

Crosstalk Inhibition Nullified by a Receiver Domain Missense Substitution

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RESPONSE REGULATOR SEQUENCE FAMILIES

Receiver domain phylogenetic analysis was conducted as described in Materials and Methods. Sequences included in these analyses are listed in **Table S1**. A representative result is presented in **Fig. S1**; trees generated from different sequence subsets and/or with different parameters differed in detail but shared similar overall topologies with respect to the major groups (clades).

Results extend conclusions drawn from prior studies (1, 7, 8, 96, 97). We sort these response regulator sequences into four groups, based on receiver domain phylogenetic affinity and conserved sequence features as well as on output domains. Note that several receiver domain features differentiate the FixJ-StyR and NarL-VraR families.

OmpR-PhoB sequence family. These response regulators mostly have the winged helix DNA binding domain. Most interact with the closely-related HPK1, HpK2 or HPK3 sequence family transmitter modules (8), excepting a few that interact with Hpt domains from phosphorelays (see below).

NtrC-DctD receiver sequence family. These response regulators have a central ATPase domain plus the Fis DNA-binding domain, and interact with the distinct HPK4 sequence family of transmitter modules (8). Receiver domain sequences are similar to those of the OmpR-PhoB family; the most obvious distinction is at CheY position 104, which is Asp or Asn in the OmpR-PhoB family but Phe or Tyr in the NtrC-DctD family (**Fig. S2**).

FixJ-StyR sequence family. These response regulators have the GerE DNA binding domain. Nevertheless, the receiver domain sequence families clearly are distinct from those of the NarL-VraR family, both in phylogenetic position (**Fig. S1**) and in conserved sequence features (**Fig. S2**). Instead, the receiver domain sequences are very similar to those of the NtrC-DctD family, as noted previously (8, 96).

NarL-VraR sequence family. These response regulators all have the GerE DNA binding domain. Our analysis consciously included more representatives from this group, in part to explore sequence diversity at position T+1 (see below). Two sets of

sequences, represented by FlhR and BvgA, clustered on the edges of the NarL phylogenetic group (**Fig. S1**); however, these did not have obvious differentiating features.

Exceptional sequences. A few proteins included in this analysis have OmpR-PhoB family receiver domain sequences and interact with HPK1-type transmitters, and evidently arose through domain shuffling (8, 96). These are AlgB.Pa, which has a central ATPase domain plus the Fis DNA-binding domain characteristic of NtrC-DctD family members, and PprB.Pa, Pden_3311, and SMb20610, all of which a GerE family DNA binding domain. Interestingly, these latter three proteins have longer interdomain linker sequences (PprB.Pa, 73 residues; Pden_3311, 129; SMb20610, 114) than is typical for NarL-VraR or FixJ-StyR family members (13-36 residues).

RECEIVER DOMAIN SEQUENCE FEATURES

Fig. S2 shows sequence logos compiled from the receiver domain sequences used to construct **Fig. S1**. Note that sequences from the OmpR-PhoB, NtrC-DctD and FixJ-StyR families are quite similar to each other, so distinctions between these three families are based mostly on their different output domains. The NarL-VraR family sequences by contrast are quite distinct. Here we focus on three positions where conserved differences may have functional consequences.

Position T+1 (CheY residue 88). As noted previously by Volz (7), the NarL-VraR family exhibits considerable sequence diversity at this position, whereas most sequences from the other families have either Gly or Ala (**Fig. S2; Table S1**). The outer ring in **Fig. S1** is colored to indicate the identity of the T+1 residue for each sequence. It is immediately apparent that diversity at this position is scattered throughout the NarL-VraR family. Thus, the identity of the T+1 residue has no ostensible value in differentiating subgroups within this family.

Position K+1 (CheY residue 110). Again as noted previously by Volz (7), the NarL-VraR family exhibits considerable sequence diversity at this position, whereas most sequences from the other families have Pro (**Fig. S2; Table S1**). In the HK853-RR468 complex, receiver residue Pro-106 (K+1) contacts DHp residues Leu-266, Thr-267 and Lys-270, just carboxyl-terminal to the active site region spanning residues His-260 (phospho-accepting) through Thr-264 (81). The corresponding region in HisKA_3 sensors displays a different sequence conservation pattern (14). Thus, the absence of a Pro residue at position K+1 may reflect differences in detail for receiver domain interaction between HisKA and HisKA_3 DHp domains. indeed, a Glu-to-Lys missense

substitution at this position in NarL (E110K) apparently causes decreased interaction with the cross-regulating NarQ sensor (16).

Position DD+1 (CheY residue 14). The NarL-VraR family sequences have a polar His or Gln residue at this position, whereas most sequences from the other families have an acidic residue (Asp or Glu) (**Fig. S2; Table S1**). A His-to-Leu missense substitution at this position in DegU (Q12L) apparently causes decreased interaction with the cognate DegS sensor (99, 100), congruent with the recessive null phenotype conferred by the analogous substitution (H15L) in NarL (16).

ASSOCIATIONS WITH HYBRID SENSORS

Phosphorelays involve His-Asp-His-Asp phosphoryl transfer from the DHp domain (H1) through receiver (D1) and Hpt (H2) domains to the response regulator receiver domain (D2) (9). Tripartite hybrid sensors contain the H1, D1 and H2 domains in a single protein, whereas bipartite hybrid sensors contain only H1 and D1 domains, and transfer the phosphoryl group to a separate Hpt (H2) protein. **Fig. S1** indicates response regulators that partner with bipartite (green lines) and tripartite (red lines) sensors. For the subset of response regulators included in this analysis, those partnered with bipartite hybrid sensors cluster at one end of the NarL-VraR family (FlhR group). However, other bipartite hybrid sensors partner with response regulators from other families; one example is the LuxN sensor, which partners through the LuxU Hpt domain protein to the LuxO response regulator, a member of the NtrC-DctD family (Ng and Bassler, 2009, *Annu. Rev. Genet.* **43**:197–222).

