

**Supporting Information**  
**for**  
**Synthesis of icariin from kaempferol through**  
**regioselective methylation and *para*-Claisen–Cope**  
**rearrangement**

Qinggang Mei<sup>1,2</sup>, Chun Wang<sup>1</sup>, Zhigang Zhao<sup>3</sup>, Weicheng Yuan<sup>2</sup> and Guolin Zhang\*<sup>1</sup>

Address: <sup>1</sup>Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu 610041, China, <sup>2</sup>Chengdu Institute of Organic Chemistry, Chinese Academy of Sciences, Chengdu 610041, China and <sup>3</sup>College of Chemistry and Environmental Protection Engineering, Southwest University for Nationalities, Chengdu 610041, China

Email: Guolin Zhang\* - zhanggl@cib.ac.cn

\*Corresponding author

**Experimental section and copies of**  
**NMR, ESI-HRMS, HMBC, HSQC, COSY and NOESY spectra**

Contents	Page No.
Experimental section	S2–S11
<sup>1</sup> H NMR spectra of <b>1, 2, 3, 7, 8, 9, 10, 12, 14</b>	S12–S20
<sup>13</sup> C NMR spectra of <b>1, 2, 3, 7, 8, 9, 10, 12, 14</b>	S21–S29
ESI-HRMS spectra of <b>1, 2, 3, 7, 8, 9, 10, 12, 14</b>	S30–S38
HMBC spectra of <b>1, 2, 3, 7, 8, 9, 12, 14</b>	S39–S46
HSQC/COSY spectra of <b>1, 2, 14</b> ; NOESY Spectrum of <b>3</b>	S47–S53

## Experimental Section

Melting points were determined by an X-6 apparatus without correction. Optical rotations were measured with a Perkin-Elmer M341 automatic polarimeter. IR spectra were recorded on a Perkin-Elmer Spectrum One spectrometer (KBr disc) and NMR spectra on a Bruker AC-400 spectrometer ( $^1\text{H}$ : 400 MHz;  $^{13}\text{C}$ : 100 MHz) with  $\text{DMSO-}d_6$  ( $\delta$  2.50/39.52) as solvent and internal standard at ambient temperature. The 2D NMR spectra (COSY, NOESY and  $^1\text{H}$ - $^{13}\text{C}$  HMBC, HSQC) were conducted using standard software. The ESI-HRMS was carried out on a Bruker Bio TOF IIIQ (quadrupole time of flight) mass spectrometer. Preparative TLC was performed using commercially available precoated glass silica gel GF<sub>254</sub> plates of 0.15–0.2 mm thickness. Column chromatography was carried out with 200–300 mesh silica gel using the flash technique.

Unless otherwise specified, chemicals and solvents were of analytical reagent grade and used as obtained from commercial sources without further purification. Kaempferol was from Yangling Dongke Maidisen Pharmaceutical Co., Ltd., China. Icariin reference substance was purchased from Aladdin Reagent (Shanghai) Co., Ltd. DMF was dried with 4 Å molecular sieves and distilled at reduced pressure. PhCl was freshly distilled from sodium benzophenone ketyl prior to use.  $\text{CH}_2\text{Cl}_2$  and  $\text{CHCl}_3$  were freshly distilled over  $\text{CaH}_2$  and acetone was freshly distilled from potassium carbonate prior to use.

Kaempferol was converted to 7-O-benzylkaempferide (**6**) following the reported procedure [1]. 2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-glucopyranosyl bromide (**15**) and 2,3,4-tri-O-acetyl- $\alpha$ -L-rhamnopyranosyl bromide (**16**) were synthesized according to literature procedures [2].

### 3-O-Methoxymethyl-4'-O-methyl-7-O-benzylkaempferol (7)

To a solution of 7-O-benzylkaempferide (**6**, 3.12 g, 8 mmol) and  $i\text{Pr}_2\text{NEt}$  (3.76 mL, 21.6 mmol) in  $\text{CH}_2\text{Cl}_2$  (80 mL), chloromethyl methyl ether (0.85 mL, 11.2 mmol) was added at 0 °C. The mixture was stirred for 6 h at room temperature, and was acidified cautiously with 0.5 M HCl (aq) until pH = 5. The organic phase was washed with brine (50 mL), dried over  $\text{MgSO}_4$ , and evaporated under reduced pressure. The dried residue was recrystallized from EtOAc/95% ethanol (1:5) to provide **7** as yellow needles. Yield: 3.24 g (93%); mp: 116–117 °C. IR ( $\text{cm}^{-1}$ ): 3444, 2919, 1664, 1607, 1497, 1351, 1306, 1265, 1221, 1174, 1082, 1025.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  12.55 (s, 1H, OH-5), 8.05 (d,  $J = 8.9$  Hz, 2H, H-2'/6'), 7.47 (d,  $J = 7.3$  Hz, 2H, H-2'''/6'''), 7.41 (t,  $J = 7.3$  Hz, 2H, H-3'''/5'''), 7.36 (d,  $J = 7.3$  Hz, 1H, H-4'''), 7.15 (d,  $J = 8.9$  Hz, 2H, H-3'/5'), 6.85 (d,  $J = 2.0$  Hz, 1H, H-8), 6.48 (d,  $J = 2.0$  Hz, 1H, H-6), 5.24 (s, 2H, H-7'''), 5.14 (s, 2H,  $\text{OCH}_2\text{O}$ ), 3.86 (s, 3H,  $\text{OCH}_3$ -4'), 3.14 (s, 3H,  $\text{CH}_2\text{OCH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ )  $\delta$  177.83 (C-4), 164.22 (C-7), 161.40 (C-4'), 160.94 (C-5), 156.40 (C-2), 156.34 (C-9), 136.06 (C-1'''), 134.72 (C-3), 130.46 (C-2'/6'), 128.50 (C-3'''/5'''), 128.10 (C-4'''), 127.81 (C-2'''/6'''), 122.08 (C-1'), 114.06 (C-3'/5'), 105.24 (C-10), 98.52 (C-6), 97.19 ( $\text{OCH}_2\text{O}$ ), 93.30 (C-8), 70.01 (C-7'''), 57.04 ( $\text{CH}_2\text{OCH}_3$ ), 55.44 ( $\text{OCH}_3$ -4'). ESI-HRMS  $m/z$ : 457.1258 [ $\text{M}+\text{Na}$ ] $^+$  (calcd for  $\text{C}_{25}\text{H}_{22}\text{O}_7\text{Na}$ : 457.1258).

### 3-O-Methoxymethyl-4'-O-methyl-5-O-isopentenyl-7-O-benzylkaempferol (8)

To a well stirred suspension of **7** (2.5 g, 5.76 mmol), 18-crown-6 (0.05 g, 0.19 mmol) and anhydrous potassium carbonate (2.07 g, 15 mmol) in dry acetone (70 mL), a solution of 3,3-dimethylallyl bromide (1.94 mL, 16.13 mmol) in dry acetone (8 mL) was added dropwise over 20 min at room temperature. The resulting suspension was continuously stirred for 19 h. After evaporation of the filtrate under reduced pressure,

the obtained yellow viscous oil was dissolved in CH<sub>2</sub>Cl<sub>2</sub> and subjected to silica gel column chromatography with EtOAc/petroleum ether (1:3) as an eluent to offer an off-white solid. Recrystallization from CH<sub>2</sub>Cl<sub>2</sub>-95% ethanol (1:9) provided the desired compound **8** as off-white crystals. Yield: 2.49 g (86%); mp: 79–81 °C. IR (cm<sup>-1</sup>): 2935, 1633, 1608, 1575, 1455, 1350, 1295, 1262, 1180, 1099, 1035, 826. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.02 (d, *J* = 8.9 Hz, 2H, H-2'/6'), 7.49 (d, *J* = 7.4 Hz, 2H, H-2'''/6'''), 7.42 (t, *J* = 7.4 Hz, 2H, H-3'''/5'''), 7.37 (t, *J* = 7.4 Hz, 1H, H-4'''), 7.12 (d, *J* = 8.9 Hz, 2H, H-3'/5'), 6.88 (d, *J* = 2.1 Hz, 1H, H-8), 6.60 (d, *J* = 2.1 Hz, 1H, H-6), 5.47 (brs, 1H, H-2''), 5.24 (s, 2H, H-7'''), 5.09 (s, 2H, OCH<sub>2</sub>O), 4.62 (d, *J* = 6.5 Hz, 2H, H-1''), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.11 (s, 3H, CH<sub>2</sub>OCH<sub>3</sub>), 1.76 (s, 3H, H-5''), 1.72 (s, 3H, H-4''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 171.94 (C-4), 162.61 (C-7), 160.83 (C-4'), 159.50 (C-5), 158.16 (C-9), 152.40 (C-2), 136.86 (C-3''), 136.75 (C-3), 136.15 (C-1'''), 129.99 (C-2'/6'), 128.50 (C-3'''/5'''), 128.12 (C-4'''), 127.93 (C-2'''/6'''), 122.53 (C-1'), 119.57 (C-2''), 113.92 (C-3'/5'), 108.67 (C-10), 97.55 (C-6), 96.68 (OCH<sub>2</sub>O), 93.83 (C-8), 69.95 (C-7'''), 65.86 (C-1''), 56.88 (CH<sub>2</sub>OCH<sub>3</sub>), 55.36 (OCH<sub>3</sub>-4'), 25.44 (C-5''), 18.10 (C-4''). ESI-HRMS *m/z*: 503.2075 [M+H]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>31</sub>O<sub>7</sub>: 503.2064).

**3-O-Methoxymethyl-7-O-benzylcaritin (9) and 4-benzyl-2,3,3-trimethyl-7-(4-methoxyphenyl)-8-methoxymethoxy-2,3-dihydrofuro[2,3-*f*]chromen-9-one (12)**

A solution of **8** (3 g, 5.98 mmol) in dry PhCl (80 mL) was mixed with Eu(fod)<sub>3</sub> (0.62 g, 0.6 mmol) and NaHCO<sub>3</sub> (0.5 g, 6 mmol) under nitrogen atmosphere. After stirring for 24 h at 85 °C, the mixture was cooled, filtered and evaporated to dryness. The dried residue was subjected to silica gel column chromatography eluting with petroleum ether/CH<sub>2</sub>Cl<sub>2</sub>/EtOAc (6:3:0.5). 8-Prenyl derivative **9** was obtained after evaporation of solvents. A diastereomeric mixture of dihydrofuro[2,3-*f*]chromone **12** was succeeded by further elution with acetone/petroleum ether (1:3).

Compound **9**: pale yellow powder, yield 1.84 g (61%), mp 151–153 °C. IR (cm<sup>-1</sup>): 3438, 2924, 2853, 1652, 1610, 1594, 1438, 1377, 1301, 1253, 1178, 1087, 840. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.61 (s, 1H, OH-5), 8.02 (d, *J* = 9.0 Hz, 2H, H-2'/6'), 7.46 (d, *J* = 7.1 Hz, 2H, H-2'''/6'''), 7.41 (t, *J* = 7.1 Hz, 2H, H-3'''/5'''), 7.35 (t, *J* = 7.1 Hz, 1H, H-4'''), 7.15 (d, *J* = 9.0 Hz, 2H, H-3'/5'), 6.66 (s, 1H, H-6), 5.26 (s, 2H, H-7'''), 5.15 (brs, 1H, H-2''), 5.12 (s, 2H, OCH<sub>2</sub>O), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.46 (d, *J* = 6.6 Hz, 2H, H-1''), 3.11 (s, 3H, CH<sub>2</sub>OCH<sub>3</sub>), 1.60 (s, 6H, H-4''/5''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 178.18 (C-4), 161.48 (C-7), 161.42 (C-4'), 159.45 (C-5), 156.44 (C-2), 152.87 (C-9), 136.34 (C-1'''), 134.34 (C-3), 131.41 (C-3''), 130.40 (C-2'/6'), 128.49 (C-3'''/5'''), 128.05 (C-4'''), 127.62 (C-2'''/6'''), 122.02 (C-1'), 121.92 (C-2''), 114.15 (C-3'/5'), 107.43 (C-8), 104.71 (C-10), 97.20 (OCH<sub>2</sub>O), 96.33 (C-6), 70.20 (C-7'''), 57.04 (CH<sub>2</sub>OCH<sub>3</sub>), 55.47 (OCH<sub>3</sub>-4'), 25.41 (C-5''), 21.35 (C-1''), 17.72 (C-4''). ESI-HRMS *m/z* 503.2079 [M+H]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>31</sub>O<sub>7</sub>: 503.2064).

Compound **12**: pale yellow glassy solid, yield 0.87 g (29%), mp 72–74 °C. IR (cm<sup>-1</sup>): 2960, 2929, 1626, 1607, 1445, 1347, 1298, 1257, 1183, 1071, 943. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.00 (d, *J* = 9.0 Hz, 2H, H-2'/6'), 7.49 (d, *J* = 7.2 Hz, 2H, H-2'''/6'''), 7.44 (t, *J* = 7.2 Hz, 2H, H-3'''/5'''), 7.36 (t, *J* = 7.2 Hz, 1H, H-4'''), 7.12 (d, *J* = 9.0 Hz, 2H, H-3'/5'), 6.90 (s, 1H, H-8), 5.27 (s, 2H, H-7'''), 5.09 (s, 2H, OCH<sub>2</sub>O), 4.52 (q, *J* = 6.6 Hz, 1H, H-2''), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.09 (s, 3H, CH<sub>2</sub>OCH<sub>3</sub>), 1.38 (s, H-4'' or 5''), 1.37 (d, *J* = 6.6 Hz, 3H, H-3''), 1.12 (s, 3H, H-4'' or 5''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 171.80 (C-4), 160.91 (C-4'), 158.89 (C-7), 157.55 (C-9), 156.76 (C-5), 153.67 (C-2), 136.37 (C-3), 136.31 (C-1'''), 130.10 (C-2'/6'), 128.59 (C-3'''/5'''), 128.06 (C-4'''), 127.44 (C-2'''/6'''), 122.63 (C-1'), 119.50 (C-6), 113.92 (C-3'/5'), 105.44 (C-10), 96.69 (OCH<sub>2</sub>O), 93.49 (C-8), 90.24 (C-2''), 69.94 (C-7'''), 56.88 (CH<sub>2</sub>OCH<sub>3</sub>), 55.39 (OCH<sub>3</sub>-4'), 42.94 (C-1''), 25.15 (C-4'' or 5''), 20.71 (C-4'' or 5''), 13.96 (C-3''). ESI-HRMS *m/z*:

525.1900 [M+Na]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>30</sub>O<sub>7</sub>Na: 525.1884).

### 7-O-Benzylcaritin (**10**)

To a solution of **9** (2.01 g, 4 mmol) in MeOH (120 mL), 3 M HCl (aq) (30 mL) was added under nitrogen atmosphere. The reaction mixture was refluxed for 2.5 h, and then half of the solvent was removed under reduced pressure. The residue was neutralized with saturated NaHCO<sub>3</sub> solution and retained for 12 h at room temperature. Abundant yellow precipitates were recovered by filtration. After washing with ethanol, the crude product was recrystallized from petroleum ether/EtOAc (4:1) to give **10** as yellow granules. Yield: 1.74 g (95%); mp: 185–187 °C. IR (cm<sup>-1</sup>): 3304, 2928, 1648, 1616, 1590, 1552, 1511, 1317, 1261, 1171, 1076, 985. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.51 (s, 1H, OH-5), 9.61 (s, 1H, OH-3), 8.14 (d, *J* = 9.1 Hz, 2H, H-2'/6'), 7.46 (d, *J* = 6.9 Hz, 2H, H-2'''/6'''), 7.42 (t, *J* = 7.3 Hz, 2H, H-3'''/5'''), 7.35 (t, *J* = 7.1 Hz, 1H, H-4'''), 7.14 (d, *J* = 9.1 Hz, 2H, H-3'/5'), 6.64 (s, 1H, H-6), 5.26 (s, 2H, H-7'''), 5.15 (t, *J* = 6.9 Hz, 1H, H-2''), 3.84 (s, 3H, OCH<sub>3</sub>-4'), 3.49 (d, *J* = 6.9 Hz, 2H, H-1''), 1.63 (s, 3H, H-4''), 1.61 (s, 3H, H-5''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 176.53 (C-4), 161.20 (C-7), 160.66 (C-4'), 158.99 (C-5), 152.67 (C-9), 146.81 (C-2), 136.46 (C-1'''), 136.03 (C-3), 131.39 (C-3'''), 129.34 (C-2'/6'), 128.52 (C-3'''/5'''), 128.07 (C-4'''), 127.65 (C-2'''/6'''), 123.48 (C-1'), 122.10 (C-2''), 114.18 (C-3'/5'), 107.21 (C-8), 103.73 (C-10), 95.79 (C-6), 70.20 (C-7'''), 55.44 (OCH<sub>3</sub>-4'), 25.46 (C-5''), 21.40 (C-1''), 17.77 (C-4''). ESI-HRMS *m/z*: 481.1620 [M+Na]<sup>+</sup> (calcd for C<sub>28</sub>H<sub>26</sub>O<sub>6</sub>Na: 481.1622).

### Icaritin (**3**)

A mixture of **10** (1.37 g, 3 mmol), 10% palladium on carbon (1.5 g) and 1,4-cyclohexadiene (2.9 mL, 30 mmol) in MeOH (110 mL) was stirred at room temperature for 2 h, followed by filtration through celite, rinsed with EtOAc (20 mL).

The filtrate was concentrated and recrystallized from MeOH to afford pure icaritin (**3**) as yellow needles. Yield: 0.93 g (84%); mp: 207–208 °C; IR (cm<sup>-1</sup>): 3319, 2927, 1626, 1603, 1536, 1422, 1379, 1318, 1257, 1178, 1149, 1036, 838; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.37 (s, 1H, OH-5), 10.74 (s, 1H, OH-7), 9.48 (s, 1H, OH-3), 8.12 (d, *J* = 9.1 Hz, 2H, H-2'/6'), 7.13 (d, *J* = 9.1 Hz, 2H, H-3'/5'), 6.29 (s, 1H, H-6), 5.17 (t, *J* = 6.9 Hz, 1H, H-2''), 3.84 (s, 3H, OCH<sub>3</sub>-4'), 3.43 (d, *J* = 6.9 Hz, 2H, H-1''), 1.75 (s, 3H, H-4''), 1.63 (s, 3H, H-5''); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 176.22 (C-4), 161.22 (C-7), 160.47 (C-4'), 158.29 (C-5), 153.51 (C-9), 146.19 (C-2), 135.90 (C-3), 131.00 (C-3''), 129.16 (C-2'/6'), 123.55 (C-1'), 122.46 (C-2''), 114.09 (C-3'/5'), 105.62 (C-8), 103.06 (C-10), 97.81 (C-6), 55.38 (OCH<sub>3</sub>-4'), 25.41 (C-5''), 21.18 (C-1''), 17.81 (C-4''); ESI-HRMS *m/z*: 369.1328 [M+H]<sup>+</sup> (calcd for C<sub>21</sub>H<sub>21</sub>O<sub>6</sub>: 369.1333). The <sup>1</sup>H and <sup>13</sup>C NMR data of icaritin (**3**) were in agreement with those reported [3,4].

### **7-O-(2''',3''',4''',6'''-Tetra-O-acetyl-β-D-glucopyranosyl)icaritin (13)**

To a stirred suspension containing **3** (0.8 g, 2.17 mmol), Ag<sub>2</sub>CO<sub>3</sub> (0.78 g, 2.83 mmol) and molecular sieves (4 Å powder) (1.2 g) in a mixed-solvent of DMF (2 mL) and CHCl<sub>3</sub> (12 mL), **15** (1.16 g, 2.83 mmol) in CHCl<sub>3</sub> (6 mL) was added. The mixture was stirred in dark at room temperature for 30 h under nitrogen, then diluted with 10 mL of CHCl<sub>3</sub> and filtered through celite eluting with CHCl<sub>3</sub>. The filtrate was extracted with 0.3 M HCl (aq) (25 mL), washed with saturated NaHCO<sub>3</sub> solution, brine and dried over MgSO<sub>4</sub>. After evaporation of solvents, **13** was collected as yellow powder (1.13 g), which was used in the next step without further purification.

### **3-O-(2''',3''',4''',6'''-Tri-O-acetyl-α-L-rhamnopyranosyl)-7-O-(2''',3''',4''',6'''-tetra-O-acetyl-β-D-glucopyranosyl)icaritin (14)**

A mixture of the above powder **13** (0.89 g), rhamnose bromide **16** (0.66 g, 1.88 mmol), and Ag<sub>2</sub>O (0.44 g, 1.88 mmol) was stirred in the presence of molecular sieves (4 Å

powder) (1.5 g) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) under nitrogen for 32 h, followed by filtration through celite and washed with CH<sub>2</sub>Cl<sub>2</sub>. The filtrate was concentrated under reduced pressure and the residue was purified over silica gel column chromatography with CH<sub>2</sub>Cl<sub>2</sub>/EtOAc (9:1). Compound **14** was collected as yellow needles. Yield: 514 mg (31%, for 2 steps); mp: 194–196 °C;  $[\alpha]_D^{20}$  -111.8 (c 0.17, CHCl<sub>3</sub>). IR (cm<sup>-1</sup>): 2925, 1756, 1652, 1599, 1434, 1372, 1222, 1180, 1045. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.41 (s, 1H, OH-5), 7.87 (d, *J* = 9.0 Hz, 2H, H-2'/6'), 7.15 (d, *J* = 9.0 Hz, 2H, H-3'/5'), 6.64 (s, 1H, H-6), 5.72 (d, *J* = 7.9 Hz, 1H, H-1'''), 5.53 (dd, *J* = 3.3, 1.6 Hz, 1H, H-2'''), 5.47 (d, *J* = 1.6 Hz, 1H, H-1'''), 5.42 (t, *J* = 9.6 Hz, 1H, H-3'''), 5.18 – 5.10 (m, 2H, H-2'''/3'''), 5.02 (t, *J* = 9.6 Hz, 2H, H-2''/4'''), 4.79 (t, *J* = 10.0 Hz, 1H, H-4'''), 4.41 – 4.33 (m, 1H, H-5'''), 4.21 (dd, *J* = 12.2, 6.4 Hz, 1H, H-6'''b), 4.09 (d, *J* = 10.2 Hz, 1H, H-6'''a), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.37 – 3.26 (m, 3H, H-1''/5'''), 2.09 (s, 3H, CH<sub>3</sub>COO), 2.04 (s, 3H, CH<sub>3</sub>COO), 2.02 (s, 6H, 2×CH<sub>3</sub>COO), 1.98 (s, 6H, 2×CH<sub>3</sub>COO), 1.96 (s, 3H, CH<sub>3</sub>COO), 1.62 (s, 3H, H-4''), 1.60 (s, 3H, H-5''), 0.77 (d, *J* = 6.2 Hz, 3H, H-6'''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 177.94 (C-4), 170.08 (CH<sub>3</sub>COO), 169.75 (CH<sub>3</sub>COO), 169.72 (CH<sub>3</sub>COO), 169.64 (CH<sub>3</sub>COO), 169.54 (CH<sub>3</sub>COO), 169.50 (CH<sub>3</sub>COO), 169.16 (CH<sub>3</sub>COO), 161.83 (C-4'), 159.15 (C-7), 159.12 (C-5), 157.99 (C-2), 153.28 (C-9), 134.02 (C-3), 131.73 (C-3''), 130.69 (C-2'/6'), 121.70 (C-1'), 121.59 (C-2''), 114.31 (C-3'/5'), 108.63 (C-8), 106.32 (C-10), 98.04 (C-6), 98.01 (C-1'''), 96.80 (C-1'''), 71.83 (C-3'''), 71.15 (C-5'''), 70.56 (C-2'''), 69.37 (C-4'''), 68.51 (C-2'''), 68.26 (C-3'''), 68.11 (C-4'''), 67.95 (C-5'''), 61.78 (C-6'''), 55.67 (OCH<sub>3</sub>-4'), 25.47 (C-5''), 21.23 (C-1''), 20.60 (CH<sub>3</sub>COO), 20.51 (CH<sub>3</sub>COO), 20.50 (2×CH<sub>3</sub>COO), 20.46 (CH<sub>3</sub>COO), 20.38 (2×CH<sub>3</sub>COO), 17.85 (C-4''), 16.86 (C-6'''). ESI-HRMS *m/z*: 971.3174 [M+H]<sup>+</sup> (calcd for C<sub>47</sub>H<sub>55</sub>O<sub>22</sub>: 971.3179).



