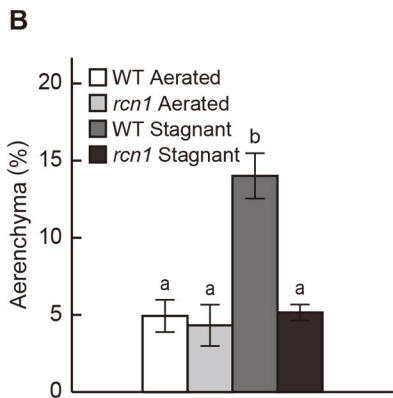
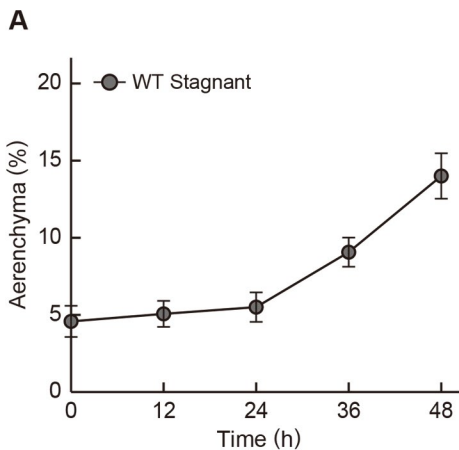
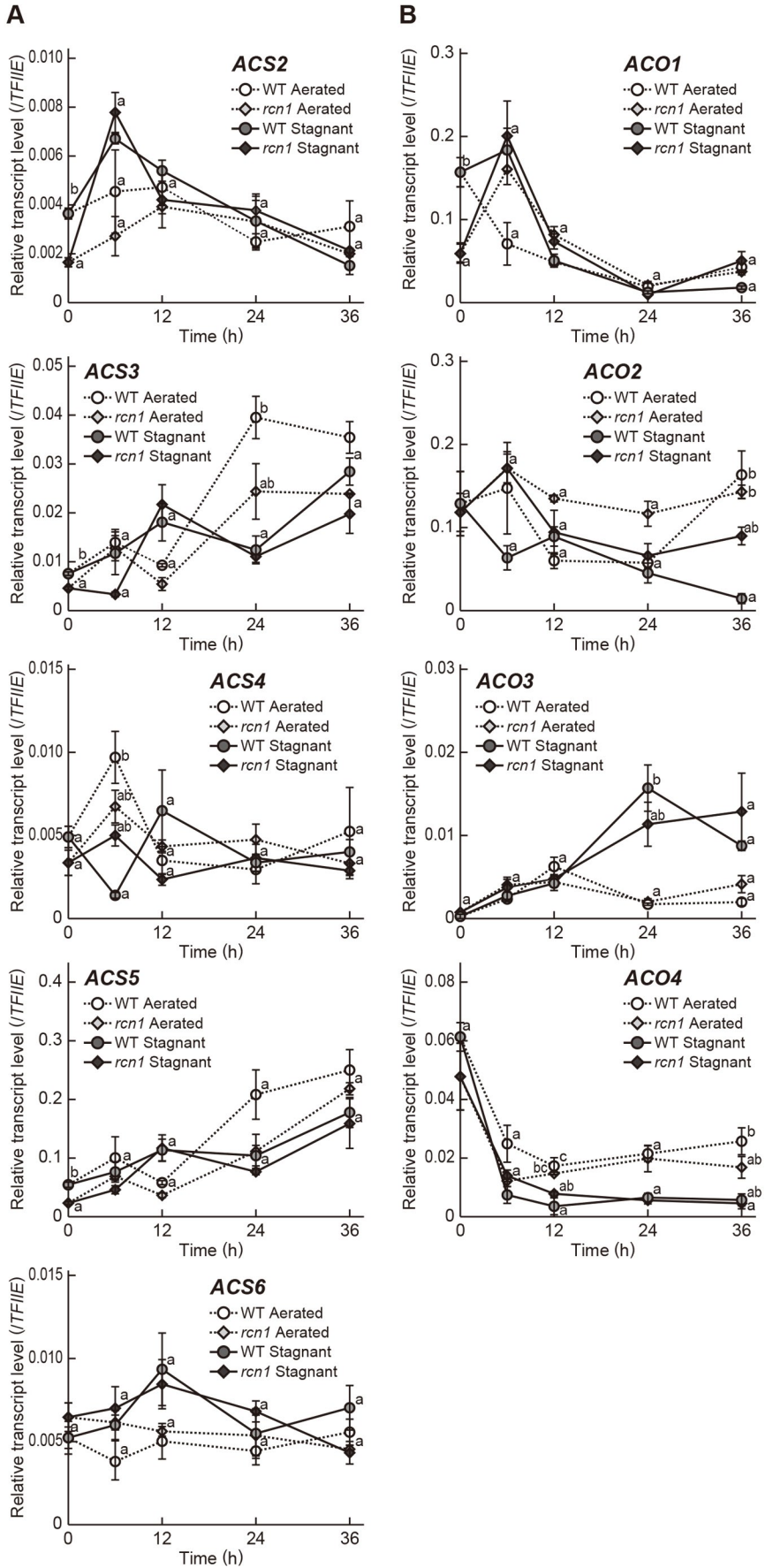


