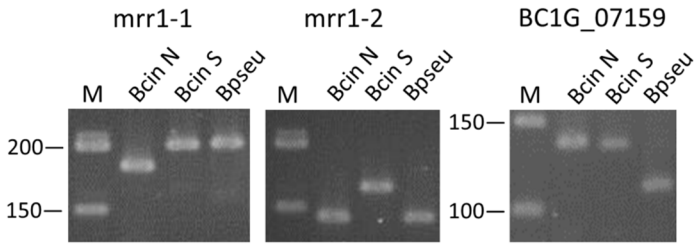
Plesken et al.: Supplemental data

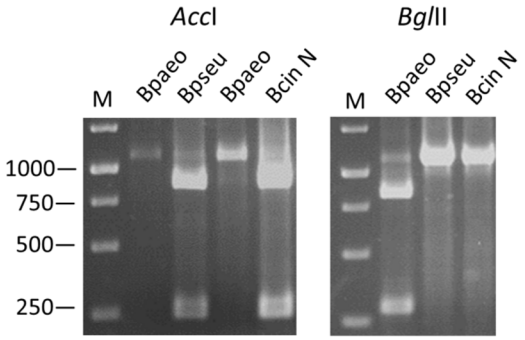
A

Primer pair	mrr1-1	mrr1-2	BC1G_07159
B. pseudocinerea	200 bp	144 bp	112bp
B. cinerea N	182 bp	144 bp	136 bp
B. cinerea S	200 bp	165 bp	136 bp



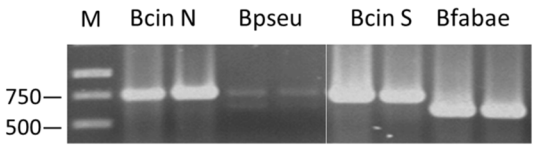
B

Primer pair	HSP60_fw/HSP60_rev	
Restriction digestion	*Bgl*II	*Acc*I
B. pseudocinerea	1111 bp	870 + 241 bp
B. cinerea N	1111 bp	870 + 241 bp
B. cinerea S	1111 bp	870 + 241 bp
B. paeoniae	830 + 281 bp	1111 bp



C

Primer pair	C729_for/ C729_rev
B. pseudocinerea	728 bp
B. cinerea N	728 bp
B. cinerea S	728 bp
B. fabae	606 bp



D

Primer pair	MS547-685_R/ MS547-201_F	
Restriction digestion	undigested	*Ear*I
B. pseudocin. group A	484 bp	205+121+158 bp
B. pseudocin. group B	484 bp	326 +158 bp

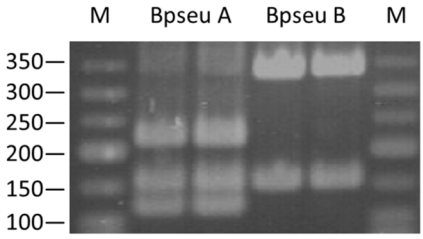


Fig. S1. PCR-based differentiation of *B. pseudocinerea* and *B. cinerea* genotypes. On the left side, primer pairs, restriction enzymes (if used), and predicted fragment sizes are indicated. On the right side, the resulting DNA fragments after gel electrophoresis are shown A: Indel PCR for differentiation of *B. cinerea* N, *B. cinerea* S and *B. pseudocinerea*, using primer pairs mrr1-1 (BcinN-in-F/ BcinN-in-R, flanking an 18 bp indel) and mrr1-2 (Mrr1-spez-F/ Mrr1-spez-R, flanking the 21 bp indel) in *mrr1* (Leroch et al., 2013). Identification of *B. pseudocinerea* is preliminary, because the same band sizes are obtained by a rare genotype of *B. cinerea*, referred to as *B. cinerea* X. Final identification of *B. pseudocinerea* is achieved by detection of a 24 bp deletion with primer pair BC1G_07159 (g2944_137_F/ g2944_273_R). B: Identification of *B. paeoniae* by PCR-RFLP. *B. paeoniae hsp60* contains a BglII site that is lacking in *B. pseudocinerea* and *B. cinerea*, and lacks an AccI site that is present in *B.*