Similarly, tripartite hybrid sensors partner with response regulators from different families; examples shown in **Fig. S1** include ArcB (partnered with the OmpR-PhoB family response regulator ArcA), and BarA (partnered with the NarL-VraR family response regulator UvrY). Thus, although there are strong associations between receiver domain and transmitter sequence families for standard two-component pairs, there is no obvious association for phosphorelays. This may reflect distinct specificity determinants for different Hpt domain families, which to our knowledge have not yet been defined in detail.

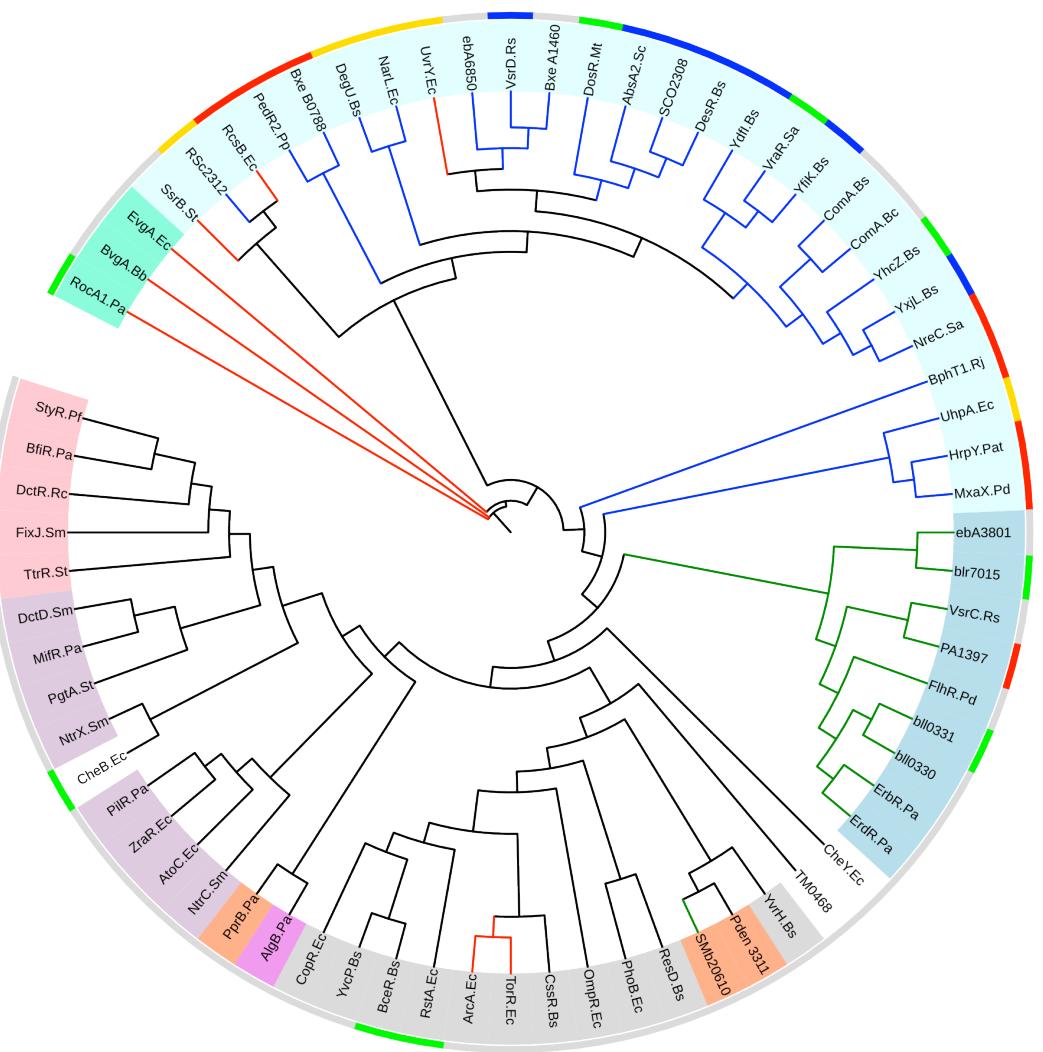
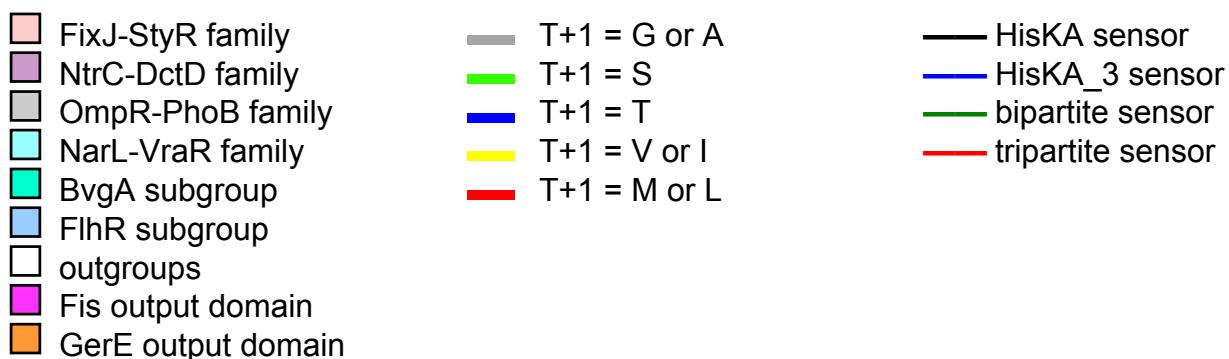
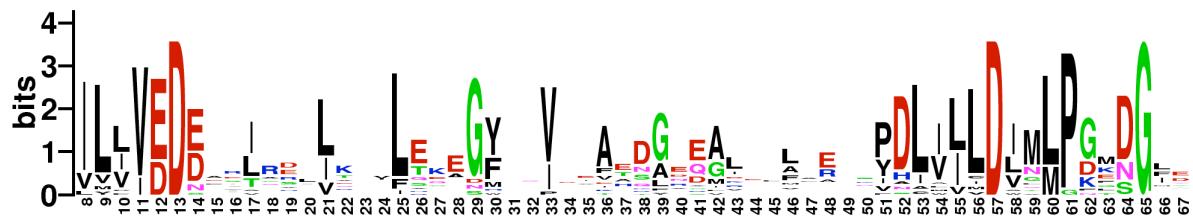


