

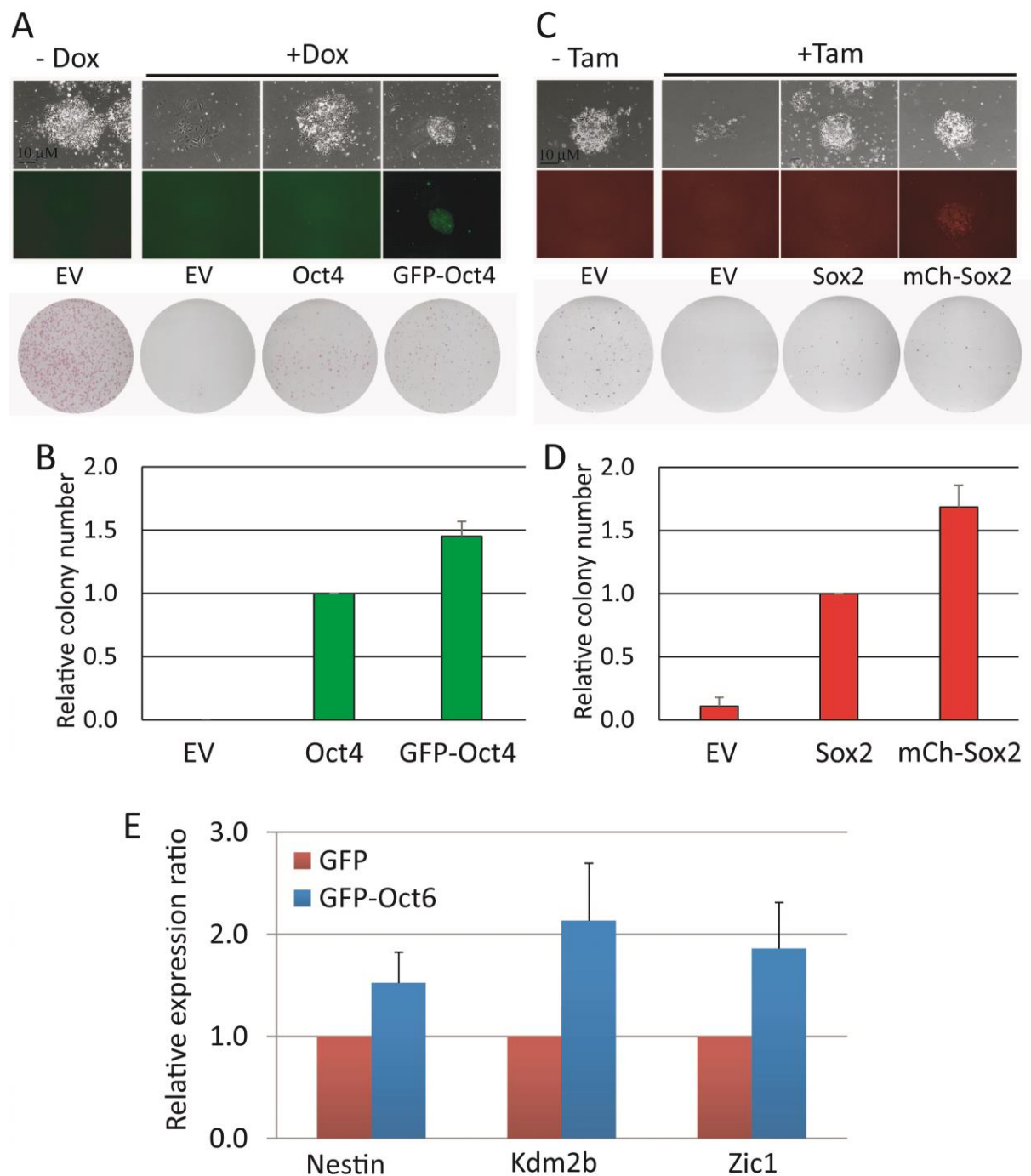
## **Selective influence of Sox2 on POU transcription factor binding in embryonic and neural stem cells**

