

Design of CID-Cleavable Protein Cross-linkers: Identical Mass Modifications for Simpler Sequence Analysis

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Supporting Information

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General Methods

¹H NMR and ¹³C NMR spectra were recorded at ambient temperature at 500 MHz and 125 MHz, respectively, on a Bruker DRX500 NMR instrument. ¹H and ¹³C NMR data is reported as follows: chemical shifts are reported in ppm on a δ scale and referenced to internal tetramethylsilane or residual solvent (TMS: δ 0.00; CHCl₃: δ 7.27(¹H), 77.23 (¹³C)), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, qu = quintet, m = multiplet), coupling constants (Hz), and integration. Infrared (IR) spectra were obtained using a FT-IR spectrometer. Accurate mass spectra were acquired on a Waters LCT Premier quadrupole time-of-flight spectrometer and were obtained by peak matching. Gas chromatography/mass spectrometry (GC/MS) was performed with a Thermo-Finnigan Trace Mass Spectrometer Plus quadrupole system with a fused silica capillary column (30 m x 0.32 mm x 0.25 mm) wall-coated with DB-5 (J & W Scientific) using electron ionization (70 eV) or a Waters GCT Premier orthogonal acceleration time-of-flight spectrometer using chemical ionization. Melting points are uncorrected and were obtained using a Büchi 510 melting point apparatus. Analytical thin layer chromatography was performed on EMD Chemicals Inc. silica gel 60 F₂₅₄ plates. Liquid chromatography was performed using forced flow (flash chromatography) of the indicated solvent system on Sorbent Technologies silica gel (SiO₂) 60 (230–400 mesh). Unless otherwise noted, all reactions were carried out under an atmosphere of argon in flame-dried glassware. Solvents were distilled from CaH₂ or filtered through alumina before use.¹

General procedure 1: Ester Hydrolysis

Starting ester was dissolved in 2:1 or 4:1 THF:H₂O, depending on solubility. The mixture was cooled to 0 °C and lithium hydroxide (98%, 5 equiv.) dissolved in minimal

H_2O was added slowly. The reaction was monitored by thin layer chromatography, while stirring vigorously. Upon completion, the mixture was diluted with diethyl ether and water before the layers were separated. Diethyl ether was added to the resulting aqueous layer, followed by addition of HCl to pH 1. The layers were separated and the aqueous layer was extracted with diethyl ether (2x). Organics were combined, washed with brine, dried over Na_2SO_4 , filtered and evaporated *in vacuo*.

General procedure 2: TBAF-Promoted Michael Additions

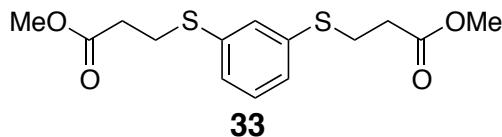
Acrylic acid (0.28 mmol), thiophenol (0.51 mmol), and THF (1 mL) were combined in a round bottom flask to which $\text{TBAF}\bullet 3\text{H}_2\text{O}$ (0.06 mmol) was added at rt. The flask was fitted with a cold-water condenser and the mixture was heated to 50° C for 16 h. The mixture was cooled to rt, evaporated *in vacuo*, diluted with EtOAc and washed with 1N HCl (2x). The organic layer was washed with brine, dried over Na_2SO_4 , filtered, and evaporated *in vacuo* to yield the crude product. In some cases, this reaction was run neat.²

General procedure 3: NHS Ester Preparation

Crude diacid (0.08 mmol) was dissolved in dry dichloromethane (0.62 mL) in a flame dried round bottom flask under argon. $\text{NHS}\bullet\text{TFA}$ (0.41 mmol) was added before a slow addition of triethylamine (0.49 mmol) at 0° C. The mixture was left to warm slowly overnight while stirring under argon. After 16 h, the mixture was diluted with dichloromethane and washed with water (2x). The organic layer was dried with Na_2SO_4 , filtered, and evaporated *in vacuo*.

General procedure 4: *m*-CPBA-Oxidation to Sulfoxide

Di-NHS ester (0.026 mmol) was dissolved in CDCl₃ (1 mL) and *m*-CPBA (77% w/w, 0.026 mmol) was added slowly while monitoring by LRMS ESI + and ¹H NMR. When this reaction was run on larger scale than 30 mg starting material, the solution was cooled in an ice bath prior to *m*-CPBA addition. Once the reaction was complete, the solution was diluted with dichloromethane (2 mL) and washed with 10% sodium bicarbonate solution (3 x 2 mL). The organic layer was dried over Na₂SO₄, filtered, and evaporated *in vacuo* to afford the product.

