10: 1 to 3: 1 hexane: ethyl acetate) to afford the desired product **2a** (775 mg, 96% yield, 99% ee) with recovery of (*R*)-C₈-TCYP (83 mg).

Transformation of the products:

N-((*1S*,*2R*)-2-hydroxy-1-phenylcyclohexyl)benzamide (**3a**)



To a solution of **2a** (130 mg, 0.322 mmol) in DCM (3 mL) was added TFA (1.5 mL) at rt. After stirring for 3 h, the mixture was concentrated to afford a residue.

To the solution of the above residue in MeOH (4 mL) was added Raney Ni (200 mg). After purging with H_2 , the mixture was stirred overnight under sonication. Then the mixture was filtered and concentrated to give a residue.

To the solution of the above residue in DCM (3 mL) was added Et₃N (134 uL, 0.97 mmol) and BzCl (75 uL, 0.64 mmol) at rt. After stirring for 3 h, satd. NaHCO₃ solution was added into the mixture, which was then extracted with DCM for 3 times. The combined organic layer was washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography (3: 1, Hexane/ ethyl acetate) to afford **3a** (66 mg, 70% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.73 (d, *J* = 7.4 Hz, 2H), 7.65 (d, *J* = 7.8 Hz, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.42 (t, *J* = 7.5 Hz, 2H), 7.36 (t, *J* = 7.6 Hz, 2H), 7.30 – 7.23 (m, 1H), 6.46 (s, 1H), 4.13 (t, *J* = 4.9 Hz, 1H), 3.45 – 2.56 (brs, 1H), 2.53 – 2.31 (m, 2H), 2.09 – 1.89 (m, 2H), 1.83 – 1.68 (m, 1H), 1.65 – 1.43 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.99, 142.50, 135.08, 131.73, 128.75, 128.61, 127.43, 127.19, 127.01, 74.64, 62.14, 31.29, 29.88, 21.61, 21.13.

m/z HRMS (ESI) found $[M+H]^+ 296.1645$, $C_{19}H_{22}NO_2^+$ requires 296.1645. $[\alpha]_D^{20} = -32.6$ (c 1.0, CHCl₃). HPLC (Chiralpak IA column, 90:10 hexanes/ isopropanol, 1 ml/min; tr= 17.1 min (minor), 18.5 min (major); 99% ee.

(S)-Ketamine:



To a solution of **2j** (255 mg, 0.58 mmol) in DCM (5 mL) was added TFA (2.5 ml) at rt. After stirring for 3h, acetone (3 mL) was added and the mixture was stirred for another 10 min before concentration *in vacuo*.

To the solution of the above residue in HOAc (5 ml) was added active zinc powder (500 mg). After purging with N_2 , the mixture was stirred overnight. After filtration, the filtrate was concentrated *in vacuo* to afford a residue, which was diluted with DCM and basified with satd. Na₂CO₃ solution. Extraction with DCM five times to afford the organic layer, which was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography (1: 1, Hexane/ EA) to afford **4j** (96 mg, 74% yield). The ¹H NMR was matched with the literature¹⁶.

To the solution of **4j** (12.0 mg, 0.54 mmol) in MeOH (1 mL) was added HCHO (5.2 uL, 37% aqueous solution, 0.065 mmol), HOAc (3.1 uL, 0.054 mmol) and NaBH(OAc)₃ (17 mg, 0.081) successively. After stirring overnight, satd. Na₂CO₃ solution was added. The mixture was extracted with Et₂O three times and the combined organic layer was then dried over Na₂SO₄, filtered and concentrated *in vacuo* to give a residue, which was purified by flash chromatography (3: 2, Hexane/ethyl acetate) to afford (*S*)-Ketamine (6.6 mg, 52% yield). The ¹H NMR was matched with the literature¹⁷. $[\alpha]_D^{20} = -55.5$ (c 0.47, EtOH), compared to $[\alpha]_D^{20} = -56.3$ (c 1.20, EtOH)¹⁷. HPLC (Chiralpak ASH column, 95:5 hexanes/ isopropanol, 1 ml/min; tr= 6.5 min (major), 7.3 min (minor); >99% ee.