## Icariin (1)

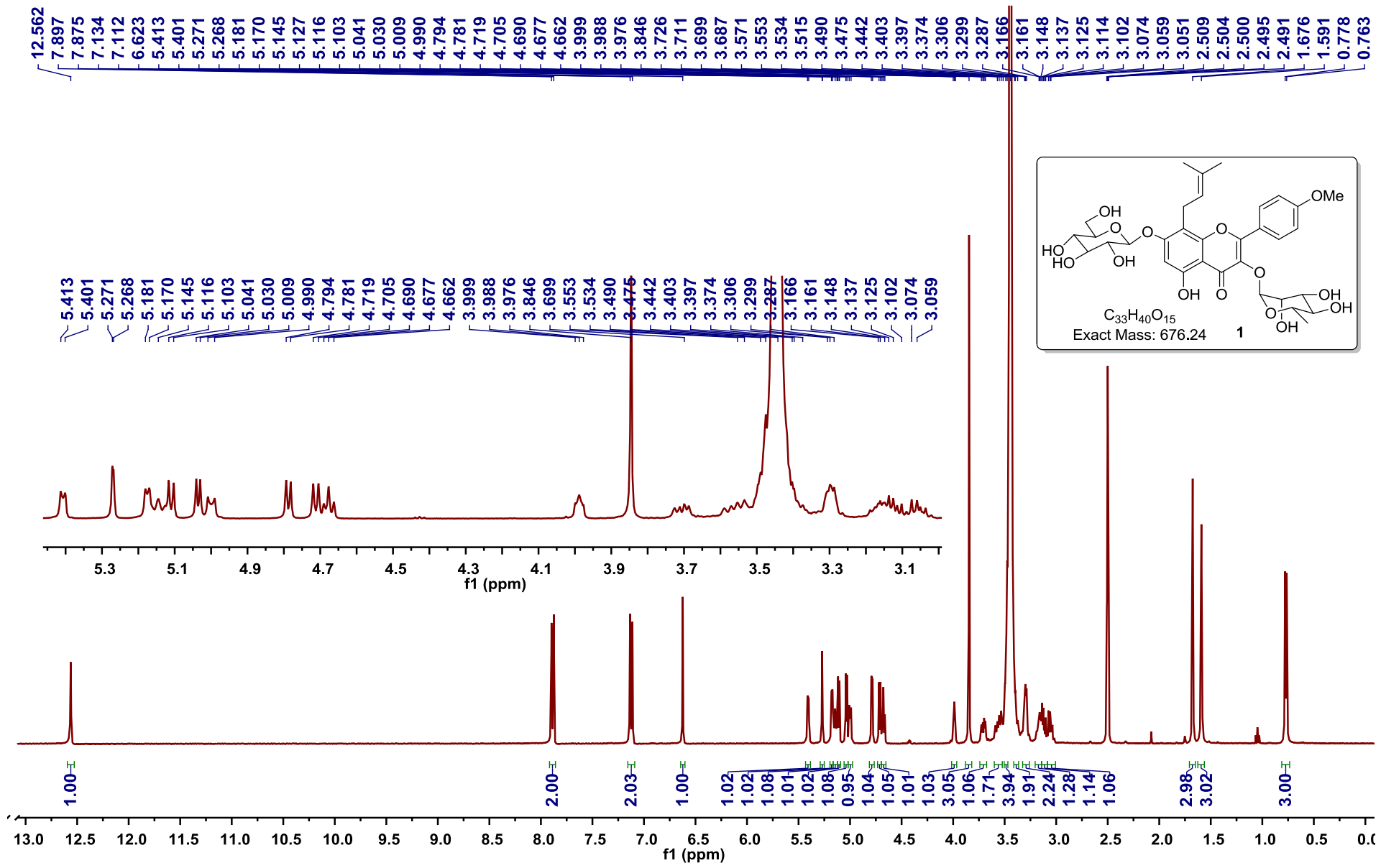
The solution of compound **14** (0.44 g, 0.45 mmol) in 7.0 M methanolic ammonia (15 mL) was stirred at room temperature for 3 h. The solution was concentrated and kept overnight at room temperature. Yellow powdery crystals of **1** were collected by filtration, washed with petroleum ether, and dried in vacuum. Yield: 286 mg (94%); mp: 224–226 °C;  $[\alpha]_D^{20}$  –120.0 (*c* 0.08, CH<sub>3</sub>OH). IR (cm<sup>-1</sup>): 3368, 2928, 1651, 1598, 1503, 1440, 1304, 1259, 1182, 1074. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.56 (s, 1H, OH-5), 7.89 (d, *J* = 9.0 Hz, 2H, H-2'/6'), 7.12 (d, *J* = 9.0 Hz, 2H, H-3'/5'), 6.62 (s, 1H, H-6), 5.41 (d, *J* = 4.7 Hz, 1H, OH-2'''), 5.27 (d, *J* = 1.3 Hz, 1H, H-1''''), 5.18 (d, *J* = 4.3 Hz, 1H, OH-3'''), 5.14 (t, *J* = 7.6 Hz, 1H, H-2''), 5.11 (d, *J* = 5.4 Hz, 1H, OH-4'''), 5.04 (d, *J* = 4.6 Hz, 1H, OH-2''''), 5.00 (d, *J* = 7.4 Hz, H-1'''), 4.79 (d, *J* = 4.9 Hz, 1H, OH-4''''), 4.71 (d, *J* = 5.8 Hz, 1H, OH-3''''), 4.68 (t, *J* = 5.5 Hz, 1H, OH-6'''), 3.99 (t, *J* = 4.6 Hz, 1H, H-2''''), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.71 (dd, *J* = 10.1, 5.3 Hz, 1H, H-6'''b), 3.56 (dd, *J* = 14.6, 7.6 Hz, 1H, H-1''b), 3.51 – 3.47 (m, 1H, H-3''''), 3.44 – 3.37 (m, 3H, H-1''a/5''''/6''''a, overlapped with H<sub>2</sub>O), 3.31 – 3.27 (m, 2H, H-2'''/3'''), 3.19 – 3.02 (m, 3H, H-4'''/4''''/5''''), 1.68 (s, 3H, H-4''), 1.59 (s, 3H, H-5''), 0.77 (d, *J* = 6.0 Hz, 3H, H-6'''''); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ 178.39 (C-4), 161.52 (C-4'), 160.60 (C-7), 159.18 (C-5), 157.49 (C-2), 153.12 (C-9), 134.72 (C-3), 131.30 (C-3''), 130.71 (C-2'/6'), 122.34 (C-1'), 122.21 (C-2''), 114.20 (C-3'/5'), 108.41 (C-8), 105.69 (C-10), 102.06 (C-1''''), 100.59 (C-1'''), 98.21 (C-6), 77.27 (C-5'''), 76.67 (C-3'''), 73.45 (C-2'''), 71.18 (C-4'''), 70.83 (C-5''''), 70.39 (C-3''''), 70.18 (C-2''''), 69.74 (C-4''''), 60.71 (C-6'''), 55.63 (OCH<sub>3</sub>-4'), 25.59 (C-5''), 21.53 (C-1''), 17.98 (C-4''), 17.56 (C-6'''). ESI-HRMS *m/z*: 699.2256 [M+Na]<sup>+</sup> (calcd for C<sub>33</sub>H<sub>40</sub>O<sub>15</sub>Na: 699.2259). The <sup>1</sup>H and <sup>13</sup>C NMR data of icariin (**1**) were in agreement with those reported [4,5].

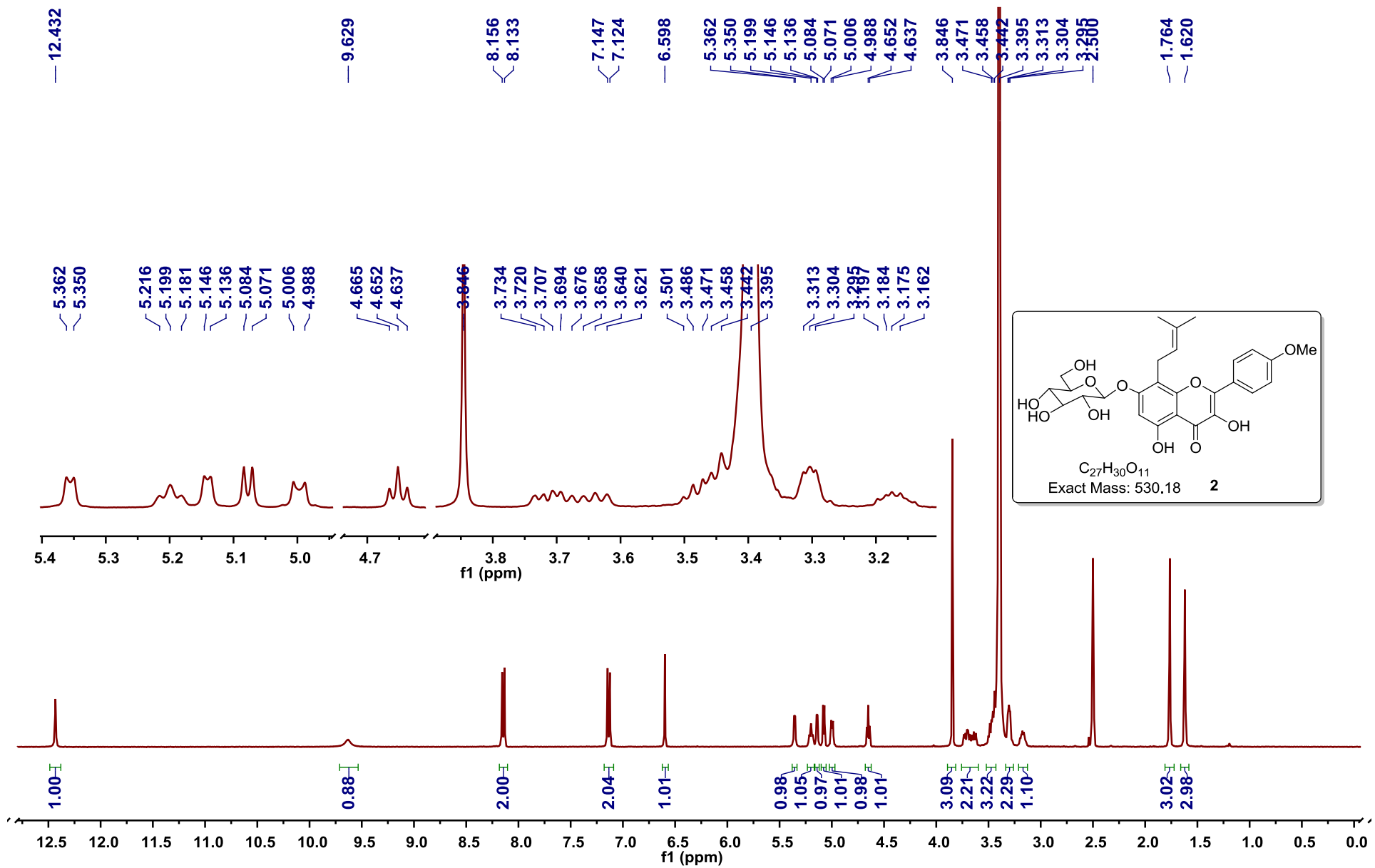
## Icariside I (2)

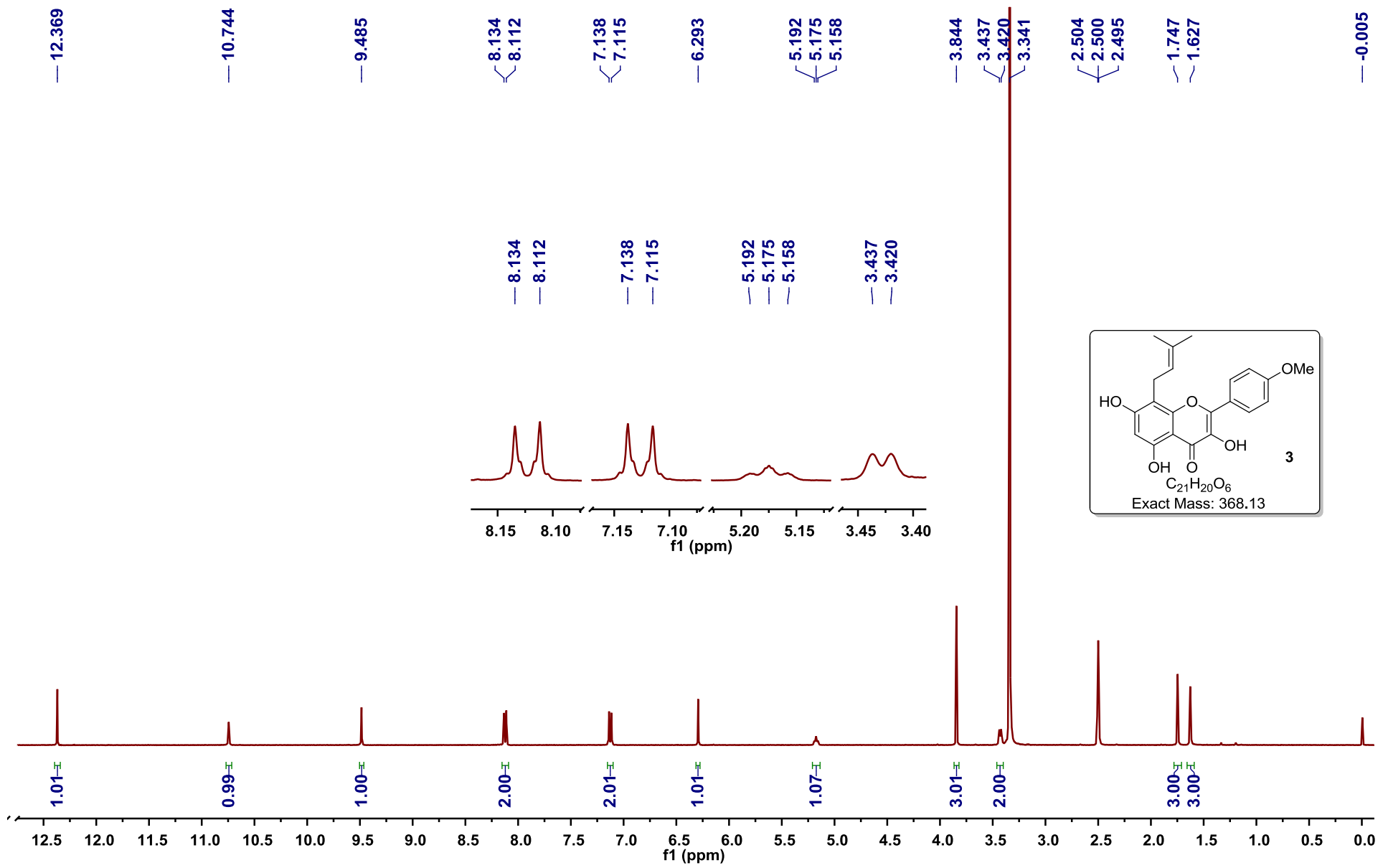
The yellow powder **13** (90 mg) was dissolved in 7.0 M methanolic ammonia (3 mL) and stirred at room temperature for 3 h. The solution was neutralized cautiously with 0.5 M HCl (aq) and left at room temperature for 12 h. The precipitates were collected by filtration and recrystallized from MeOH to furnish **2** as yellow needles. Yield: 58 mg (63%, for 2 steps); mp: 253–255 °C;  $[\alpha]_D^{20} = -6.0$  (c 0.1, CH<sub>3</sub>OH). IR (cm<sup>-1</sup>): 3393, 2918, 1651, 1597, 1557, 1512, 1312, 1260, 1183, 1098. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 12.43 (s, 1H, OH-5), 9.63 (s, 1H, OH-3), 8.14 (d, *J* = 9.0 Hz, 2H, H-2'/6'), 7.14 (d, *J* = 9.0 Hz, 2H, H-3'/5'), 6.60 (s, 1H, H-6), 5.36 (d, *J* = 4.6 Hz, 1H, OH-2'''), 5.20 (t, *J* = 6.9 Hz, 1H, H-2''), 5.14 (d, *J* = 3.9 Hz, 1H, OH-3'''), 5.08 (d, *J* = 5.3 Hz, 1H, OH-4'''), 5.00 (d, *J* = 7.1 Hz, 1H, H-1'''), 4.65 (t, *J* = 5.6 Hz, 1H, OH-6'''), 3.85 (s, 3H, OCH<sub>3</sub>-4'), 3.73 – 3.62 (m, 2H, H-1''b/6'''b), 3.50 – 3.44 (m, 3H, H-1''a/5'''/6'''a, overlapped with H<sub>2</sub>O), 3.31 – 3.27 (m, 2H, H-2'''/3'''), 3.20 – 3.16 (m, 1H, H-4'''), 1.76 (s, 3H, H-4'''), 1.62 (s, 3H, H-5'''). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 176.58 (C-4), 160.71 (C-4'), 160.19 (C-7), 158.63 (C-5), 152.79 (C-9), 146.99 (C-2), 136.27 (C-3), 131.25 (C-3''), 129.44 (C-2'/6'), 123.47 (C-1'), 122.38 (C-2''), 114.21 (C-3'/5'), 108.11 (C-8), 104.55 (C-10), 100.50 (C-1'''), 97.50 (C-6), 77.22 (C-5'''), 76.66 (C-3'''), 73.43 (C-2'''), 69.71 (C-4'''), 60.70 (C-6'''), 55.48 (OCH<sub>3</sub>-4'), 25.56 (C-5''), 21.50 (C-1''), 17.99 (C-4''). ESI-HRMS *m/z*: 531.1852 [M+H]<sup>+</sup> (calcd for C<sub>27</sub>H<sub>31</sub>O<sub>11</sub>: 531.1866). The <sup>1</sup>H and <sup>13</sup>C NMR data of icariside I (**2**) were in agreement with those reported [6].