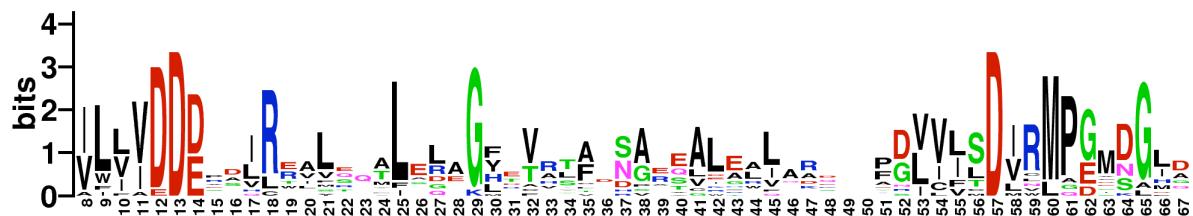
Fig. S1. Representative phylogram for receiver domains. The input file contained 86 sequences, and was constructed through the subtree pruning and regrafting method (see Materials and Methods for details). The resulting log likelihood value is $-19,646.6$. For presentation clarity, the tree was manually "pruned" in iTOL to remove closely-similar sequences, leaving the 70 sequences shown (see **Table S2**).



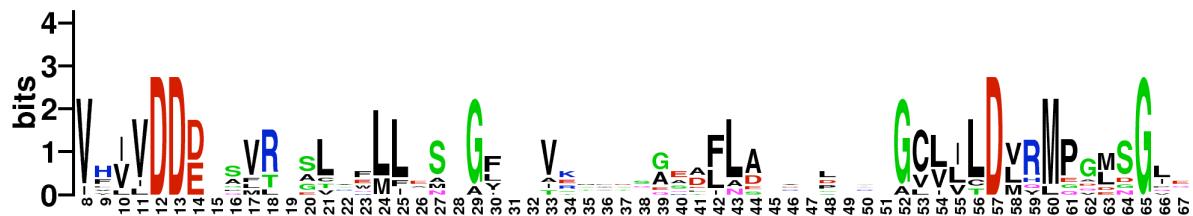
A. OmpR-PhoB Family (N-terminal half)



B. NtrC-DctD Family (N-terminal half)



C. FixJ-StyR Family (N-terminal half)



D. NarL-VraR Family (N-terminal half)

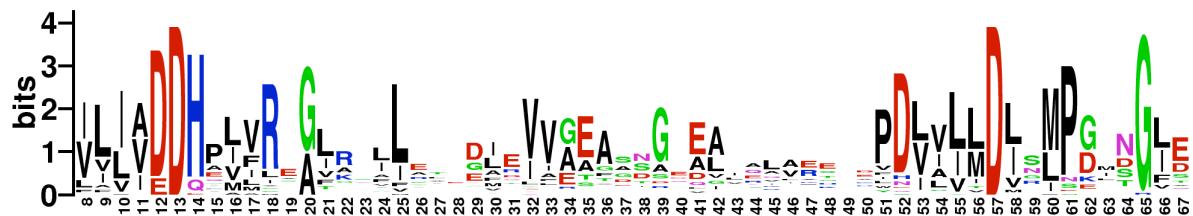
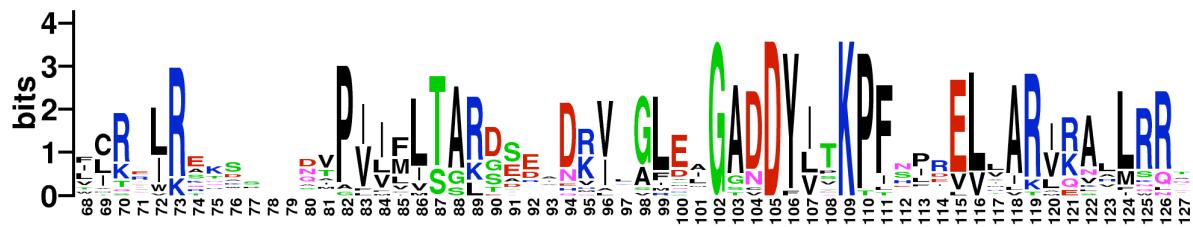
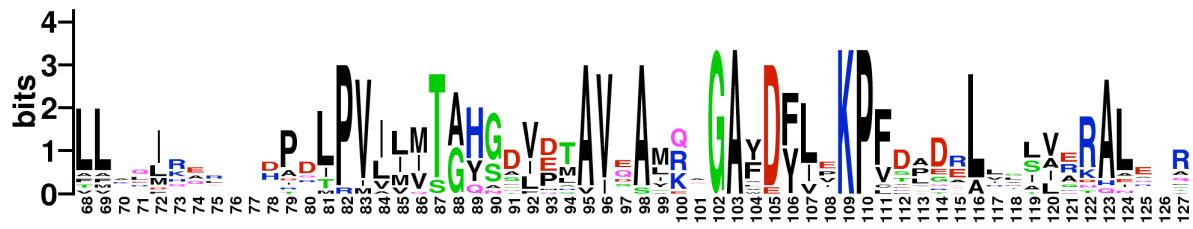


Fig. S2. Receiver domain sequence logos. Sequences are those depicted in **Fig. S1**. Numbering corresponds to that for CheY.Ec. See Materials and Methods for details of logo construction.

