Supplemental Materials Molecular Biology of the Cell

Kubo et al.







Supplemental Figure S1. The *tpg2-3* mutation facilitates the flagellar growth of *pf23*.

pf23 (A(a-d)) and pf23tpg2-3 (B(a-c)) in M-medium, or in M-medium containing autolysin (C) observed by DIC microscopy. Bars, 10 µm. (D) Presence or absence of flagella on pf23 and pf23tpg2-3 cells in M-medium (Normal condition) or M-medium containing autolysin. Biflagellated (cells with two flagella), uniflagellated (one flagella), bald (no flagella), and palmelloid cells were counted.



Supplemental Figure S2. The polyE antibody recognizes only tubulins in the axoneme.

Western blot analyses of axonemes using polyE antibody (A) and B3 antibody (B). Images obtained from the same membranes were used in Figure 1B except that the entire blots are shown here and the membranes were exposed for a longer time to detect any minor signals that might be present. The polyE antibody was produced against Cys-E₉ [1] and should recognize polyglutamate side chains of at least three or more glutamates [2], whereas the B3 antibody recognizes tubulins with two or more glutamate side chains [3]. The result that the polyE antibody recognized only tubulin indicates that there are no other major substrates of polyglutamylation in the axoneme.



Supplemental Figure S3. Flagellar length of mutants lacking various components of the axoneme.

(A) Flagellar length of wild type, *tpg1*, and *tpg2-1* cultured in TAP medium. (B) Flagellar length in wild type (WT), *ida3*, *ida3tpg2-1*, *ida5*, *ida5tpg2-1*, *ida6*, *ida6tpg2-1*, *ida9*, and *ida9tpg1*. (C) Flagellar length in *oda2* and *oda2tpg1*. (D) Flagellar length in *pf18*, *pf18tpg1*, *pf17*, and *pf17tpg1*. Cells in B, C, and D were cultured in M-medium. Data for wild type are from Figure 3. Standard deviations for each measurement are shown as bars. Asterisks indicate statistically significant differences (t-test, *P<0.001 or **P<0.05).



Supplemental Figure S4. Tubulin polyglutamylation deficiency inhibits flagellar shortening after mating.

Histograms of dikaryon flagellar lengths after the mating between CC124 and CC125, *tpg1* and *tpg1*, *tpg2-1* and *tpg2-1*, and *tpg1* and *tpg2-1*. The same data sets were used in Figure 4B. Non-flagellated cells were excluded from the flagellar length measurements. Note that the flagellar-length peaks of wild-type and *tpg1-tpg2-1* dikaryons gradually shift toward shorter lengths during the time course, reflecting the rate of the slow flagellar-shortening phase. Those of the *tpg1* and *tpg2-1* dikaryons did not undergo this shift. The arrows indicate the average lengths of the flagella at each time point.



Supplemental Figure S5. Generation of triple mutants lacking outer arm dynein, inner arm dynein f/I1, and tubulin polyglutamylation.

Western blot analysis of isolated axonemes to confirm the production of triple mutants. Western blots of axonemes of wild type (WT), the double mutants *pf28tpg1*, *pf28tpg2-1*, and *oda3tpg2-1*, and candidate strains for *pf28pf30tpg1*, *pf28pf30tpg2-1*, and *oda3ida1tpg2-1*, probed with anti-TTLL9, anti-polyglutamylated tubulin (polyE), anti-IC140 (dynein f/11 intermediate chain), anti-IC2 (outer arm dynein intermediate chain), and anti-PACRG (for loading control) antibodies. For experiments with *pf28pf30tpg1* and *pf28pf30tpg2-1* (Figure 2), 1G and 5E strains were used respectively.

