This S1 File contains the following:

Fig A Fig B Table A Table B Table C



Supplemental Fig A. Expression of the small heat shock protein gene *AtHsp15.7* in response to abiotic stresses. FC, fold change.



Supplemental Fig B. Total number of peroxisomal genes with significantly changed expression levels in response to stresses. Left, abiotic stresses. Right, biotic stresses. Genes that have log2 normalized fold change >1 or <-1 are considered as significantly regulated.

Abbreviation in heatmap/figure legend	Dataset series number	Experiment design and description of used dataset	Reference or link
Developmental			
Abbreviation in Heatmap	Stage#	Stage description	All data for Developmental stages were downloaded from AtGenExpress: http://arabidopsis.org/servlets/TairObject?type=expres sion_set&id=1006710873
seedling_cotyledon	1	ATGE_1 development baseline Wt cotyledons 7 days continuous light soil	
seedling_hypocotyl	2	ATGE_2 development baseline Wt hypocotyl 7 days continuous light soil	
seeding_leaves1+2	5	ATGE_5 development baseline Wt leaves 1 + 2 7 days continuous light soil	
adult_leaves	15	ATGE_15 development baseline Wt rosette leaf # 8 17 days continuous light soil	
senescing leaves	25	ATGE_25 development baseline Wt senescing leaves 35 days continuous light soil	
flower	39	ATGE_39 development baseline Wt flowers stage 15 21+ days continuous light soil	
silique_stage3	76	ATGE_76 seed & silique development Wt siliques, w/ seeds stage 3; mid globular to early heart embryos 8 wk long day (16/8) soil	
silique_stage4	77	ATGE_77 seed & silique development Wt siliques, w/ seeds stage 4; early to late heart embryos 8 wk long day (16/8) soil	
silique_stage5	78	ATGE_78 seed & silique development Wt siliques, w/ seeds stage 5; late heart to mid torpedo embryos 8 wk long day (16/8) soil	
seed_stage6	79	ATGE_79 seed & silique development Wt seeds, stage 6, w/o siliques; mid to late torpedo embryos 8 wk long day (16/8) soil	

Table A. Microarray Datasets Used in This Study.

seed_stage7 81		ATGE_81 seed & silique development Wt seeds,	
		stage 7, w/o siliques; late torpedo to early walking-	
		stick embryos 8 wk long day (16/8) soil	
seed_stage8	82	ATGE_82 seed & silique development Wt seeds,	
		stage 8, w/o siliques; walking-stick to early curled	
		cotyledons embryos 8 wk long day (16/8) soil	
seed_stage9	83	ATGE_83 seed & silique development Wt seeds,	
		stage 9, w/o siliques; curled cotyledons to early	
		green cotyledonsembryos 8 wk long day (16/8) soil	
seed_stage10	84	ATGE_84 seed & silique development Wt seeds,	
		stage 10, w/o siliques; green cotyledons embryos 8	
		wk long day (16/8) soil	
abiotic stress			
HL_0.5hr, HL_2hr	E-MTAB-	Col-0 plants were grown in growth chambers for 3.5	Jung, H.S., Crisp, P.A., Estavillo, G.M., Cole, B.,
	403	weeks. Plant leaf tissue was incubated for at least 6	Hong, F., Mockler, T.C., Pogson, B.J., and Chory, J.
		h in a LL growth chamber (light intensity was 65	(2013). Subset of heat-shock transcription factors
		µmol photons /m ² ·s, 22 °C) and then collected as	required for the early response of Arabidopsis to
		control sample. The leaf tissue was exposed for 0.5	excess light. Proc Natl Acad Sci USA 110, 14474-
		hr and 2 hr to light with an intensity of 1,300 µmol	14479.
		photons /m ² ·s, (22 °C).	
cold_3hr, cold_6hr,	GSE3326	The wild-type seeds were plated on MS agar plates	Lee, B.H., Henderson, D.A., and Zhu, J.K. (2005). The
cold_24hr		supplemented with 3%	Arabidopsis cold-responsive transcriptome and its
		sucrose. Seedlings were grown at 22°C with 16-h-	regulation by ICE1. Plant Cell 17, 3155-3175.
		light and 8-h-dark	
		cycles for 2 weeks before being harvested. To	
		avoid variations due to	
		circadian rhythm, all cold treatments were started at	
		12 PM at 0°C under	
		light and continued for 0 (untreated control), 3, 6,	
		and 24 h.	
hypoxia_2hr,	GSE9719	Seedlings grown in vertical orientation for 7–14 d	Sorenson, R., and Bailey-Serres, J. (2014). Selective
hypoxia_9hr		on solid Murashige–Skoog medium containing 1%	mRNA sequestration by OLIGOURIDYLATE-BINDING
		sucrose were treated with mixed gases in	PROTEIN 1 contributes to translational control during
		humidified chambers. Seedlings were deprived of	hypoxia in Arabidopsis. Proc Natl Acad Sci USA 111,
		O_2 as well as CO_2 in chambers which were purged	2373-2378.
		with 99.99% Ar(gas), as control. Alternatively,	
1	1		