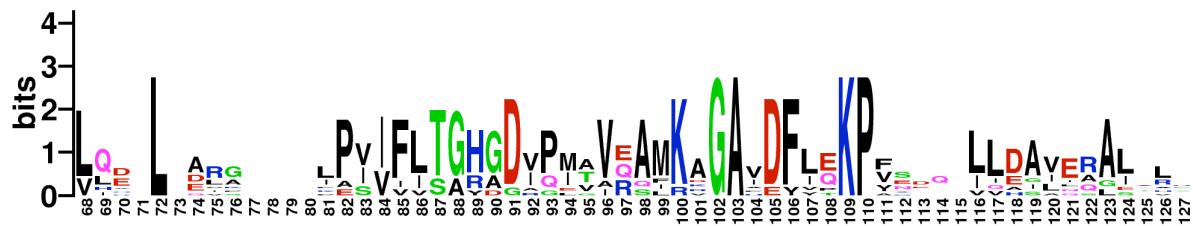
A. OmpR-PhoB Family (C-terminal half)



B. NtrC-DctD Family (C-terminal half)



C. FixJ-StyR Family (C-terminal half)



D. NarL-VraR Family (C-terminal half)

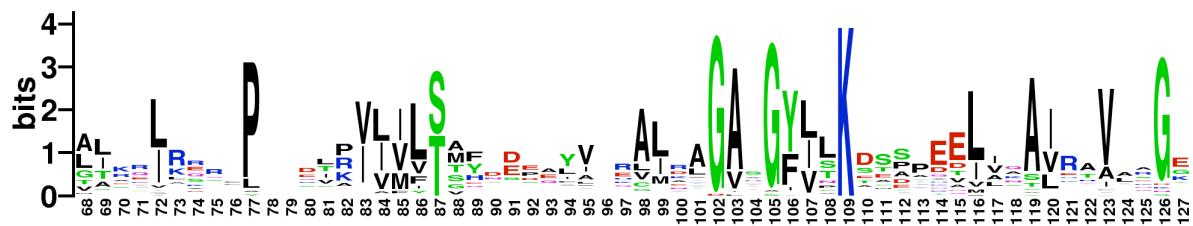


Fig. S2 (continued). Receiver domain sequence logos.