Table S1

Name	Strain	Missing component/protein	Mutated protein	Mutated gene	Reference
Wild type	CC124, CC125				
pf23		Inner-arm dynein a, c, d, f	DYX1C1	DYXICI	[4]
ssh1	A kind gift from Dr.	FAP267 (TTLL9),	FAP234	TPG2	[5]
(<i>tpg2-3</i>)	Gianni Piperno	FAP234			
tpg1		FAP267 (TTLL9),	FAP267	TPG1	[6]
2 1		FAP234	EL DOO (TD CO	[7]
tpg2-1		FAP26/(TTLL9),	FAP234	TPG2	[7]
nf?&nf30cch1	WA a kind gift from	ГАР234 Inner arm dynein	DHC1	DHCI	[8]
<i>pj20pj50ssn1</i>	Dr. David Mitchell	f/I1 outer arm	vHC	DHC15	[0]
	Di. Duvia Mitchell	dynein, FAP267	FAP234	TPG2	
		(TTLL9), FAP234	1111 23 1		
oda2	CC2230, CC2231	Outer-arm dynein	γHC	DHC15	[9-11]
ida l	CC2664, CC2665	Dynein f	DHC1	DHC1	[12-17]
ida3	CC2668, CC2669	Dynein f			[12]
ida4	CC2670, CC2671	Dynein a, c, d	p28	DII1	[12], [13], [18]
ida5	CC3420, CC3421	Dynein a, c, d, e	Actin	DII4	[19], [20]
ida6	CC3090, CC3091	Dynein e			[20]
ida9	CC4074	Dynein c	DHC9	DHC9	[21]
pf17	CC262	RSP1, 4, 6, 9, 10	RSP9	RSP9	[22]
pf18	CC1036	Central pair			[23], [24]
fla10-1	CC1919	Anterograde IFT	FLA10	FLA10	[25-27]
		lost at 32°C			

Table S2.

Antibody	Dilution for WB	Dilution for IFM	Animal/type	Source
Anti-α-tubulin (B-5-1-2)	1:5,000	N/A	Mouse-monoclonal IgG1	Sigma-Aldrich
Anti-acetylated tubulin (6-11B-1)	N/A	1:500	Mouse-monoclonal IgG2b	Abcam
Anti-polyglutamylated tubulin (B3)	1:2,000	N/A	Mouse-monoclonal IgM	Abcam
Anti-polyglutamylated tubulin (polyE)	1:2,000	N/A	Rabbit-polyclonal	[1]
Anti-FAP234C	1:1,000	N/A	Rabbit-polyclonal	[7]
Anti-TTLL9	1:1,000	N/A	Rabbit-polyclonal	[6]
Anti-IFT172	1:20	N/A	Mouse-monoclonal IgG1	[28]
Anti-IFT139	1:100	N/A	Mouse-monoclonal IgG2a	[28]
Anti-IFT81	1:100	N/A	Mouse-monoclonal IgG1	[28]
Anti-IFT57	1:20	N/A	Mouse-monoclonal IgG2a	[28]
Anti-IFT46	N/A	1:2000	Rabbit-polyclonal	[29]
Anti-DHC1b	1:2,000	N/A	Rabbit-polyclonal	[30]
Anti-D1bLIC	1:2,000	N/A	Rabbit-polyclonal	[31]
Anti-IC2	1:50	N/A	Mouse monoclonal IgG	[32]
Anti-IC140	1:10,000	N/A	Rabbit-polyclonal	[33]
Anti-PACRG	1:5,000	N/A	Rabbit-polyclonal	[34]
Anti-HA tag (3F10)	1:2,000	1:100	Rat-monoclonal IgG1	Roche
Anti-βF1-ATPase	1:80,000	N/A	Rabbit-polyclonal	Agrisera

N/A; not applicable

References for Supplemental data

[1] Shang Y, Li B, Gorovsky MA (2002). *Tetrahymena thermophila* contains a conventional gamma-tubulin that is differentially required for the maintenance of different microtubule-organizing centers. J Cell Biol 158, 1195-1206.