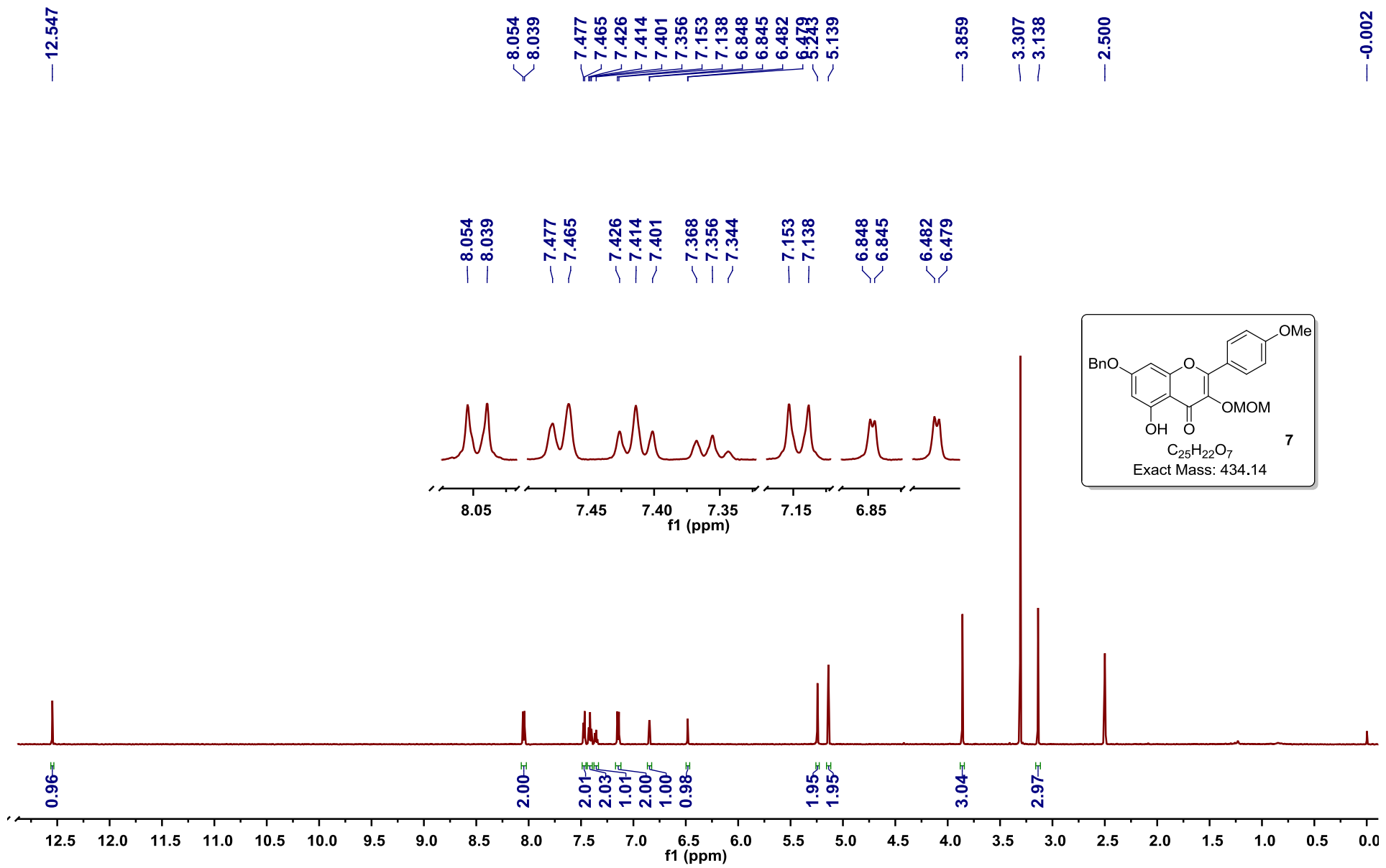
## References

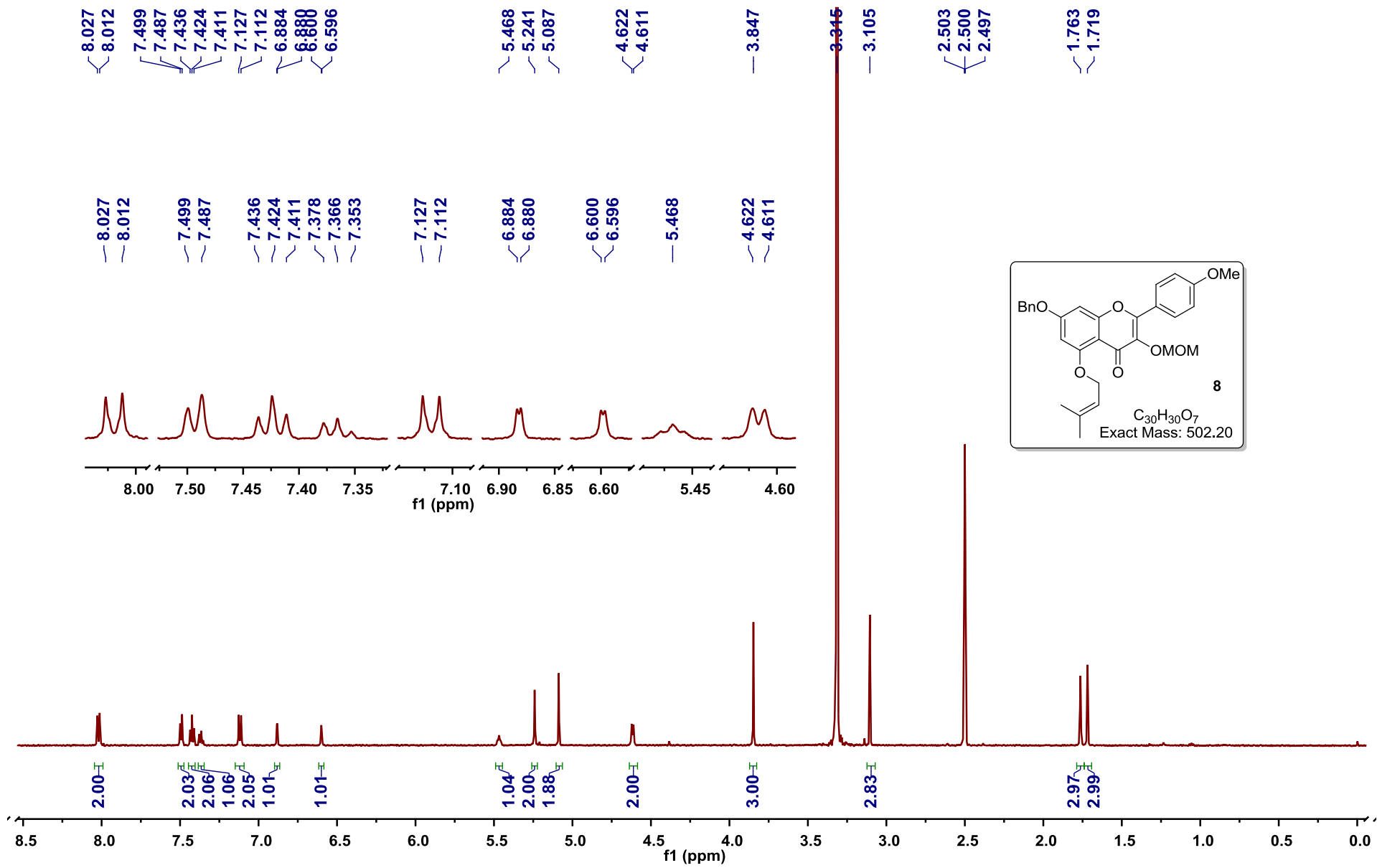
1. Mei, Q. G.; Wang, C.; Yuan, W. C.; Zhang, G. L. *Beilstein J. Org. Chem.* **2015**, *11*, 288–293.
2. Li, Y.-W.; Li, Y.-X.; Zhang, W.; Guan, H.-S. *Chin. J. Org. Chem.* **2004**, *24*, 438–439.
3. Nguyen, V.-S.; Shi, L.; Li, Y.; Wang, Q.-A. *Lett. Org. Chem.* **2014**, *11*, 677–681.
4. Dell’Agli, M.; Galli, G. V.; Dal Cero, E.; Belluti, F.; Matera, R.; Zironi, E.; Pagliuca, G.; Bosisio, E. *J. Nat. Prod.* **2008**, *71*, 1513–1517.
5. Liu, R.; Li, A.; Sun, A.; Cui, J.; Kong, L. *J. Chromatogr. A* **2005**, *1064*, 53–57.
6. Han, B.; Shen, T.; Liu, D.; Yang, J. *Chin. Pharm. J.* **2002**, *37*, 740–742.



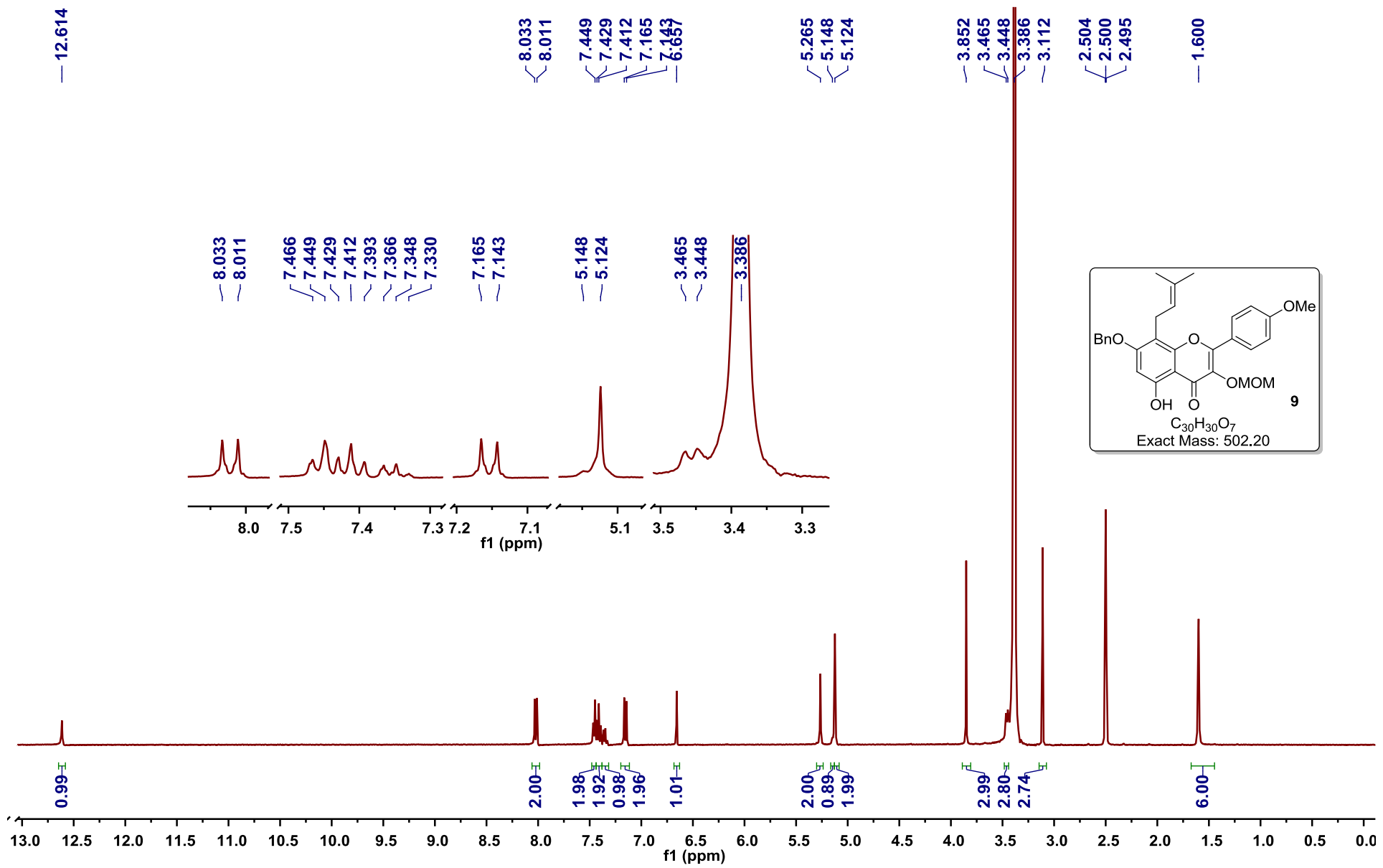


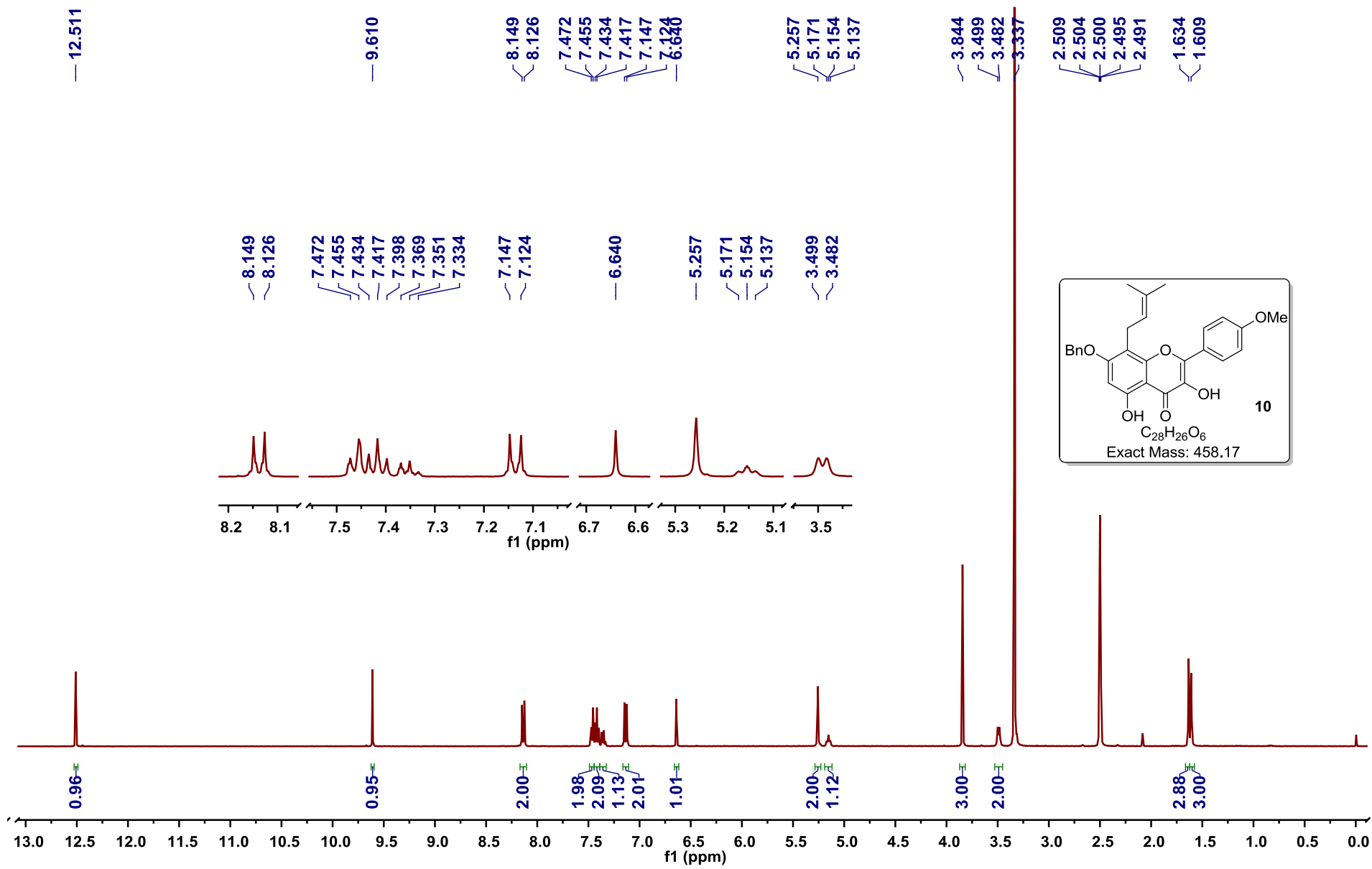


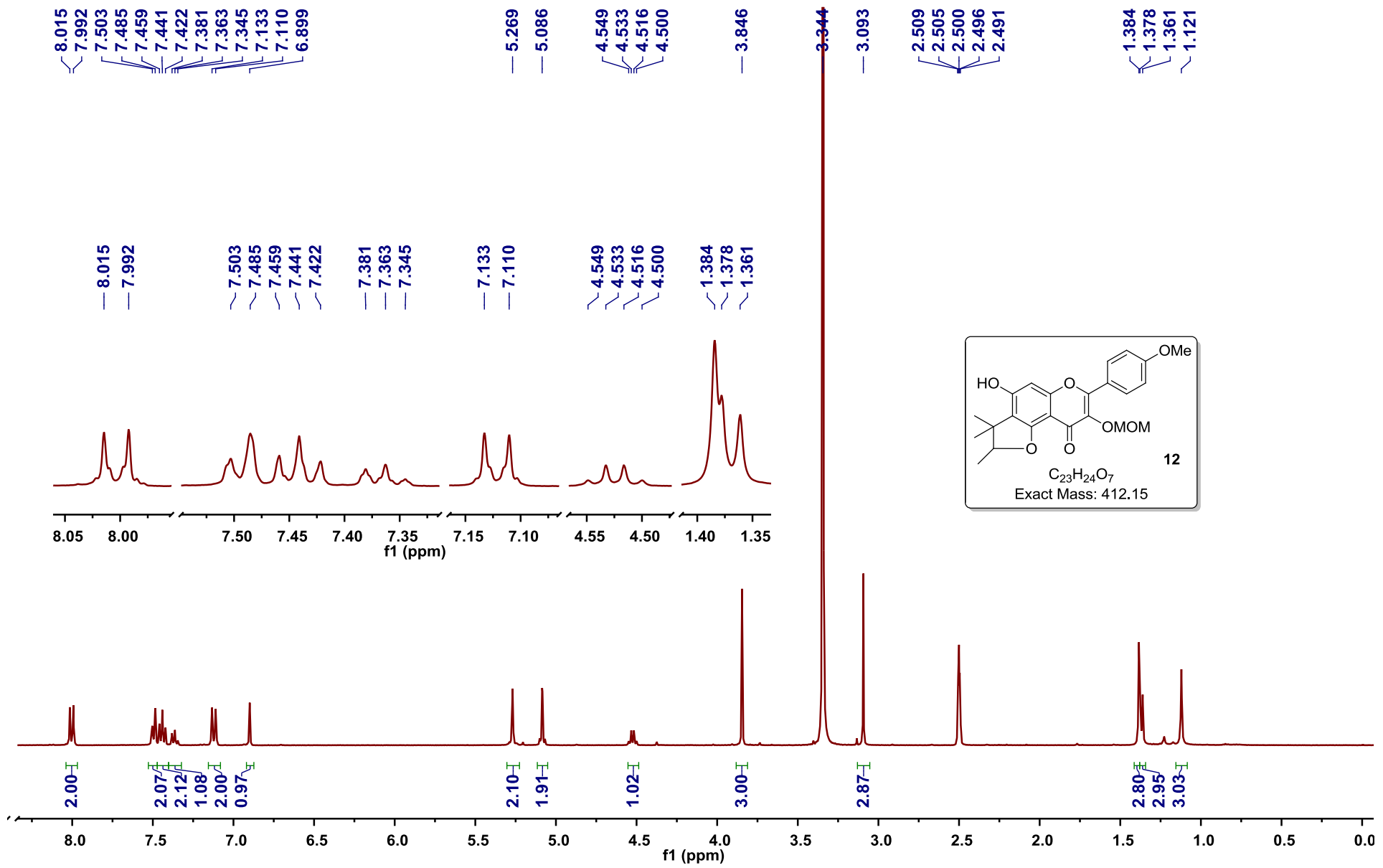


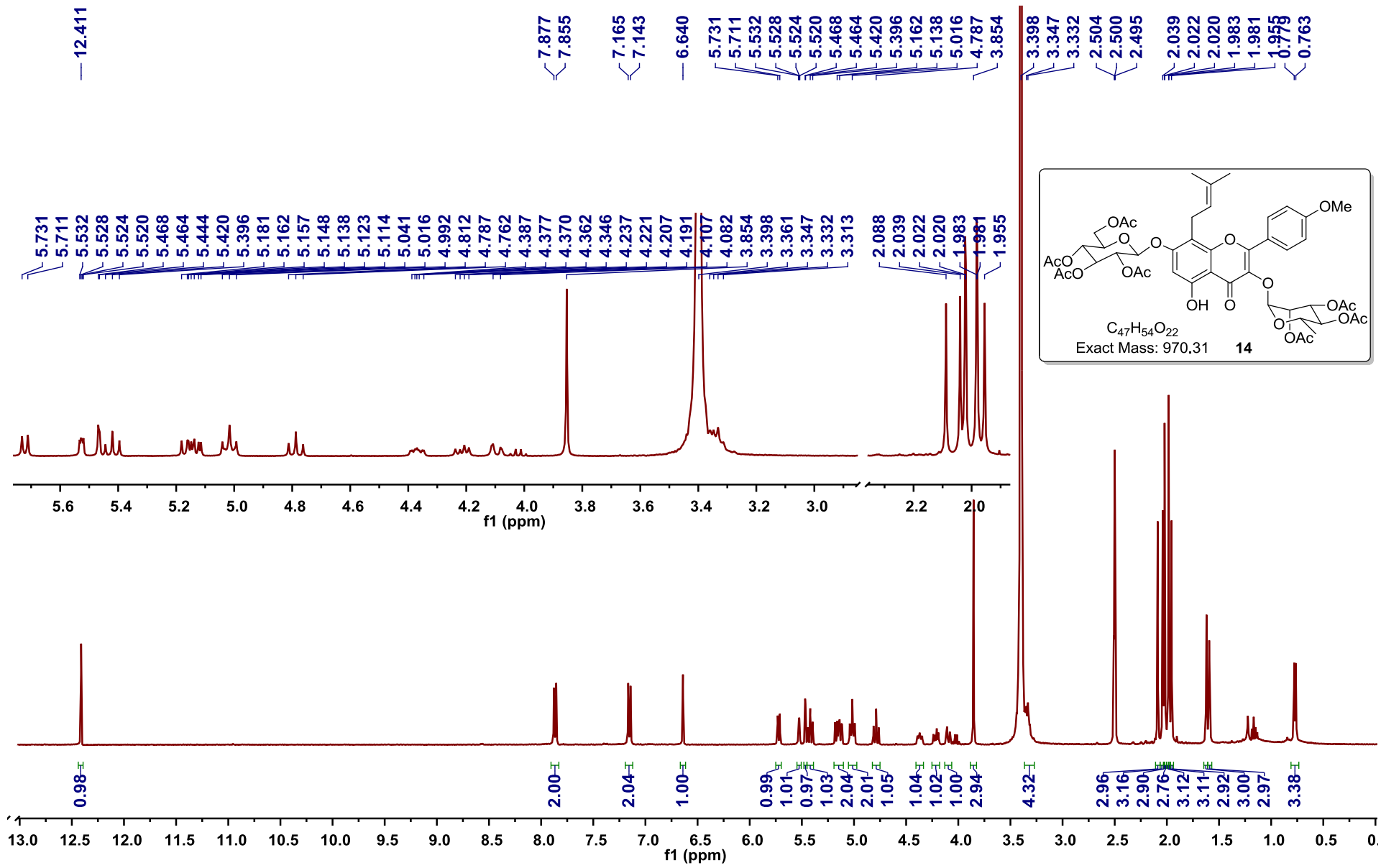












— 178.395

161.522  
160.603  
159.183  
157.486  
153.117

134.721  
131.296  
130.708

122.342  
122.207

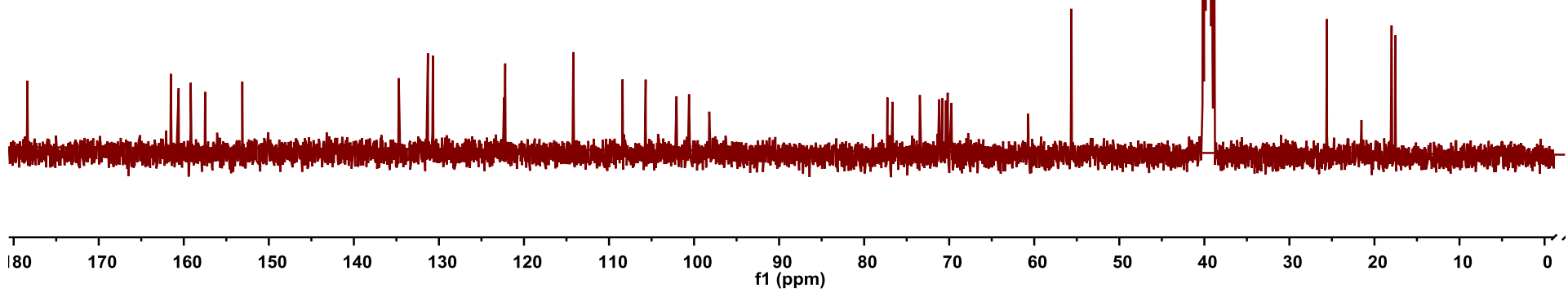
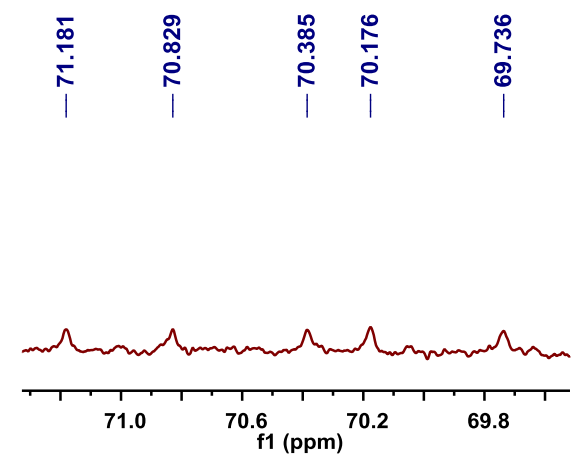
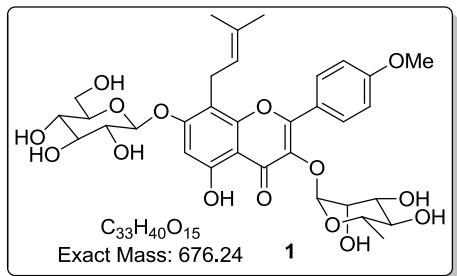
— 114.200