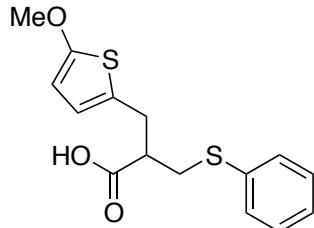


Dimethyl 3,3'-(1,3-phenylenebis(sulfanediyl))dipropanoate (33): 1,3-benzenedithiol (0.500 g, 3.52 mmol) and methyl acrylate (0.632 mL, 7.02 mmol) were dissolved in THF (6 mL), and Et₃N (0.980 mL, 7.03 mmol) was added to the solution. After 5 h, the reaction mixture was partitioned between H₂O (10 mL) and EtOAc (10 mL). The EtOAc layer was collected, washed with brine (10 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude oil was purified by silica gel chromatography (5:1 hexanes:EtOAc) to afford 0.550 g (50%) of the sulfide as a clear, colorless oil: ¹H (500 MHz, CDCl₃) δ 7.38 (s, 1H), 7.29–7.22 (m, 3H), 3.73 (s, 6H), 3.22 (t, J = 7.5 Hz, 4H), 2.69 (t, J = 7.5 Hz, 4H); ¹³C (125 MHz, CDCl₃) δ 172.2, 136.6, 130.5, 129.6, 127.0, 52.0, 34.2, 28.8; IR (neat) 2997, 2951, 2846, 1739, 1570 cm⁻¹; Accurate Mass (ES+/MeOH) m/z calcd for C₁₄H₁₈O₄S₂Na [M+Na]⁺ 337.0544, found 337.0547.

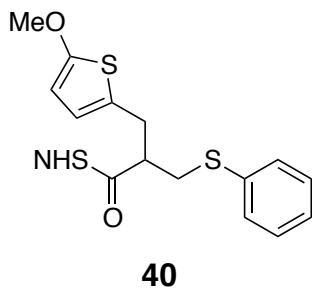
Bis(2,5-dioxopyrrolidin-1-yl) 3,3'-(1,3-phenylenedisulfinyl)dipropanoate (2).

Bis(2,5-dioxopyrrolidin-1-yl) 3,3'-(1,3-phenylenebis(sulfanediyl))dipropanoate (0.110 g, 0.229 mmol) was dissolved in CHCl₃ (7 mL), and the reaction mixture was cooled to 0°C.

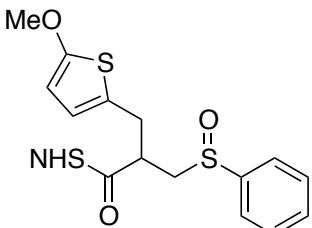
A solution of *m*-CPBA (0.105 g, 0.457 mmol) in CHCl₃ (2 mL) was added drop-wise, and the reaction mixture was stirred for 10 min. The reaction mixture was then partitioned between CHCl₃ (10 mL) and saturated aqueous NaHCO₃ (10 mL). The CHCl₃ layer was collected, dried over MgSO₄, filtered, and concentrated to afford 0.080 g (75%) of sulfoxide **2** as a white solid: ¹H (500 MHz, CDCl₃) δ 7.93–7.90 (m, 1H), 7.80–7.78 (m, 3H), 3.46–3.38 (m, 2H), 3.17–3.10 (m, 4H), 2.82 (brs, 10H); ¹³C (125 MHz, CDCl₃) δ 169.0, 167.0, 144.6, 144.5, 131.0, 127.1, 120.3, 49.7, 49.5, 25.7, 22.9, 22.7; IR (KBr) 2943, 2935, 1786, 1736, 1427 cm⁻¹; Accurate Mass (ES+/MeOH) m/z calcd for C₂₀H₂₀N₂₀O₁₀S₂Na [M+Na]⁺ 535.0457, found 535.0457.

**31**

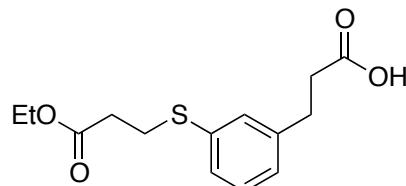
3-(5-methoxythiophen-2-yl)-2-((phenylthio)methyl)propanoic acid (31). Acrylic acid **29** (0.073 g, 0.25 mmol) and thiophenol (0.039 mL, 0.38 mmol) were subjected to general procedure 2 without solvent. The crude product was purified by flash column chromatography (30% EtOAc/hexanes) to yield desired product (0.018 g, 23%). ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, *J* = 7.9, 2H), 7.29 (t, *J* = 7.6, 2H), 7.22 (t, *J* = 7.6, 1H), 6.42 (d, *J* = 3.6, 1H), 5.99 (d, *J* = 3.8, 1H), 3.84 (s, 3H), 3.23 (dd, *J* = 7.7, 13.7, 1H), 3.16–3.04 (m, 3H), 2.88 (quintet, *J* = 6.8, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 179.2, 165.3, 135.2, 130.5, 129.3, 127.0, 126.0, 123.8, 103.4, 60.4, 47.0, 34.6, 31.6; cm⁻¹; Accurate Mass ES- *m* / *z* calcd for C₁₅H₁₅O₃S₂ (M-H)⁻ 307.0463, found 307.0466.