A	Class	Genus	Species	Strain	Sensor	Regulator	Alias Sensor	Alias Regul.	Locus Tag Sensor	Locus Tag Regulator	Accession Regul.	B	C	D	E	F	G	H	I	J	K	L
☒	Gamma proteobact.	<i>Escherichia coli</i>		K-12	CheA	CheY.Ec	—	—	b1888	b1882	NP_416396	D D F	T A	S G Y	K P F							
☒	Gamma proteobact.	<i>Escherichia coli</i>		K-12	CheA	CheB.Ec	—	—	b1888	b1883	NP_416397	D D S	S S	I D F	K P L							
☒	Thermotogales	<i>Thermotoga maritima</i>		MSB8	RR468	—	—	TM0853	TM0468	NP_228278	D D S	T A	R K V	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	BceS.Bs	Bce.Rs	YtsB	YtsA	BSU30390	BSU30400	NP_390918	E D D	S S	D D F	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	CssS.Bs	Css.Rs	YvqB	YvqA	BSU33020	BSU33010	NP_391181	E D E	S A	N D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	Pho.R.Bs	Pho.P.Bs	—	—	BSU29100	BSU29110	NP_390789	D D E	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	ResE.Bs	ResD.Bs	—	—	BSU23110	BSU23120	NP_390193	D D E	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	WalK.Bs	WalR.Bs	YycG	YycF	BSU40400	BSU40410	NP_391921	D D E	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YbdK.Bs	YbdJ.Bs	—	—	BSU02010	BSU02000	NP_388082	E D D	S A	E D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YcbM.Bs	YcbL.Bs	—	—	BSU02560	BSU02550	NP_388137	E D D	S A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YclK.Bs	YclJ.Bs	—	—	BSU03760	BSU03750	NP_388257	E D N	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YkO.Bs	YkG	—	—	BSU13260	BSU13250	NP_389208	E D E	T A	N D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YrkQ.Bs	YrkP.Bs	—	—	BSU26420	BSU26430	NP_390520	E D D	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YvcQ.Bs	YvcP.Bs	—	—	BSU34710	BSU34720	NP_391352	E D D	S A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YvrG.Bs	YvrH.Bs	—	—	BSU33210	BSU33221	NP_391202	D D E	T A	D D Y	K P F								
☒	Firmicutes	<i>Bacillus subtilis</i>	168	YxdK.Bs	YxJ.Ds	—	—	BSU39650	BSU39660	NP_391845	E D S	S A	D D Y	K P F								
☒	Alphaproteobact.	<i>Paracoccus denitrificans</i>	Pd1222	Pden_3312	Pden_3311	—	—	Pden_3312	Pden_3311	ABL71387	E D E	T A	D D Y	K P I								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	SMB20609 [REC]	SMB20610	—	—	SMB20609	SMB20610	NP_438062	D D S	T G	V D Y	K P I								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	ArcB.Ec [Hpt]	ArcA.Ec	—	—	b2310	b2401	NP_418818	E D E	T G	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	BaeS.Ec	BaeR.Ec	—	—	b2078	b2079	NP_416583	E D E	T A	D D Y	K P Y								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	CopS.Ec	CopR.Ec	YedV	YedW	b1968	b1969	NP_416478	E D N	T A	N D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	CpxA.Ec	CpxR.Ec	—	—	b3911	b3912	NP_418348	D D D	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	CreC.Ec	CreB.Ec	—	—	b4399	b4398	NP_418815	E D E	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	CusS.Ec	CusR.Ec	YbcZ	YlcA	b0570	b0571	NP_415103	E D E	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	KdpD.Ec	KdpE.Ec	—	—	b0695	b0694	NP_415222	E D E	S A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	EnvZ.Ec	OmpR.Ec	—	—	b3404	b3405	NP_417664	D D D	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	PhoR.Ec	PhoB.Ec	—	—	b0400	b0399	NP_414933	E D E	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	PhoQ.Ec	PhoP.Ec	—	—	b1129	b1130	NP_415648	E D N	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	PmrB.Ec	PmrA.Ec	BasS	BasR	b4112	b4113	NP_418537	E D D	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	QseC.Ec	QseB.Ec	YgiY	YgiX	b3026	b3025	NP_417497	E D D	T A	D D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	RstB.Ec	RstA.Ec	—	—	b1609	b1608	NP_416125	E D D	T S	C D Y	K T T								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	TorS.Ec [Hpt]	TorR.Ec	—	—	b0993	b0995	NP_415155	E D E	T G	D D Y	K P L								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	KinB.Pa	AlgB.Pa	—	—	PA5484	PA5483	NP_254170	D D E	T A	V D Y	K P C								
☒	Alphaproteobact.	<i>Bradyrhizobium japonicum</i>	USDA 110	FixL.Bj	FixJ.Bj	—	—	b112760	b112759	NP_769399	D D D	T G	V D F	K P F								