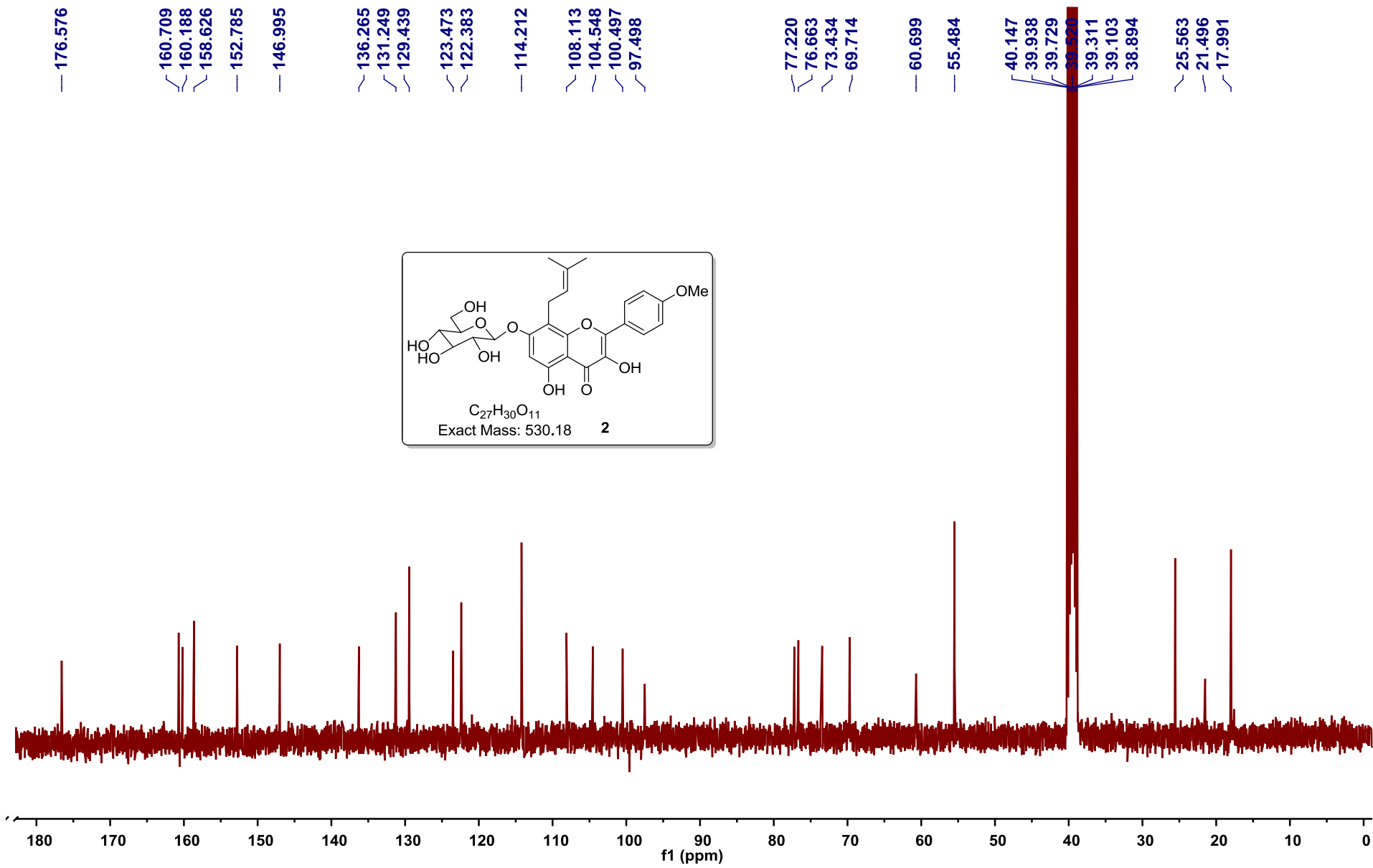
108.406  
105.689  
102.062  
100.594  
98.212

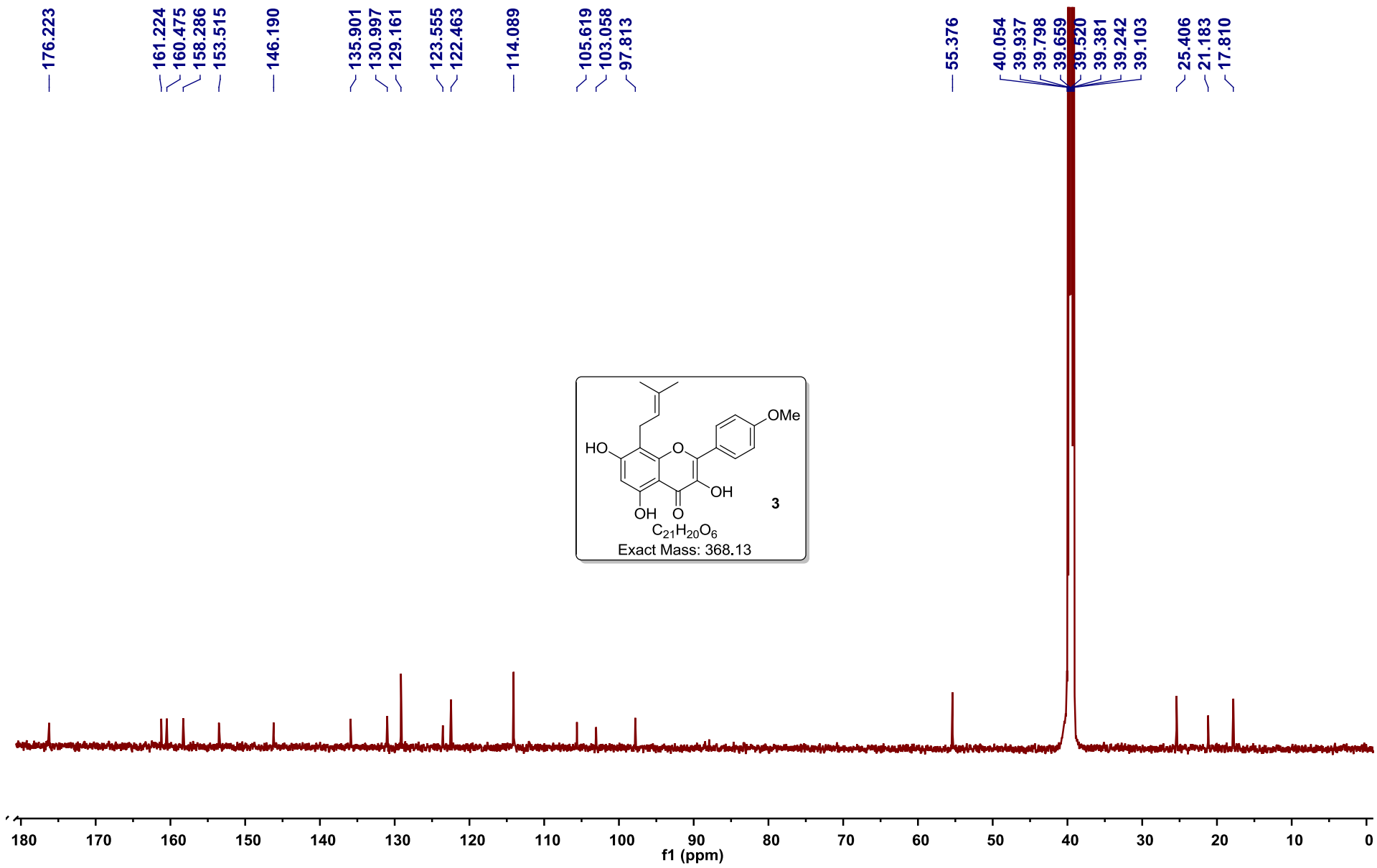
77.269  
76.675  
73.451  
71.181  
70.829  
70.385  
70.176  
69.736  
— 60.709  
— 55.625

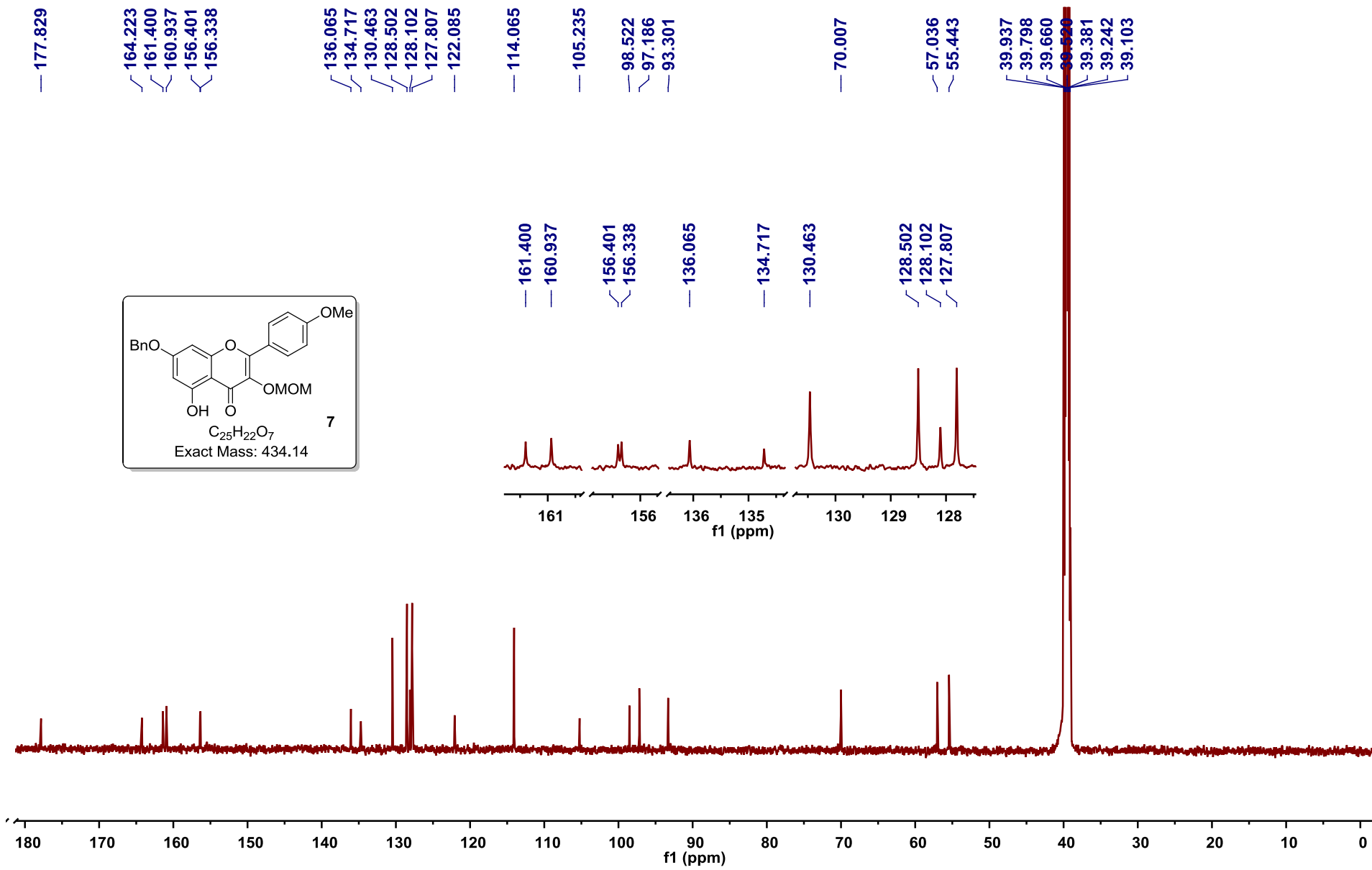
40.146  
39.938  
39.729  
39.520  
39.311  
39.103  
38.894

— 25.594  
21.526  
17.980  
17.562

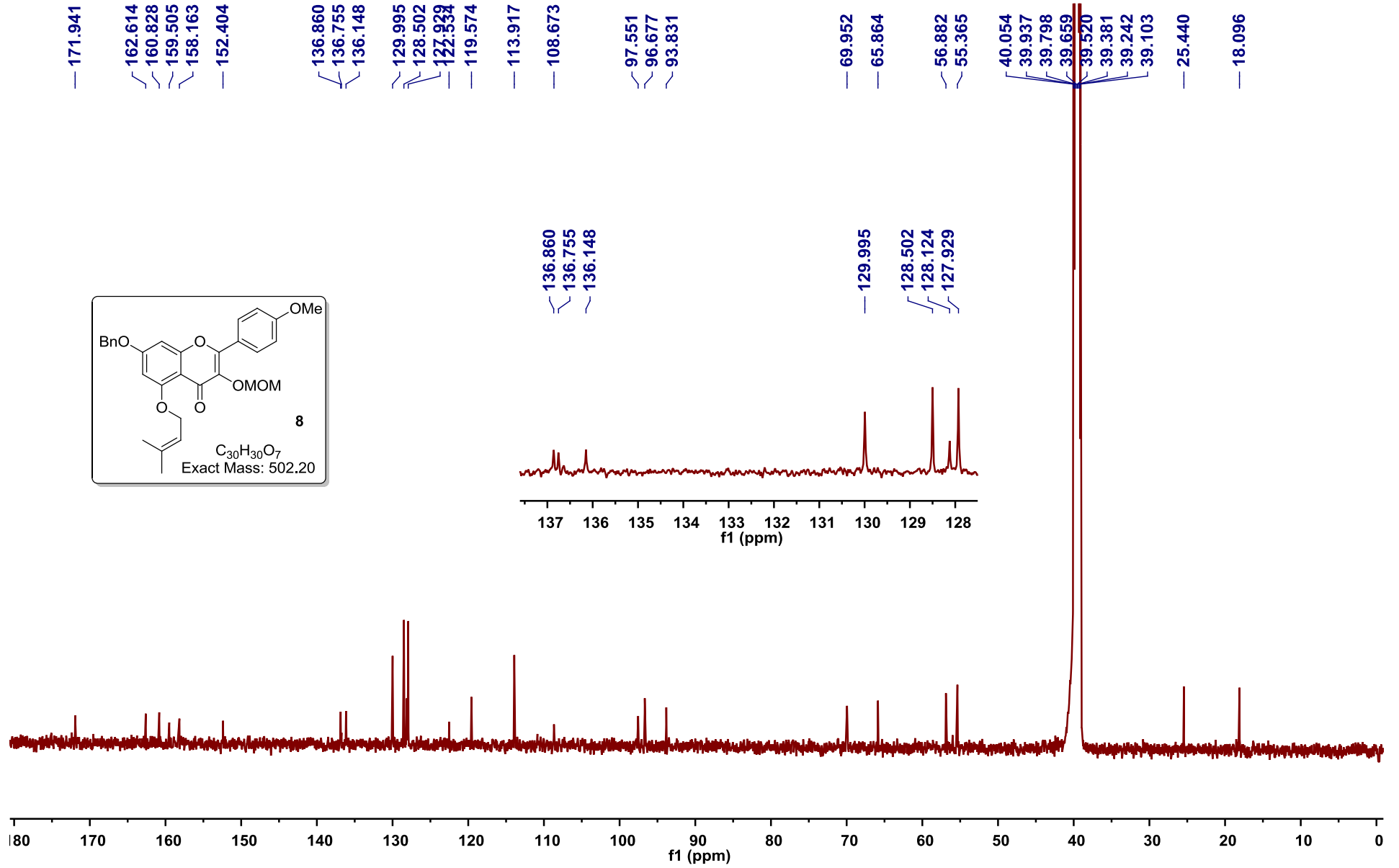
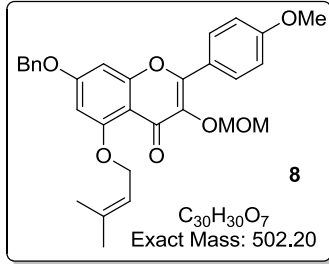