[2] Wloga D, Rogowski K, Sharma N, Van Dijk, Janke C, Eddé B, Bré MH, Levilliers N, Redeker V, Duan J, Gorovsky MA, Jerka-Dziadosz M, Gaertig J (2008). Glutamylation on alpha-tubulin is not essential but affects the assembly and functions of a subset of microtubules in *Tetrahymena thermophile*. Eukaryot Cell 7, 1362-1372.

[3] Gagnon C, White D, Cosson J, Huitorel P, Eddé B, Desbruyéres E, Paturle-Lafanechére L, Multigner L, Job D, Cibert C (1996). The polyglutamylated lateral chain of alpha-tubulin plays a key role in flagellar motility. J Cell Sci 109, 1545-1553.

[4] Huang B, Piperno G, Luck DJ (1979). Paralyzed flagella mutants of *Chlamydomonas reinhardtii* defective for axonemal doublet microtubule arms. J Biol Chem 254, 3091-3099.

[5] LeDizet M, Piperno G (1995). The Light Chain p28 Associates with a Subset of Inner Dynein Arm Heavy Chains in *Chlamydomonas* Axonemes. Mol Biol Cell 6, 697-711.

[6] Kubo T, Yanagisawa HA, Yagi T, Hirono M, Kamiya R (2010). Tubulin polyglutamylation regulates axonemal motility by modulating activities of inner-arm dyneins. Curr Biol 20, 441-445.

[7] Kubo T, Yanagisawa HA, Liu Z, Shibuya R, Hirono M, Kamiya R (2014). A conserved flagella-associated protein in *Chlamydomonas*, FAP234, is essential for axonemal localization of tubulin polyglutamylase TTLL9. Mol Biol Cell 25, 107-117.

[8] Freshour J, Yokoyama R, Mitchell DR (2007). *Chlamydomonas* flagellar outer row dynein assembly protein ODA7 interacts with both outer row and I1 inner row dyneins. J Biol Chem 282, 5404-5412.

[9] Kamiya R (1988). Mutations at twelve independent loci result in absence of outer dynein arms in *Chlamydomonas reinhardtii*. J Cell Biol 107, 2253-2258.

[10] Mitchell DR, Rosenbaum JL (1985). A motile *Chlamydomonas* flagellar mutant that lacks outer dynein arms. J Cell Biol 100, 1228-1234.

[11] Wilkerson CG, King SM, Witman GB (1994). Molecular analysis of the gamma heavy chain of *Chlamydomonas* flagellar outer-arm dynein. J Cell Sci 107, 497-506.

[12] Kamiya R, Kurimoto E, Muto E (1991). Two types of *Chlamydomonas* flagellar mutants missing different components of inner-arm dynein. J Cell Biol 112, 441-447.

[13] Kagami O, Kamiya R (1992). Translocation and rotation of microtubules caused by multiple species of *Chlamydomonas* inner-arm dynein. J Cell Sci 103, 653-664.

[14] Piperno G, Ramanis Z, Smith EF, Sale WS (1990). Three distinct inner dynein arms in *Chlamydomonas* flagella: molecular composition and location in the axoneme. J Cell Biol 110, 379-389.

[15] Myster SH, Knott JA, O'Toole E, Porter ME (1997). The *Chlamydomonas Dhc1* gene encodes a dynein heavy chain subunit required for assembly of the I1 inner arm complex. Mol Biol Cell 8, 607-620.

[16] Myster SH, Knott JA, Wysocki KM, O'Toole E, Porter ME (1999). Domains in the 1α dynein heavy chain required for inner arm assembly and flagellar motility in *Chlamydomonas*. J Cell Biol 146, 801-818.

[17] Luck DJL, Piperno G (1989). Dynein arm mutants of Chlamydomonas. In: Cell

Movement: the Dynein ATPases, vol. 1, eds. Warner FD, Satir P, Gibbons IR, New York Alan R Liss Inc, 49-60.