☒	Alphaproteobact.	<i>Bradyrhizobium japonicum</i>	USDA 110	NovD.Bj	NovW.Bj	—	—	b111715	b111714	NP_768354	E D D	T G	V D F	K P F								
☒	Alphaproteobact.	<i>Bradyrhizobium japonicum</i>	USDA 110	NwsA.Bj	NwsB.Bj	—	—	blr4773	blr4774	NP_771414	D D D	T G	V D F	K P V								
☒	Alphaproteobact.	<i>Rhodobacter capsulatus</i>	—	DctS.Rc	DctR.Rc	—	—	—	—	PT3740	D D E	T G	F D F	K P F								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	FixL.Sm	FixJ.Sm	—	—	SMa1229	SMa1227	NP_435915	D D E	T G	V D F	K P F								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	SMa0939	SMa0940	—	—	SMa0939	SMa0940	NP_435754	D D D	T G	H D F	K P A								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	SMB20668	SMB20869	—	—	SMB20868	SMB20869	NP_437703	D D D	T G	V D F	K P V								
☒	Betaproteobact.	<i>Aromatoleum aromaticum</i>	EbN1	DctA.Aa	DctR.Aa	—	—	eb4156	eb4154	CAI04876	D D D	T G	F D F	K P F								
☒	Betaproteobact.	<i>Aromatoleum aromaticum</i>	EbN1	eba126	eba125	—	—	eba126	eba125	CAI06190	D D D	T G	V D F	K P F								
☒	Betaproteobact.	<i>Aromatoleum aromaticum</i>	EbN1	Tcs2.Aa	Tcr2.Aa	—	—	c1a81	c1a82	CAI07439	D D E	S A	M D F	K P F								
☒	Betaproteobact.	<i>Aromatoleum aromaticum</i>	EbN1	Tcs3.Aa	Tcr3.Aa	TdiS	TdiR	c2A301	c2A300	CAI07155	D D E	S A	I D F	K P Y								
☒	Betaproteobact.	<i>Aromatoleum aromaticum</i>	EbN1	Ttr.S.Aa	Ttr.R.Aa	—	—	ebA3063	ebA3060	CAI07852	D D D	T G	S D F	K P F								
☒	Betaproteobact.	<i>Burkholderia xenovorans</i>	LB400	Bxe_A1540	Bxe_A1539	—	—	Bxe_A1540	Bxe_A1539	ABE31416	D D D	T G	M D F	K P F								
☒	Betaproteobact.	<i>Ralstonia solanacearum</i>	GMI1000	RSc1598	RSc1597	—	—	RSc1598	RSc1597	CD15299	D D D	T G	V D F	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	Pf1.S.Pa	Bf1.Pa	—	—	PA1497	PA1496	NP_252885	D D D	S G	V D F	K P Y								
☒	Gamma proteobact.	<i>Pseudomonas putida</i>	KT2440	PP_3552	PP_3551	—	—	PP_3552	PP_3551	NP_745688	D D D	S G	C D F	K P Y								
☒	Gamma proteobact.	<i>Pseudomonas putida</i>	KT2440	PP_5242	PP_5241	—	—	PP_5242	PP_5241	NP_747342	D D D	T G	V E F	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	PprA.Pa	PprB.Pa	—	—	PA4293	PA4296	NP_252986	D D E	T G	A D Y	K P V								
☒	Gamma proteobact.	<i>Pseudomonas fluorescens</i>	ST	StyS.Pf	StyR.Pf	—	—	—	—	YI0_A	D D D	T A	I E F	K P F								
☒	Gamma proteobact.	<i>Salmonella typhimurium</i>	LT2	Ttr.S.St	Ttr.R.St	—	—	STM1386	STM1387	NP_460352	D D D	T G	V D F	K P V								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	DctB.Sm	DctD.Sm	—	—	SMB20612	SMB20613	NP_438065	D D D	T G	Y D F	K P F								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	NtrB.Sm	NtrC.Sm	—	—	SMc01042	SMc01043	NP_385565	D D D	S A	Y D Y	K P F								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	NtrY.Sm	NtrZ.Sm	—	—	SMc01044	SMc01045	NP_385567	D D E	S G	Y D F	K P F								
☒	Alphaproteobact.	<i>Sinorhizobium meliloti</i>	1021	SMb21201	SMb21200	—	—	SMb21201	SMb21200	NP_437405	D D E	T G	V D F	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	AtoS.Ec	AtoC.Ec	—	—	b2219	b2220	NP_416724	D D E	T A	F D Y	K P F								
☒	Gamma proteobact.	<i>Escherichia coli</i>	K-12	NtrB.Ec	NtrC.Ec	—	—	b3869	b3868	NP_418304	D D D	T A	F D Y	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	FleS.Pa	ZraE.Rc	—	—	b4003	b4004	NP_418432	D D D	T A	L D Y	K P L								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	MifS.Pa	MifR.Pa	—	—	PA1098	PA1099	NP_249790	E D D	T A	A D Y	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	NtrB.Pa	NtrC.Pa	—	—	PA5112	PA5111	NP_254198	D D E	T G	Y D F	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	PA5165	PA5166	—	—	PA5165	PA5166	NP_253853	D D D	T G	Y D F	K P F								
☒	Gamma proteobact.	<i>Pseudomonas aeruginosa</i>	PAO1	PilS.Pa	PilR.Pa	—	—	PA4546	PA4547	NP_253237	D D E	T A	F D F	K P V								
☒	Gamma proteobact.	<i>Salmonella typhimurium</i>	LT2	PgtB.St	PgtA.St	—	—	STM2397	STM2396	NP_461337	D D D	T G	W D F	K P V								
☒	Alphaproteobact.	<i>Bradyrhizobium japonicum</i>	USDA 110	blr0329 [REC]	blr0330	—	—	blr0329	blr0330	NP_766970	E D H	S G	S G Y	K S S								
☒	Alphaproteobact.	<i>Bradyrhizobium japonicum</i>	USDA 110	blr0329 [REC]	blr0331	—	—	blr0329	blr0331	NP_7												