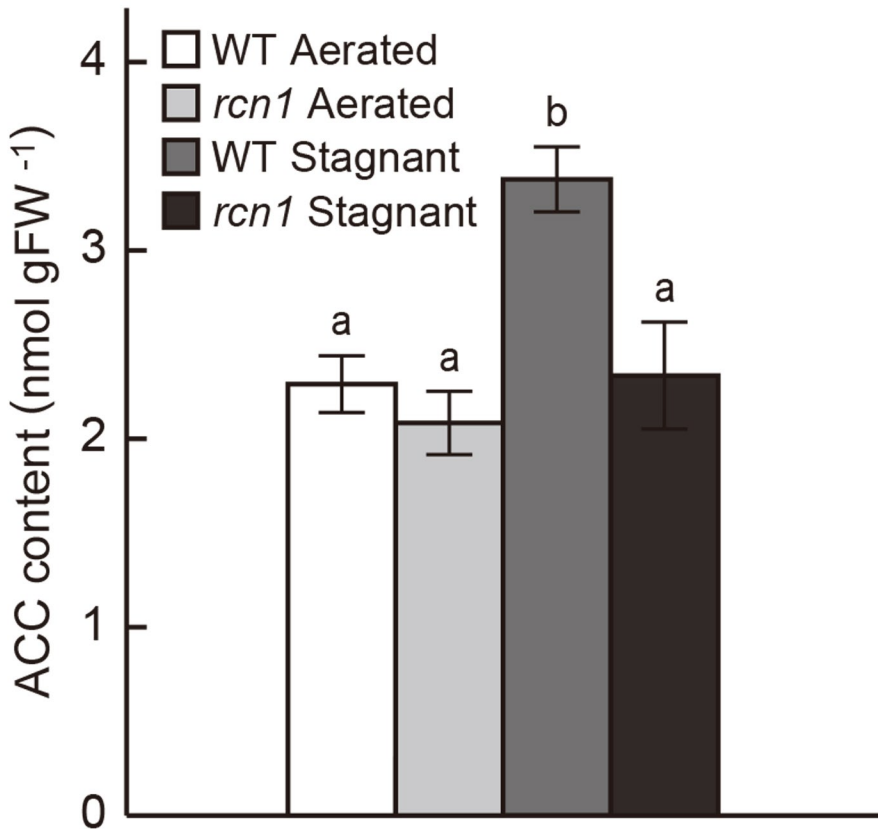
**Supplemental Figure S1.** Emergence of adventitious roots of the wild type and the *rcn1* mutant grown under aerated conditions. Emerged adventitious root numbers of the wild type and the *rcn1* mutant grown under aerated conditions were counted every day until day 10. Values are means ( $n = 15$ )  $\pm$  SE. No Significant differences were observed between the wild type and the *rcn1* mutant at  $P < 0.01$  (two sample *t* test).



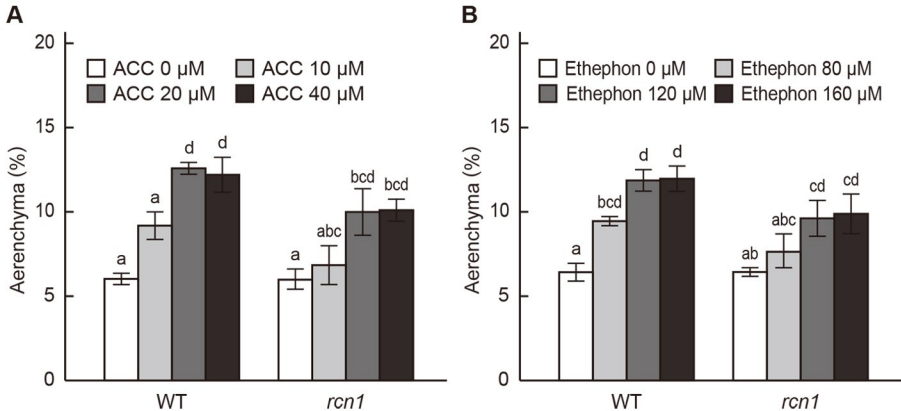
**Supplemental Figure S2.** Aerenchyma formation at 20 mm from the tips of adventitious roots in the wild type and the *rcn1* mutant. Twenty-d-old aerobically grown rice seedlings were transferred to aerated or stagnant conditions. Time-course of aerenchyma formation at 20 mm from the tips of adventitious roots in the wild type under stagnant conditions (A). The percentage of aerenchyma of root-cross-sectional area at 20 mm from the tips grown under aerated or stagnant conditions for 48 h (B). Values are means ( $n = 6$ )  $\pm$  SE. Different lower-case letters denote significant differences between the wild type and the *rcn1* mutant in each condition ( $P < 0.01$ , one-way ANOVA and then Tukey's test for multiple comparisons).