		treatment was with 2% O2, 370 ppm CO2, in a balance of N2 for 2hr and 9hr.	
drought	GSE10643	Wild-type plants were grown under normal watering conditions for 24 days and then stressed by completely depriving of irrigation for 10 days.	Zhang, Y., Xu, W., Li, Z., Deng, X.W., Wu, W., and Xue, Y. (2008). F-box protein DOR functions as a novel inhibitory factor for abscisic acid-induced stomatal closure under drought stress in Arabidopsis. Plant Physiol 148, 2121-2133.
salt_4day	GSE53308	Wild type Arabidopsis Col-0 plants were grown hydroponically and treated with or without 150mM NaCl and harvested after 4 days of treatment.	Allu, A.D., Soja, A.M., Wu, A., Szymanski, J., and Balazadeh, S. (2014). Salt stress and senescence: identification of cross-talk regulatory components. J Exp Bot 65, 3993-4008.
salt_6day	GSE16765	Arabidopsis Col-0 were grown in the growth chamber in the absence and presence of salt stress. Plants of 2 weeks were subject to salt treatment for 6 days and were used for RNA extraction.	Chan, Z., Grumet, R., and Loescher, W. (2011). Global gene expression analysis of transgenic, mannitol-producing, and salt-tolerant Arabidopsis thaliana indicates widespread changes in abiotic and biotic stress-related genes. J Exp Bot 62, 4787-4803.
ABA	RIKEN- GODA17& 21	Wild-type seedlings were treated with ABA and mock for 3 hr.	Nemhauser, J.L., Hong, F., and Chory, J. (2006). Different plant hormones regulate similar processes through largely nonoverlapping transcriptional responses. Cell <i>126</i> , 467-475. http://arabidopsis.org/servlets/TairObject?type=hyb_d escr_collection&id=1007964750
biotic stress			
flg22_4hr	GSE11807	Six-week old Col-0 plants were infiltrated with 1 µM flg22 or ddH2O and harvested 4 hours later. Total RNA was extracted, biotinlabeled and hybridized to the Affymetrix ATH1 chip. Gene expression values from treated sample were compared to that of mock-treated sample	Bethke G, Unthan T, Uhrig JF, Poschl Y, Gust AA, et al. (2009) Flg22 regulates the release of an ethylene response factor substrate from MAP kinase 6 in Arabidopsis thaliana via ethylene signaling. Proc Natl Acad Sci USA 106: 8067-8072.

elf18_2hr	GSE34047	3-week old Col-0 plants were infiltrated with 10 µM elf18 or mock and harvested after 2 hrs. Three biological replicates were included	Pajerowska-Mukhtar KM, Wang W, Tada Y, Oka N, Tucker CL, et al. (2012) The HSF-like transcription factor TBF1 is a major molecular switch for plant growth-to-defense transition. Curr Biol 22: 103-112	
chitin_30min	GSE28227	2-week old MS medium-grown seedlings were treated with chitooctaose at a final concentration of 1 μM for 30 minutes. The controls were similarly treated with an equivalent amount of ddH2O. Seedlings were harvested for RNA isolation. Three biological replicates were conducted for the experiment	http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=G SE28227	
DC3K_7hr; DC3K_24hr	GSE5520	4-week old Arabidopsis leaves were treated with bacterial pathogen Pseudomonas syringae pv. Tomato DC3000. The first leaf sample was treated with bacteria at the concentration of 10*8 bacteria/ml and sampled 7 hours later, and the second leaf sample was treated with bacteria at the concentration of 10*6 bacteria/ml and sampled 24 hours later	Thilmony R, Underwood W, He SY (2006) Genome- wide transcriptional analysis of the Arabidopsis thaliana interaction with the plant pathogen Pseudomonas syringae pv. tomato DC3000 and the human pathogen Escherichia coli O157:H7. Plant J 46: 34-53.	
BC_18hr; BC_48hr	GSE5684	Adult Col-0 leaves were inoculated by placing 4 5- µl drops of 5x10*5 Botrytis cinerea spore solution. Control leaves were spotted with droplets of 24g/L potato dextrose broth medium. Samples were collected at 18 hrs and 48 hrs after inoculation.	http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=G SE5684	

