Supplementary Material

Supplementary Fig. 1. DNA fragments accelerate dissociation of fluorescent-labeled-Fis in a direct visualization experiment.

Left panel: Sequential images of DNA tether during exchange of pre-bound GFP-Fis with 20 ng/µl herring sperm DNA (HS-DNA) fragments. Initial binding to a phage λ chromosomal DNA (49152 bp, Promega) was done with 200 nM Fis in 100 mM KGlu buffer, pH 7.6. Middle and bottom panels are images 180 s and 690 s, respectively, post addition of 5 ng/µl HS-DNA.

Right panel: Plot of control data (blue squares), exchange of GFP-Fis with 5 ng/µl HS-DNA fragments (green diamonds) and 50 ng/µl HS-DNA fragments (red circles). Dissociation rates are 0.0033 +/- 0.00081 (s⁻¹) for 5 ng/µl HS-DNA and 0.0062 +/- 0.0018 (s⁻¹) for 50 ng/µl HS-DNA, from N=4 trials for each case.

Supplementary Fig. 2. Comparison of HS-DNA-facilitated dissociation of Fis from DNA at 100 mM and 200 mM KGIu concentration

Off-rate of Fis from DNA as a function of HS-DNA concentration at fixed salt concentration of 100 mM KGlu (black squares) and 200 mM KGlu (gray triangles). The off-rates are uniformly faster at the higher salt concentration, with a higher initial exchange rate (initial slope) and a higher saturation value of off-rate, reflecting the weakening of electrostatic interactions between Fis and DNA with increased salt concentration.

Mean reaction time for the facilitated dissociation model

We consider the reaction $0 \leftrightarrow 1 \leftrightarrow 2 \rightarrow 3$ with rates k_{ij} for transitions from $i \rightarrow j$. The objective is to compute the mean time for the transition from 0 to 3 to occur, given that the final reaction from 2 to 3 is irreversible. Following the general approach of Ref. ⁴⁵, we compute the mean first passage time to state 3 by enumerating all possible sequences of states leading from 0 to 3, appropriately averaging their mean transit times based on the reaction rates, using the branching probability at each step from state i to state j, $p_{ij} = k_{ij}/(\sum_l k_{il})$:

$$\begin{split} \langle \tau_{03} \rangle &= \frac{1}{k_{01}} + \sum_{n=1}^{\infty} p_{01}^n \left(\frac{1+p_{12}}{k_{10}+k_{12}} + \frac{1}{k_{01}} \right) + \frac{p_{12}}{k_{10}+k_{12}} + \frac{p_{23}}{k_{21}+k_{23}} \\ &+ \sum_{n=1}^{\infty} p_{21}^n \left[\frac{1+p_{23}}{k_{21}+k_{23}} + \frac{p_{12}}{k_{10}+k_{12}} + \sum_{m=1}^{\infty} p_{10}^m \left(\frac{1+p_{12}}{k_{10}+k_{12}} + \frac{1}{k_{01}} \right) \right] \\ &= \frac{k_{01}(k_{12}+k_{21}+k_{23}) + k_{10}(k_{21}+k_{23}) + k_{21}k_{23}}{k_{01}k_{12}k_{23}} \end{split}$$

For this paper we make the symmetry assumption $k_{21} = k_{23}$, (which does not affect the fit function in the main text but only the interpretation of the fit parameters) plus we include the bulk DNA concentration *c* in the rate k_{12} , by replacing $k_{12} \rightarrow k_{12}c$ in the above equation (see Fig. 7). The result is Eq. 1 of the main text. We note that addition of a "direct" dissociation transition from state 0 to 3 is possible with only a slight complication of the results, but is not needed for this paper.