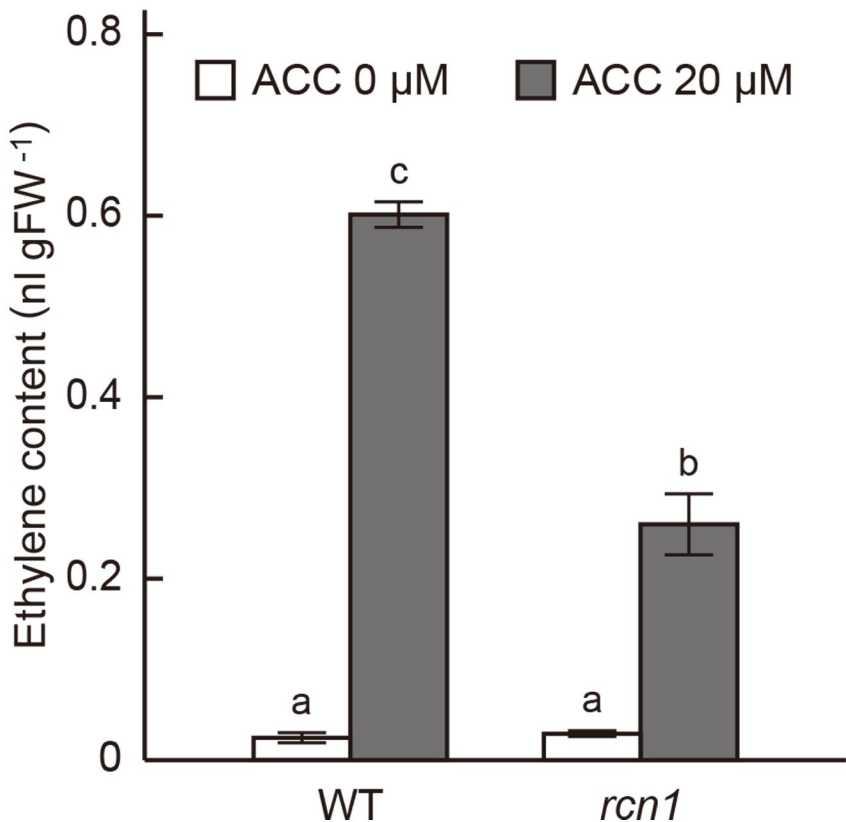
**Supplemental Figure S3.** Expression of genes encoding ethylene biosynthesis enzymes in adventitious roots of the wild type and the *rcn1* mutant grown under aerated or stagnant conditions. Time-course qRT-PCR analyses of the *ACS* genes (A) and the *ACO* genes (B) using RNAs extracted from adventitious roots at 10 mm ( $\pm 2$  mm) from the root tips grown under aerated or stagnant conditions. The transcription initiation factor *TFIIIE* gene was used as a control. Values are means ( $n = 3$ )  $\pm$  SE. Different lower-case letters denote significant differences between the wild type and the *rcn1* mutant in each condition ( $P < 0.01$ , one-way ANOVA and then Tukey's test for multiple comparisons).