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### **Table of contents:**

<b>Content</b>	<b>Page(s)</b>
Appendix Figures S1-S3 and legends	2-5
1. Functional characterization of fluorescent protein fusions of Oct4, Sox2 and Oct6	2-3
2. Comparison of theoretical models for best fit to FCS data	4
3. Enriched Sox2 and POU TF motifs in ChIP-Seq data	5
Appendix Table S1: Oligonucleotides for analysis of Sox/Oct, PORE, and MORE motifs	6
Appendix Table S2: Oligonucleotides used for qPCR	7
References	8

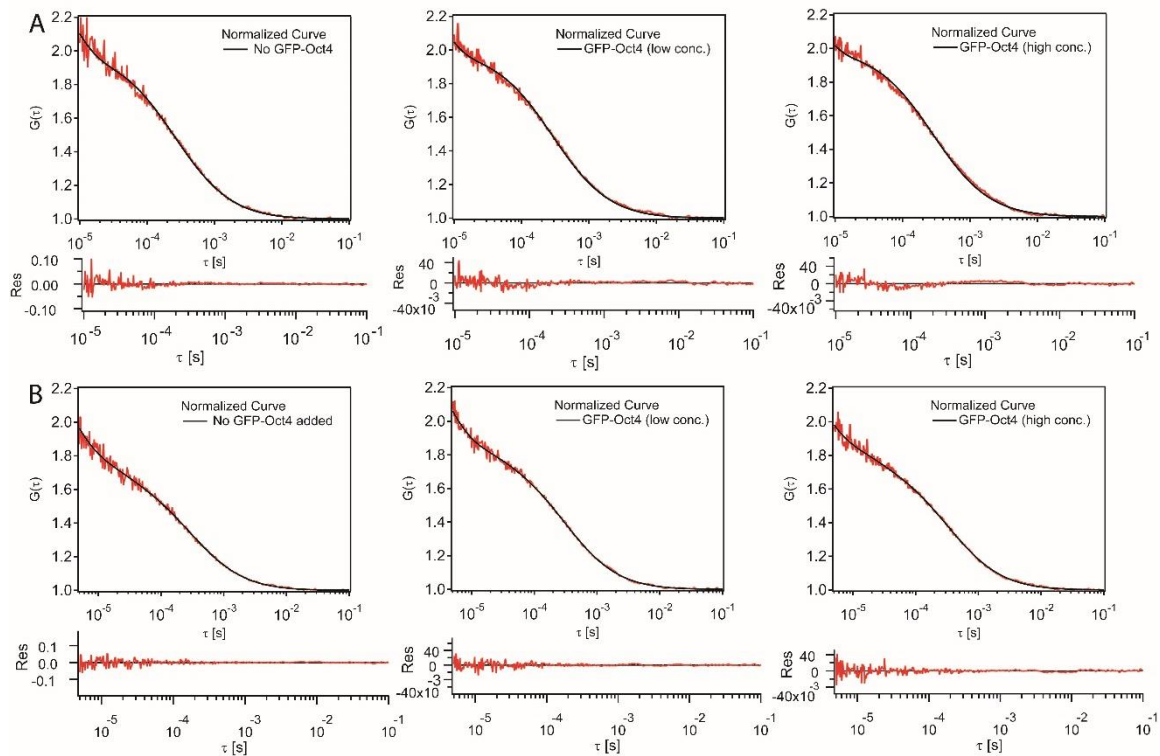
## Appendix Figure S1



**Appendix Figure S1: Functional characterization of fluorescent protein fusions of Oct4, Sox2 and Oct6.** (A) ZHBTc4 ES cells, which lack endogenous Oct4 but contain a doxycycline (Dox) suppressible Oct4 transgene, were used to functionally test the GFP-Oct4 fusion protein. In the presents of Dox ES cells differentiate would normally differentiate. Representative morphologies of colonies