[18] LeDizet M, Piperno G (1995). *ida4-1*, *ida4-2*, and *ida4-3* are intron splicing mutations affecting the locus encoding p28, a light chain of *Chlamydomonas* axonemal inner dynein arms. Mol Biol Cell 6, 713-723.

[19] Kato T, Kagami O, Yagi T, Kamiya R (1993). Isolation of two species of *Chlamydomonas reinhardtii* flagellar mutants, ida5 and ida6, that lack a newly identified heavy chain of the inner dynein arm. Cell Struct Funct 18, 371-377.

[20] Kato-Minoura T, Hirono M, Kamiya R (1997). *Chlamydomonas* inner-arm dynein mutant, *ida5*, has a mutation in an actin-encoding gene. J Cell Biol 137, 649-656.

[21] Yagi T, Minoura I, Fujiwara A, Saito R, Yasunaga T, Hirono M, Kamiya R (2005). An axonemal dynein particularly important for flagellar movement at high viscosity: Implications from a new *Chlamydomonas* mutant deficient in the dynein heavy chain gene DHC9. J Biol Chem 280, 41412-41420.

[22] Huang B, Piperno G, Ramanis Z, Luck DJ (1981). Radial spokes of *Chlamydomonas* flagella: genetic analysis of assembly and function. J Cell Biol 88, 80-88.

[23] Starling D, Randall J (1971). The flagella of temporary dikaryons of *Chlamydomonas reinhardtii*. Genet Res Camb 18, 107-113.

[24] Adams GMW, Huang B, Piperno G, Luck DJL (1981). Central-pair microtubular complex of *Chlamydomonas* flagella: polypeptide composition as revealed by analysis of mutants. J Cell Biol 91, 69-76.

[25] Huang B, Rifkin MR, Luck DJL (1977). Temperature-sensitive mutations affecting flagellar assembly and function in *Chlamydomonas reinhardtii*. J Cell Biol 72, 67-85.

[26] Adams GMW, Huang B, Luck DJL (1982). Temperature-sensitive, assembly defective flagella mutants of *Chlamydomonas reinhardtii*. Genetics 100, 579-586.

[27] Walther Z, Vashishtha M, Hall JL (1994). The *Chlamydomonas FLA10-1* gene encodes a novel kinesin-homologous protein. J Cell Biol 126, 175-188.

[28] Cole DG, Diener DR, Himelblau AL, Beech PL, Fuster JC, Rosenbaum JL (1998). *Chlamydomonas* kinesin-II dependent intraflagellar transport (IFT): IFT particles contain proteins required for ciliary assembly in *Caenorhabditis elegans* sensory neurons. J Cell Biol 141, 993-1008.

[29] Hou Y, Qin H, Follit JA, Pazour GJ, Rosenbaum JL, Witman GB (2007). Functional analysis of an individual IFT protein: IFT46 is required for transport of outer dynein arms into flagella. J Cell Biol 176, 653-665.

[30] Pazour GJ, Dickert BL, Witman GB (1999). The DHC1b (DHC2) isoform of cytoplasmic dynein is required for flagellar assembly. J Cell Biol 144, 473-481.

[31] Hou Y, Pazour GJ, Witman GB (2004). A dynein light intermediate chain, D1bLIC, is required for retrograde intraflagellar transport. J Cell Biol 15, 4382-4394.

[32] King SM, Witman GB (1990). Localization of an intermediate chain of outer arm dynein by immunoelectron microscopy. J Biol Chem 265, 19807-19811.

[33] Yang P, Sale WS (1998). The Mr 140,000 intermediate chain of *Chlamydomonas* flagellar inner arm dynein is a WD-repeat protein implicated in dynein arm anchoring. Mol Biol Cell 9, 3335-3349.

[34] Ikeda K, Ikeda T, Morisawa K, Kamiya R (2007). Axonemal localization of *Chlamydomonas* PACRG, a homologue of the human Parkin-coregulated gene product. Cell Motil Cytoskeleton 64, 814-821.