Table B. Arabidopsis Peroxisomal Gene List.

160 peroxisomal genes have expression data and were used for co-expression analysis. The last eight genes do not have microarray probes.

Order	Gene name	A.t Locus	Microarray probe ID	Annotation
1	ACH2	AT1G01710	261560_at	acyl-CoA thioesterase family protein
2	PEX11c	AT1G01820	261534_at	PEX11c, peroxisome elongation
3	PEX6	AT1G03000	263170_at	peroxin6
4	st4	AT1G04290	263661_at	thioesterase family protein
5	KAT1	AT1G04710	264608_at	3-keto-acyl-CoA thiolase 1
6	ACX3	AT1G06290	260789_s_at	acyl-CoA oxidase 3
7	ACX6	AT1G06310	259419_at	acyl-CoA oxidase 6
8	ACD31.2	AT1G06460	262629_at	alpha-crystallin domain 31.2
9	NDA1	AT1G07180	256057_at	alternative NAD(P)H dehydrogenase 1
10	GLX1	AT1G11840	264372_at	glyoxalase i homolog 1
11	GAPC2	AT1G13440	259361_at	glyceraldehyde-3-phosphate dehydrogenase
12	UP6	AT1G16730	255763_at	unknown protein 6
13	DHAR	AT1G19570	261149_s_at	dehydroascorbate reductase 1
14	4Cl3	AT1G20480	259569_at	4-coumaroyl-CoA synthase family protein
15	OPCL1	AT1G20510	259518_at	opc-8:0 -CoA ligase1
16	AAE1	AT1G20560	259545_at	acyl-activating enzyme 1
17	CAT3	AT1G20620	259544_at	catalase 3
18	CAT1	AT1G20630	259517_at	catalase 1
19	ATF1	AT1G21770	262499_at	acyl-CoA n-acyltransferases
20	GGT1	AT1G23310	262988_at	alanine-2-oxoglutarate aminotransferase 1
21	DEG15	AT1G28320	245687_at	endopeptidase
22	UP9	AT1G29120	260889_at	unknown protein 9
23	PEX7	AT1G29260	260844_at	peroxin 7

24	AAE14	AT1G30520	261801_at	acyl-activating enzyme 14
25	PEX11a	AT1G47750	261739_at	PEX11a, peroxisome elongation
26	st1	AT1G48320	262237_at	thioesterase family protein
27	pxPfkB	AT1G49350	262398_at	pfkb-type carbohydrate kinase family protein
28	NQR	AT1G49670	261601_at	involved in oxidative stress tolerance.
29	ICDH	AT1G54340	262962_at	isocitrate dehydrogenase
30	NS	AT1G60550	264920_at	naphthoate synthase
31	ECI	AT1G65520	264627_at	enoyl-CoA hydratase/isomerase family protein
32	PAO4	AT1G65840	262933_at	polyamine oxidase 4
33	BZO1	AT1G65880	261915_at	benzoate-CoA ligase
34	AAE12	AT1G65890	261922_at	acyl-activating enzyme 12
35	HPR1	AT1G68010	260014_at	hydroxypyruvate reductase 1
36	GGT2	AT1G70580	260309_at	glutamate:glyoxylate aminotransferase 2
37	ECH2	AT1G76150	261771_at	monofunctional enoyl-CoA hydratase 2
38	ATF2	AT1G77540	259706_at	histone acetyltransferase
39	PEX2	AT1G79810	261348_at	peroxin2
40	SMP2	AT2G02510	267239_at	NADH dehydrogenase
41	OPR3	AT2G06050	265530_at	OPDA-reductase 3
42	AGT1	AT2G13360	263350_at	alanine:glyoxylate aminotransferase
43	DRP3B	AT2G14120	263278_at	dynamin-related protein 3B
44	MDH1	AT2G22780	266457_at	peroxisomal NAD-malate dehydrogenase 1
45	SOX	AT2G24580	263788_at	sarcosine oxidase family protein
46	Uri	AT2G26230	267374_at	uricase/urate oxidase putative
47	PEX10	AT2G26350	267433_at	peroxin 10
48	CoAE	AT2G27490	265637_at	dephospho-CoA kinase
49	st5	AT2G29590	266298_at	thioesterase family protein
50	NDA2	AT2G29990	266835_at	alternative NAD(P)H dehydrogenase 2