Asymmetric Mannich reaction example:



To a solution of **1n** (60 mg, 0.3 mmol) in xylene (3.0 mL) was added **imine-1** (92 mg, 0.45 mmol), 5 Å MS (50 mg), and (*R*)-C₈-TCYP (36 mg, 0.03 mmol). After stirring for 40h at rt, the mixture was purified by fast column chromatography (10: 1, Hexane/ EA) to afford the desired product **5n** (87 mg, 72% yield, 96% ee). Data for **5n**: ¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.21 (m, 10H), .43 – 6.13 (m, 3H), 4.76 (d, *J* = 10.4 Hz, 1H), 2.60 (ddd, *J* = 15.3, 11.6, 6.2 Hz, 1H), 2.42 (dt, *J* = 15.5, 4.7 Hz, 1H), 2.07 – 1.95 (m, 1H), 1.92 – 1.69 (m, 5H), 1.25 (s, 9H). ¹³C NMR (150 MHz, CDCl₃) δ 213.35, 155.48, 139.71, 136.74, 133.22, 131.75, 129.28, 128.66, 128.19, 127.97, 127.42, 126.56, 79.25, 60.51, 59.85, 40.55, 35.97, 32.00, 28.34, 25.93, 21.33. m/z HRMS (ESI) found [M+H]⁺ 406.2382, C₂₆H₃₂NO₃⁺ requires 406.2377. HPLC (Chiralpak IC column, 97:3 hexanes/ isopropanol, 1 ml/min; tr= 11.4 min (major), 19.0 min (minor); 96% ee.

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X-Ray Crystal Structure Data for 2v



A colorless block 0.13 x 0.06 x 0.03 mm in size was mounted on a Cryoloop with Paratone oil. Data were collected in a nitrogen gas stream at 100(2) K using phi and omega scans. Crystal-to-detector distance was 60 mm and exposure time was 5 seconds per frame using a scan width of 1.0°. Data collection was 99.7% complete to 67.00° in θ . A total of 21859 reflections were collected covering the indices, $-12 \le h \le 10$, $-12 \le k \le 12$, $-13 \le 1 \le 13$. 4110 reflections were

found to be symmetry independent, with an R_{int} of 0.0338. Indexing and unit cell refinement indicated a primitive, orthorhombic lattice. The space group was found to be P2(1) (No. 4). The data were integrated using the Bruker SAINT software program and scaled using the SADABS software program. Solution by direct methods (SHELXL-2013) produced a complete heavyatom phasing model consistent with the proposed structure. All non-hydrogen atoms were refined anisotropically by full-matrix least-squares (SHELXL-2013). All hydrogen atoms were placed using a riding model. Their positions were constrained relative to their parent atom using the appropriate HFIX command in SHELXL-97. Absolute stereochemistry was unambiguously determined to be *S* at C1.

Table 1.	Crystal	data and	l structure	refinement	for xy	001.

Identification code	XY001		
Empirical formula	C1.77 H2.46 F0 N0.15 O0.38		
Formula weight	32.04		
Temperature	100(2) K		
Wavelength	1.54178 Å		
Crystal system	Monoclinic		
Space group	P 2(1)		
Unit cell dimensions	a = 10.2527(6) Å	α= 90°.	
	b = 10.1344(6) Å	β= 95.063(2)°.	
	c = 11.1386(6) Å	$\gamma = 90^{\circ}$.	
Volume	1152.84(11) Å ³		
Z	26		
Density (calculated)	1.200 Mg/m ³		
Absorption coefficient	0.686 mm ⁻¹		
F(000)	448		