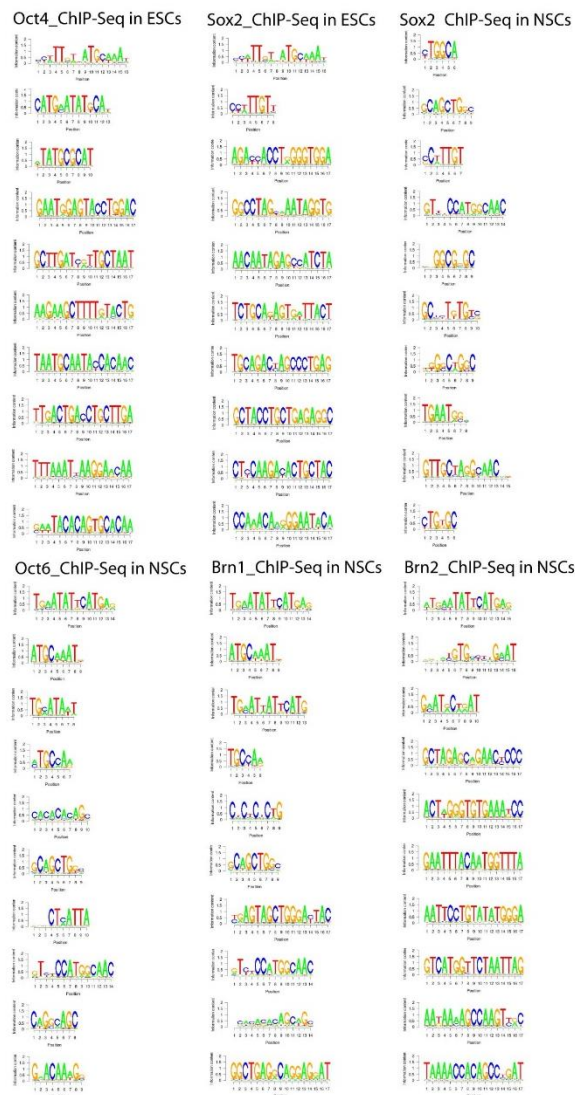
formed in the presence of Dox (1 $\mu$ g/ml) and after transfecting with Oct4 (pPyCAG-Oct4-IP), GFP-Oct4 (pPyCAG-GFP-Oct4-IP), or empty vector (pPyCAG-IP) are shown (top). Colonies were stained for alkaline phosphate activity (bottom). **(B)** Quantitation of colony morphologies from alkaline phosphate stained plates shown in (A). **(C)** SCKO cells, in which loxP-flanked Sox2 can be deleted in response to Tamoxifen (Tam)-inducible Cre-ERT<sup>2</sup> activity and led to differentiation, are assessed for their ES cell colony forming ability after transfection. Representative morphologies of colonies formed in the presence of Tam (1 $\mu$ g/ml) by cells transfected with Sox2 (pPyCAG-Sox2-IH), mCherry-Sox2 (pPyCAG-mCherry-Sox2-IH), or empty vector (pPyCAG-IH) are shown (top). Colonies were stained for alkaline phosphate activity (bottom). 'mCh' stands for mCherry. **(D)** Quantitation of colony morphologies from alkaline phosphate stained plates shown in (C). **(E)** Relative expression level of two key Oct6 target genes (*Kdm2b* and *Zic1*) and the neural lineage marker *Nestin* by E14Tg2a cells transfected with GFP or GFP-Oct6. Expression level was normalised to *TBP*. All experiments above were performed in triplicate. (n = 3, mean  $\pm$  SD).

## Appendix Figure S2



**Appendix Figure S2: Comparison of theoretical models for best fit to FCS data.** (A) A one-component coupled with a triplet state model (3D-1P-1T) used to fit three different ACFs generated from different protein concentration starting from zero (left panel), low (middle) to high (right panel) in a titration reaction with DNA. The attached residual illustrates the goodness of fitting by the model. (B) A two-component model (3D-2P-1T) used for fitting the above same ACFs resulting a better residual. The improved residual confirms that 3D-2P-1T is the better model for an ACF generated when protein was added to DNA. Protein-DNA Complex formation is well noticeable from the increased average diffusion time from  $273.0 \pm 2.9 \mu\text{s}$  in the absence of protein to  $320 \pm 5.2 \mu\text{s}$  at the maximum protein concentration.