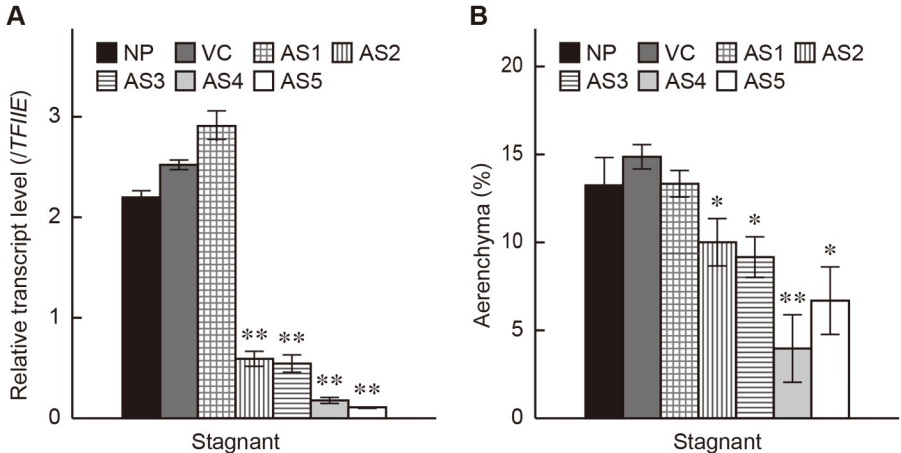
**Supplemental Figure S4.** ACC contents in adventitious roots of the wild type and the *rcn1* mutant. ACC content was measured at 10 to 30 mm from the root tips grown under aerated or stagnant conditions for 12 h. Values are means ( $n = 3$ )  $\pm$  SE. Different lower-case letters denote significant differences between the wild type and the *rcn1* mutant in each condition ( $P < 0.01$ , one-way ANOVA and then Tukey's test for multiple comparisons).



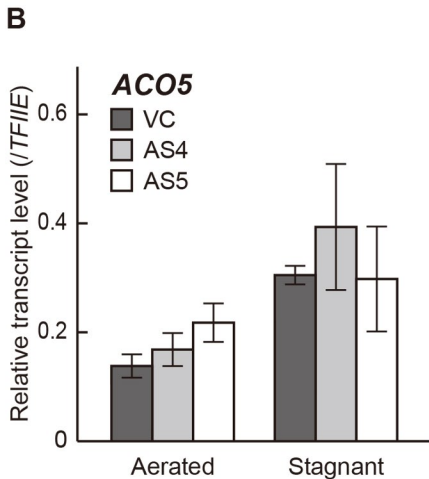
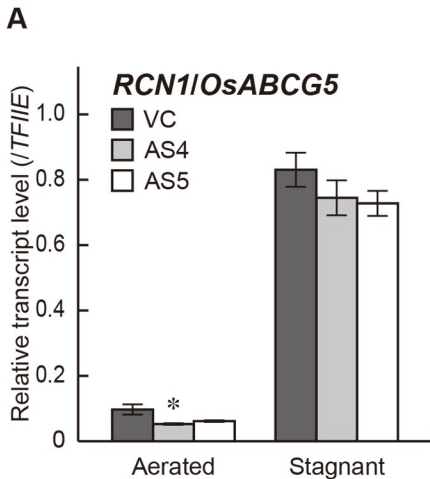
**Supplemental Figure S5.** Effects of an ethylene precursor and an ethylene-releasing compound on aerenchyma formation at 20 mm from the tips of adventitious roots of the wild type and the *rcn1* mutant. The percentage of aerenchyma of root-cross-sectional area at 20 mm from the tips of adventitious roots grown under aerated conditions for 48 h with 0, 10, 20 or 40  $\mu\text{M}$  ACC treatment (A), and grown under aerated conditions for 48 h with 0, 80, 120 or 160  $\mu\text{M}$  ethephon treatment (B). Values are means ( $n = 6$ )  $\pm$  SE. Different lower-case letters denote significant differences between the wild type and the *rcn1* mutant in each condition ( $P < 0.01$ , one-way ANOVA and then Tukey's test for multiple comparisons).



**Supplemental Figure S6.** Ethylene contents in roots of the wild type and the *rcn1* mutant grown under aerated conditions with or without 20 μM ACC treatment. Ethylene content was measured in roots at 24 h after initiation of growth under aerated conditions with or without 20 μM ACC treatment. Values are means ( $n = 3$ )  $\pm$  SE. Different lower-case letters denote significant differences between the wild type and the *rcn1* mutant in each condition ( $P < 0.01$ , one-way ANOVA and then Tukey's test for multiple comparisons).



**Supplemental Figure S7.** Expression of the *CUTIL* gene and aerenchyma formation in adventitious roots of the *pACT1::CUTIL*-antisense  $T_0$  transgenic lines. The percentage of aerenchyma of root-cross-sectional area at 10 mm (A) and at 20 mm (B) from the tips of adventitious roots of the wild type (Nipponbare; NP), the *pACT1::CUTIL*-antisense  $T_0$  transgenic lines (AS1 to AS5) and the vector control (VC) grown under stagnant conditions for 48 h. All transgenic lines had a cv. Nipponbare background. Values are means ( $n = 3$ )  $\pm$  SE. Significant differences between each of the transgenic lines and the vector control at  $P < 0.01$  or at  $P < 0.05$  (two sample  $t$  test) are denoted by \*\* or \*, respectively.