Crystal size	0.130 x 0.060 x 0.030 mm ³
Theta range for data collection	3.984 to 68.228°.
Index ranges	-12<=h<=10, -12<=k<=12, -13<=l<=13
Reflections collected	21859
Independent reflections	4110 [R(int) = 0.0338]
Completeness to theta = 67.000∞	99.7 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.7531 and 0.7121
Refinement method	Full-matrix least-squares on F ²
Refinement method Data / restraints / parameters	Full-matrix least-squares on F ² 4110 / 1 / 277
Refinement method Data / restraints / parameters Goodness-of-fit on F ²	Full-matrix least-squares on F ² 4110 / 1 / 277 1.027
Refinement method Data / restraints / parameters Goodness-of-fit on F ² Final R indices [I>2sigma(I)]	Full-matrix least-squares on F ² 4110 / 1 / 277 1.027 R1 = 0.0253, wR2 = 0.0649
Refinement method Data / restraints / parameters Goodness-of-fit on F ² Final R indices [I>2sigma(I)] R indices (all data)	Full-matrix least-squares on F ² 4110 / 1 / 277 1.027 R1 = 0.0253, wR2 = 0.0649 R1 = 0.0255, wR2 = 0.0651
Refinement method Data / restraints / parameters Goodness-of-fit on F ² Final R indices [I>2sigma(I)] R indices (all data) Absolute structure parameter	Full-matrix least-squares on F ² 4110 / 1 / 277 1.027 R1 = 0.0253, wR2 = 0.0649 R1 = 0.0255, wR2 = 0.0651 0.04(3)
Refinement method Data / restraints / parameters Goodness-of-fit on F ² Final R indices [I>2sigma(I)] R indices (all data) Absolute structure parameter Extinction coefficient	Full-matrix least-squares on F ² 4110 / 1 / 277 1.027 R1 = 0.0253, wR2 = 0.0649 R1 = 0.0255, wR2 = 0.0651 0.04(3) n/a

Table 2. Atomic coordinates ($x \ 10^4$) and equivalent isotropic displacement parameters (Å²x 10³)

	х	У	Z	U(eq)	
C(14)	5333(2)	7760(2)	5606(1)	18(1)	
C(15)	7142(2)	7625(2)	7186(2)	21(1)	
C(16)	8144(2)	7042(2)	6415(2)	34(1)	
C(17)	6463(2)	6577(2)	7874(2)	32(1)	
C(18)	7743(2)	8678(2)	8036(2)	25(1)	
C(19)	3752(2)	7923(2)	3126(1)	18(1)	
N(2)	4432(1)	8656(1)	5130(1)	18(1)	
O(3)	5331(1)	6604(1)	5339(1)	22(1)	
O(4)	4936(1)	8358(1)	2923(1)	22(1)	
O(5)	6161(1)	8382(1)	6411(1)	20(1)	
C(1)	2291(2)	7599(2)	4723(1)	18(1)	
C(2)	2293(2)	6084(2)	4724(2)	22(1)	
C(3)	835(2)	5700(2)	4530(2)	26(1)	
C(4)	234(2)	6731(2)	3647(2)	27(1)	
C(5)	1060(2)	7955(2)	3855(1)	21(1)	
C(6)	2039(2)	8219(2)	5914(1)	19(1)	

for xy001. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

C(7)	1827(2)	7548(2)	6908(1)	21(1)
C(8)	1535(2)	8094(2)	8077(1)	21(1)
C(9)	1287(2)	7219(2)	9004(2)	26(1)
C(10)	1027(2)	7685(2)	10127(2)	31(1)
C(11)	990(2)	9023(2)	10352(2)	31(1)
C(12)	1235(2)	9904(2)	9442(2)	27(1)
C(13)	1510(2)	9444(2)	8318(2)	22(1)
C(20)	5512(2)	7966(2)	1802(2)	26(1)
C(21)	4749(2)	8584(2)	713(2)	36(1)
C(22)	5560(3)	6473(2)	1724(2)	45(1)
C(23)	6871(2)	8552(3)	1995(2)	45(1)
N(1)	3456(1)	8230(1)	4280(1)	18(1)
O(1)	813(1)	9040(1)	3466(1)	25(1)
O(2)	2973(1)	7359(1)	2424(1)	22(1)

