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Supplemental Data

Symmetrical Dose-Dependent DNA-Methylation Profiles in Children with Deletion or Duplication of 7q11.23

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Figure S1: Volcano plots pre- and post-adjustment for cell type heterogeneity. (A) Methylation data for the WS cohort pre-adjustment for cell type. Blue data points depict probes that are both significant ($P \le 0.05$) and have a delta beta of $\ge 10\%$ relative to TD controls. Pre-adjustment resulted in 87391 significant probes, of which 7466 had a delta beta of $\ge 10\%$ (6260 hypermethylated and 1206 hypomethylated relative to TD). (B) Adjusting for cell type heterogeneity within the WS cohort resulted in 5154 significant probes, of which 1413 had a delta beta of $\ge 10\%$ relative to TD controls (1032 hypermethylated and 381 hypomethylated). (C) Pre-adjustment within the Dup7 cohort relative to TD) controls resulted in 35789 significant probes, of which 1333 had a delta beta of $\ge 10\%$ (779 hypermethylated and 554 hypomethylated relative to TD). (D) Adjusting for cell type heterogeneity within the Dup7 cohort resulted in 3126 significant probes relative to TD controls, of which 508 had a delta beta $\ge 10\%$ (306 hypermethylated, 202 hypomethylated).



Figure S2: Quantile-quantile (QQ) and Manhattan plots of methylation data. QQ-

plots depict significance values after adjusting for cell type heterogeneity. Red lines show the threshold for genome-wide significance and blue lines the threshold for a suggestive association. QQ-plot and Manhattan plot of (**A**) WS methylation data relative to TD controls, (**B**) Dup7 methylation data relative to TD controls and (**C**) WS methylation data relative to Dup7.





D





Figure S3: Differential methylation alters expression of RGS2 and HDAC4. (A)

Average methylation level at *RGS2* cg05374271 showed a significant effect of genotype (P < 0.001, Kruskal-Wallis) with levels of 39.7% in the WS group, 64.9% in the TD group and 79.9% in the Dup7 group, resulting in a WS DM of -25.2% (adj. P = 9.96E-07) and Dup7 DM of 15% (adj. P = 1.1E-05) (**B**) Expression analysis of *RGS2* across the WS, TD and Dup7 cohorts revealed a significant decrease in expression within the WS group only (P < 0.01, Kruskal-Wallis). (**C**) Average methylation level at *HDAC4* cg08177681 showed a significant effect of genotype (P < 0.0001, Kruskal-Wallis) with levels of 21.9% in the WS group, 11.1% in the TD group and 8.7% in the Dup7 group, resulting in a WS DM of 10.8% and a non-significant change in Dup7 methylation. (**D**) Expression analysis of *HDAC4* across WS, TD controls and Dup7 identified a significant decrease in expression within the WS group (P < 0.01, Kruskal-Wallis). * P < 0.05, *** P < 0.001 using Dunn's post-hoc analysis.





Dupl

60

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0

NS

contri

Μ

20

40

Array methylation level (%)

r²=0.9845

100

80









Array methylation level (%)

Figure S4: Pyrosequencing validation of DM probes in WS, Dup7 and TD controls confirms dose-dependent changes in DNA methylation. Methylation level of 4 WS, 4 TD and 4 Dup7 samples were analyzed using secondary pyrosequencing assays and the methylation level at each CpG is shown. Significance testing between the three groups was performed using the Kruskal-Wallis test. (A) Average methylation levels across ANKRD30B cg21293934 were 70.3% in the WS group, 21.28% in the TD group and 11.65% in the Dup7 group, corresponding to a WS DM of 49.02% and Dup7 DM of -9.63%. (B) Comparison of pyrosequencing and array methylation levels resulted in a significant correlation of R^2 =0.9947 (P < 0.0001) using linear regression. (**C**) Average methylation levels across ANKRD30B cg13266435 were 55% in the WS group, 14.11% in the TD group and 9.26% in the Dup7 group, resulting in a WS DM of 40.9% and Dup7 DM of -4.85%. (D) Comparative analysis of ANKRD30B cg13266435 showed a significant correlation of R^2 =0.9825 (*P* < 0.0001). (**E**) Average methylation levels across ANKRD30B cg23703062 were 48.33% in the WS group,14.17% in the TD group and 8.22% in the Dup7 group, resulting in a WS DM of 34.16% and Dup7 DM of -5.95%. (F) Comparative analysis of ANKRD30B cg23703062 showed a significant correlation of R^2 =0.9881 (P < 0.0001). (G) Average methylation levels across RFPL2 cg12906381 were 80.8% in the WS group, 53.62% in the TD group and 36.1% in the Dup7 group, resulting in a WS DM of 27.18% and Dup7 DM of -17.52%. (H) Comparative analysis of *RFPL2* cg12906381 showed a significant correlation of R^2 =0.9919 (*P* < 0.0001). (I) Average methylation levels across RFPL2 cg01124132 were 65.17% in the WS group, 33.95% in the TD group and 19.87% in the Dup7 group, resulting in WS DM of 31.22% and Dup7 DM of -14.08%. (J) Comparative analysis of RFPL2 cg01124132 revealed a

significant correlation of R²=0.9845 (P < 0.0001). (**K**) Average methylation levels across *RGS2* cg05374271 were 25.63% in the WS group, 48.05% in the TD group and 63.87% in the Dup7 group, resulting in a WS DM of -22.42% and Dup7 DM of 15.82%. (**L**) Comparative analysis of *RGS2* cg05374271 showed a significant correlation of R²=0.9825 (P < 0.0001). (**M**) A CpG upstream of cg05374271 in RGS2 was captured in the same pyrosequencing assay, however it was not captured on the array. Average methylation levels from pyrosequencing across cg05374271 were 21.09% in the WS group, 47.48% in the TD group and 72.65% in the Dup7 group, resulting in a WS DM of -26.39% and Dup7 DM of 25.17%. Significance testing for average methylation changes was performed using the Kruskal-Wallis test.