**Supplemental Figure S8.** Expression of *RCN1/OsABCG5* and *ACO5* in adventitious roots of the *pACT1::CUTIL*-antisense T<sub>1</sub> transgenic lines. qRT-PCR analyses of the *RCN1/OsABCG5* gene (A) and the *ACO5* gene (B) using RNAs extracted from adventitious roots at 10 mm ( $\pm 2$  mm) from the root tips of the *pACT1::CUTIL*-antisense T<sub>1</sub> transgenic lines (AS4 and AS5) and the vector control (VC) grown under aerated or stagnant conditions for 36 h. All transgenic lines had a cv. Nipponbare background. The transcription initiation factor *TFIIE* gene was used as a control. Values are means (n = 3)  $\pm$  SE. Significant differences between each of the transgenic lines and the vector control at  $P < 0.05$  (two sample *t* test) are denoted by \*.



**Supplemental Table S1.** List of primers used for qRT-PCR analysis

<b>Gene Name</b>	<b>ID (RAP-DB)</b>	<b>Forward primer sequence</b>	<b>Reverse primer sequence</b>
<b>ACC synthase (ACS)</b>			
<i>ACS1</i>	Os03g0727600	5'-ACAAAACCACACCATGTCCA-3'	5'-CGAAAGGAATCTGCTACTGCTGC-3'
<i>ACS2</i>	Os04g0578000	5'-CACCACCACCACCTCAGC-3'	5'-GACGTAGTAAGGCGCAGCAT-3'
<i>ACS3</i>	Os05g0196600	5'-GAGGCGAAGCTGAACATCTC-3'	5'-CATGTTGTTCTTGCTCCCATT-3'
<i>ACS4</i>	Os05g0319200	5'-AGCTGAGGCTGTGGGACA-3'	5'-GTGGCCAGGCTCATGTTC-3'
<i>ACS5</i>	Os01g0192900	5'-GCTTGGACACGCTGGATCTT-3'	5'-TTATTGCTGTTCTTGCTGCTG-3'
<i>ACS6</i>	Os06g0130400	5'-GGATGGTTCAGGTGTTGCTT-3'	5'-CCTGGCAAAGCAGTTATTCC-3'
<b>ACC oxidase (ACO)</b>			
<i>ACO1</i>	Os09g0451000	5'-GATAGCGTGTGTACCACAGCGACC-3'	5'-AGGTAGAAAACGCGAGCTGA-3'
<i>ACO2</i>	Os09g0451400	5'-AAGTCCATGGAAACCGAGAC-3'	5'-CCACAGTTCATGCACACACA-3'
<i>ACO3</i>	Os02g0771600	5'-GAGGTTCGTGTTTCGAGGACT-3'	5'-CGCAGCCGTAGCTAGTGAAG-3'
<i>ACO4</i>	Os11g0186900	5'-GCATGGCCAACATTGCTC-3'	5'-GTTCGCCAGGGCTGCGAACC-3'
<i>ACO5</i>	Os05g0149400	5'-CGAGTACCCGGAGTACGTGTT-3'	5'-ATTTTGGCGCCTTGACGGCC-3'
<i>ACO7</i>	Os01g0580500	5'-GGACTACTACCAGGGCACCA-3'	5'-GATTAGCGCACGCGATTTTA-3'
<b>G-type ABC transporter</b>			
<i>RCN1/OsABCG5</i>	Os03g0281900	5'-AAGTGGGAGTGCCTCTGGAT-3'	5'-GGTGAGACGAGGTGACGAT-3'
<b>Fatty acid elongase (<math>\beta</math>-ketoacyl-CoA synthase; KCS)</b>			
<i>CUTIL</i>	Os03g0220100	5'-AAGTGCCTCCGCACAGTC-3'	5'-ACCATGGATTTCGATCACAGC-3'
<b><i>CUTIL</i> 5'-UTR primers for the <i>CUTIL</i>- antisense transgenic lines</b>			
<i>CUTIL5UTR</i>	Os03g0220100	5'-CCATCACCAACACACCACTG-3'	5'-TCCTACGTACGACTTTAGTTTCTTC-3'
<b>Control</b>			
<i>TFIIE</i>	Os10g0397200	5'-GTGCAGCCCAAGGCTAAG-3'	5'-CGTCGAATAAGCGTAGAGCA-3'