C(14)-O(3)	1.209(2)	N(2)-H(2)	0.8800
C(14)-O(5)	1.337(2)	O(4)-C(20)	1.482(2)
C(14)-N(2)	1.369(2)	C(1)-N(1)	1.477(2)
C(15)-O(5)	1.4802(19)	C(1)-C(6)	1.510(2)
C(15)-C(17)	1.515(3)	C(1)-C(2)	1.536(2)
C(15)-C(16)	1.515(3)	C(1)-C(5)	1.563(2)
C(15)-C(18)	1.521(2)	C(2)-C(3)	1.541(2)
C(16)-H(16A)	0.9800	C(2)-H(2A)	0.9900
C(16)-H(16B)	0.9800	C(2)-H(2B)	0.9900
С(16)-Н(16С)	0.9800	C(3)-C(4)	1.528(3)
C(17)-H(17A)	0.9800	C(3)-H(3A)	0.9900
C(17)-H(17B)	0.9800	C(3)-H(3B)	0.9900
С(17)-Н(17С)	0.9800	C(4)-C(5)	1.508(3)
C(18)-H(18A)	0.9800	C(4)-H(4A)	0.9900
C(18)-H(18B)	0.9800	C(4)-H(4B)	0.9900
C(18)-H(18C)	0.9800	C(5)-O(1)	1.201(2)
C(19)-O(2)	1.211(2)	C(6)-C(7)	1.333(2)
C(19)-O(4)	1.330(2)	C(6)-H(6)	0.9500
C(19)-N(1)	1.382(2)	C(7)-C(8)	1.470(2)
N(2)-N(1)	1.3839(18)	C(7)-H(7)	0.9500

Table 3.	Bond lengths [Å	and angles	[°] for xy001.

C(8)-C(13)	1.395(3)		
C(8)-C(9)	1.400(2)	O(3)-C(14)-O(5)	127.54(15)
C(9)-C(10)	1.385(3)	O(3)-C(14)-N(2)	124.10(15)
C(9)-H(9)	0.9500	O(5)-C(14)-N(2)	108.34(13)
C(10)-C(11)	1.380(3)	O(5)-C(15)-C(17)	109.82(14)
С(10)-Н(10)	0.9500	O(5)-C(15)-C(16)	109.35(14)
C(11)-C(12)	1.390(3)	C(17)-C(15)-C(16)	112.22(16)
С(11)-Н(11)	0.9500	O(5)-C(15)-C(18)	102.48(13)
C(12)-C(13)	1.388(2)	C(17)-C(15)-C(18)	110.85(15)
С(12)-Н(12)	0.9500	C(16)-C(15)-C(18)	111.66(15)
С(13)-Н(13)	0.9500	С(15)-С(16)-Н(16А)	109.5
C(20)-C(23)	1.512(3)	С(15)-С(16)-Н(16В)	109.5
C(20)-C(22)	1.516(3)	H(16A)-C(16)-H(16B)	109.5
C(20)-C(21)	1.519(3)	С(15)-С(16)-Н(16С)	109.5
C(21)-H(21A)	0.9800	H(16A)-C(16)-H(16C)	109.5
C(21)-H(21B)	0.9800	H(16B)-C(16)-H(16C)	109.5
C(21)-H(21C)	0.9800	С(15)-С(17)-Н(17А)	109.5
C(22)-H(22A)	0.9800	С(15)-С(17)-Н(17В)	109.5
C(22)-H(22B)	0.9800	H(17A)-C(17)-H(17B)	109.5
C(22)-H(22C)	0.9800	С(15)-С(17)-Н(17С)	109.5
C(23)-H(23A)	0.9800	H(17A)-C(17)-H(17C)	109.5
C(23)-H(23B)	0.9800	H(17B)-C(17)-H(17C)	109.5
C(23)-H(23C)	0.9800	C(15)-C(18)-H(18A)	109.5