NHS ester 40. Carboxylic acid **31** (0.018 g, 0.058 mmol) was combined with dry DMF (0.3 mL), NHS (0.013 g, 0.12 mmol), pyridine (0.019 mL, 0.23 mmol), and, lastly, trifluoroacetic anhydride (0.016 mL, 0.12 mmol). After 2.5 h stirring at rt, the mixture was diluted with dichloromethane and 1M HCl. After the layers were separated, the organic layer was washed with 1 M HCl (2x) and dilute NaHCO₃ solution (2x). The organic layer was dried with Na₂SO₄, filtered, and evaporated *in vacuo*. This procedure is adapted from a published procedure.³ The crude product was purified by flash column chromatography (2% EtOAc/CH₂Cl₂) to yield desired product (0.015 g, 63%). ¹H NMR (500 MHz, CDCl₃) δ 7.26-7.20 (m, 3H), 7.07 (d, *J* = 6.5, 1H), 4.15 (q, *J* = 7.2, 2H), 3.17 (t, *J* = 7.4, 2H), 3.04 (t, *J* = 7.5, 2H), 2.92 (t, *J* = 7.7, 2H), 2.85 (s, 4H), 2.62 (t, *J* = 7.4, 2H), 1.26 (t, *J* = 7.1, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.0, 169.3, 168.0, 140.2, 136.0, 129.9, 129.5, 128.4, 126.7, 60.9, 34.6, 32.7, 30.5, 29.1, 25.8, 14.4; IR (liquid) 2928, 1814, 1784, 1737, 1591 cm⁻¹; Accurate Mass ES+ *m/z* calcd for C₁₉H₁₉O₅S₂NNa (M+Na)⁺ 428.0602, found 428.0591.

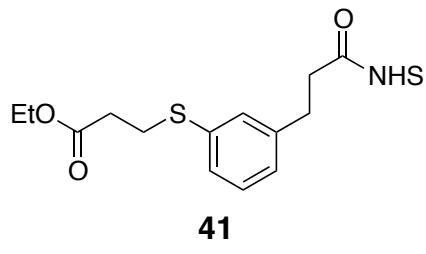


Sulfoxide 10. NHS ester **40** (0.012 g, 0.03 mmol) was subjected to general procedure 4. The crude product was purified by flash column chromatography (10–20% EtOAc/CH₂Cl₂) to yield desired product as a mixture of diastereomers (0.003 g, 26%). ¹H NMR (500 MHz, CDCl₃) δ 7.68–7.63 (m, 2H), 7.53 (m, 3H), 6.61 (d, *J* = 3.8, 0.6H), 6.49 (d, *J* = 3.7, 0.5H), 6.04 (d, *J* = 3.8, 0.6H), 5.98 (d, *J* = 3.8, 0.5H), 3.86 (s, 1.8H), 3.84 (s, 1.3H), 3.60–3.53 (m, 0.5H), 3.47–3.40 (m, 0.6H), 3.39 (s, 0.6H), 3.38 (s, 0.5H), 3.30–3.24 (m, 1.5H), 3.10 (dd, *J* = 7.2, 15.3, 0.5H), 3.03 (dd, *J* = 8.0, 13.2, 0.6H), 2.96 (dd, *J* = 4.0, 13.4, 0.5H), 2.87 (s, 2.1H), 2.84 (s, 2.3H); ¹³C NMR (125 MHz, CDCl₃) δ 168.7, 168.6, 168.3, 165.7, 165.6, 143.5, 143.0, 131.6, 131.4, 129.6, 129.5, 125.3, 124.8, 124.2, 124.0, 123.5, 123.2, 103.6, 103.5, 60.2, 60.2, 57.8, 56.6, 39.5, 39.0, 32.5, 31.0, 29.8, 25.7, 25.6; IR (thin film) 2924, 1810, 1782, 1739, 1507, cm⁻¹; Accurate Mass ES+ *m/z* calcd for C₁₉H₁₉NO₆S₂Na (M+Na)⁺ 444.0551, found 444.0548.

