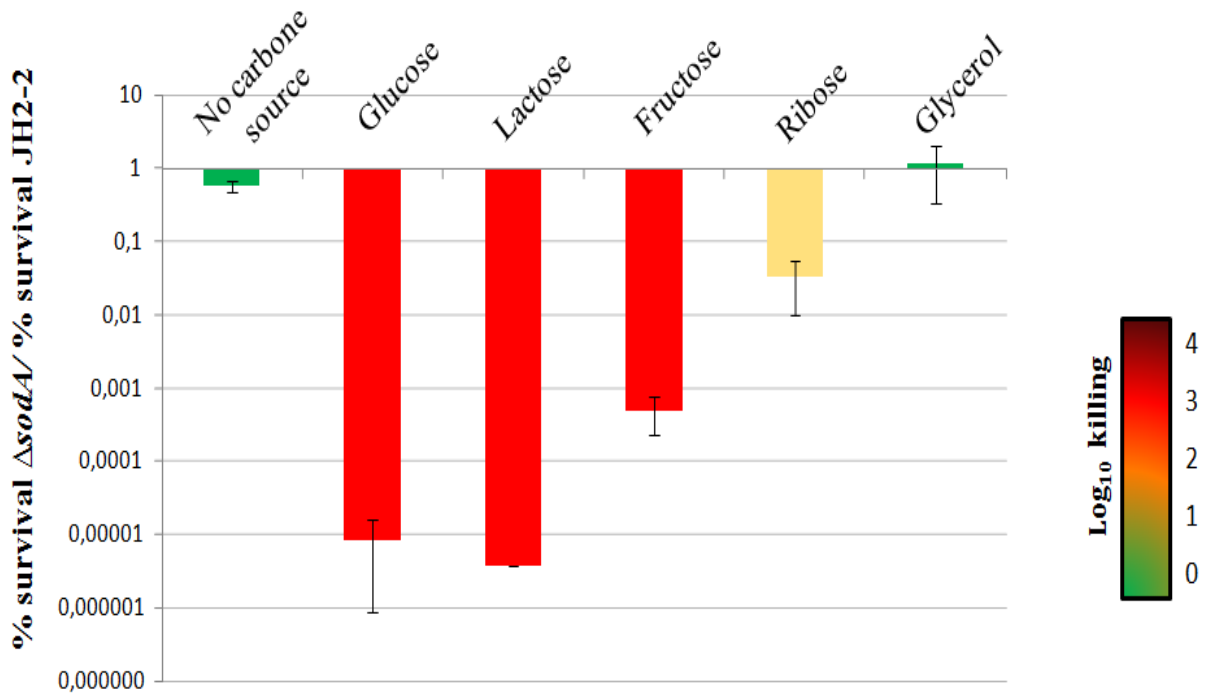
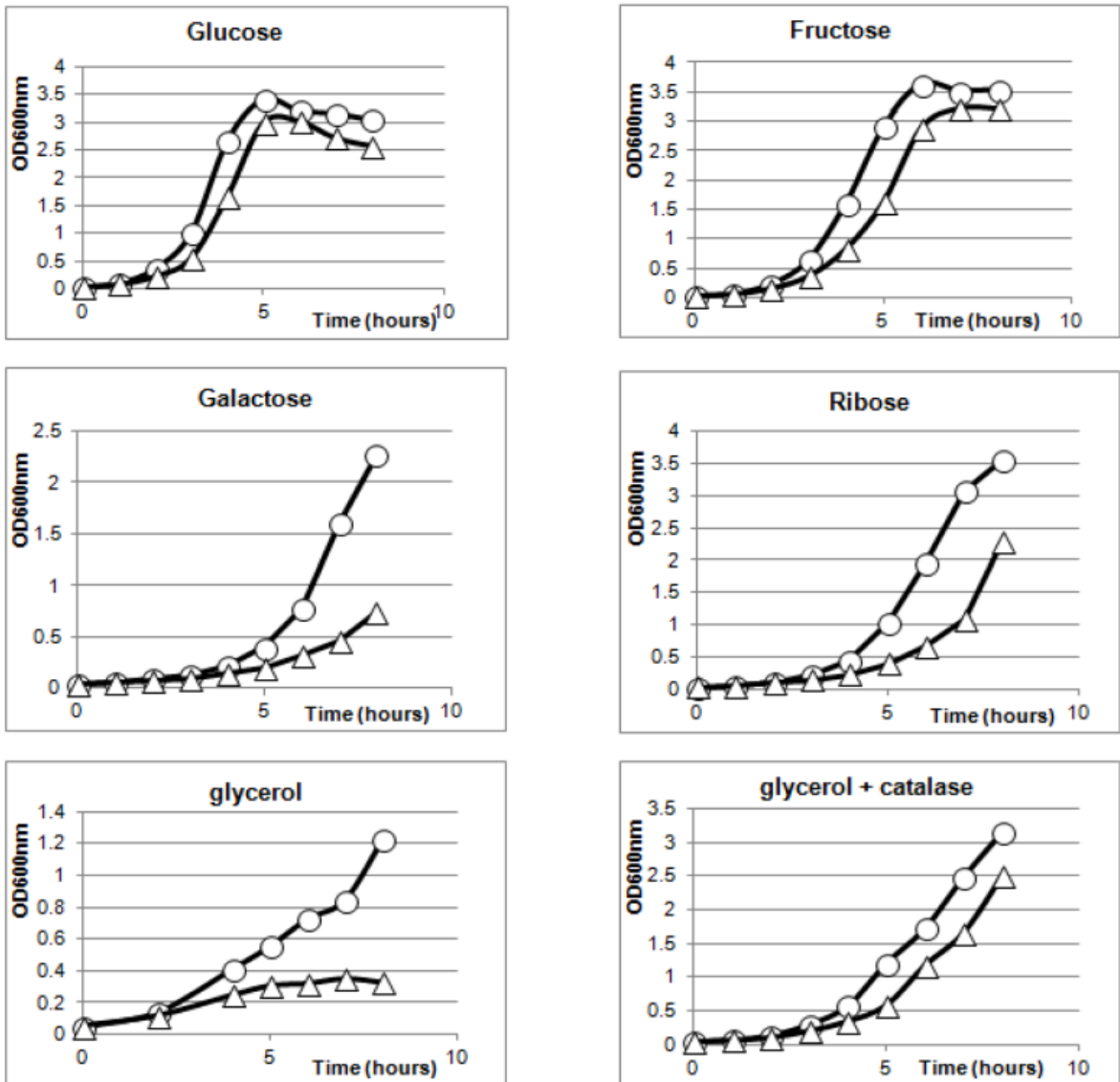