WS ● TD ◎ Dup7 ●

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Figure S5: Differential methylation identified across *MKRN3* and *NDN*, within the imprinted Prader-Willi and Angelman syndrome region at 15q11-13. (A) Average DNA methylation levels in WS (blue) and Dup7 (green) spanning the imprinted Prader-Willi and Angelman syndrome region. Significantly DM probes with a change in methylation of >10% are plotted from the symmetrically opposite WS v Dup7 dataset (B) A DMR representing significant hypomethylation of 10 probes within *MKRN3* was identified in children with Dup7 (mean DMR *P* = 4.94e-07, chr15:23810163-23812334). Children with WS showed a non-significant gain of methylation. (C) A DMR representing significant hypomethylation. (C) A DMR representing significant hypomethylation methylation. (C) A DMR representing significant hypomethylation of 5 CG probes within *NDN* was found in children with Dup7 (mean DMR *P* = 9.86e-04, chr15:23932370-23933620). A non-significant increase in methylation was detected for children with WS.



Figure S6: Cohort specific DMRs identified across *HOXA4* and *HOXB7*. Scatter plots fitted with Loess curves show DNA methylation changes across children with WS (blue), TD children (grey) and children with Dup7 (green). (**A**) Hypermethylation of *HOXA4* identified in the WS cohort exclusively. Of the 32 probes spanning *HOXA4*, 17 probes showed a gain of methylation of $\geq 10\%$ (mean DMR *P* = 1.15e-03, chr7:27,168,962-27,171,528). DM probes are highlighted in the line plot, showing the selective gain of methylation across the promoter region of *HOXA4* in the WS cohort only. (**B**) Hypomethylation of *HOXB7* was identified specifically within the Dup7 cohort. 3 probes across this region were significantly DM by at least 10% relative to TD controls (mean DMR *P* = 1.13e-07, chr17:46,684,750-46,685,448).



В



Figure S7: Dose-dependent DNA methylation identified across EIF4E and DPP6.

(A) A DMR representing dose-dependent methylation changes identified across the promoter region of *EIF4E*. Of the 24 probes that span *EIF4E*, 15 probes that span the region of differential methylation are shown in the line graph. 8 probes show a DM of \geq 10% (mean DMR *P* = 8.6e-04, chr4:99,849,709-99,851,281). The Dup7 group showed a non-significant trend of hypomethylation relative to TD. (**B**) A DMR representing dose-dependent methylation changes identified in *DPP6*. Of 8 DM probes, 3 probes showed DM of >10% in the WS group relative to the Dup7 group (cg09506081, cg16115627, cg04086391; mean DMR *P* = 4.54e-03, chr7:154,683,249-154,684,963).

B WS hypo DMPs



Figure S8: Relative position of significantly differentially methylated CGs (beta ≥

10%). Position of DM probes using annotations from UCSC RefGene Group of (**A**) WS hypermethylated DM probes, (**B**) WS hypomethylated DM probes, (**C**) background probes on the array calculated as all probes retained after filtering out SNPs and cross-reactive probes, (**D**) Dup7 hypermethylated DM probes and (**E**) Dup7 hypomethylated probes. Positions outside UCSC Refgene Group annotations are listed as 'other' and include distal and intergenic regions.



Figure S9: Enrichment analysis of DM probes using ENCODE data reveals significant enrichment in H3K9me3 and CTCF that spans many different cell types. Top 50 most significant enrichments from (A) WS hypermethylated probes (B) WS hypomethylated probes (C) Dup7 hypermethylated probes and (D) Dup7 hypomethylated probes. Random intersections are scaled by mean and standard deviation. The blue line represents the Z-score associated with a Bonferroni adjusted q = 0.05





Z Score

6

Figure S10: Enrichment analysis using ENCODE data identifies significant and dose-dependent enrichment in histone marks and transcription factor binding sites in H1-hESCs. (A) Significant enrichment in H3K9me3, CTCF and Rad21binding sites were identified in WS hypermethylated (blue) and Dup7 hypomethyated (green) probes, using ENCODE H1-hESCS data. Additional enrichment in binding sites for developmentally important transcription factors ZNF143, RFX5 and TCF12 were identified within the WS hypermethylated group (blue). (B) Significant enrichment for several transcriptional regulator binding sites and histone marks were identified in WS hypomethylated (blue) and Dup7 hypermethylated (green) probes, using ENCODE H1-hESCS data.

Group	Mean age (years) ± SD	Range (years)	Sex	Ethnicity
WS	6.1±1.58	2.8-8.5	14 F 6 M	19 participants were of European ancestry. One participant's mother was of Asian ancestry
Dup7	7.8±2.44	4.4-10.7	5 F 5 M	Eight participants were of European ancestry. Two participants' fathers were of mixed European and Native American ancestry
TD controls	5.5±1.43	2.4-7.4	12 F 3 M	All 15 participants were of European ancestry.

 Table S1. Demographic information for participants at time of blood draw