## Appendix Figure S3



**Appendix Figure S3: Enriched Sox2 and POU TF motifs in CHIP-Seq data.** The top 10 motifs from Oct4, Sox2, Oct6, Brn1, and Brn2 CHIP-Seq data from E14 ESCs and NS5 NSCs are shown with WebLogo.

Name	Sequence	Figure	
SO wt	TCCACCATGGACATTGTAATGCAAAAGAAGCTGTAAG	1, 2, 3, 7, EV1, EV2, EV3	
PORE wt	AAGTTAAAATCACATTTGAAATGCAAATGGAAAAGC	1, 2, 4, 7, EV1,	
PORE mt	AAGTTAAAATCACtgccaAAATGCAAATGGAAAAGC	1, 2, 4, 7, EV1,	
MORE wt	TGATCACAGCCTCATGCATATGCATATATCGTTGTTG	4, 7	
MORE mt	TGATCACAGCCTCATGCATAcctgTATATCGTTGTTG	4	
Artificial motif	AAATAATGGAAACCTTTGTATTTATGCAAATAAACAAAGGTC ATTT	1	
fl mut	TcgAccTCACATTTGAAATGCAAAT	EV1C	
p1 mut	TTAAAATCACAgacGAAATGCAAAT		
p2 mut	TTAAAATCACATTTGAAATGCgctT		
fl p1 mut	TcgAccTCACAgacGAAATGCAAAT		
fl p2 mut	TcgAccTCACATTTGAAATGCgctT		
p1 p2 mut	TTAAAATCACAgacGAAATGCgctT		
fl p1 p2 mut	TcgAccTCACAgacGAAATGCgctT		
PORE	TTAAAATCACATTTGAAATGCAAAT		
5L>0	TTAAAATCACATTTGCAAAT		EV1D
5L>1	TTAAAATCACATTTG <u>A</u> CAAAT		
5L>2	TTAAAATCACATTTG <u>AA</u> CAAAT		
5L>3	TTAAAATCACATTTG <u>AAA</u> CAAAT		
5L>4	TTAAAATCACATTTG <u>AAAT</u> CAAAT		
PORE	TTAAAATCACATTTGAAATGCAAAT		
L1,3 mut	TTAAAATCACATTTGcAcTGCAAAT		
L3,5 mut	TTAAAATCACATTTGAAgTaCAAAT		
L1-5 mut	TTAAAATCACATTTGcgccaCAAAT		

**Appendix Table S1: Oligonucleotides for analysis of Sox/Oct, PORE, and MORE motifs.** DNAs used are shown 5'-3'. The red colour indicates the Sox/Oct, PORE, and MORE motifs; mutations are marked by lower case (substitutions) or underling (insertion). The top six DNAs are Cy5 labeled at the 5' end (unless otherwise mentioned in the Figure legend); DNAs used in Figure 2 panels C, D are untagged. fl, P1 and P2 indicate oligonucleotides with mutations in the flanking sequence, palindromic repeat 1 and repeat 2, respectively.

Primers	Sequence
Nestin-Frw	CTGCAGGCCACTGAAAAGT
Nestin-Rev	TTCCAGGATCTGAGCGATCT
Zic1-Frw	AACCTCAAGATCCACAAAAGGA
Zic1-Rev	CCTCGAACTCGCACTTGAA
Kdm2b-Frw	TCCACCCTGGATGCCTTA
Kdm2b-Rev	GCATGGTTACACTTCGGACA
TBP-Frw	GGGGAGCTGTGATGTGAAGT
TBP-Rev	CCAGGAAATAATTCTGCCTCA

**Appendix Table S2: Oligonucleotides used for qPCR.** DNAs used are shown 5'-3'. Primers for Nestin and Zic1 were described previously [1]. Primers for Kdm2b were designed using the *ROCHE Universal Probe Library Assay DesignCenter*.

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