51	CHYH1	AT2G30650	267571_at	3-hydroxyisobutyryl-coenzyme a hydrolase
52	CHYH2	AT2G30660	267572_at	3-hydroxyisobutyryl-coenzyme a hydrolase
53	UP3	AT2G31670	263449_at	unknown protein 3
54	KAT2	AT2G33150	245168_at	3-keto-acyl-CoA thiolase 2
55	ACX5	AT2G35690	265843_at	acyl-CoA oxidase 5
56	GLH	AT2G38180	267096_at	gdsl-motif lipase/hydrolase family protein
57	PXN/PMP38/PMP36	AT2G39970	267363_at	peroxisomal membrane protein 36
58	PM16	AT2G41790	260554_at	peptidase m16 family protein
59	CuAO	AT2G42490	265882_at	copper amine oxidase
60	CSY3	AT2G42790	263986_at	citrate synthase 3
61	PAO2	AT2G43020	265244_at	polyamine oxidase 2
62	PEN2	AT2G44490	267392_at	o-glycosyl compounds hydrolase
63	PEX16	AT2G45690	267512_at	peroxin 6
64	PEX11d	AT2G45740	266925_at	PEX11d, peroxisome elongation
65	SO	AT3G01910	258948_at	sulfite oxidase
66	SDRc	AT3G01980	258976_at	short-chain dehydrogenase/reductase (sdr) family protein
67	6PGDH	AT3G02360	256328_at	6-phosphogluconate dehydrogenase family protein
68	PEX19A	AT3G03490	259052_at	peroxin 19-1
69	PEX12	AT3G04460	258627_at	peroxin 22
70	PNC1	AT3G05290	259306_at	peroxisomal adenine nucleotide carrier 1
71	LACS6	AT3G05970	258563_at	long-chain acyl-CoA synthetase 6
72	IBR3	AT3G06810	258525_at	acyl-CoA dehydrogenase
73	MFP2	AT3G06860	258555_at	multifunctional protein 2
74	CML3	AT3G07490	259064_at	calcium ion binding
75	PEX13	AT3G07560	259068_at	peroxin 13
76	SDRb	AT3G12800	257687_at	short-chain dehydrogenase
77	HAOX1/2	AT3G14130, AT3G14150	257004_s_at	putative glycolate oxidase

78	GOX1/2	AT3G14415, AT3G14420	258359_s_at	glycolate oxidase
79	HBCDH	AT3G15290	257052_at	3-hydroxybutyryl-CoA dehydrogenase, putative
80	AAE7	AT3G16910	257880_at	acyl-activating enzyme 7
81	GPK1	AT3G17420	257295_at	glyoxysomal protein kinase 1
82	PEX3A	AT3G18160	258150_at	peroxin 3a
83	SCO3	AT3G19570	257047_at	snowy cotyledon3
84	DRP5B	AT3G19720	257045_at	dynamin-related protein 5B
85	ICL	AT3G21720	257947_at	isocitrate lyase
86	PEX22	AT3G21865	257953_at	peroxin 22
87	GR1	AT3G24170	257252_at	glutathione-disulfide reductase 1
88	MDAR4	AT3G27820	257227_at	monodehydroascorbate reductase 4
89	PEX11b	AT3G47430	252411_at	PEX11b, peroxisome elongation
90	B12D1	AT3G48140	252348_at	senescence-associated protein
91	BADH	AT3G48170	252354_at	betaine aldehyde dehydrogenase
92	MIF	AT3G51660	252076_at	macrophage migration inhibitory factor family protein
93	ACX4	AT3G51840	246304_at	acyl-CoA oxidase 4
94	MDAR1	AT3G52880	252024_at	monodehydroascorbate reductase
95	SDRd	AT3G55290	251780_s_at	short-chain dehydrogenase/reductase (sdr) family protein]
96	CDC	AT3G55640	251757_at	mitochondrial substrate carrier family protein
97	ZnDH	AT3G56460	251687_at	zinc-binding dehydrogenase
98	HIT3	AT3G56490	251707_at	histidine triad family protein
99	FIS1A	AT3G57090	251659_at	fission protein 1A
100	СР	AT3G57810	251558_at	OTU-like cysteine protease
101	CSY1	AT3G58740	251540_at	citrate synthase 1
102	CSY2	AT3G58750	251541_at	citrate synthase 2
103	PMD1	AT3G58840	251556_at	peroxisome and mitochodria division protein
104	PAO3	AT3G59050	251505_at	polyamine oxidase 3