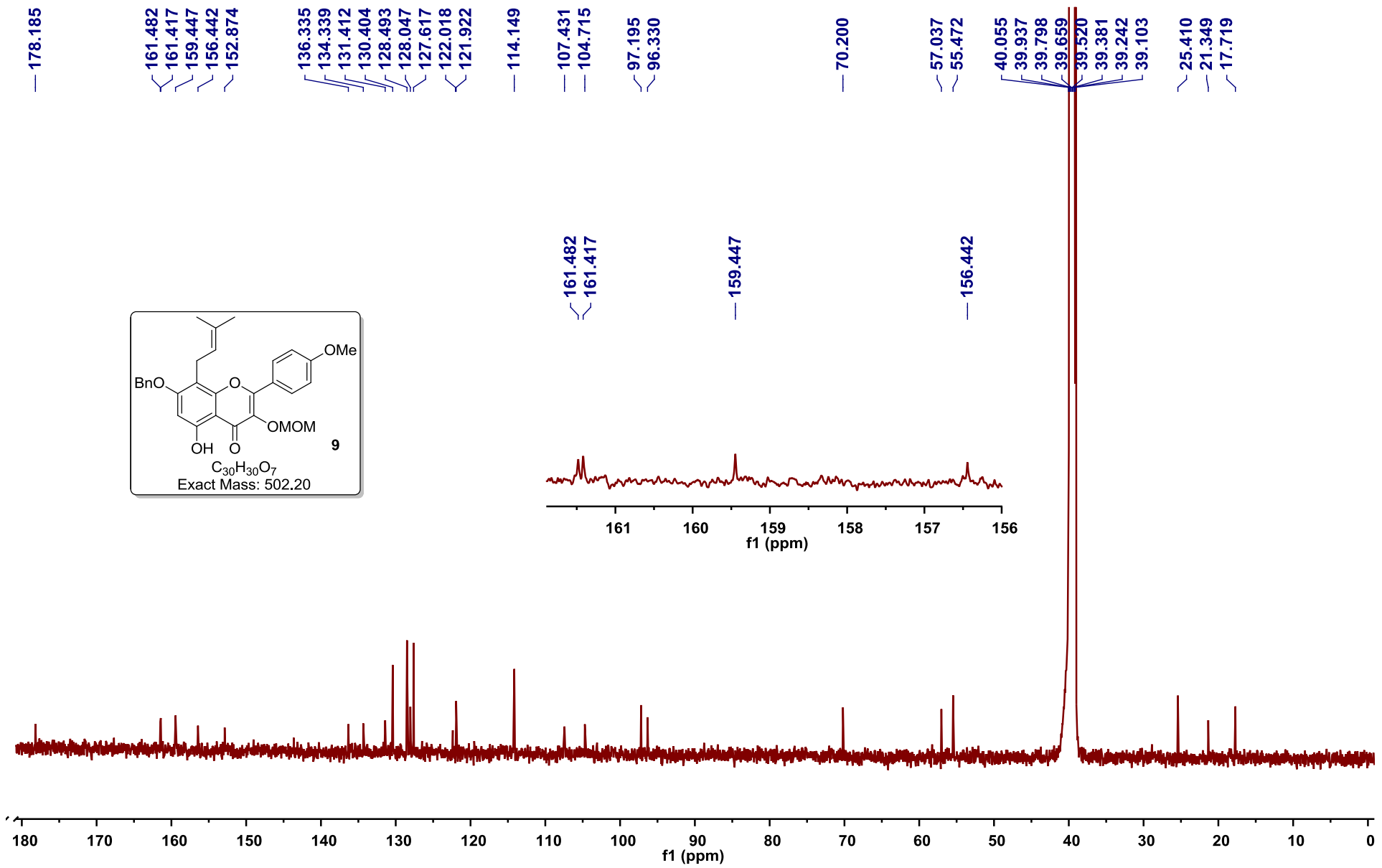


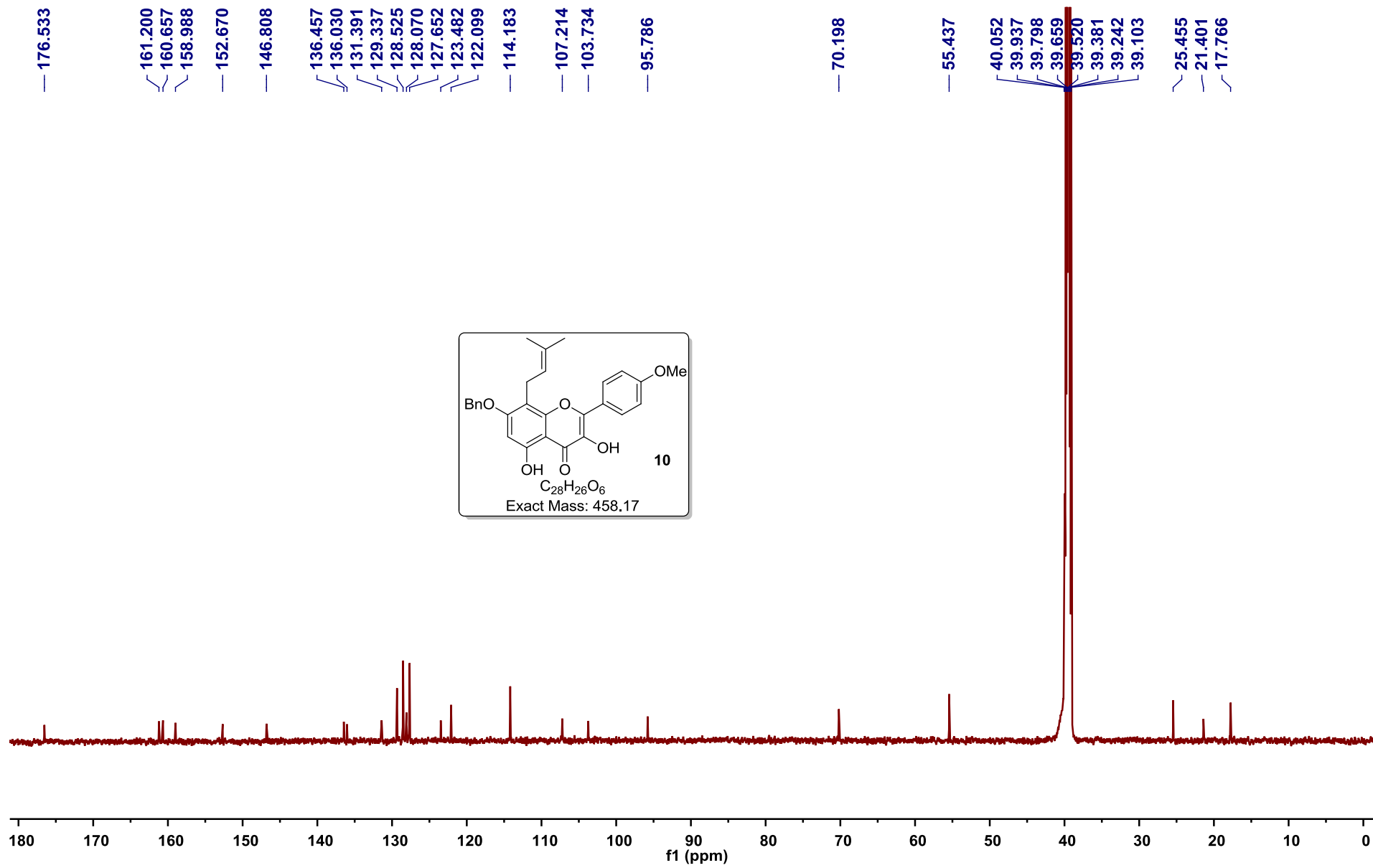


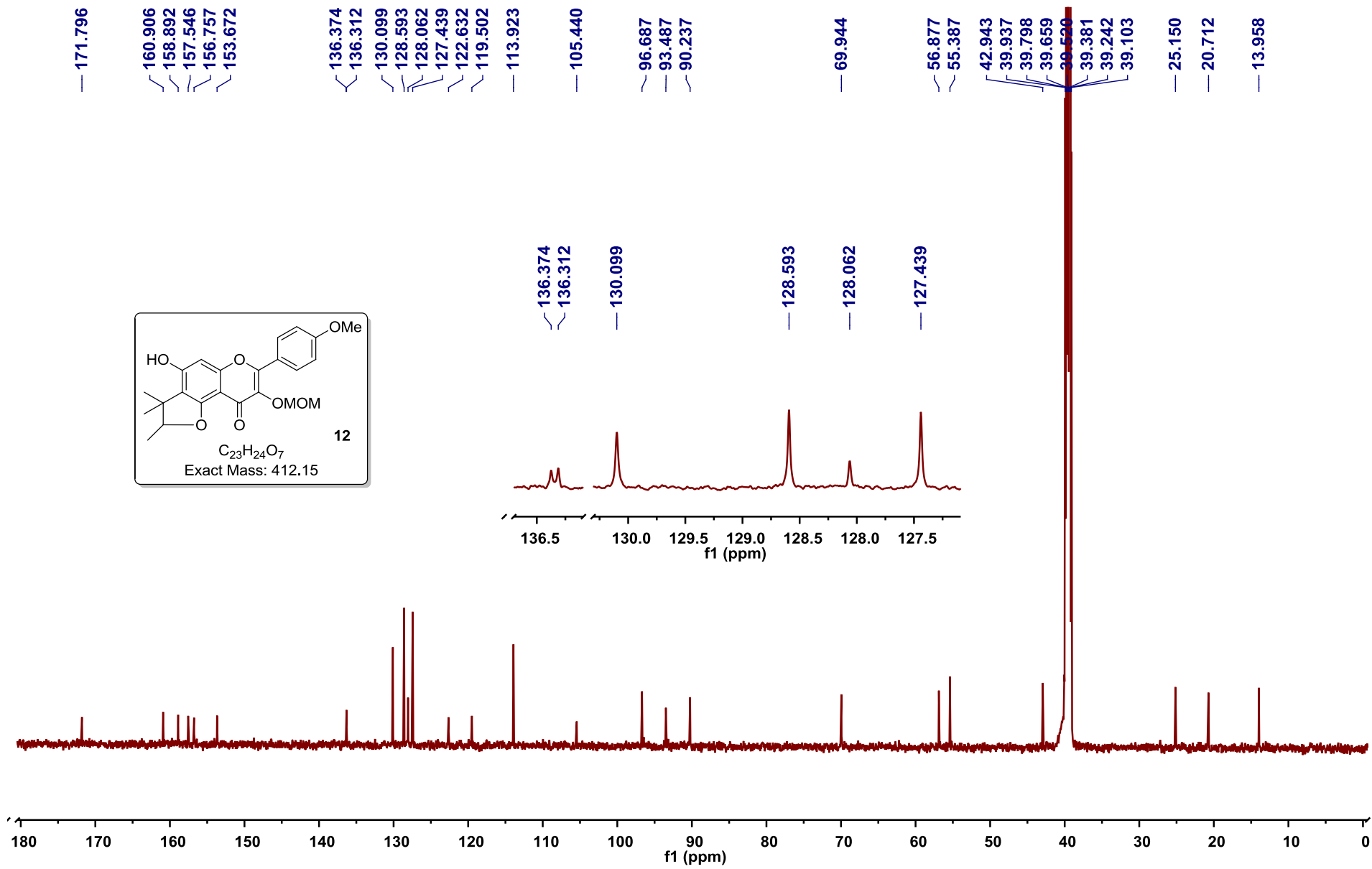


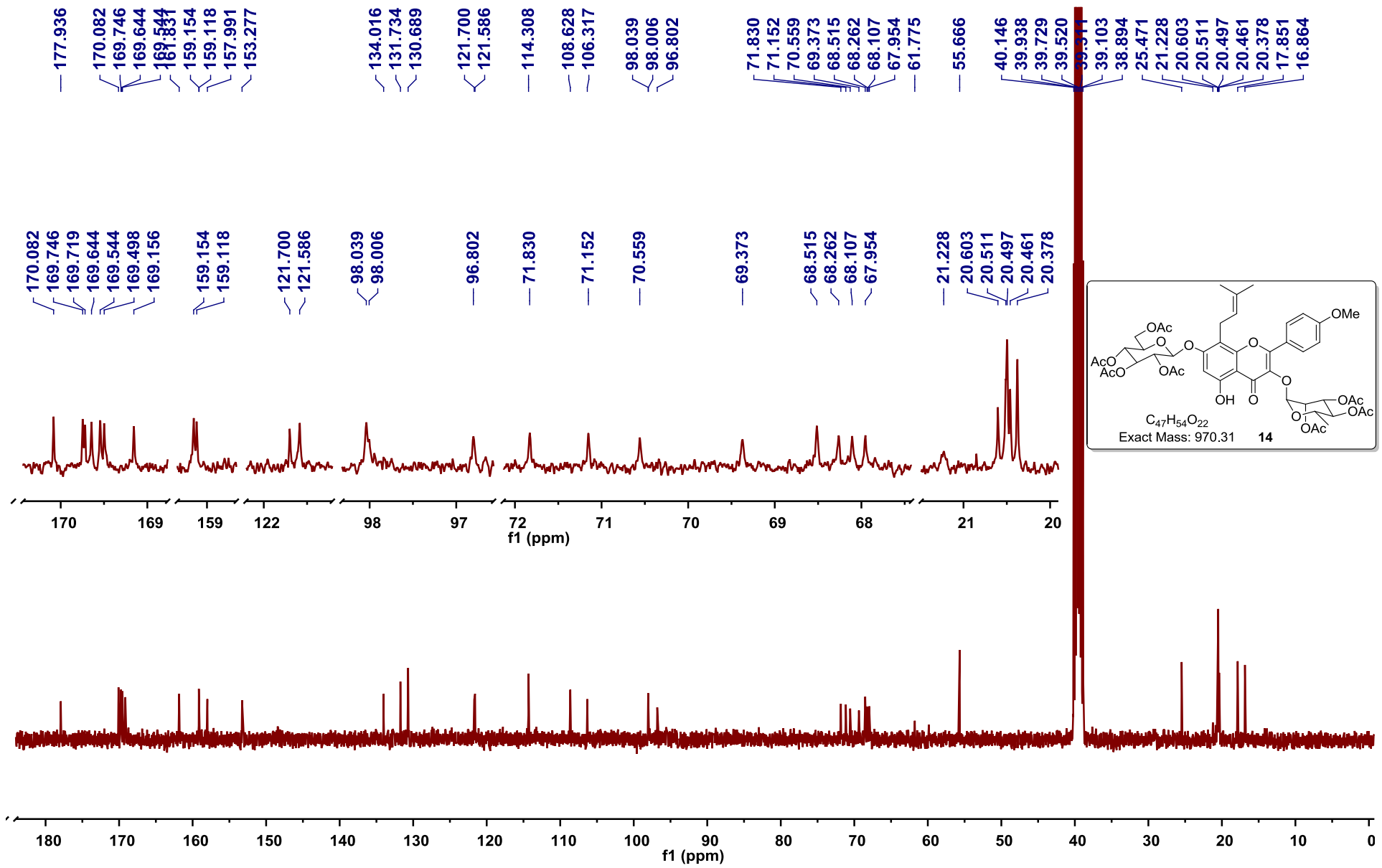












# Mass Spectrum SmartFormula Report

**Analysis Info**

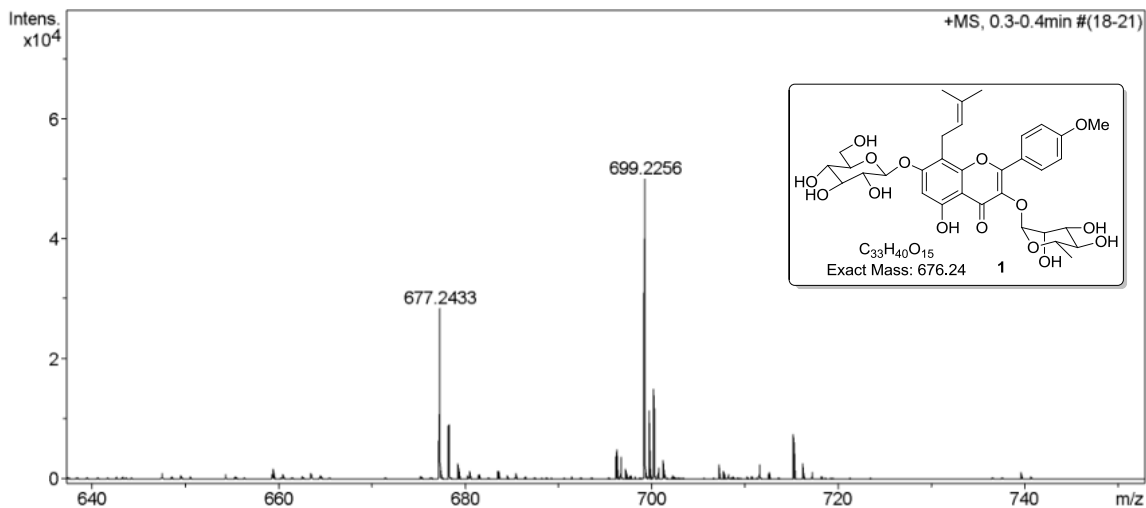
Analysis Name D:\Data\USER-2015\m4.d  
 Method lc-ms-ljr-20111104.m  
 Sample Name m4  
 Comment

Acquisition Date 1/21/2015 3:40:00 PM

Operator Ma  
 Instrument / Ser# microTOF-Q II 10203

**Acquisition Parameter**

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1500 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
677.2433	1	C <sub>33</sub> H <sub>41</sub> O <sub>15</sub>	100.00	677.2440	1.0	1.3	25.7	13.5	even	ok
699.2256	1	C <sub>33</sub> H <sub>40</sub> NaO <sub>15</sub>	100.00	699.2259	0.5	0.8	38.1	13.5	even	ok

# Elemental Composition Report

## Single Mass Analysis

Sample Name: mei 530

Tolerance = 5.0 PPM

RDB: min = -10.0, max = 120.0

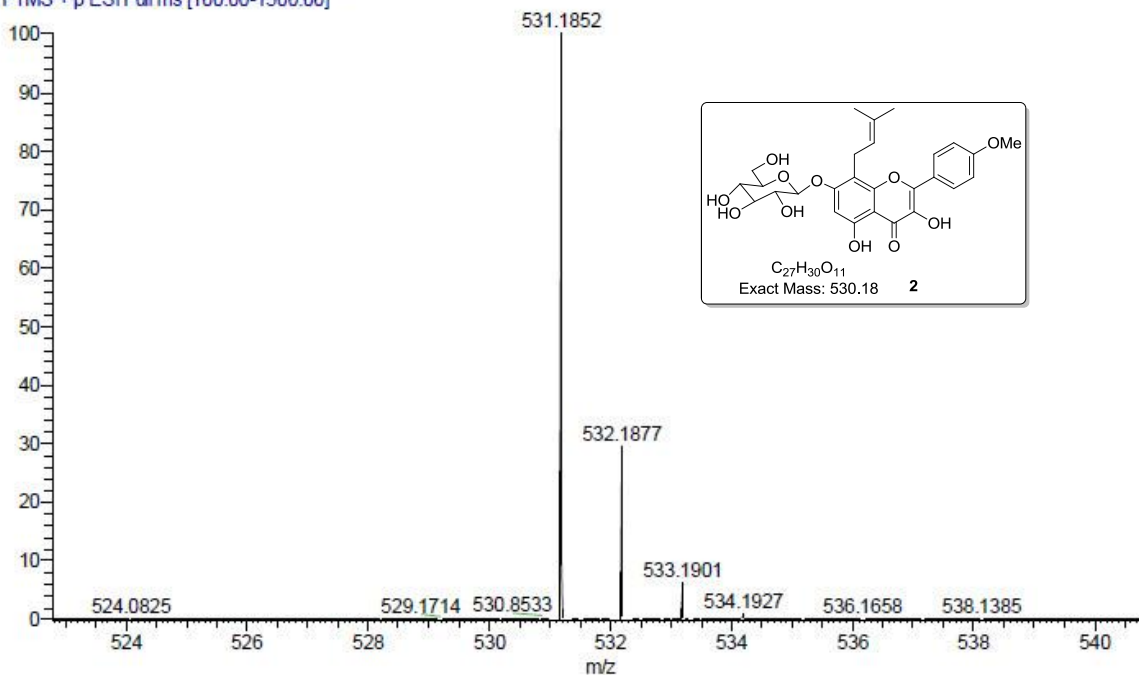
Selected filters: None

## Monoisotopic Mass, Odd and Even Electron Ions

Elements Used:

C: 0-40 H: 0-100 O: 0-20 N: 0-10

MEI530 #391 RT: 3.92 AV: 1 NL: 6.07E8  
T: FTMS + p ESI Full ms [100.00-1500.00]



Measured Mass: 531.1852

Idx	Formula	RDB	Delta ppm
1	C27 H30 O11	12.5	-1.170

# Mass Spectrum SmartFormula Report

## Analysis Info

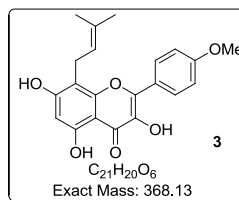
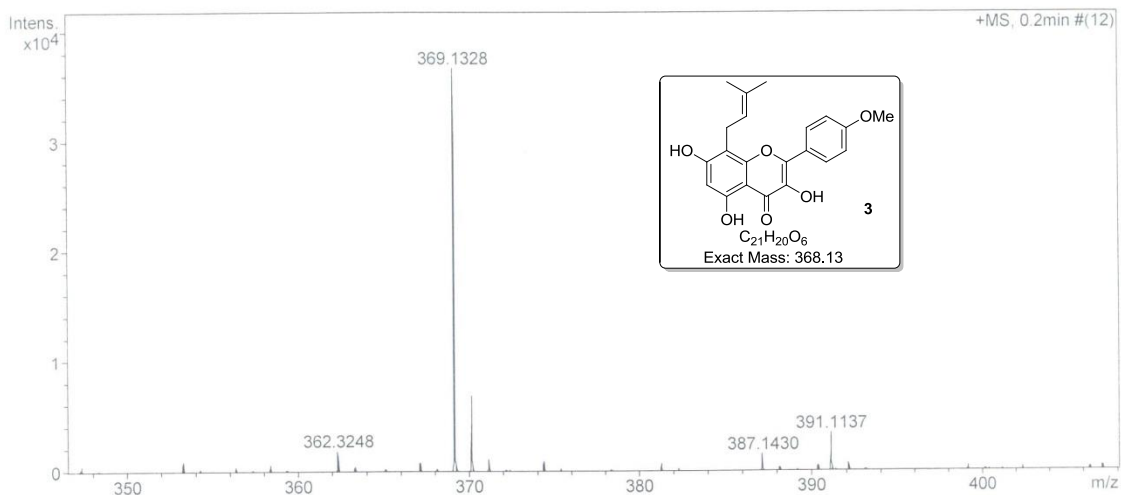
Analysis Name D:\Data\USER-2013\mei368.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name mei368  
 Comment

Acquisition Date 10/15/2013 11:43:16 AM

Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	100 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-R rule
369.1328	1	C <sub>21</sub> H <sub>21</sub> O <sub>6</sub>	100.00	369.1333	1.4	1.7	31.4	11.5	even	ok



# Mass Spectrum SmartFormula Report

## Analysis Info

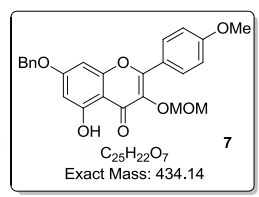
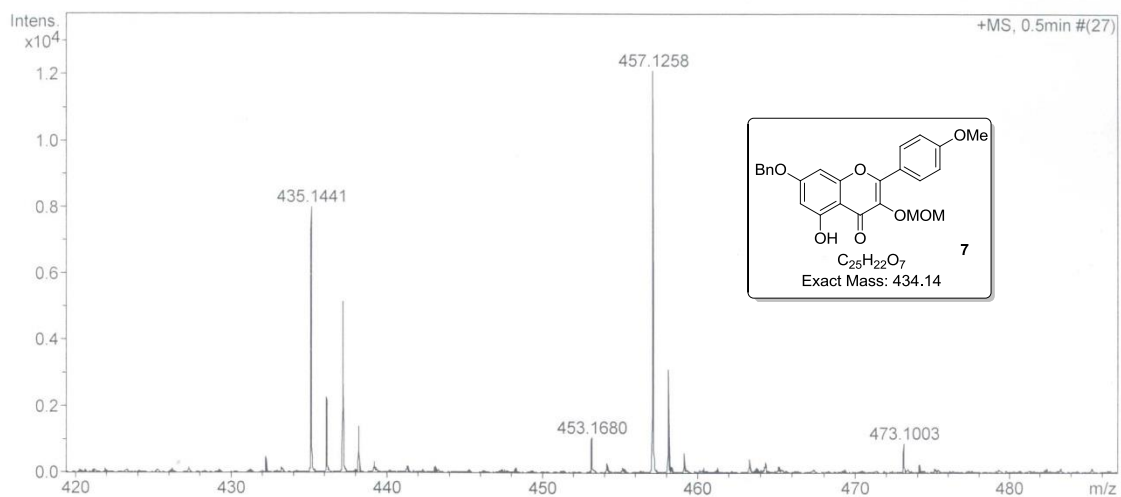
Analysis Name D:\Data\USER-2013\W-434.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name M-434  
 Comment

Acquisition Date 5/14/2013 4:04:15 PM

Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
457.1258	1	C <sub>25</sub> H <sub>22</sub> NaO <sub>7</sub>	100.00	457.1258	-0.1	0.1	7.0	14.5	even	ok

# Mass Spectrum SmartFormula Report

## Analysis Info

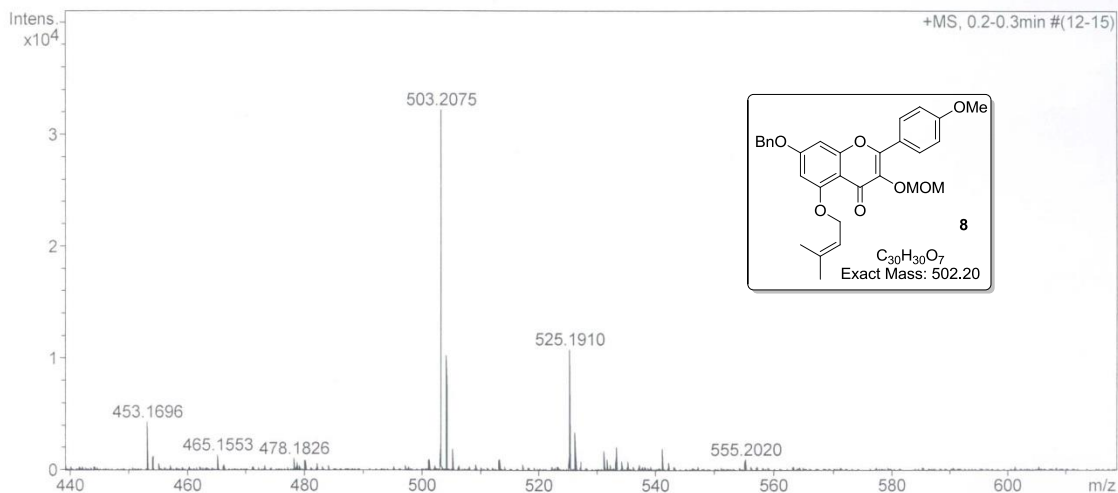
Analysis Name D:\Data\USER-2013\MEI502W.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name MEI502W  
 Comment

Acquisition Date 5/28/2013 4:33:47 PM

Operator Ma  
 Instrument / Ser# microTOF-Q II 10203

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSi gma	rdb	e <sup>-</sup> Conf	N-R uler
503.2075	1	C <sub>30</sub> H <sub>30</sub> O <sub>7</sub>	100.00	503.2064	-2.1	-2.9	6.8	15.5	even	ok