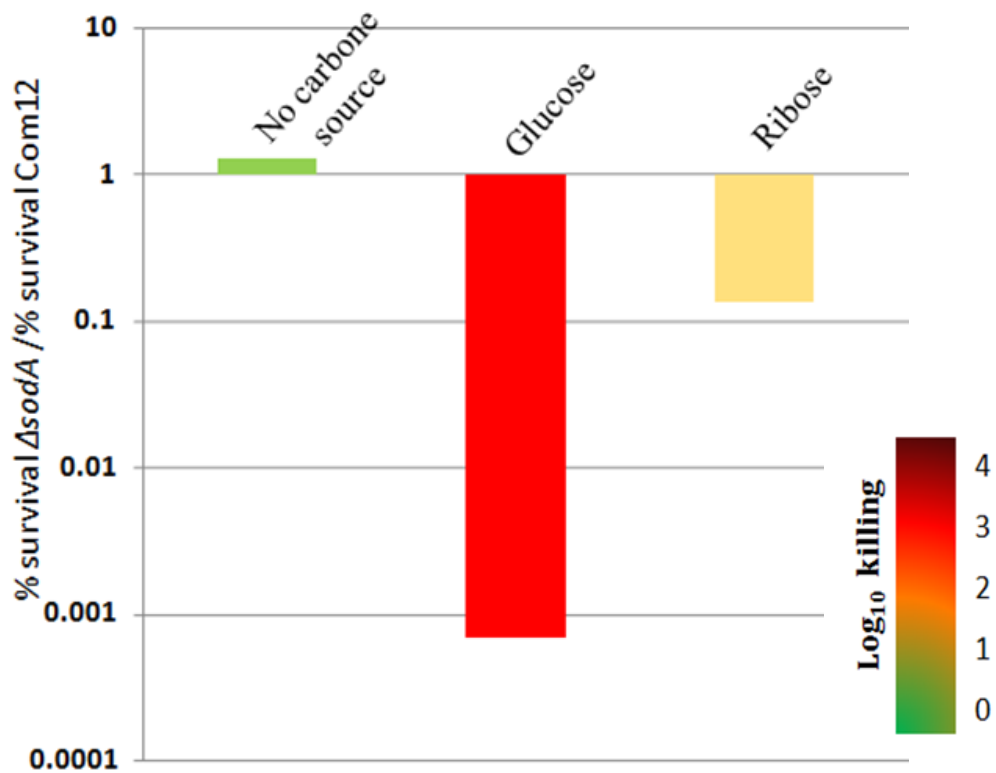
**Fig. S1. Schematic presentation of the mapping method for the transposon insertion sites.** To map the sites of transposon insertion, genomic DNA of mutants with partially restored tolerance screened from the transposon library was extracted and digested with *HaellI* endonuclease. The chromosomal parts attached to the transposon were amplified by PCR using the transposon specific primer pair LTn/RTn after circularization of the restriction fragments by ligation. The PCR products were purified and sequenced and the obtained sequences were blasted against the genome of *E. faecalis* strain OG1RF to locate the transposon sites.