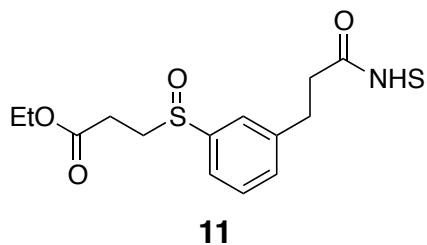
**32**

3-((3-Ethoxy-3-oxopropyl)thio)phenylpropanoic acid (32). Carboxylic acid **15** (0.10 g, 0.55 mmol) was combined with ethyl acrylate (0.088 mL, 0.82 mmol) in ethanol (5.5 mL) in a scintillation vial. Triethylamine (0.15 mL, 1.1 mmol) was added and the mixture was capped and stirred 14 days. The solvent was removed and the crude product was purified by flash column chromatography (50% EtOAc/hexanes) to yield desired product (0.022 g, 14%). ¹H NMR (500 MHz, CDCl₃) δ 7.23 (br s, 3H), 7.07 (d, *J* = 6.7, 1H), 4.16 (q, *J* = 7.2, 2H), 3.17 (t, *J* = 7.3, 2H), 2.95 (t, *J* = 7.6, 2H), 2.68 (t, *J* = 7.6, 2H), 2.61 (t, *J* = 7.5, 2H), 1.25 (t, *J* = 7.2, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.7, 172.2, 141.3, 135.6, 130.0, 129.4, 128.2, 126.8, 61.1, 35.5, 34.7, 30.7, 29.2, 14.4; IR

(thin film) 3200, 2982, 2661, 2361, 1732 cm^{-1} ; Accurate Mass ES- m / z calcd for $\text{C}_{14}\text{H}_{17}\text{O}_4\text{S} (\text{M}-\text{H})^-$ 281.0847, found 281.0844.



NHS ester 41. Carboxylic acid **32** (0.022 g, 0.078 mmol) was combined with dry DMF (0.4 mL), NHS (0.018 g, 0.16 mmol), pyridine (0.025 mL, 0.31 mmol), and, lastly, trifluoroacetic anhydride (0.022 mL, 0.16 mmol). After 2.5 h stirring at rt, the mixture was diluted with dichloromethane and 1M HCl. After the layers were separated, the organic layer was washed with 1 M HCl (2x) and dilute NaHCO_3 solution (2x). The organic layer was dried with Na_2SO_4 , filtered, and evaporated *in vacuo*. This procedure is adapted from a published procedure.³ The crude product was purified by flash column chromatography (10% EtOAc/ CH_2Cl_2) to yield desired product (0.009 g, 30%). ^1H NMR (500 MHz, CDCl_3) δ 7.40 (d, $J = 7.4$, 2H), 7.31 (t, $J = 7.5$, 2H), 7.26 (t, $J = 7.3$, 1H), 6.50 (d, $J = 3.7$, 1H), 6.00 (d, $J = 3.7$, 1H), 3.85 (s, 3H), 3.33–3.05 (m, 5H), 2.84 (s, 4H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.1, 169.0, 165.5, 134.6, 131.1, 129.4, 127.3, 124.7, 124.6, 103.6, 60.4, 44.9, 34.7, 31.4, 25.8; IR (liquid) 3057, 2944, 1810, 1737, 1562 cm^{-1} ; Accurate Mass ES+ m / z calcd for $\text{C}_{18}\text{H}_{21}\text{O}_6\text{SNa} (\text{M}+\text{Na})^+$ 402.0987, found 402.0979.



Sulfoxide 11. NHS ester **41** (0.007 g, 0.02 mmol) was subjected to general procedure 4 to yield desired product (0.005 g, 60%). ^1H NMR (500 MHz, CDCl_3) δ 7.54–7.46 (m,

3H), 7.39 (d, J = 3.8, 1H), 4.18–4.07 (m, 2H), 3.28–3.20 (m, 1H), 3.15 (t, J = 7.5, 2H), 3.00–2.93 (m, 3H), 2.90–2.79 (m, 5H), 2.56–2.45 (m, 1H), 1.25 (t, J = 7.0, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.5, 169.2, 167.8, 143.7, 140.8, 131.4, 129.9, 124.0, 122.8, 61.3, 51.3, 32.6, 30.6, 26.4, 25.8, 14.4; IR (thin film) 2930, 2360, 1813, 1783, 1737 cm^{-1} ; Accurate Mass ES+ m/z calcd for $\text{C}_{18}\text{H}_{21}\text{NO}_7\text{SNa}$ ($\text{M}+\text{Na}$) $^+$ 418.0937, found 418.0917.