# Mass Spectrum SmartFormula Report

## Analysis Info

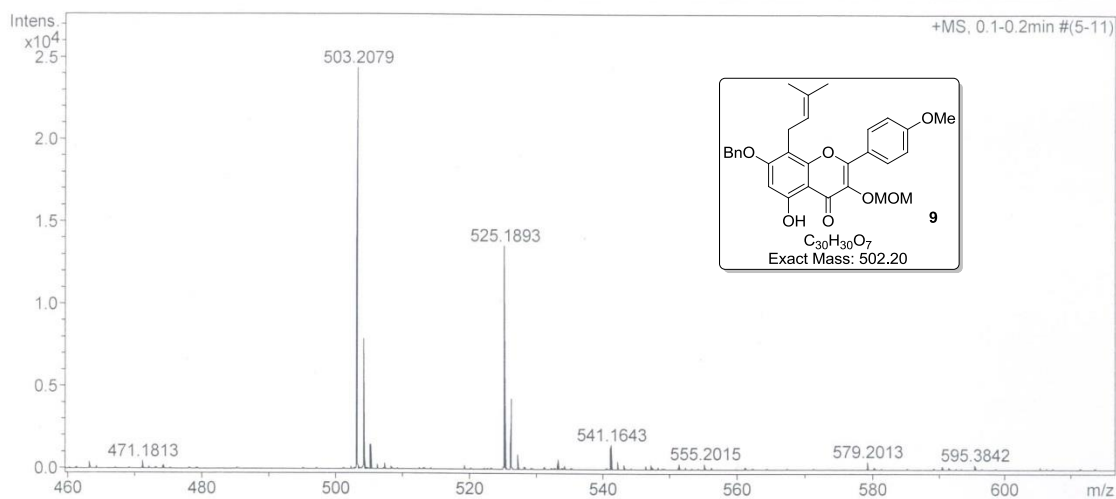
Analysis Name D:\Data\USER-2013\MEI5022.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name MEI5022  
 Comment

Acquisition Date 7/24/2013 5:40:20 PM

Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mea n err [ppm]	mSi gma	rdb	e <sup>-</sup> Conf	N-Rul e
503.2079	1	C <sub>30</sub> H <sub>31</sub> O <sub>7</sub>	100.00	503.2064	-3.0	-2.7	3.3	15.5	even	ok
525.1893	1	C <sub>30</sub> H <sub>30</sub> NaO <sub>7</sub>	100.00	525.1884	-1.7	-1.6	6.2	15.5	even	ok

# Mass Spectrum SmartFormula Report

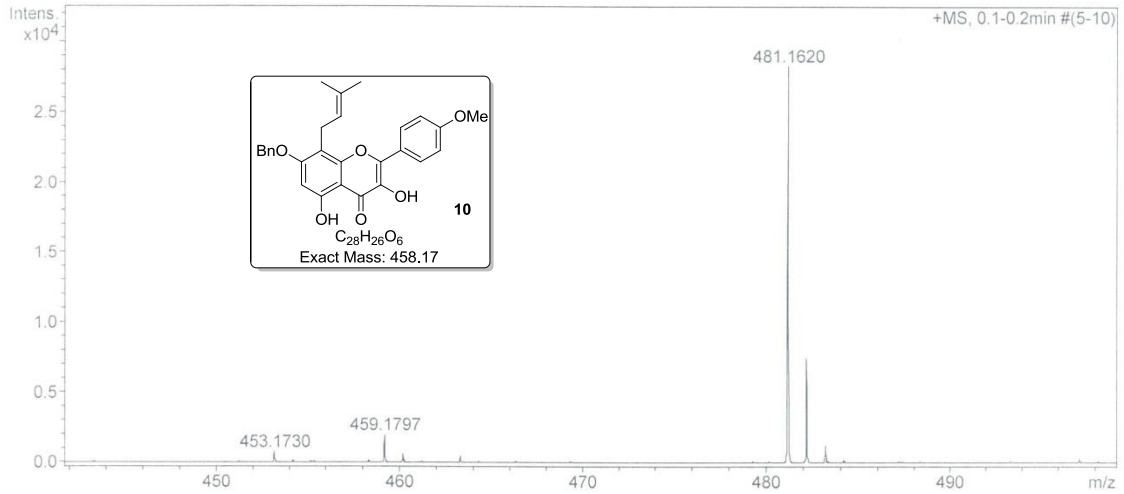
**Analysis Info**

Analysis Name D:\Data\USER-2013\mei458.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name mei458  
 Comment

Acquisition Date 12/17/2013 6:34:30 PM  
 Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

**Acquisition Parameter**

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Meas. n err [ppm]	mSig ma	rdb	e <sup>-</sup> Conf	N-Rule
481.1620	1	C <sub>28</sub> H <sub>26</sub> NaO <sub>6</sub>	100.00	481.1622	0.4	0.6	20.7	15.5	even	ok

# Mass Spectrum SmartFormula Report

**Analysis Info**

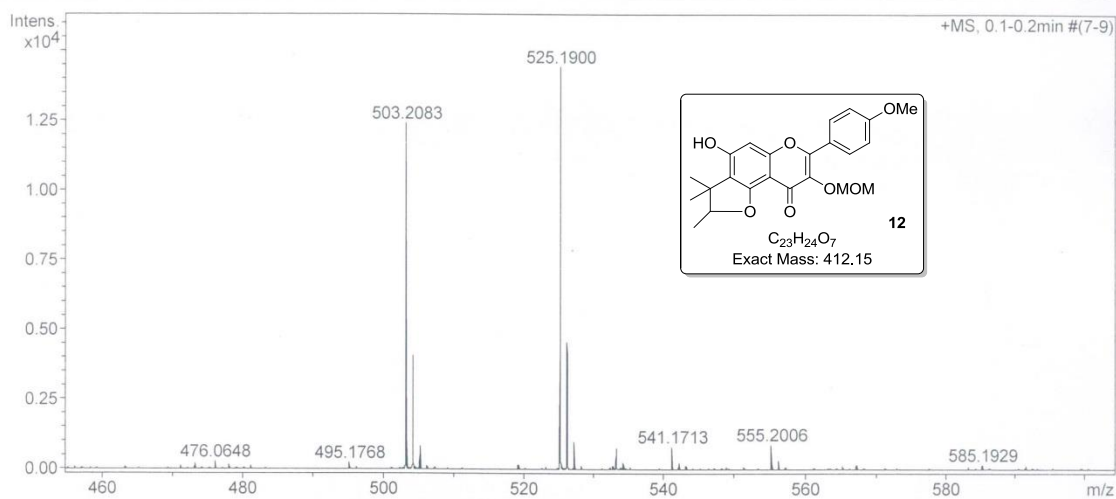
Analysis Name D:\Data\USER-2013\MEI5021.d  
 Method WU\_tune\_low\_20121222.m  
 Sample Name MEI5021  
 Comment

Acquisition Date 7/24/2013 5:38:11 PM

Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

**Acquisition Parameter**

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSi gma	rdb	e <sup>-</sup> Conf	N-Rule
503.2083	1	C <sub>30</sub> H <sub>31</sub> O <sub>7</sub>	100.00	503.2064	-3.7	-3.3	0.9	15.5	even	ok
525.1900	1	C <sub>30</sub> H <sub>30</sub> NaO <sub>7</sub>	100.00	525.1884	-3.0	-2.6	4.0	15.5	even	ok

# Mass Spectrum SmartFormula Report

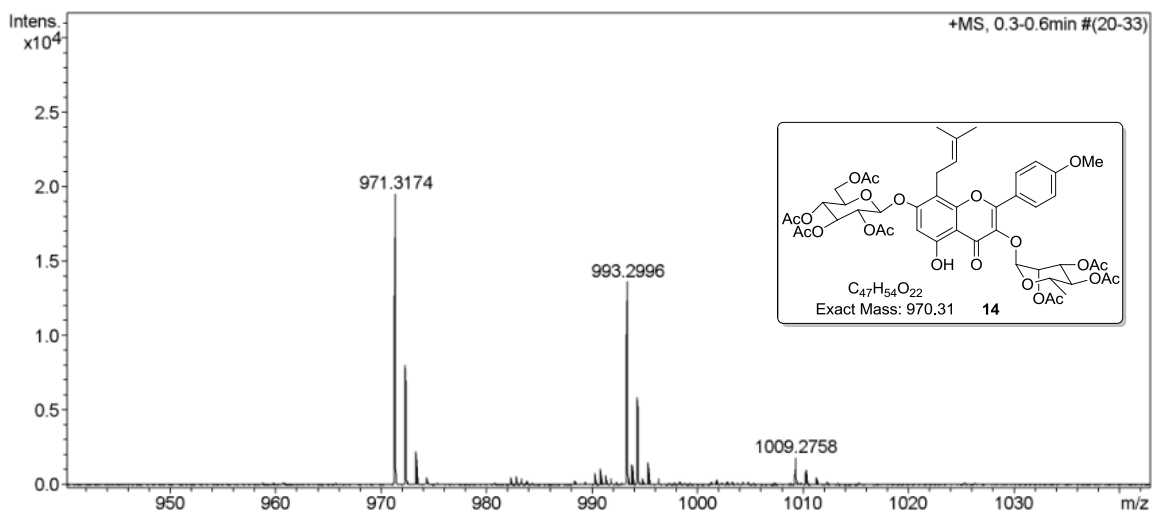
**Analysis Info**

Analysis Name D:\Data\USER-2015\m970.d  
 Method tune\_wide.m  
 Sample Name m970  
 Comment

Acquisition Date 2/4/2015 4:27:19 PM  
 Operator Ma  
 Instrument / Ser# micrOTOF-Q II 10203

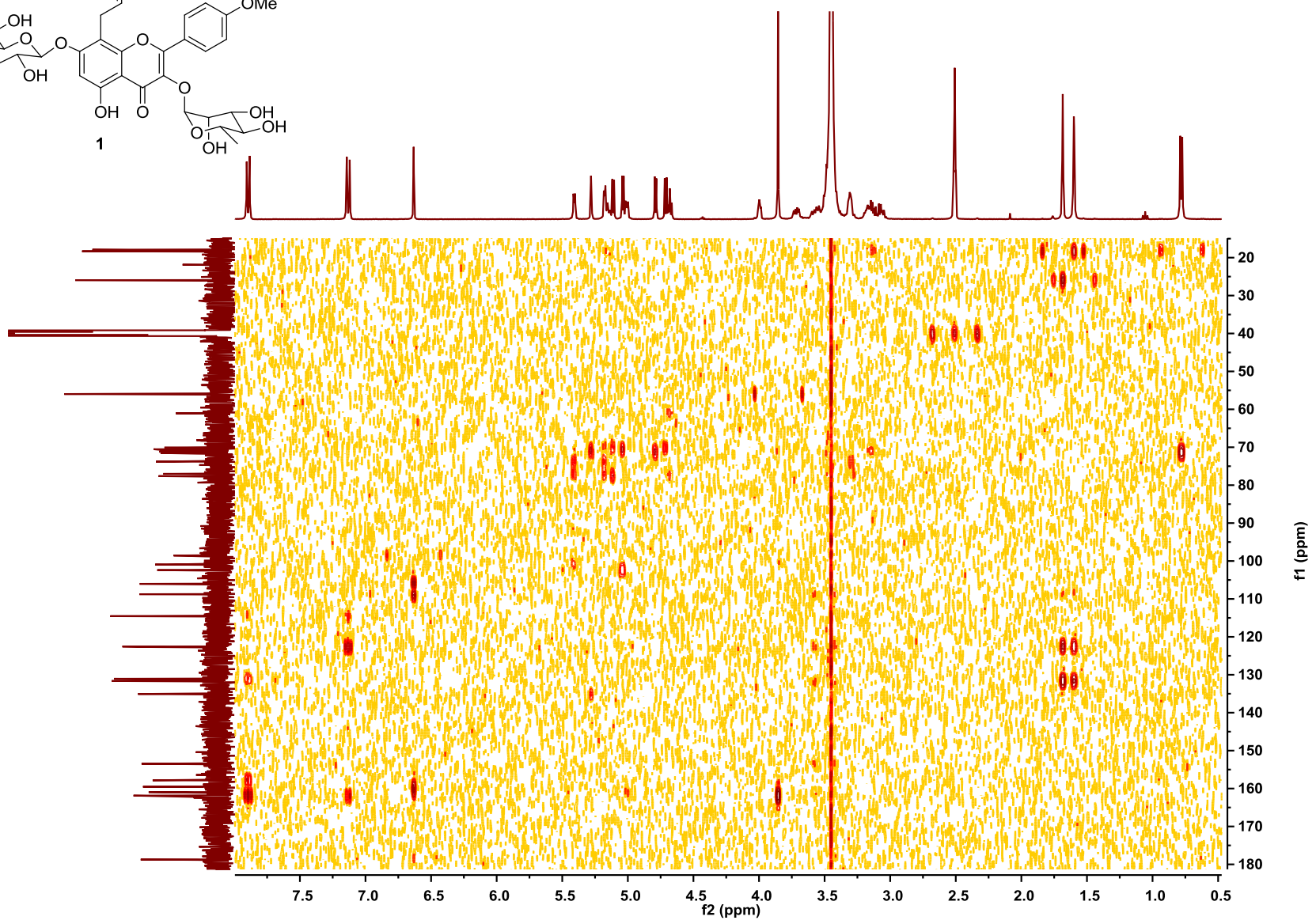
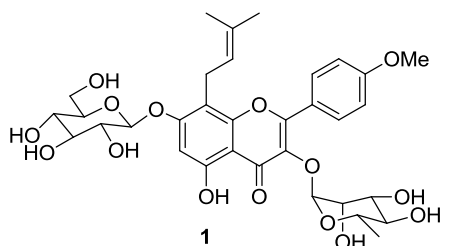
**Acquisition Parameter**

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	650.0 Vpp	Set Divert Valve	Source

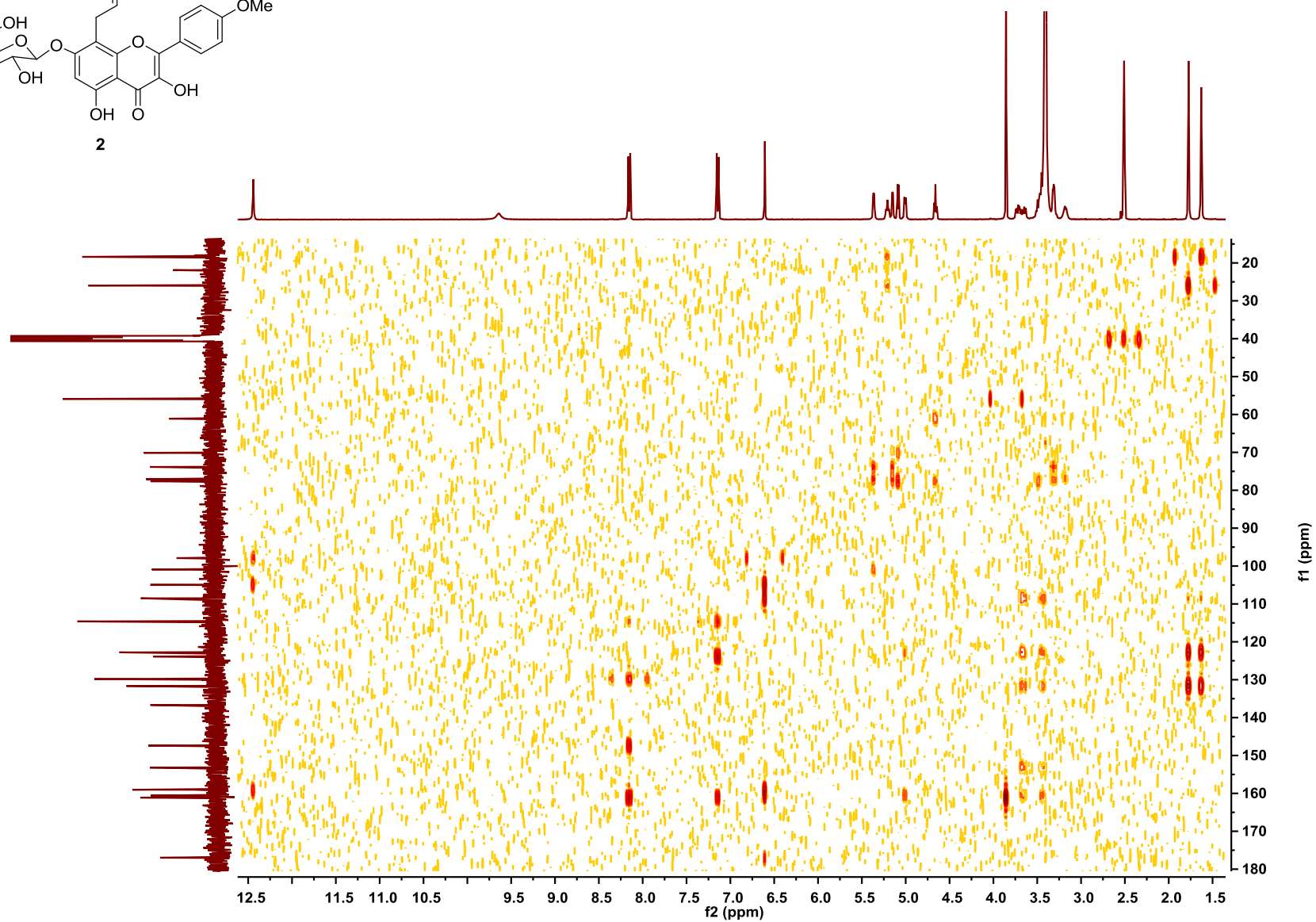
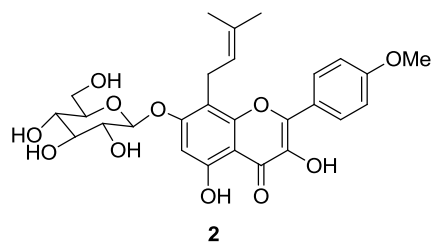


Meas. m/z	#	Formula	Score	m/z	err [ppm]	Mea n err [ppm]	mSig ma	rdb	e <sup>-</sup> Conf	N-Rule
971.3174	1	C 47 H 55 O 22	100.00	971.3179	0.6	0.1	59.6	20.5	even	ok
993.2996	1	C 47 H 54 Na O 22	100.00	993.2999	0.3	-0.4	52.6	20.5	even	ok
1009.2758	1	C 47 H 54 K O 22	100.00	1009.2738	-2.0	-1.4	38.5	20.5	even	ok

# HMBC Spectrum of Compound 1

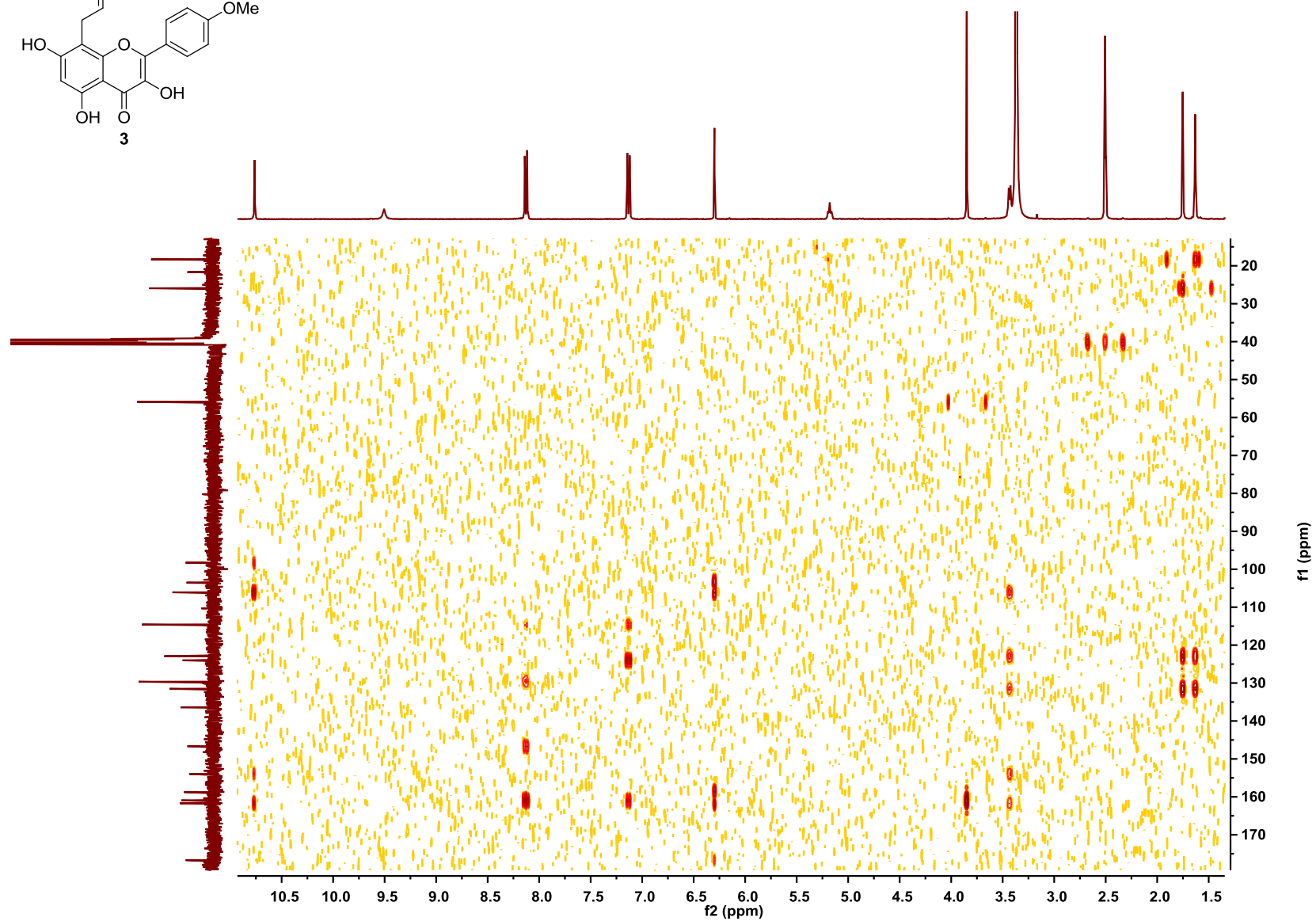
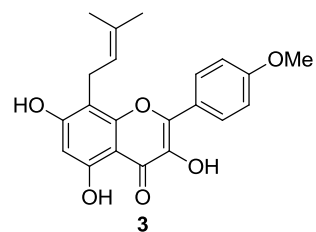