C(15)-C(18)-H(18B)	109.5	C(3)-C(2)-H(2B)	110.8
H(18A)-C(18)-H(18B)	109.5	H(2A)-C(2)-H(2B)	108.9
С(15)-С(18)-Н(18С)	109.5	C(4)-C(3)-C(2)	104.37(14)
H(18A)-C(18)-H(18C)	109.5	C(4)-C(3)-H(3A)	110.9
H(18B)-C(18)-H(18C)	109.5	C(2)-C(3)-H(3A)	110.9
O(2)-C(19)-O(4)	127.31(14)	C(4)-C(3)-H(3B)	110.9
O(2)-C(19)-N(1)	121.40(15)	C(2)-C(3)-H(3B)	110.9
O(4)-C(19)-N(1)	111.27(13)	H(3A)-C(3)-H(3B)	108.9
C(14)-N(2)-N(1)	118.72(13)	C(5)-C(4)-C(3)	105.94(14)
C(14)-N(2)-H(2)	120.6	C(5)-C(4)-H(4A)	110.5
N(1)-N(2)-H(2)	120.6	C(3)-C(4)-H(4A)	110.5
C(19)-O(4)-C(20)	119.50(13)	C(5)-C(4)-H(4B)	110.5
C(14)-O(5)-C(15)	120.35(12)	C(3)-C(4)-H(4B)	110.5
N(1)-C(1)-C(6)	108.63(13)	H(4A)-C(4)-H(4B)	108.7
N(1)-C(1)-C(2)	115.59(14)	O(1)-C(5)-C(4)	126.85(16)
C(6)-C(1)-C(2)	114.58(14)	O(1)-C(5)-C(1)	124.62(15)
N(1)-C(1)-C(5)	109.02(12)	C(4)-C(5)-C(1)	108.48(14)
C(6)-C(1)-C(5)	104.80(13)	C(7)-C(6)-C(1)	124.78(15)
C(2)-C(1)-C(5)	103.41(13)	C(7)-C(6)-H(6)	117.6
C(1)-C(2)-C(3)	104.56(14)	C(1)-C(6)-H(6)	117.6
C(1)-C(2)-H(2A)	110.8	C(6)-C(7)-C(8)	127.24(16)
C(3)-C(2)-H(2A)	110.8	C(6)-C(7)-H(7)	116.4
C(1)-C(2)-H(2B)	110.8	C(8)-C(7)-H(7)	116.4
C(13)-C(8)-C(9)	118.15(16)	C(22)-C(20)-C(21)	112.56(18)
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C(13)-C(8)-C(7)	123.21(15)	C(20)-C(21)-H(21A)	109.5
C(9)-C(8)-C(7)	118.64(16)	C(20)-C(21)-H(21B)	109.5
C(10)-C(9)-C(8)	120.77(18)	H(21A)-C(21)-H(21B)	109.5
С(10)-С(9)-Н(9)	119.6	C(20)-C(21)-H(21C)	109.5
C(8)-C(9)-H(9)	119.6	H(21A)-C(21)-H(21C)	109.5
C(11)-C(10)-C(9)	120.62(17)	H(21B)-C(21)-H(21C)	109.5
С(11)-С(10)-Н(10)	119.7	C(20)-C(22)-H(22A)	109.5
C(9)-C(10)-H(10)	119.7	C(20)-C(22)-H(22B)	109.5
C(10)-C(11)-C(12)	119.34(17)	H(22A)-C(22)-H(22B)	109.5
С(10)-С(11)-Н(11)	120.3	C(20)-C(22)-H(22C)	109.5
С(12)-С(11)-Н(11)	120.3	H(22A)-C(22)-H(22C)	109.5
C(13)-C(12)-C(11)	120.34(18)	H(22B)-C(22)-H(22C)	109.5
С(13)-С(12)-Н(12)	119.8	C(20)-C(23)-H(23A)	109.5
С(11)-С(12)-Н(12)	119.8	C(20)-C(23)-H(23B)	109.5
C(12)-C(13)-C(8)	120.79(17)	H(23A)-C(23)-H(23B)	109.5
С(12)-С(13)-Н(13)	119.6	C(20)-C(23)-H(23C)	109.5
C(8)-C(13)-H(13)	119.6	H(23A)-C(23)-H(23C)	109.5
O(4)-C(20)-C(23)	101.83(14)	H(23B)-C(23)-H(23C)	109.5
O(4)-C(20)-C(22)	109.42(16)	C(19)-N(1)-N(2)	120.11(13)
C(23)-C(20)-C(22)	111.46(19)	C(19)-N(1)-C(1)	117.61(13)
O(4)-C(20)-C(21)	110.26(14)	N(2)-N(1)-C(1)	117.61(12)
C(23)-C(20)-C(21)	110.78(17)		