pseudocinerea and *B. cinerea*. C: Indel PCR for identification of *B. fabae*. *B. fabae* contains a unique 122 bp deletion in a gene corresponding to *B. cinerea* BC1G_06014 (Rigotti et al., 2002), amplified with primer pair C729_for/ C719_rev. With *B. pseudocinerea* strains, only weak duplex bands are obtained. D: PCR-RFLP based differentiation of *B. pseudocinerea* groups A and B. The *ms547* sequence of *B. pseudocinerea* group A contains two EarI site one of which is missing in group B.

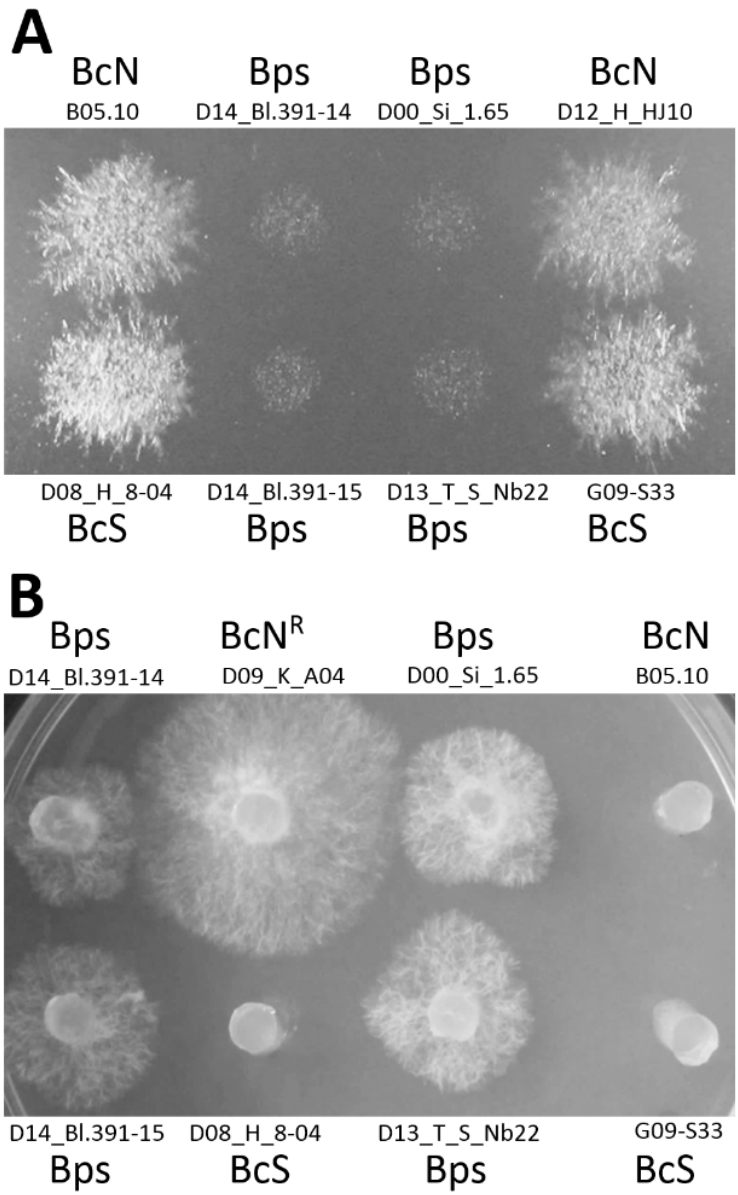


Fig. S2. Differentiation of *B. cinerea* and *B. pseudocinerea* by growth tests on 0.5% sucrose agar containing discriminatory fungicide concentrations. A: Agar with 1 mg fenpropidin l^{-1}, inoculated with conidia. B: Agar with 5 mg fenhexamid l^{-1}, inoculated with agar discs with 24 h old germlings. Pictures were taken 3 days after inoculation. Bps: *B. pseudocinerea* strains; BcN: *B. cinerea* N strains; BcS: *B. cinerea* S strains. BcN^R: Fenhexamid resistant *B. cinerea* N strain. Strains B05.10, D08_H_8-04 and G09-S33 are described in Leroch et al. (2013).