# HMBC Spectrum of Compound 2

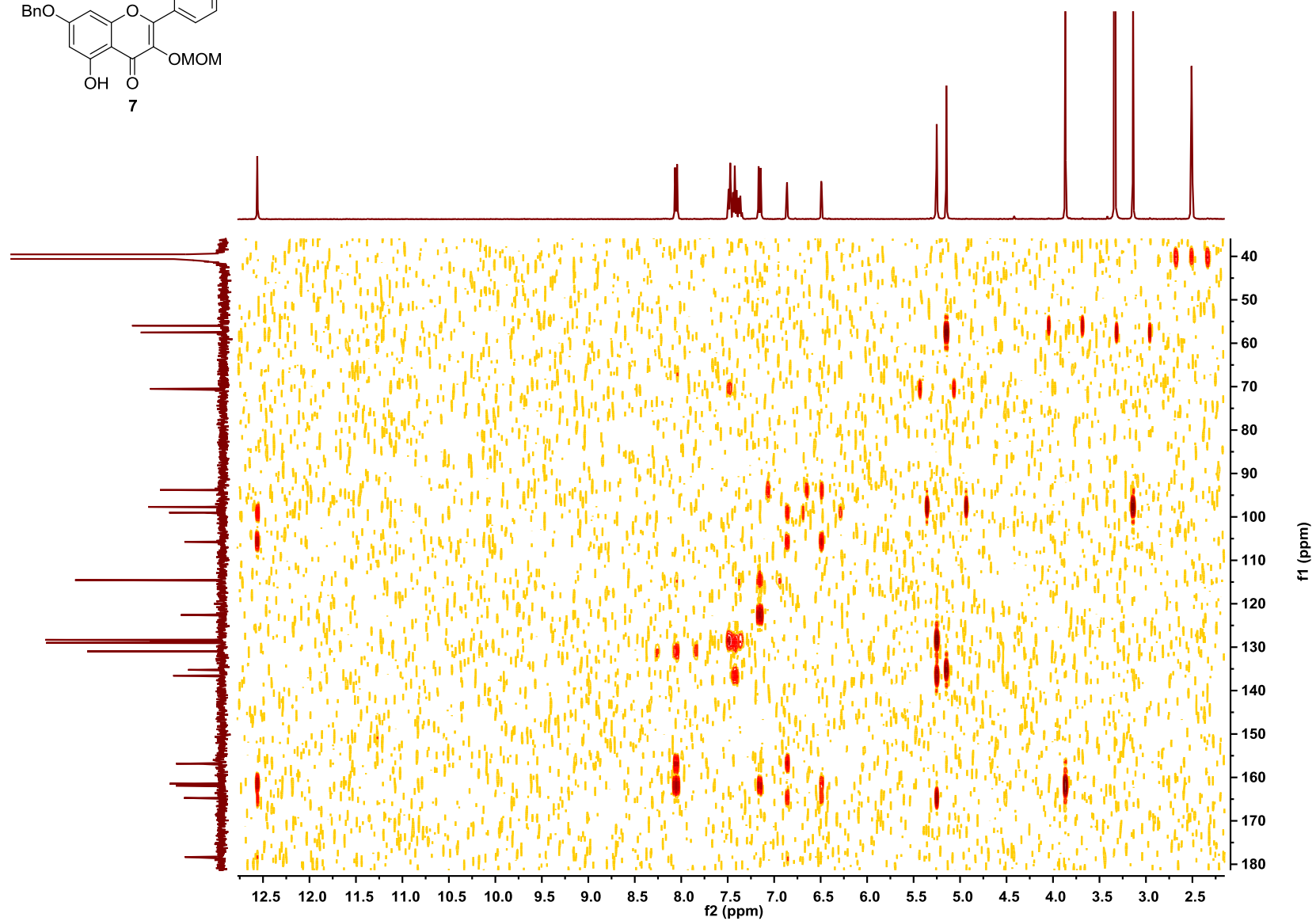
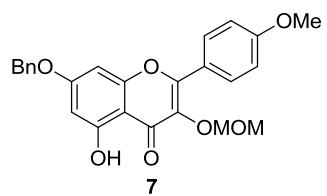




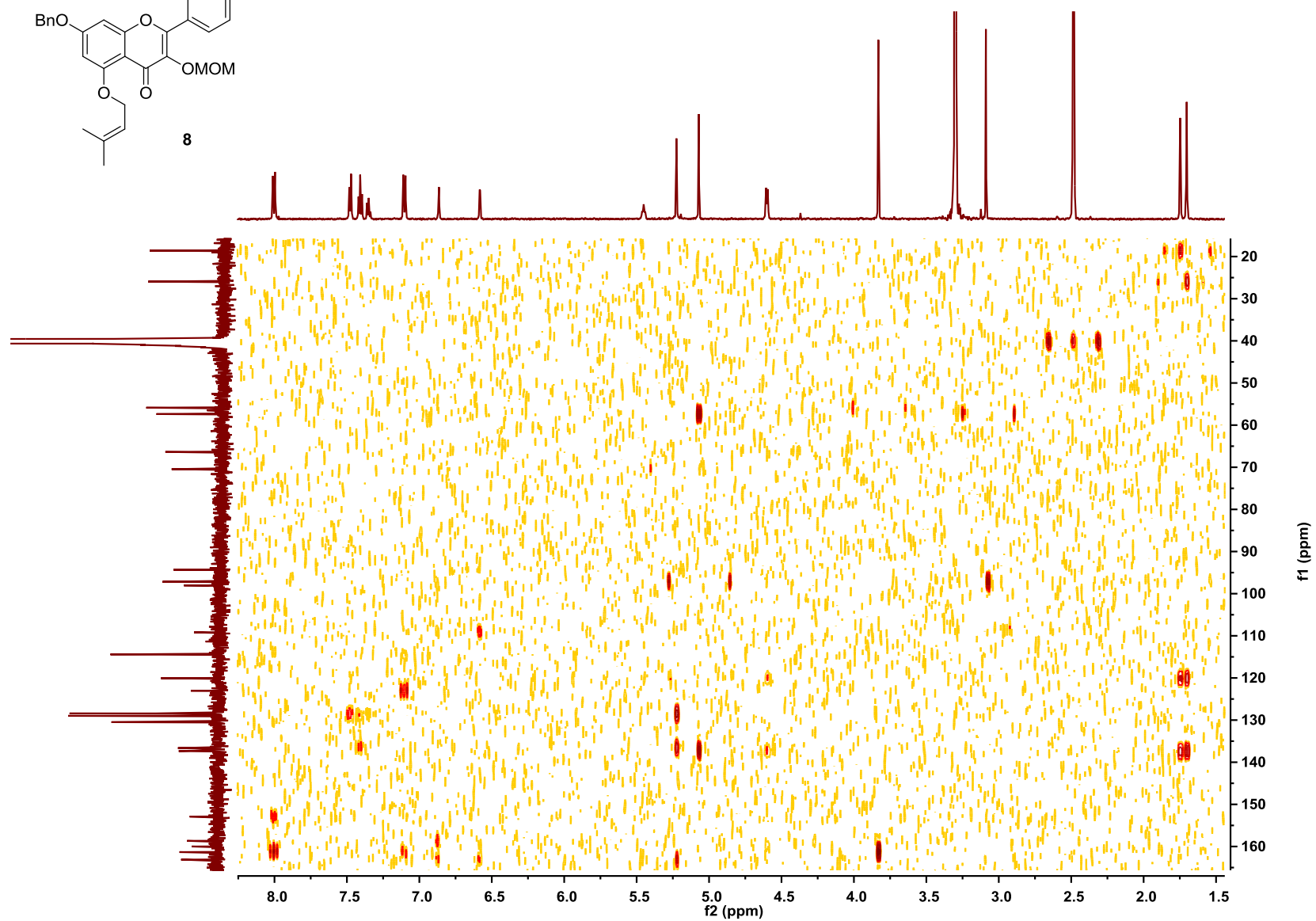
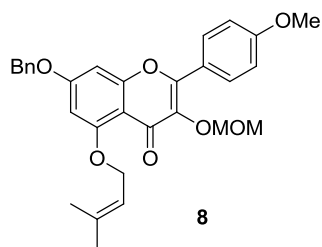
# HMBC Spectrum of Compound 3



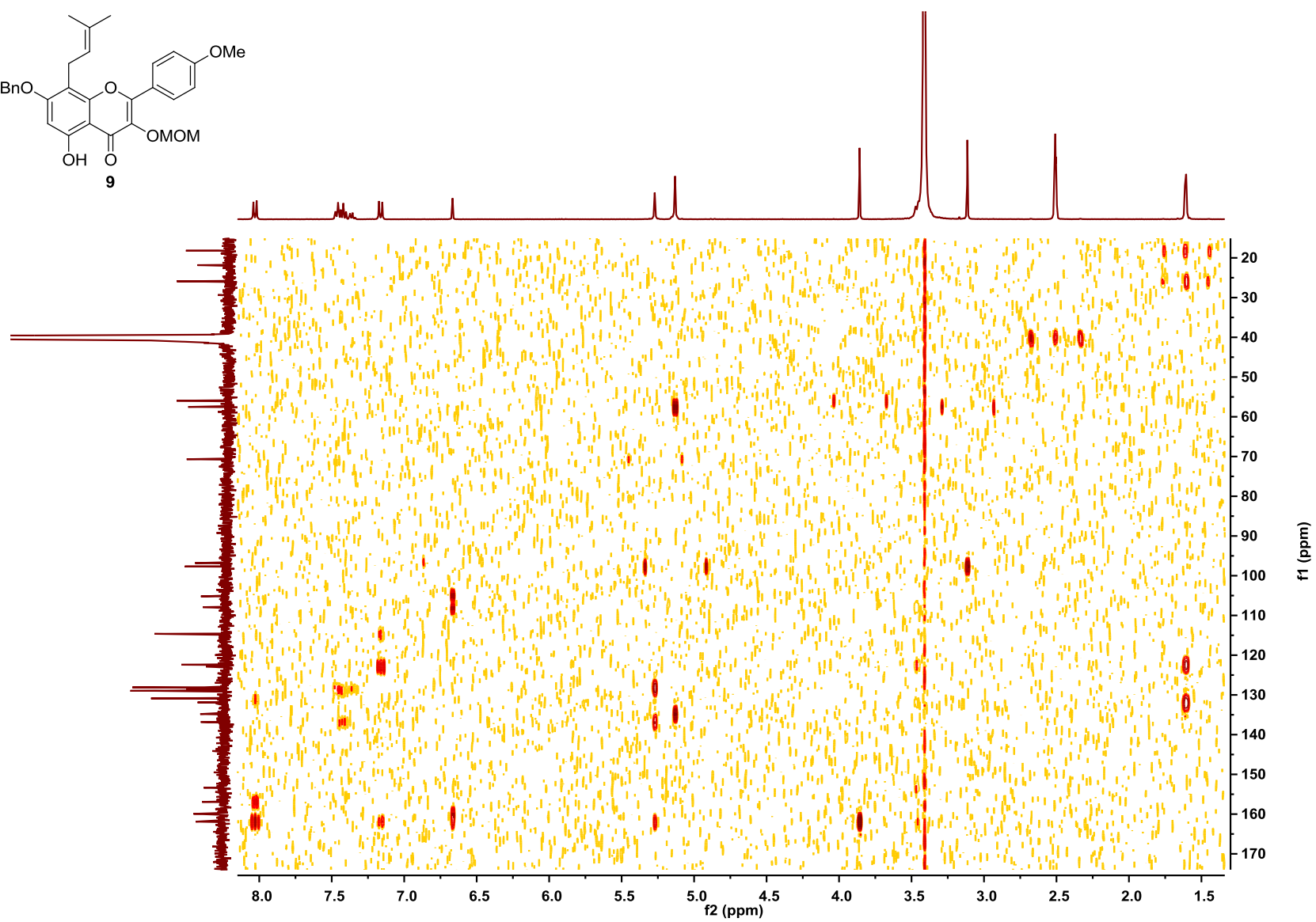
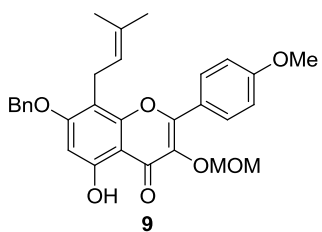
# HMBC Spectrum of Compound 7



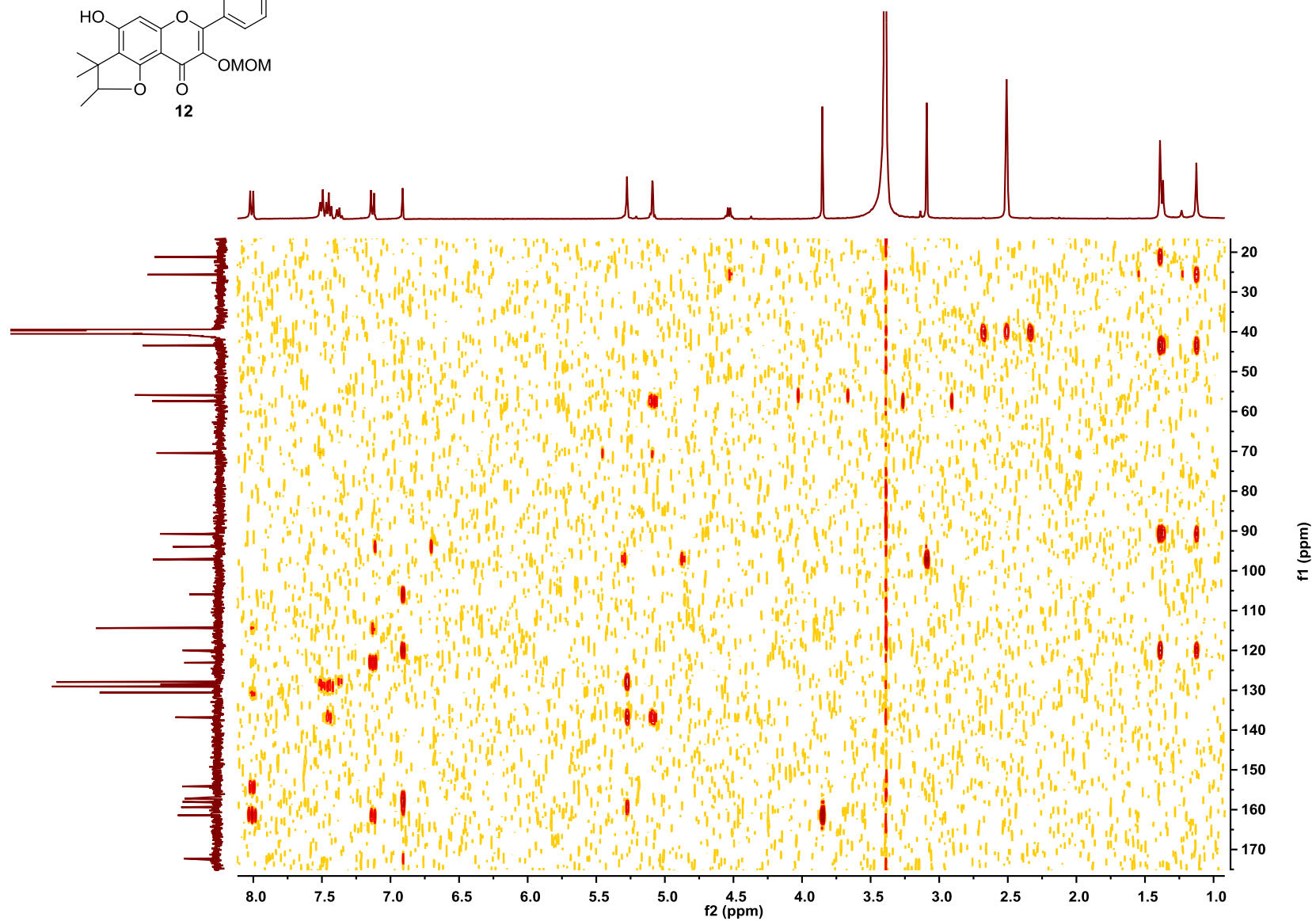
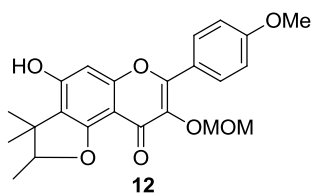
# HMBC Spectrum of Compound 8



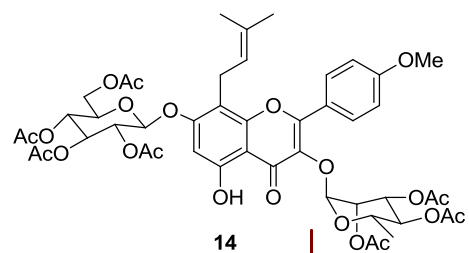
# HMBC Spectrum of Compound 9



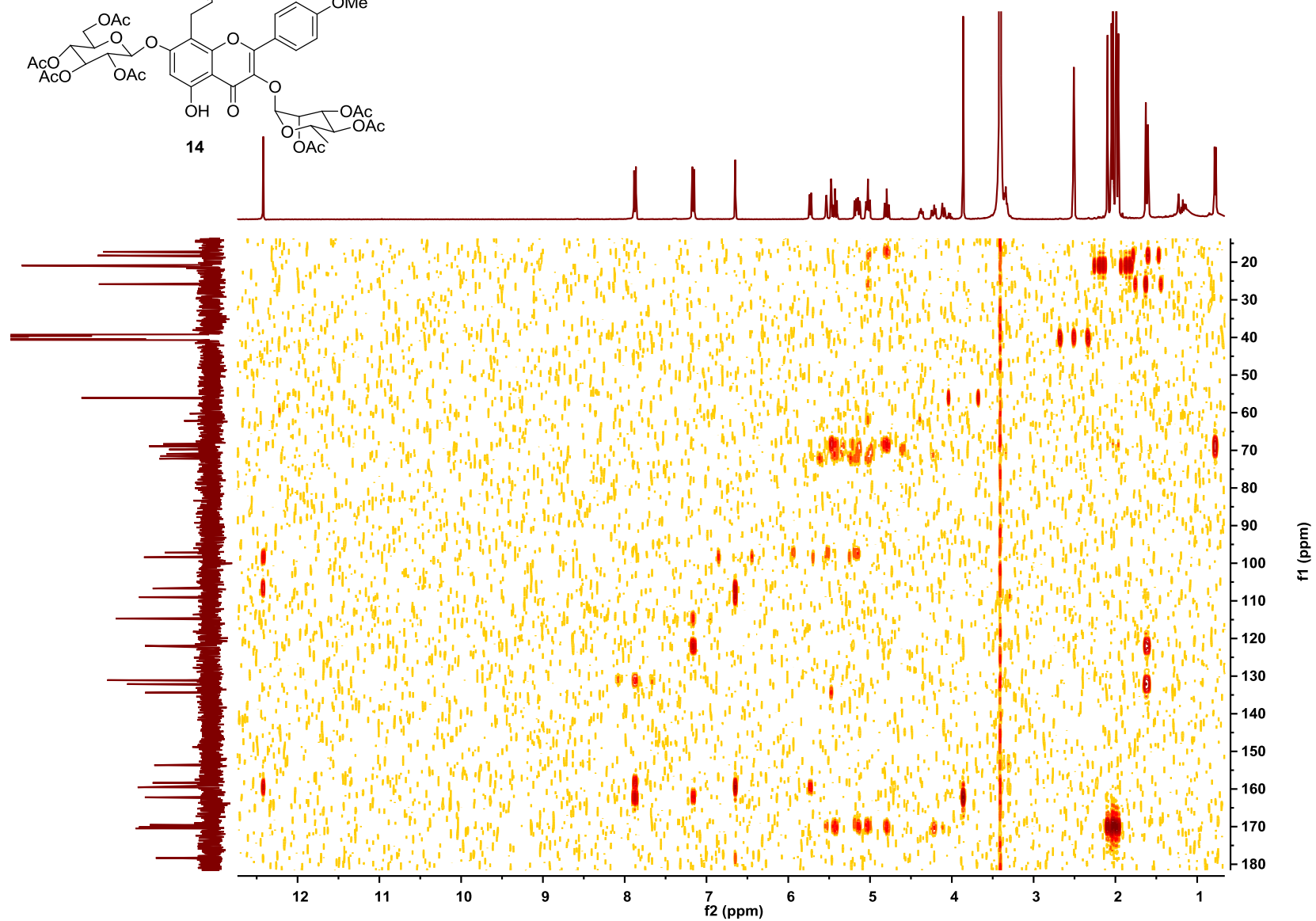
# HMBC Spectrum of Compound 12



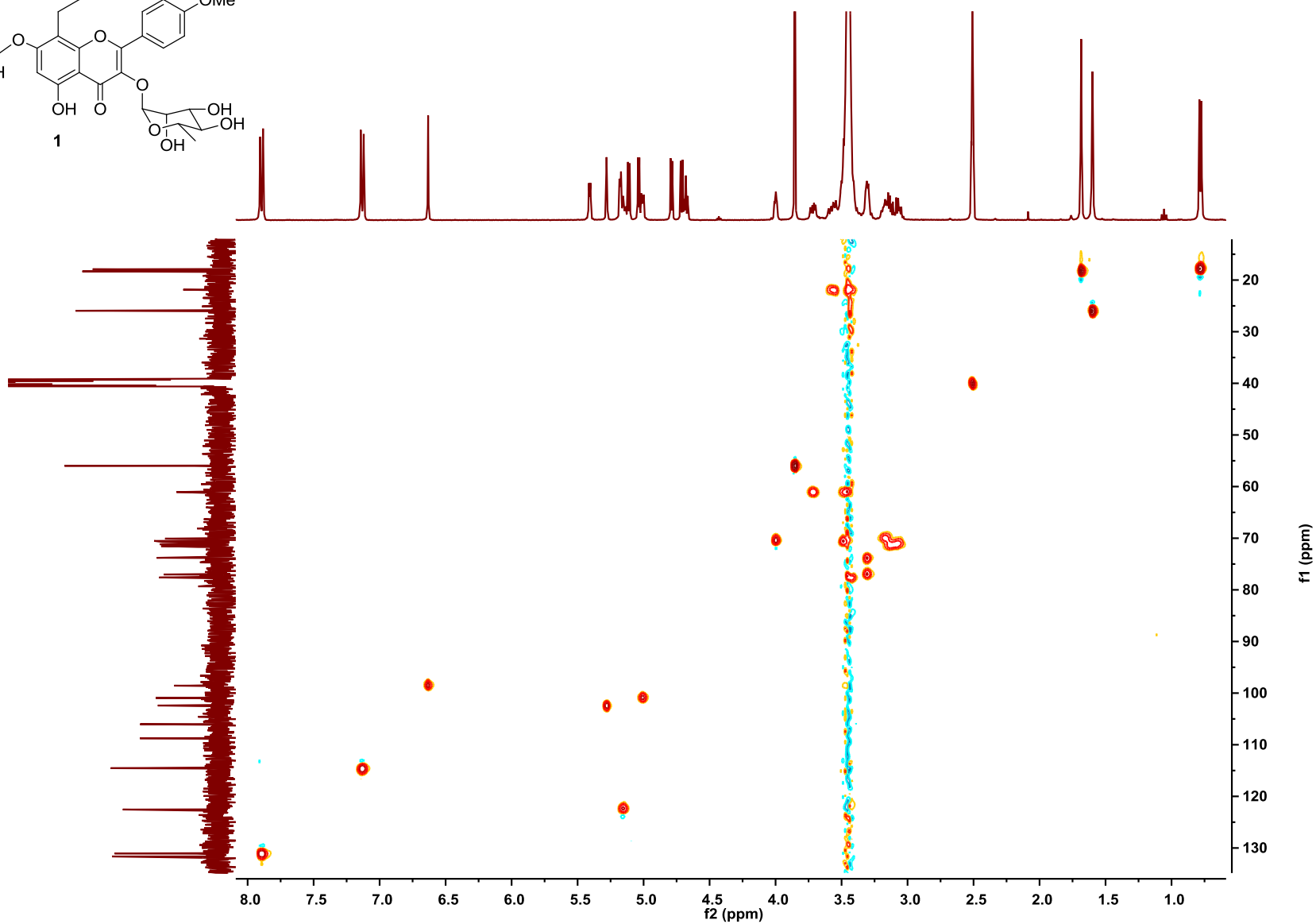
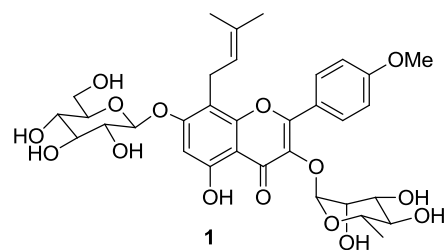
# HMBC Spectrum of Compound 14