Symmetry transformations used to generate equivalent atoms: -x, y+1/2, -z

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²	
C(14)	20(1)	19(1)	16(1)	1(1)	2(1)	-2(1)	
C(15)	20(1)	21(1)	22(1)	2(1)	-4(1)	1(1)	
C(16)	22(1)	31(1)	49(1)	-12(1)	2(1)	3(1)	
C(17)	42(1)	28(1)	26(1)	8(1)	-7(1)	-8(1)	
C(18)	25(1)	26(1)	23(1)	1(1)	-5(1)	-2(1)	
C(19)	19(1)	16(1)	19(1)	2(1)	2(1)	3(1)	
N(2)	20(1)	15(1)	18(1)	-2(1)	-3(1)	-1(1)	
O(3)	25(1)	16(1)	26(1)	-2(1)	-2(1)	0(1)	
O(4)	19(1)	26(1)	20(1)	-1(1)	4(1)	-2(1)	
O(5)	23(1)	16(1)	21(1)	0(1)	-5(1)	1(1)	
C(1)	20(1)	17(1)	18(1)	1(1)	2(1)	0(1)	
C(2)	24(1)	18(1)	23(1)	-1(1)	5(1)	-2(1)	
C(3)	29(1)	23(1)	28(1)	-5(1)	9(1)	-7(1)	
C(4)	22(1)	35(1)	25(1)	-6(1)	3(1)	-5(1)	
C(5)	19(1)	28(1)	16(1)	-3(1)	4(1)	2(1)	
C(6)	18(1)	18(1)	19(1)	-1(1)	0(1)	1(1)	
C(7)	21(1)	21(1)	20(1)	1(1)	0(1)	0(1)	
C(8)	16(1)	28(1)	18(1)	3(1)	-1(1)	0(1)	

Table 4. Anisotropic displacement parameters (Å²x 10³) for xy001. The anisotropic displacement factor exponent takes the form: $-2\pi^2$ [h² a^{*2}U¹¹ + ... + 2 h k a^{*} b^{*} U¹²]

C(9)	26(1)	32(1)	21(1)	4(1)	-1(1)	-5(1)
C(10)	26(1)	49(1)	17(1)	7(1)	-1(1)	-10(1)
C(11)	22(1)	54(1)	16(1)	-5(1)	1(1)	-2(1)
C(12)	21(1)	36(1)	24(1)	-5(1)	-2(1)	3(1)
C(13)	19(1)	30(1)	18(1)	1(1)	0(1)	2(1)
C(20)	22(1)	30(1)	26(1)	-3(1)	10(1)	2(1)
C(21)	36(1)	49(1)	23(1)	2(1)	12(1)	7(1)
C(22)	49(1)	32(1)	58(1)	-4(1)	24(1)	12(1)
C(23)	26(1)	69(2)	42(1)	-8(1)	14(1)	-8(1)
N(1)	18(1)	20(1)	15(1)	-1(1)	0(1)	-2(1)
O(1)	26(1)	28(1)	21(1)	0(1)	0(1)	7(1)
O(2)	21(1)	27(1)	19(1)	-3(1)	1(1)	0(1)

_	Х	у	Z	U(eq)	
H(16A)	8476	7736	5910	52	
H(16B)	7732	6347	5900	52	
H(16C)	8870	6667	6936	52	
H(17A)	7094	6177	8479	49	
H(17B)	6107	5896	7313	49	
H(17C)	5750	6981	8275	49	
H(18A)	7062	9059	8495	37	
H(18B)	8129	9373	7568	37	
H(18C)	8426	8281	8593	37	
H(2)	4471	9487	5360	22	
H(2A)	2695	5737	5501	26	
H(2B)	2783	5737	4064	26	
H(3A)	731	4801	4186	32	
H(3B)	421	5730	5300	32	
H(4A)	252	6419	2806	33	
H(4B)	-685	6912	3802	33	

Table 5. Hydrogen coordinates ($x \ 10^4$) and isotropic displacement parameters (Å²x 10³) for xy001.