A	Class	Genus	Species	Strain	Sensor	Regulator	Alias Sensor	Alias Regul.	Locus Tag Sensor	Locus Tag Regulator	Accession Regul.	B	C	D	E	F	G	H	I	J	K	L	
☒	Actinobacteria	<i>Mycobacterium</i>	<i>tuberculosis</i>	H37Rv	DosS.Mt	DosR.Mt	—	—	Rv3132c	Rv3133c	NP_217649	D D H	T S	S G Y	K D I								
☒	Actinobacteria	<i>Mycobacterium</i>	<i>smeqmatis</i>	MC2 155	MSMEG_6238	MSMEG_6236	—	—	MSMEG_6238	MSMEG_6236	YP_890456	D D H	T T	R G Y	K D V								
☒	Actinobacteria	<i>Rhodococcus</i>	<i>jostii</i>	RHA1	BphS1.Rj	BphT1.Rj	—	—	RHA1_r08052	RHA1_r08051	ABC99098	D D H	S M	R G Y	K E C								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	AbsA1.Sc	Abs2.Sc	—	—	SCO3225	SCO3226	NP_627440	D D E	T T	A G F	K A S								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO1369	SCO1370	—	—	SCO1369	SCO1370	NP_625654	D D H	T T	T G Y	K D A								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO2307	SCO2308	—	—	SCO2307	SCO2308	NP_626556	E D Q	T T	A G F	K D G								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO4073	SCO4072	—	—	SCO4073	SCO4072	NP_628253	D D E	T S	R G F	K T V								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO5824	SCO5825	—	—	SCO5824	SCO5825	NP_629948	D D Q	T T	R G Y	K D A								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO6139	SCO6140	—	—	SCO6139	SCO6140	NP_630245	D D Q	T T	S G F	K D A								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO6163	SCO6162	—	—	SCO6163	SCO6162	NP_630267	D D H	T M	C G Y	K S V								
☒	Actinobacteria	<i>Streptomyces</i>	<i>coelicolor</i>	A3(2)	SCO6668	SCO6667	—	—	SCO6668	SCO6667	NP_630742	D D Q	T T	S G F	K D V								
	Firmicutes	<i>Bacillus</i>	<i>anthraeis</i>	Ames Ancest.	GBAA1792	GBAA1791	—	—	GBAA1792	GBAA1791	AT30904	D D N	T T	K G Y	K N N								
☒	Firmicutes	<i>Bacillus</i>	<i>cereus</i>	ATCC 14579	ComP.Bc	ComA.Bc	—	—	BC0881	BC0882	NP_830668	D D H	T G	L G V	K T S								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	ComP.Bs	ComA.Bs	—	—	BSU13690	BSU13680	NP_391046	D D H	T G	H G A	K T E								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	DegS.Bs	DegU.Bs	—	—	BSU35500	BSU35490	NP_391429	D D H	S I	R G Y	K E M								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	DesK.Bs	DesR.Bs	YocF	YocG	BSU19190	BSU19200	NP_389801	E D Q	T T	K G Y	K D S								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	LiaS.Bs	LiaR.Bs	YvqE	YvqC	BSU33080	BSU33080	NP_391188	D D H	T S	L S Y	K T S								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	YdfH.Bs	YdfI.Bs	—	—	BSU05410	BSU05420	NP_388423	D D H	T T	K G Y	K D T								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	YfiJ.Bs	YfiK.Bs	—	—	BSU08290	BSU08300	NP_388711	D D Q	T T	S G Y	K D M								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	YhcY.Bs	YhcZ.Bs	—	—	BSU09320	BSU09330	NP_388814	D D H	T S	K A Y	K D T								
☒	Firmicutes	<i>Bacillus</i>	<i>subtilis</i>	168	YxjM.Bs	YxjL.Bs	—	—	BSU38900	BSU38910	NP_391770	D D Q	T T	V G Y	K D T								
	Firmicutes	<i>Clostridium</i>	<i>acetobutylicum</i>	ATCC 824	CA_C0173	CA_C0174	—	—	CA_C0173	CA_C0174	NP_346816	D D Q	T T	D G Y	K D A								
	Firmicutes	<i>Clostridium</i>	<i>acetobutylicum</i>	ATCC 824	CA_C1454	CA_C1455	—	—	CA_C1454	CA_C1455	NP_348083	D D Q	T T	S G Y	K G V								
	Firmicutes	<i>Clostridium</i>	<i>acetobutylicum</i>	ATCC 824	CA_C2253	CA_C2254	—	—	CA_C2253	CA_C2254	NP_348871	D D Q	T T	S G Y	K D S								
	Firmicutes	<i>Clostridium</i>	<i>acetobutylicum</i>	ATCC 824	CA_C2940	CA_C2939	—	—	CA_C2940	CA_C2939	NP_349541	D D E	T T	N G Y	K D T								
	Firmicutes	<i>Clostridium</i>	<i>acetobutylicum</i>	ATCC 824	CA_C3430	CA_C3429	—	—	CA_C3430	CA_C3429	NP_350019	D D H	T T	K G Y	K D T								
☒	Firmicutes	<i>Staphylococcus</i>	<i>aureus</i>	Mu50	NreB.Sa	NreC.Sa	—	—	SAV2392	SAV2391	BAB58553	D D H	T M	K G Y	K N A								
☒	Firmicutes	<i>Staphylococcus</i>	<i>aureus</i>	Mu50	SAV1321	SAV1322	—	—	SAV1321	SAV1322	BAB57484	E D Q	T T	D A Y	K E R								
☒	Firmicutes	<i>Staphylococcus</i>	<i>aureus</i>	Mu50	SAV1849	SAV1848	—	—	SAV1849	SAV1848	BAB58010	D D H	T S	D G Y	K D V								
☒	Firmicutes	<i>Staphylococcus</i>	<i>aureus</i>	Mu50	VraS.Sa	VraR.Sa	—	—	SAV1885	SAV1884	BAB58046	D D H	T S	D S Y	K T T								
☒	Alphaproteobact.	<i>Paracoccus</i>	<i>denitrificans</i>	Pd1222	MxaY.Pd	MxaX.Pd	—	—	Pden_2991	Pden_2990	ABL71073	D D H	T M	N G I	K D S								
	Alphaproteobact.	<i>Sinorhizobium</i>	<i>meliloti</i>	1021	SMB20077	SMB20078	—	—	SMB20077	SMB20078	NP_436618	D D H	T A	S G L	K E Y								