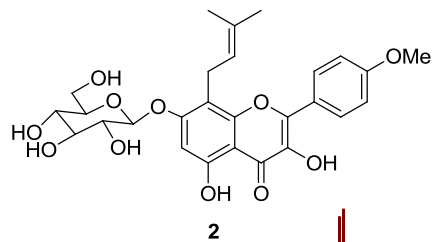


14

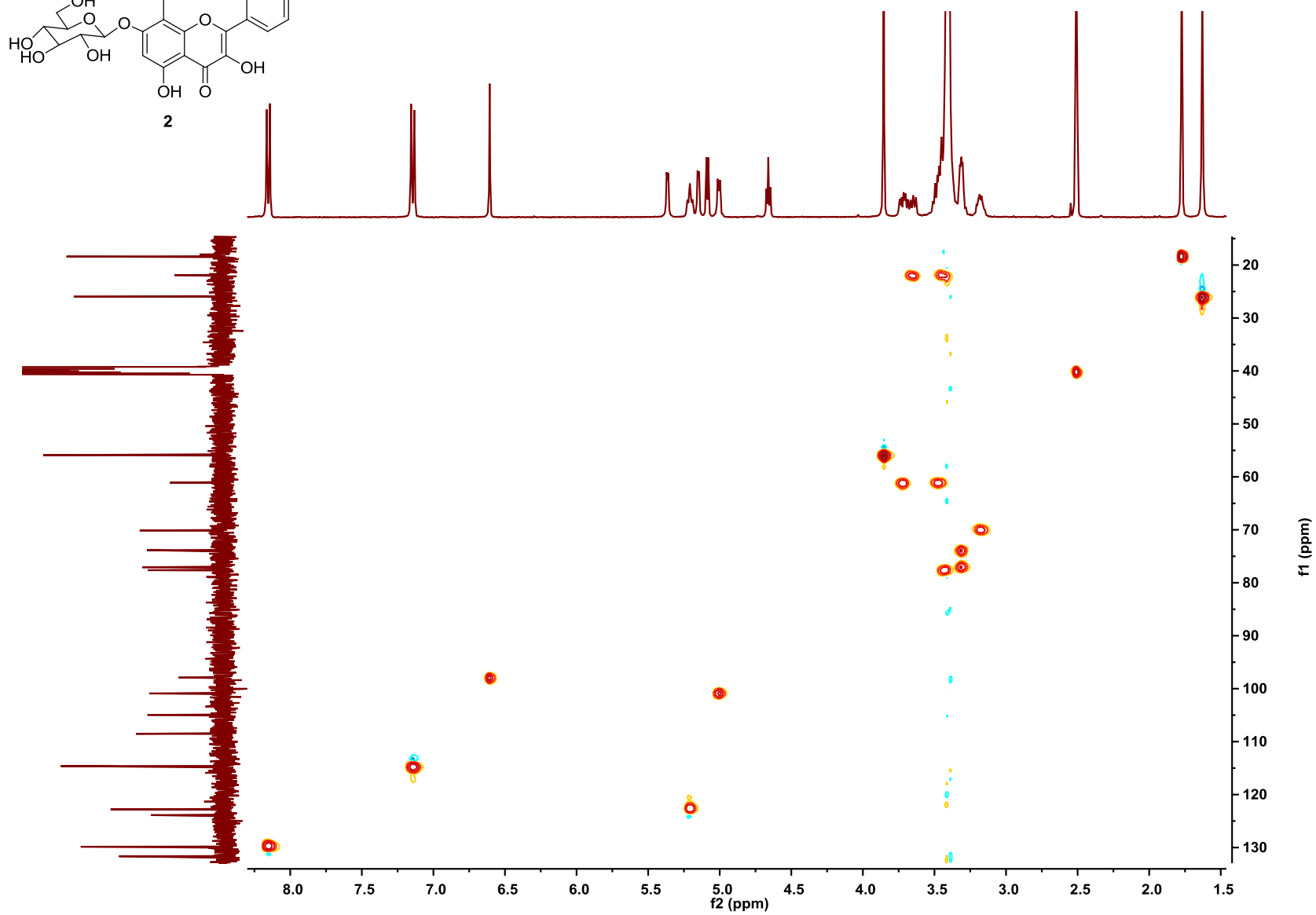


# HSQC Spectrum of Compound 1



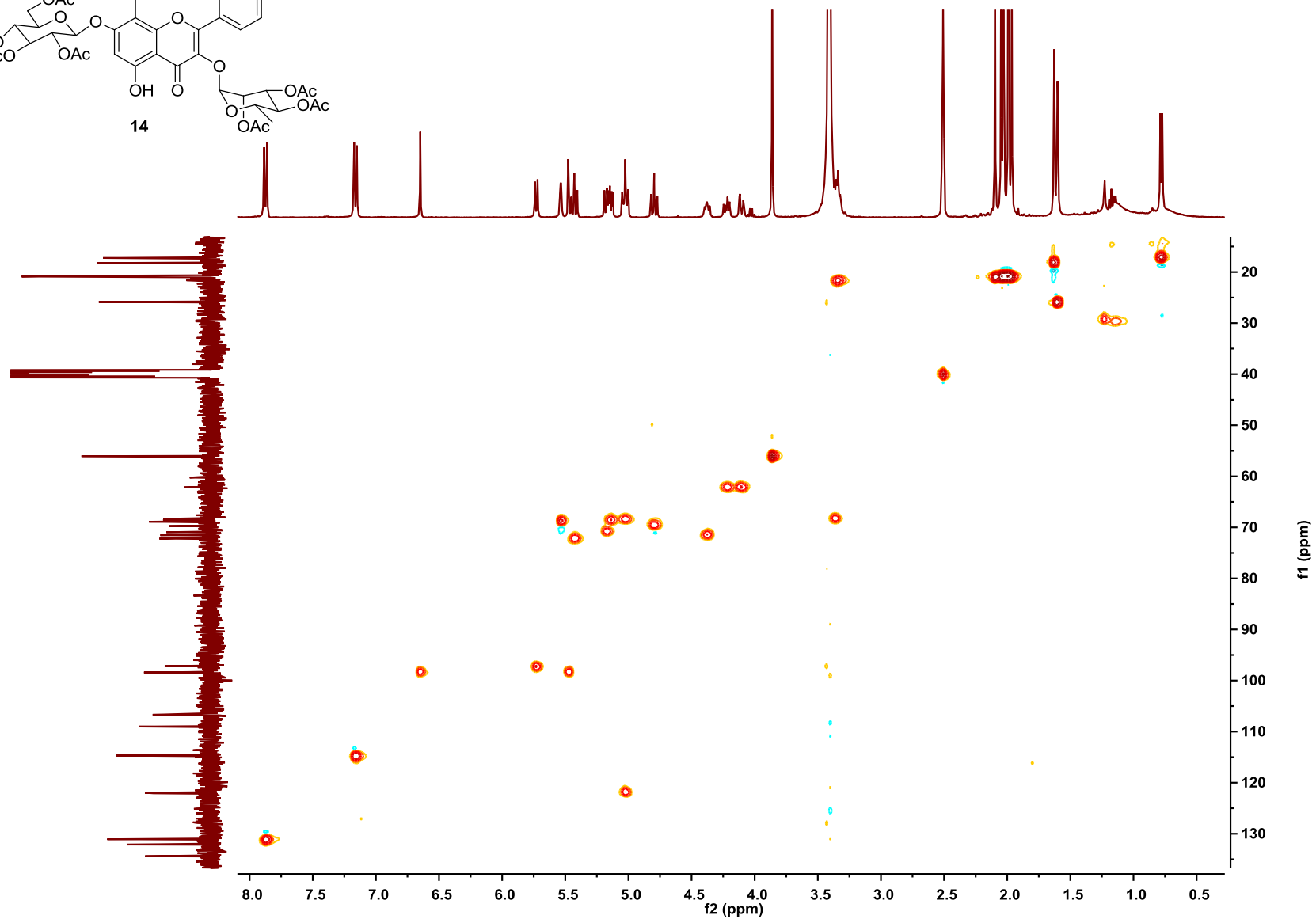
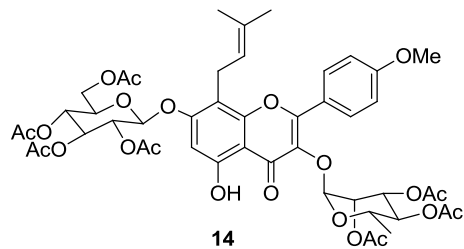


HSQC Spectrum of Compound 2

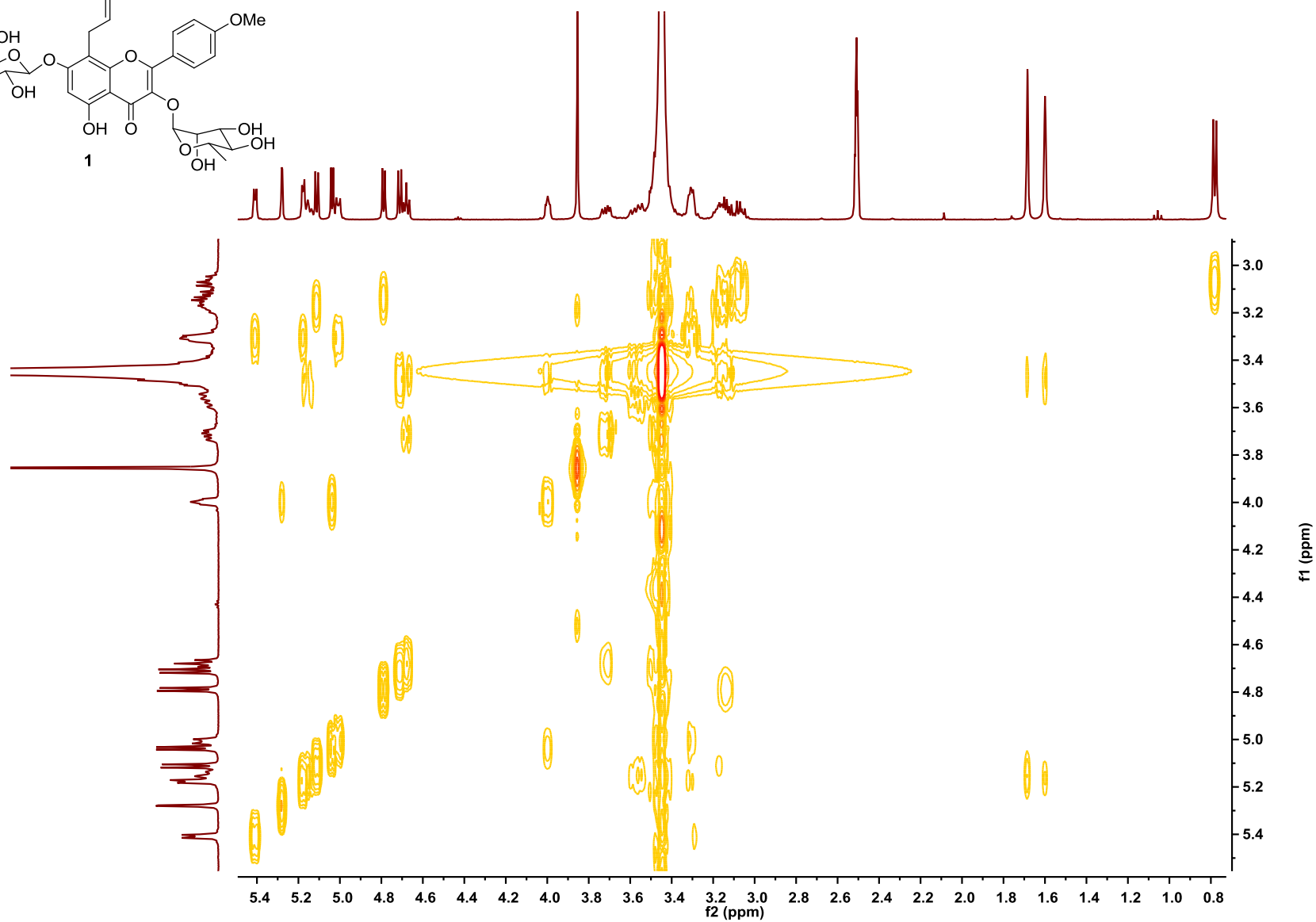
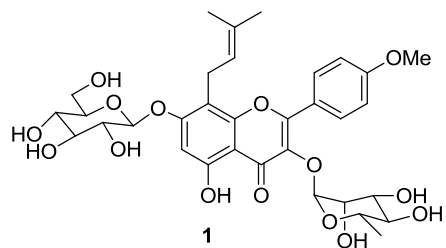


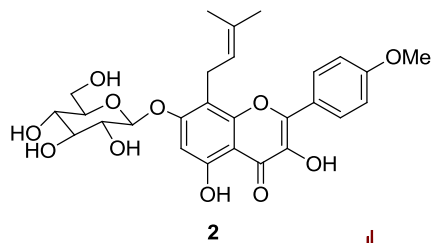


# HSQC Spectrum of Compound 14

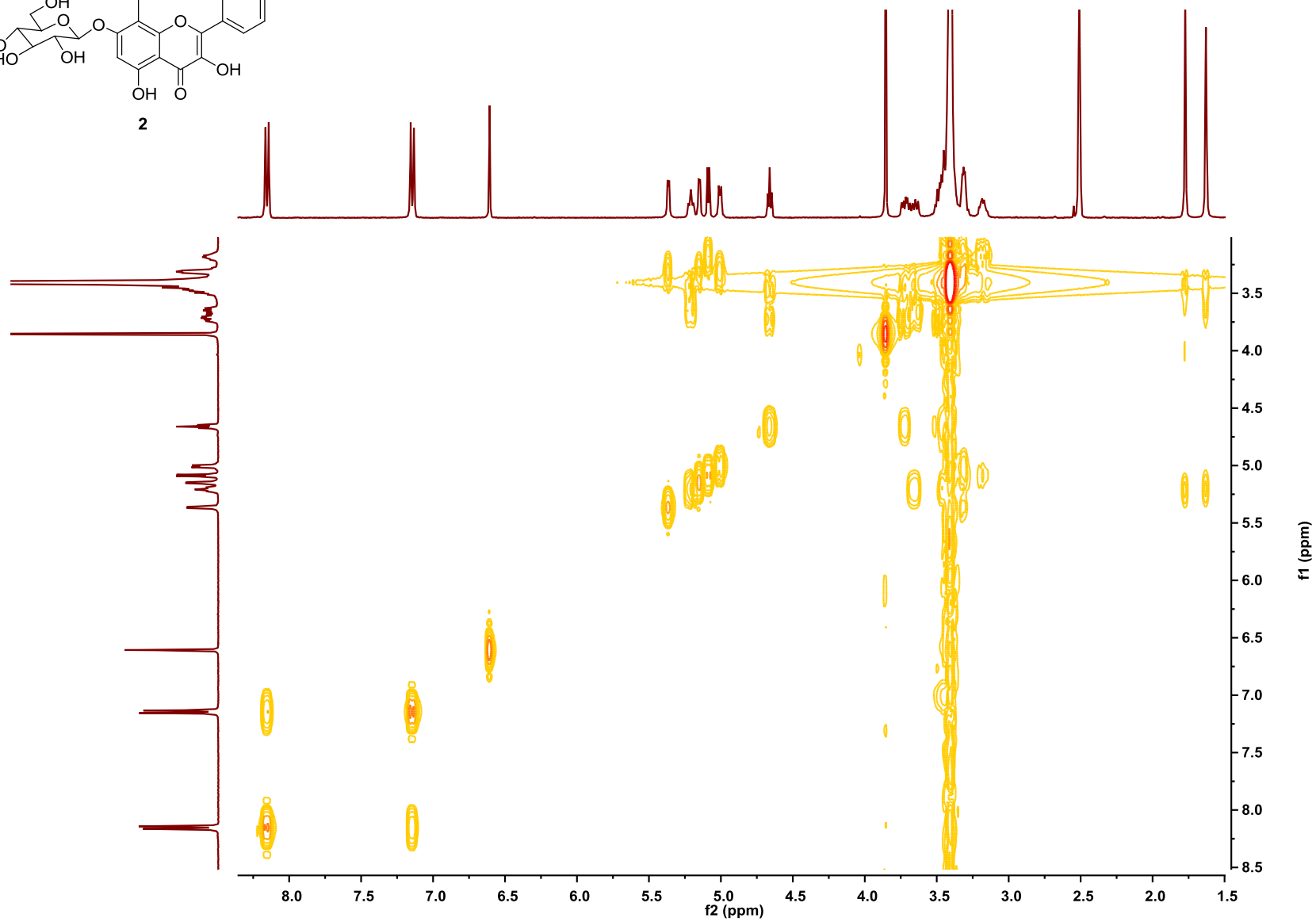


# COSY Spectrum of Compound 1

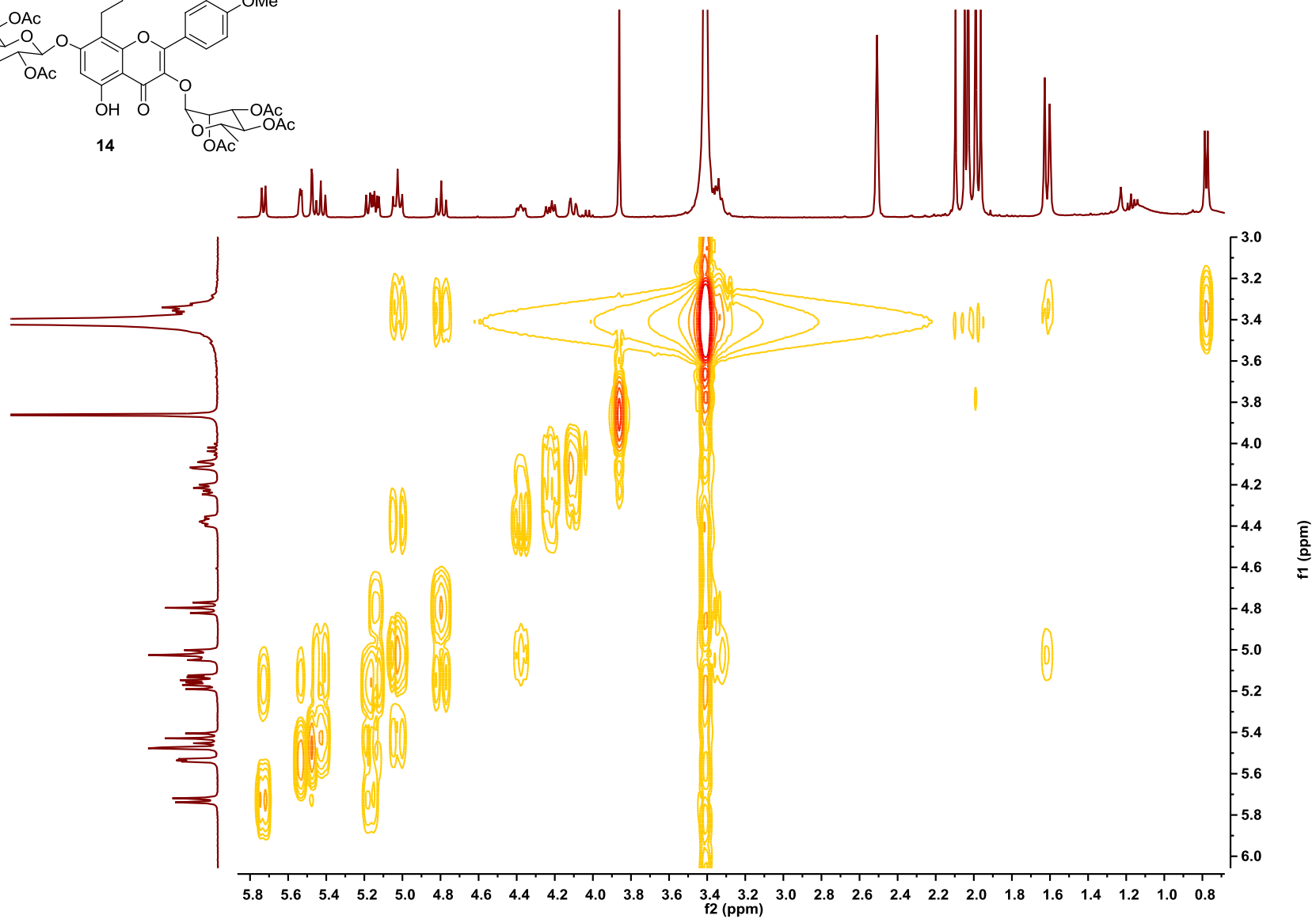
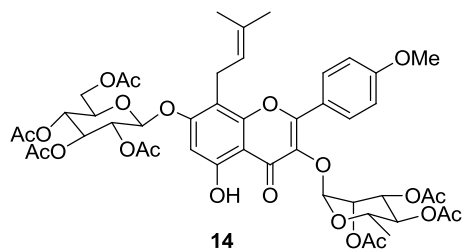




COSY Spectrum of Compound 2



# COSY Spectrum of Compound 14



### NOESY Spectrum of Compound 3