105	PEX11e	AT3G61070	251352_at	PEX11e, peroxisome elongation
106	st3	AT3G61200	251307_at	thioesterase family protein
107	ACH	AT4G00520	255679_at	acyl-CoA thioesterase family protein
108	EH3	AT4G02340	255525_at	putative epoxide hydrolase
109	MCD	AT4G04320	255327_at	malonyl-CoA decarboxylase family protein
110	PMP22	AT4G04470	255338_at	peroxisomal membrane protein 22
111	4CL1	AT4G05160	255263_at	putative 4-coumaroyl-CoA synthase
112	IBR1	AT4G05530	255240_at	short-chain dehydrogenase
113	NDPK1	AT4G09320	255089_at	nucleoside diphosphate kinase 1
114	SCPL20	AT4G12910	254791_at	serine carboxypeptidase-like 20
115	IBR10	AT4G14430	245359_at	enoyl-CoA hydratase/isomerase family protein
116	ECHIA	AT4G16210	245484_at	enoyl-CoA hydratase/isomerase family protein
117	HIT1	AT4G16566	245337_at	histidine triad family protein
118	ACX1	AT4G16760	245249_at	acyl-CoA oxidase 1
119	GOX3	AT4G18360	254630_at	putative glycolate oxidase
120	4Cl5	AT4G19010	254600_at	4-coumaroyl-coa synthase family protein
121	NDB1	AT4G28220	253810_at	NAD(P)H dehydrogenase b
122	AIM1	AT4G29010	253759_at	abnormal inflorescence meristem1, enoyl-CoA hydratase
123	DRP3A	AT4G33650	253306_at	dynamin-related protein 3A
124	APX3	AT4G35000	253223_at	ascorbate peroxidase 3
125	CAT2	AT4G35090	253174_at	catalase 2
126	RDL1	AT4G36880	246250_at	cysteine-type peptidase
127	AGT2	AT4G39660	252855_at	alanine:glyoxylate aminotransferase 2
128	PXA1/CTS	AT4G39850	252830_at	peroxisomal ABC transporter 1
129	MLS	AT5G03860	250868_at	malate synthase
130	BIOTIN_F	AT5G04620	250837_at	7-keto-8-aminopelargonic acid synthase
131	CPK1	AT5G04870	246955_at	calcium-dependent protein kinase isoform