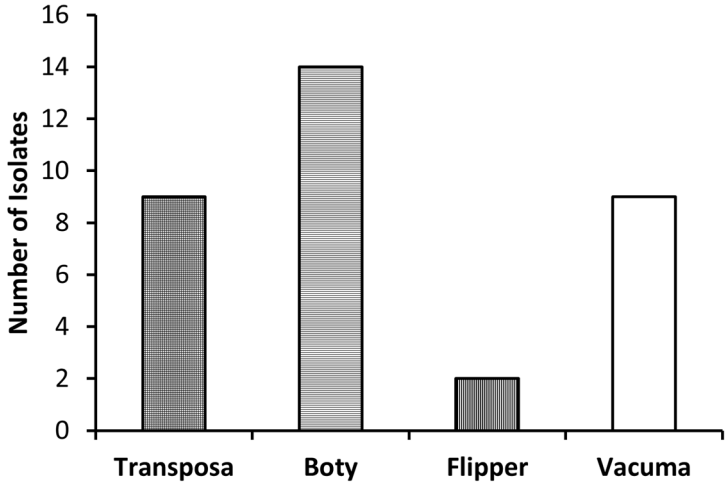


Fig. S3. Frequency of occurrence of the transposable elements Boty and Flipper in *B. pseudocinerea*, determined by PCR with Boty- and Flipper-specific primers. Transposa strains contain both elements, Vacuma strains none of these elements.

	PCR product size [bp]	
	158 bp	173 bp
B. pseudocinerea	25	6
B. cinerea	9	1

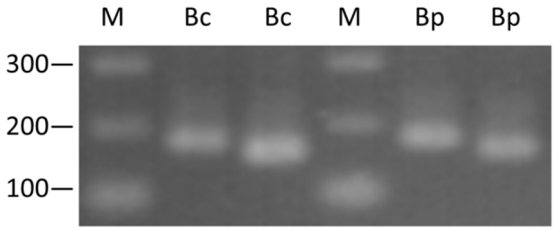


Fig. S4. PCR detection of a 15 bp indel in the mitochondrial *rpS3* coding region in strains of *B. pseudocinerea* (Bp) and B. *cinerea* (Bc). Left: Numbers of strains yielding the larger and the smaller PCR product. Right: Ethidium-bromide stained gel with PCR products from strains of both species. Fungal DNA was amplified with primers rps3_55_fw/ rps3_213_rev.

Table S2: Growth inhibition of *B. pseudocinerea* (Bps) and *B. cinerea* (Bcin) strains by different concentrations of fenpropidin in liquid and on solid medium. SA: Agar containing 0.5% sucrose. The tests were performed twice, with two replicates each. (+) Weak growth.

Species/ strain	Growth in liquid medium with fenpropidin (ppm)								Growth on SA with 1 mg fenpropidin l^{-1}
	0	0.01	0.03	0.1	0.3	1	2	4	
Bps VD110	+	+	(+)	-	-	-	-	-	-
Bps D11_T_B42	+	+	-	-	-	-	-	-	-
Bps D00-Si-1.65	+	+	(+)	-	-	-	-	-	-
Bps D00_R0_1.41	+	+	+	(+)	-	-	-	-	-
Bps D97_Ve_1.13	+	+	+	(+)	-	-	-	-	-
Bps D12_Gö_1361	+	+	-	-	-	-	-	-	-
Bps D13_K_Mai_Mu14	+	+	+	(+)	-	-	-	-	
Bps D11_T_B41 (group B)	+	+	+	+	-	-	-	-	(+)
Bcin B05.10	+	+	+	+	+	+	(+)	-	+
Bcin G09-S33	+	+	+	+	+	+	+	-	+
Bcin D09_K_A04	+	+	+	+	+	+	(+)	-	+
Bcin D08_H_8-07a	+	+	+	+	+	+	+	-	+
Bcin D13_K_S_Ge1	+	+	+	+	+	+	(+)	-	+