	Alphaproteobact.	<i>Sinorhizobium</i>	<i>meliloti</i>	1021	SMB20467	SMB20468	—	—	SMB20467	SMB20468	NP_436990	D D H	T A	S G I	K E Q								
	Alphaproteobact.	<i>Sinorhizobium</i>	<i>meliloti</i>	1021	SMB21519	SMB21520	—	—	SMB21519	SMB21520	NP_437890	D D H	T V	K G Y	K G I								
	Betaproteobact.	<i>Acidovorax</i>	<i>ebreus</i>	TPSY	Dtpsy_2833	Dtpsy_2832	—	—	Dtpsy_2833	Dtpsy_2832	ACM34266	D D H	S G	S G Y	K E C								
	Betaproteobact.	<i>Aromatoleum</i>	<i>aromaticum</i>	EbN1	ebaA3729	ebaA3727	—	—	ebaA3729	ebaA3727	CAI08225	D D H	T G	R G V	K E E								
☒	Betaproteobact.	<i>Aromatoleum</i>	<i>aromaticum</i>	EbN1	ebaA5715	ebaA5719	—	—	ebaA5715	ebaA5719	CAI09371	D D H	S M	N A Y	K D V								
☒	Betaproteobact.	<i>Aromatoleum</i>	<i>aromaticum</i>	EbN1	ebaA6844	ebaA6850	—	—	ebaA6844	ebaA6850	CAI10037	D D H	S A	T G Y	K A E								
☒	Betaproteobact.	<i>Aromatoleum</i>	<i>aromaticum</i>	EbN1	Tcs1.Aa	Tcr1.Aa	—	—	c1A75	c1A77	CAI07437	D D H	T M	A G Y	K G K								
☒	Betaproteobact.	<i>Bordetella</i>	<i>bronchiseptica</i>	RB50	BB1122	BB1120	—	—	BB1122	BB1120	CAE31618	D D H	S M	S G Y	K D M								
☒	Betaproteobact.	<i>Burkholderia</i>	<i>xenovorans</i>	LB400	Bxe_A1456	Bxe_A1460	—	—	Bxe_A1456	Bxe_A1460	ABE31494	D D H	S G	N G Y	K D C								
☒	Betaproteobact.	<i>Burkholderia</i>	<i>xenovorans</i>	LB400	Bxe_A1814	Bxe_A1815	—	—	Bxe_A1814	Bxe_A1815	ABE31147	D D H	T S	H S F	K S M								
☒	Betaproteobact.	<i>Burkholderia</i>	<i>xenovorans</i>	LB400	Bxe_A1851	Bxe_A1850	—	—	Bxe_A1851	Bxe_A1850	ABE31115	D D H	T M	S G Y	K E S								
☒	Betaproteobact.	<i>Burkholderia</i>	<i>xenovorans</i>	LB400	Bxe_B0789	Bxe_B0788	—	—	Bxe_B0789	Bxe_B0788	ABE35166	D D H	S M	S G Y	K D S								
☒	Betaproteobact.	<i>Burkholderia</i>	<i>xenovorans</i>	LB400	Bxe_B2472	Bxe_B2471	—	—	Bxe_B2472	Bxe_B2471	ABE33520	D D H	S V	L G Y	K A S								
☒	Betaproteobact.	<i>Ralstonia</i>	<i>solanacearum</i>	GMI1000	RSc0191	RSc0192	—	—	RSc0191	RSc0192	CAD13720	D D H	T M	A A Y	K E S								
☒	Betaproteobact.	<i>Ralstonia</i>	<i>solanacearum</i>	GMI1000	RSc2311	RSc2312	—	—	RSc2311	RSc2312	CAD16019	E D H	T V	D G Y	K D A								
☒	Betaproteobact.	<i>Ralstonia</i>	<i>solanacearum</i>	GMI1000	VsrA.Rs	VsrD.Rs	—	—	VsrA.Rs	VsrD.Rs	RSc0292	CAD13820	D D H	S T	S G Y	K E S							
☒	Betaproteobact.	<i>Rhodoferrax</i>	<i>ferrireducens</i>	T118	Rfer_0781	Rfer_0782	—	—	Rfer_0781	Rfer_0782	ABD68532	D D H	T A	R G V	K S E								
	Deltaproteobact.	<i>Anaeromyxobacter</i>	<i>sp.</i>	Fw109-5	Anae109_3354	Anae109_3355	—	—	Anae109_3354	Anae109_3355	ABS27539	D D H	T A	R G L	K D A								
	Gammaproteobact.	<i>Azotobacter</i>	<i>vinelandii</i>	DJ	Avin_17340	Avin_17330	—	—	Avin_17340	Avin_17330	AC077946	D D H	S M	S G Y	K D V								
	Gammaproteobact.	<i>Azotobacter</i>	<i>vinelandii</i>	DJ	Avin_41030	Avin_41040	—	—	Avin_41030	Avin_41040	AC080238	D D H	S M	R G Y	K D A								
☒	Gammaproteobact.	<i>Escherichia</i>	<i>coli</i>	K-12	NarX.Ec	NarQ.Ec	—	—	NarX.Ec	NarQ.Ec	b1222	b1221	NP_415739	D D H	S V	D G Y	K D M						
☒	Gammaproteobact.	<i>Escherichia</i>	<i>coli</i>	K-12	UhpB.Ec	UhpA.Ec	—	—	UhpB.Ec	UhpA.Ec	b2193	b2193	NP_416697	D D H	T V	D G Y	K D S						
☒	Gammaproteobact.	<i>Escherichia</i>	<i>coli</i>	K-12	BarA [Hpt]	UvrY.Ec	GacS	GacA	BarA [Hpt]	UvrY.Ec	b216	b216	NP_416721	D D H	T M	E G I	K Q G						
☒	Gammaproteobact.	<i>Pectobacterium</i>	<i>atrosepticum</i>	SCRI1043	HrpX.Pat	HrpY.Pat	—	—	HrpX.Pat	HrpY.Pat	ECA2088	ECA2089	CAG74991	D D H	S M	R G Y	K D K						
☒	Gammaproteobact.	<i>Pseudomonas</i>	<i>putida</i>	KT2440	Pp_0420	PP_0410	—	—	Pp_0420	PP_0410	PP_2671	PP_2671	NP_744816	D D H	S M	S G Y	K N S						
	Gammaproteobact.	<i>Pseudomonas</i>	<i>putida</i>	KT2440	PP_2127	PP_2126	—	—	PP_2127	PP_2126	PP_0409	PP_0409	NP_742576	E D H	S M	A G Y	K D S						
☒	Gammaproteobact.	<i>Pseudomonas</i>	<i>putida</i>	KT2440	PP_2127	PP_2126	—	—	PP_2127	PP_2126	PP_2126	PP_2126	NP_744275	D D H	S M	H G Y	K D T						
☒	Gammaproteobact.	<i>Salmonella</i>	<i>typhimurium</i>	LT2	SsrA [Hpt]	SsrB	—	—	STM1392	STM1391	NP_460356	D D H	T A	N G Y	K S S								
	Gammaproteobact.	<i>Xanthomonas</i>	<i>campesidis</i>	ATCC 33913	XCC1127	XCC1128	—	—	XCC1127	XCC1128	NP_636503	D D Q	T T	K G Y	K D V								
	Gammaproteobact.	<i>Xanthomonas</i>	<i>campesidis</i>	ATCC 33913	XCC2179	XCC2180	—	—	XCC2179	XCC2180	NP_637536	D D H	T T	C G Y	K S A								
	Gammaproteobact.	<i>Xanthomonas</i>	<i>campesidis</i>	ATCC 33913	XCC3910	XCC3909	—	—	XCC3910	XCC3909	NP_639249	E D Q	T T	A G Y	K D A	</							