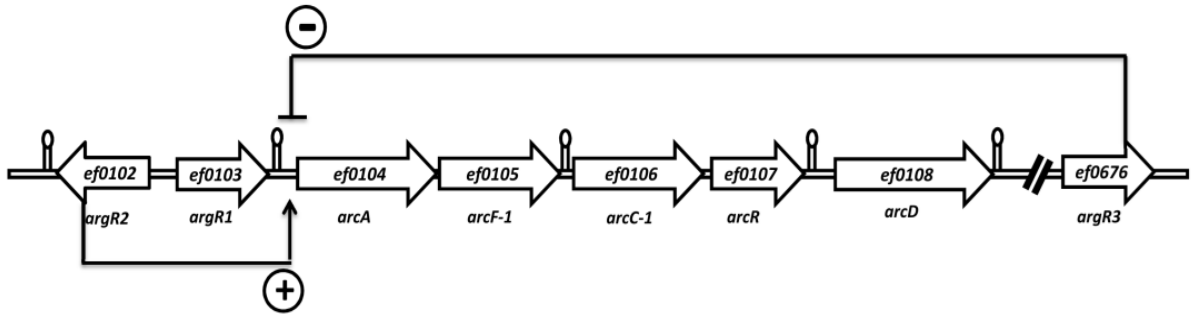
**Fig. S2. Bactericidal activity of penicillin is dependent on the energy source in *E. faecalis* JH2-2  $\Delta$ sodA mutant.** Ratio of relative survival of *E. faecalis* JH2-2 and its derivative JH2-2  $\Delta$ sodA mutant after 24 hours of exposure to 20  $\mu$ g/mL of penicillin in ccM17 MOPS medium supplemented with glucose, lactose, fructose, ribose or glycerol. Mean values of at least three different experiments are represented with error bars indicating standard deviations..



**Fig. S3. Growth of *E. faecalis* strains on different substrates.** Growth curves of *E. faecalis* JH2-2 wild-type (O) and its isogenic  $\Delta sodA$  mutant ( $\Delta$ ) under aerobic conditions (60 rpm) on ccM17MOPS medium supplemented with 0.5% of glucose, fructose, galactose, ribose or glycerol in presence or absence of catalase from bovine liver added to the medium at a final concentration of 500 U/ml are shown.



**Fig. S4. Bactericidal activity of vancomycin is dependent on the energy source in *E. faecium* Com12  $\Delta sodA$  mutant.** Ratio of relative survival of *E. faecium* Com12 and its derivative Com12  $\Delta sodA$  mutant after 24 hours of exposure to 20  $\mu\text{g/mL}$  of vancomycin in ccM17 MOPS medium supplemented with glucose or ribose. Only one experiment has been carried out for these results.



**Fig S5. Organization of ADI operon and its regulation in *E. faecalis*.** Structure and proposed regulation of expression by ArgR2 and ArgR3 of the arginine deiminase operon in *E. faecalis*.

**Table S1. Primers used in this study**

Primer pair	Sequence (5' – 3')*		Use
	Forward	Reverse	
PU/ PR	GTAAACGACGGCCAGT	CAGGAAACAGCTATGAC	Cloning verification
pZXL5 for/pZXL5 Rev	TTCAAGCGTGGTGAAATGAG	TACTTTGCCAGCGGACTTTT	Screen the presence of pZXL5
LTn/RTn	GCCCCCTGAAATCCTTACAT	AAACAGGAATTTATCGAAAATGGT	Reverse PCR for transposon mapping
104ForPst/104RevEco	<u>AAAAAACTGCAGCACAAGCAGAA</u> CATGATGCA ( <i>Pst</i> I)	<u>AAAAAAGAATTC</u> AATTGACGCAGC ATCCGTTC ( <i>Eco</i> R1)	Cloning in pUCB30
104verifF/104verifR	GCCAGATTATCTGGAGAGAC	GAATTTACGATGTTCCCGA	Cloning verification
ef0102L/ef0102R	TTCTCGTGACATTCGTGAGC	TATTTTGGACGAAGGCAGA	RT-qPCR
ef0103L/ef0103R	GCGAGATAAATGTCGCACAA	AAATCGTTAACCGCATCTCG	RT-qPCR
ef0104L/ef0104R	CGGTGAACACCGTAAATTCAT	AAACAACCAACCACCTTCG	RT-qPCR
ef0105L/ef0105R	ACTGTTTGCCAGCCTTTCAT	GCGGAAGACTTCATCCGTAA	RT-qPCR
ef0106L/ef0106R	TTGTCCAATGCGCTTAATCA	ATGCCTCATCTGCTGGATCT	RT-qPCR
ef0107L/ef0107R	CATTACGACTCCGCGATTG	TGAATCAATTGCTGGGGATT	RT-qPCR
ef0676L/ef0676R	CAGAAGTGGCTGGTACAGTGG	TCATATTTTCGATGCGTTCG	RT-qPCR

\*Underlined sequences correspond to the recognition sites of the restriction endonucleases reported in parentheses