H(6)	2030	9155	5958	22	
H(7)	1869	6614	6854	25	
H(9)	1297	6295	8860	32	
H(10)	872	7078	10748	37	
H(11)	800	9338	11120	37	
H(12)	1214	10827	9590	33	
H(13)	1682	10055	7705	27	
H(21A)	4710	9542	823	54	
H(21B)	3859	8224	628	54	
H(21C)	5186	8386	-13	54	
H(22A)	5986	6116	2478	68	
H(22B)	6058	6213	1051	68	
H(22C)	4667	6124	1594	68	
H(23A)	6805	9511	2084	68	
H(23B)	7359	8348	1300	68	
H(23C)	7329	8175	2725	68	



(S)-di-*tert*-butyl 1-(2-oxo-1-phenylcyclohexyl)hydrazine-1,2-dicarboxylate (2a)



(S)-di-*tert*-butyl 1-(2-oxo-1-(p-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2b)



(S)-di-*tert*-butyl 1-(1-(naphthalen-2-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2c)

(S)-di-*tert*-butyl 1-(1-(benzo[d][1,3]dioxol-5-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2d)





(S)-di-*tert*-butyl 1-(1-(3-methoxyphenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2e)



(S)-di-*tert*-butyl 1-(1-(4-bromophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2f)

(S)-di-*tert*-butyl 1-(2-oxo-1-(4-(trifluoromethyl)phenyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2g)



(*S*)-di-*tert*-butyl 1-(1-(4-(tert-butoxycarbonyl)phenyl)-2-oxocyclohexyl)hydrazine-1,2dicarboxylate (**2h**)





(S)-di-*tert*-butyl 1-(2-oxo-1-(o-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2i)



(S)-di-*tert*-butyl 1-(1-(2-chlorophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2j)



(S)-di-*tert*-butyl 1-(2-oxo-1-phenylcyclopentyl)hydrazine-1,2-dicarboxylate (2k)



(*R*)-di-*tert*-butyl 1-(4-oxo-3-phenyltetrahydro-2H-pyran-3-yl)hydrazine-1,2-dicarboxylate (2l)

(*R*)-di-*tert*-butyl 1-(1-(tert-butoxycarbonyl)-4-oxo-3-phenylpiperidin-3-yl)hydrazine-1,2-dicarboxylate (**2m**)





(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2n**)



(S)-di-*tert*-butyl 1-(2-oxo-1-vinylcyclohexyl)hydrazine-1,2-dicarboxylate (20)



(*S*,*Z*)-di-*tert*-butyl 1-(2-oxo-1-(prop-1-en-1-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2p**)



(*S*,*Z*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2q**)





(*S*,*E*)-di-*tert*-butyl 1-(8-oxo-7-styryl-1,4-dioxaspiro[4.5]decan-7-yl)hydrazine-1,2-dicarboxylate (2s)





(*S*,*E*)-di-*tert*-butyl 1-(5,5-dimethyl-2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2t**)



(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcycloheptyl)hydrazine-1,2-dicarboxylate (**2u**)



(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclopentyl)hydrazine-1,2-dicarboxylate (**2v**)



(*S*)-di-tert-butyl 1-(2-oxo-1-(phenylethynyl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2w**)

 \cap ^wNHBz 200 200 nm, 4 nr Retention Time Area Percent 100 100 ШĄШ ШÂ 8 Û ۰Ô g 5 20 10 5 15 25 30 Ó Mhutes 1: 234 nm, 4 nm Results Pk # Retention Time Area Percent Lambda Max 1 12.716 48.251 202 2 51.749 15.780 202 4 nm Retention Time Area Percent 200 200 ШÂ 12.684 7.206 ۳Å 100 100 5.692 92.79 ٥ 0 20 ó 5 10 15 25 30 Mhutes 1: 234 nm, 4 nm Results Pk # Retention Time Area Percent Lambda Max 7.206 1 12.684 203 2 15.692 92.795 202











(*R*)-di-*tert*-butyl 1-(2-oxo-1-(thiophen-2-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2z**)

(*R*)-2-(thiophen-2-yl)cyclohexanone (1z)





(*S*,*E*)-2-styrylcyclohexanone (**1n**)





(*S*,*E*)-4,4-dimethyl-2-styrylcyclohexanone (**1t**)



N-((*1S*,*2R*)-2-hydroxy-1-phenylcyclohexyl)benzamide (**3a**)
(S)-Ketamine:





(*E*)-tert-butyl ((2-oxo-1-styrylcyclohexyl)(phenyl)methyl)carbamate (**5n**)