Table S3: Primers used in this study

Name	**Sequence**	**Gene**	**Reference**
BcinN-in-F	GCGACCTCATCGTTCTTTCAC	*mrr1*	This work
BcinN-in-R	GGCTCTCGATGAGCTGTTTC		
Mrr1-spez-F	TATCGGTCTTGCAGTCCGC	*mrr1*	Leroch et al., 2013
Mrr1-spez-R	TTCCGTACCCCGATCTTCGGAA		
g2944_137_F	GCAGATGAGGCGGATGATAG	BC1G_07159	This work
g2944_273_R	TCCACCCAAGCATCATCTTC		
BC-hch262	AAGCCCTTCGATGTCTTGGA	*hch*	Fournier et al., 2005
BC-hch520L	ACGGATTCCGAACTAAGTAA		
C729_for	AGCTCGAGAGAGATCTCTGA	BC1G_06014	(20)
C729_rev	CTGCAATGTTCTGCGTGGAA		(20)
PIRA-fw-P225L	CGAATGTATTCTCTGCGCATGCTGCTCGACATCTTGTC	*sdhB*	This work
PIRA-rev-P225L	GCAAGCCATCTGTAACTCTG		
Bc_Tub_for	AAGATCCGCGAGGAGTTCCC	*tubA*	This work
Bc_Tub_rev	GGCGGTTTGGACGTTGTTAG		
155_betaTub_F	CAACCTTCAAAATGCGTGAG	*tubA*	Fournier et al., 2005
1174_betaTub_R	AGATGGGTTGCTGAGCTTCA		
PIRA-Tub1	GCCTCGTTATCGATACAGAAGATC	*tubA*	This work
PIRA-Tub2	TATGATGGCTACCTTCTCCGTC		
Bc-Boty1	CTTTACCGGAACACAAGCCAT	Boty *pol*	Kretschmer and Hahn, 2008
Bc-Boty2	GGTCTTCCATTCTTCGCCTTC		
F300	GCACAAAACCTACAGAAGA	Flipper	Levis et al., 1997
F1550	ATTCGTTTCTTGGACTGTA		
Qo13ext	GGTATAACCCGACGGGGTTATAGAATAG	*cytB*	Leroux et al., 2010
Qo14ext	AACCATCTCCATCCACCATACCTACAAA		
mito_55_fw	TTGCTCAGGGCTCCGGTAAC	*rps3* (mitoch.)	This work
mito_213_rev	GCTGGTCCAGAGTGTAAGG		
HSP60_fw	CAACAATTGAGATTTGCCCACAAG	*hsp60*	Staats et al., 2005
HSP60_rev	GATGGATCCAGTGGTACCGAGCAT		
G3PDH_fw	ATTGACATCGTCGCTGTCAACGA	*g3pdh*	Staats et al., 2005
G3PDH_rev	ACCCCACTCGTTGTCGTACCA		
MS547-F	AAGGAGGACGTTGGAAGGAT	*ms547*	Walker et al., 2011
MS547-R	AAGTCCAGAATCTCGATGTATTTGT		
ms547_685_R	GTCGATAGATGGGCTGCAAG	*ms547*	This work
ms547_201_F	GTTGGTGGAGGTAGGAAGAC		
Mat1-F	GTGACCAGGAAACAGCTATGACCGGAGTGTGTTGATCGTGGAGCCGAG	(MAT1-1)	Amselem et al., 2011
Mat1-R	GTGACTGTAAAACGACGGCCAGTCCACACATACATCATGACGGCTCCC	(MAT1-2)	

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