132	PEX1	AT5G08470	250520_at	peroxin 1, ATPase
133	MDH2	AT5G09660	250498_at	peroxisomal NAD-malate dehydrogenase 2
134	ASP3	AT5G11520	250385_at	aspartate aminotransferase 3
135	ELT1	AT5G11910	250299_at	esterase/lipase/thioesterase family protein
136	FIS1B	AT5G12390	245178_at	mitochodria and peroxisome fission protein
137	AAE5	AT5G16370	250114_s_at	acyl-activating enzyme 5
138	ATMS1	AT5G17920	259343_s_at	methionine synthesis 1
139	CSD3	AT5G18100	250016_at	copper superoxide dismutase 3
140	NUDT19	AT5G20070	246126_at	nudix hydrolase homolog 19
141	AAE17	AT5G23050	249869_at	acyl-activating enzyme 17
142	6PGL	AT5G24400	249733_at	6-phosphoglucunolactonase
143	PEX4	AT5G25760	246862_at	peroxin4, ubiquitin-protein ligase
144	PNC2	AT5G27520	246779_at	peroxisomal adenine nucleotide carrier 2
145	LACS7	AT5G27600	246789_at	long-chain acyl-CoA synthetase 7
146	AtHsp15.7	AT5G37670	249575_at	15.7 kda class i-related small heat shock protein-like
147	GSTT1	AT5G41210	249291_at	glutathione s-transferase (class theta) 1
148	SCP2	AT5G42890	249178_at	sterol carrier protein 2
149	AtDCI	AT5G43280	249145_at	delta(3,5),delta(2,4)-dienoyl-CoA isomerase 1
150	UP5	AT5G44250	249064_at	unknown protein 5
151	LON2	AT5G47040	248818_at	lon protease homolog 2
152	ACAT1.3	AT5G47720	248779_at	putative acetyl-CoA c-acyltransferase
153	ACAT2	AT5G48230	248690_at	acetoacetyl-CoA thiolase 2
154	KAT5	AT5G48880	248625_at	3-keto-acyl-CoA thiolase 5
155	PEX5	AT5G56290	248010_at	peroxin 5, peroxisome matrix targeting signal-1 binding
156	TLP	AT5G58220	247858 at	transthyretin-like protein
157	PEX14	AT5G62810	 247422 at	peroxin 14
158	4CL2	AT5G63380	 247380_at	4-coumarate-CoA ligase family protein
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159	ACX2	AT5G65110	247176_at	acyl-CoA oxidase 2
160	CHY1	AT5G65940	247117_at	beta-hydroxyisobutyryl-CoA hydrolase 1
161	AAE18	AT1G48635	no probe	acyl-activating enzyme 18
162	PEX3B	AT1G50510	no probe	peroxin 3B
163	IndA	AT1G55320	no probe	indigoidine synthase a family protein
164	NADK3	AT1G78590	no probe	NADH kinase
165	APEM9	AT3G10572	no probe	required for both pts1- and pts2-dependent protein transport
166	PEX19B	AT5G17550	no probe	peroxin 19B
167	MIA40	AT5G23395	no probe	mitochondrial intermembrane space assembly machinery 40
168	HIT2	AT5G48545	no probe	histidine triad nucleotide-binding 3

Table C. Mutants Used in the Primary Screen for Drought Tolerance.

KO, knock-out; KD, knock-down。

Mutant	SALK_ID	Gene expression	Transcriptional regulation by drought, log ₂ (FC)	Reference
aba1	SALK_059469	КО	Control	[1]
hpr1-1	SALK_067724	КО	limited up regulation 0.44	[0]
hpr1-2	SALK_143584	КО		[2]
cat2	SALK_076998	КО	up-regulated by drought, 1.70	[3]
gox1	SAIL_177_G11	КО	limited up-regulation by drought, 0.42	[4]
gox3	SALK_020909	КО	up-regulated by drought, 1.10	[5]
hsp15.7-1	SALK_038951	KD	up regulated by draught 2.05	[6]
hsp15.7-2	SALK_107711	КО		[0]
pao2-1	SALK_046281	КО	down requilated by drought 4.97	[7]
pao2-2	SALK_062035		down-regulated by drought, -1.37	
pxn-1	SALK_038951	КО	limited up regulation by draught 0.02	[0]
pxn-2	SALK_107711	KD		[o]
csy3	SALK_076319	КО	up-regulated by drought, 2.23	
mif	SALK_037373		up-regulated by drought, 1.79	
lon2-2	SALK_043857	КО	up-regulated by ABA/drought, 1.98/0.51	[9]
kat5-1	SALK_132871		down requilated by drought 1.02	
kat5-2	SALK_144464	KD	down-regulated by drought, - 1.95	[10]
acx3-1	SALK_128947		limited up regulation by draught 0.05	
acx3-2	SALK_044956	КО		[11]
acx4-1	SALK_000879	КО	up regulated by draught 1 21	[44]
acx4-2	SALK_065013	КО		[11]
асх6	SALK_023093	КО	limited up-regulation by drought, 0.01	[11]

acx1/5	SALK_041464/SALK_009998	КО	up-regulated by drought, 1.38/0.51	[12]
aae14	SALK_038308		down-regulated by drought, -2.27	
chy1-1	SALK_025417	КО	limited up-regulation by drought, 0.82	[13]
chy1-2	SALK_102725			

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