(*R*)-6,6'-dioctyl-3,3'-bis(2,4,6-tricyclohexylphenyl)-[1,1'-binaphthalene]-2,2'-diol (S2)

(*R*)-C₈_TCYP





(R)-H₈_TCYP







tert-butyl 4-(2-oxocyclohexyl)benzoate (1h)

(*Z*)-2-styrylcyclohexanone (**1q**)





(*E*)-7-styryl-1,4-dioxaspiro[4.5]decan-8-one (1t)

(*E*)-4,4-dimethyl-2-styrylcyclohexanone (1u)





(S)-di-*tert*-butyl 1-(2-oxo-1-phenylcyclohexyl)hydrazine-1,2-dicarboxylate (2a)



(S)-di-*tert*-butyl 1-(2-oxo-1-(p-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2b)



(S)-di-*tert*-butyl 1-(1-(naphthalen-2-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2c)



(S)-di-*tert*-butyl 1-(1-(benzo[d][1,3]dioxol-5-yl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2d)



(S)-di-*tert*-butyl 1-(1-(3-methoxyphenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2e)



(S)-di-*tert*-butyl 1-(1-(4-bromophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2f)



(S)-di-*tert*-butyl 1-(2-oxo-1-(4-(trifluoromethyl)phenyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2g)



(S)-di-*tert*-butyl 1-(1-(4-(tert-butoxycarbonyl)phenyl)-2-oxocyclohexyl)hydrazine-1,2dicarboxylate (**2h**)



(S)-di-tert-butyl 1-(2-oxo-1-(o-tolyl)cyclohexyl)hydrazine-1,2-dicarboxylate (2i)



(S)-di-*tert*-butyl 1-(1-(2-chlorophenyl)-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2j)



(S)-di-*tert*-butyl 1-(2-oxo-1-phenylcyclopentyl)hydrazine-1,2-dicarboxylate (2k)



(*R*)-di-*tert*-butyl 1-(4-oxo-3-phenyltetrahydro-2H-pyran-3-yl)hydrazine-1,2-dicarboxylate (2l)

(*R*)-di-*tert*-butyl 1-(1-(tert-butoxycarbonyl)-4-oxo-3-phenylpiperidin-3-yl)hydrazine-1,2dicarboxylate (**2m**)





(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2n**)



(S)-di-tert-butyl 1-(2-oxo-1-vinylcyclohexyl)hydrazine-1,2-dicarboxylate (20)



(*S*,*Z*)-di-*tert*-butyl 1-(2-oxo-1-(prop-1-en-1-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2**p)



(S,Z)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (2q)



(S)-di-*tert*-butyl 1-(2-oxo-[1,1'-bi(cyclohexan)]-1'-en-1-yl)hydrazine-1,2-dicarboxylate (2r)



(*S*,*E*)-di-*tert*-butyl 1-(8-oxo-7-styryl-1,4-dioxaspiro[4.5]decan-7-yl)hydrazine-1,2-dicarboxylate (2s)



(*S*,*E*)-di-*tert*-butyl 1-(5,5-dimethyl-2-oxo-1-styrylcyclohexyl)hydrazine-1,2-dicarboxylate (**2t**)



(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcycloheptyl)hydrazine-1,2-dicarboxylate (**2u**)



(*S*,*E*)-di-*tert*-butyl 1-(2-oxo-1-styrylcyclopentyl)hydrazine-1,2-dicarboxylate (**2v**)



(S)-di-tert-butyl 1-(2-oxo-1-(phenylethynyl)cyclohexyl)hydrazine-1,2-dicarboxylate (**2w**)







(*R*)-*N*-(1-methyl-2-oxocyclohexyl)benzamide (S4)



(S)-di-tert-butyl 1-(1-benzyl-2-oxocyclohexyl)hydrazine-1,2-dicarboxylate (2y)


(*R*)-di-*tert*-butyl 1-(2-oxo-1-(thiophen-2-yl)cyclohexyl)hydrazine-1,2-dicarboxylate (2z)



N-((*1S*,*2R*)-2-hydroxy-1-phenylcyclohexyl)benzamide (**3a**)



(*E*)-tert-butyl ((2-oxo-1-styrylcyclohexyl)(phenyl)methyl)carbamate (**5n**)