# Supplemental Materials Molecular Biology of the Cell

Awata et al.



**FIGURE S1. DRC3 is not expressed in the** *drc3* **mutant.** Western blot of whole-cell lysates probed with anti-DRC3 antibody. The antibody recognized a band of the predicted size for DRC3 (60 kDa) in wild type (WT) and the rescued strain (DRC3-R), but not in *drc3* cells.  $\alpha$ -tubulin served as a loading control.



**FIGURE S2.** Length of flagella in wild-type (WT) and *drc3* mutant cells. Means ± SDs of flagella length were determined for 30 cells of each strain. Statistical significance was determined by Student's t-test.



**FIGURE S3. Identification of DRC9 and DRC10 in** *drc3* **axonemes.** 0.4 M KI extracts of wild-type (WT) and *drc3* axonemes were separated by SDS-PAGE and then stained with Coomassie brilliant blue R-250. Rectangles with dashed lines indicate slices excised for analysis by MS.



**FIGURE S4. Identification of structural defects in** *drc3* **axonemes by comparison of the axonemal repeats from wild-type (WT) and** *drc3* **axonemes.** Longitudinal (A, C) and cross-sectional (B, D) tomographic slices of the averaged 96-nm axonemal repeats from wild type (A, B) and *drc3* (C, D) axonemes reveal the density of the L1 protrusion that directly connects to inner dynein arm g in wild type (red arrowheads), but is missing in *drc3* axonemes (white arrowheads). Note that the densities of other major axonemal structures, such as the outer dynein arms and inner arm dyneins, are not changed in *drc3*. Scale bar: 20 nm.



## FIGURE S5. Anti-DRC3 immunoprecipitates from KI extracts of isolated axonemes of

**wild-type (WT) and** *drc3* **cells.** (A) Silver-stained SDS-polyacrylamide gel of proteins pulled down with anti-DRC3. Rectangles with dashed lines indicate slices excised for analysis by MS; DRC3, 4 and 11 were identified in the gel slices (see Table S1). An arrowhead indicates a band likely to be DRC7 based on size. (B) The same samples as in (A) were analyzed in western blots probed with anti-DRC3, 7, and 11 antibodies. DRC7 and DRC11 were specifically co-precipitated with DRC3. The band present in both lanes at ~90 kDa represents anti-DRC3 antibody that was not completely denatured and was recognized by the secondary antibody.



#### FIGURE S6. DRC3-HA strain used for the anti-HA immunoprecipitation experiment.

Western blot of isolated flagella of wild type (WT) and the DRC3-HA strain, probed with antibodies against DRC3 (upper panel) and the HA peptide (lower panel). DRC3-HA is present at wild-type levels in the DRC3-HA flagella; in that sample, the band recognized by the anti-DRC3 antibody is shifted upward because of the addition of the 3xHA tag (4.4 kDa ).

# Table S1

			Number of	
	ProteinID	Protein description	Wild type	drc3
Slice			Α	В
	524469	DRC11 (FAP82)	9	5
	523721	RNI-like protein	3	5
	513888	β-tubulin	2	2
	515476	FAP99	1	0
	518501	Dihydroorotate dehydrogenase	1	0
	518407	Kinesin heavy chain	1	0
	523861	DNA helicase	1	0
	517045	F1-ATPase	0	2
	515031	Actin	0	1
Slice			С	D
	513508	DRC3 (FAP134)	17	0
	518854	Thioredoxin-like protein	2	0
	513514	FAP7	1	1
	510598	IFT57	1	0
	524750	ABC transporter	1	0
	513771	RSP5	0	1
	516210	TRP10	0	1
Slice			Е	F
	512343	DRC4	19	3
	513888	β-tubulin	10	4
	516143	Tetkin	6	3
	521991	α-tubulin	2	0
	513127	Enolase	1	0
	524918	cAMP kinase	1	1
	519178	Phosphodiestrase	0	1

Table S1. Complete list of proteins identified by MS analysis of selected slices of SDSpolyacrylamide gel of proteins immunoprecipitated from wild-type and *drc3* axonemal extracts by anti-DRC3 antibody. All proteins identified from slices A-F labeled in Figure S5 are listed with total number of peptides found for each. Protein IDs are from the Augustus 5 database. Peptides from DRC3, DRC4, and DRC11 (red) are much more abundant in the wild-type immunoprecipitate. Table S2Primers used in this study

Name	Sequence
Primers used to map the deletion in the <i>drc3</i> mutant	
DRC3Ex1F	5'-GAATGCATCCAGGTGTGTTG-3'
DRC3Ex2R	5'-ACGCCAGGTTTTTGAAGCTA-3'
DRC3Int3F	5'-TAGCATGAGACGTGTGTTGC-3'
DRC3Ex4R	5'-ACGTAGGACCTGTAATCGTG-3'
DRC3Ex6F	5'-TCAACACCTACACAGACGAG-3'
DRC3Ex7R	5'-TGTTGTGCTGCTCCAGAATG-3'
DRC3Ex7F	5'-TTCTGGAGCAGCACAACAAG-3'
DRC3Int7R	5'-CACTACCGTATCACAACTTCC-3'
DRC3Int7F	5'-AGTGACGGTGGCATGATGG-3'
DRC3Ex8R	5'-GTGAAGTACCCGTTGTACTG-3'
DRC3Ex8F	5'-CAGTACAACGGGTACTTCAC-3'
DRC3Ex9R	5'-CTTCTCGAACACGGTCATGG-3'
DRC3Ex10F	5'-ACATGGGCAAGATTGACTCC-3'
DRC3Ex10R	5'-ACTGCTTGAATGTCCGCTTT-3'
Primers used to screen <i>drc3/pf14</i> strains	
531300f	5'-CTGCTATGGCTCGTGTTGTG-3'
521200	

531300r	5'-GCACATACGGTAGCGAACCT-3'
RSP3up	5'-ATGATGAGCGACGTTAACGC-3'
RSP3down	5'-ACAGGAGAACACACCGATCC-3'
DRC3Ex1F	5'-GAATGCATCCAGGTGTGTTG-3'
DRC3Ex2R	5'-ACGCCAGGTTTTTGAAGCTA-3'
NPHP4-F1	5'-GCGCTCCACTCAATAACCAT-3'
NPHP4-R1	5'-AGGTAAGTCCCCTCCTGGAA-3'
NPHP4-F2	5'-AGGTAGATGGGGTGCTGGTG-3'
NPHP4-R2	5'-TAGAATTGGGAGCCTTGACG-3'

### Primers used to generate pDRC3-BgIII

DRC3PFL	5'-GACGGTGGCATGATGGACGGTGC-3'
DRC3stopSap	5'-AAGCTCTTCTATCTCCCATCTCGCCGCCCTCCTCG-3'

DRC3BglIISap DRC3EcoNIDN

## 5'-AAGCTCTTCTGATCTGGGGGGCTGAGCGGGCGTGC-3' 5'-TGTGGCTCTGGGGGCTCGGGC-3'

## Primers used to amplify partial cDNA of DRC3

DRC3cDNA-5'	5'-GGATCCCAGCATCAGGACGAGATGATTG-3'
DRC3cDNA-3'	5'-AAGCTTAGTCAAACTCCTGCACCAGCC-3'