IML 2 MS Data:

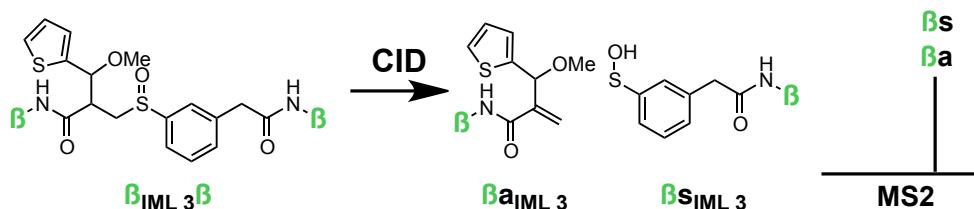


Figure 1. Expected fragmentation of IML 3 in single peptide crosslinking

Initial testing of IML 3 with synthetic peptide Ac-IR7 in the MS instrument showed significant loss of the methoxy group during CID (Figure 2). The MS^2 spectrum shows three peaks representing the IML 3-modified peptides. The first peak, m/z 496.73, represents a doubly charged IML 3-modified peptide that has lost a methoxy substituent during CID fragmentation (Figure 2, top left). This peak could only represent the alkene half of the linker, since it contains a labile methoxy in the benzylic position. When this peak was subjected to further CID, sequence data was acquired (Figure 2, top right). Similarly, the MS^2 peak with m/z 1024.48 represents a singly charged ion with the expected half IML modification. This peak could represent either the sulfenic acid-modified or the alkene-modified peptide, or it could represent a mixture of the two. Further CID of the m/z 1024.48 peak results in sequence data acquisition as well (Figure 2, lower right). The third peak in MS^2 , with m/z 595.75, represents a dead end-modified peptide that has lost the methoxy substituent during the first CID and has not undergone sulfoxide elimination. This dead end could be linked to the peptide on either end, so it is assumed the peak represents a mixture of both possible peptide linkages. When the m/z

595.75 peak is subjected to CID, the result is the expected products of sulfoxide fragmentation of the two differently-linked dead ends: one peak is the sulfenic acid modified peptide (1^+ with m/z 1024.48), and one is the alkene-modified peptide without a methoxy substituent (2^+ with m/z 496.73) (Figure 2, lower left). This data strongly supports the notion that the m/z 595.75 peak represents a mixture of both dead ends. With sulfoxide fragmentation occurring between MS² and MS³, sequence data could not be gathered for the m/z 595.75 peptides.

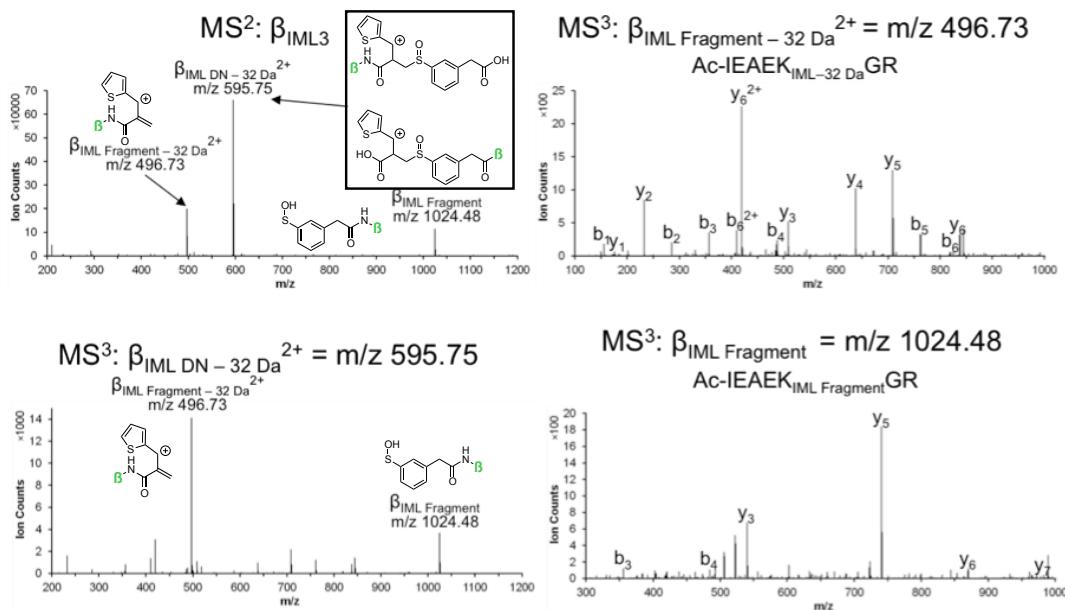
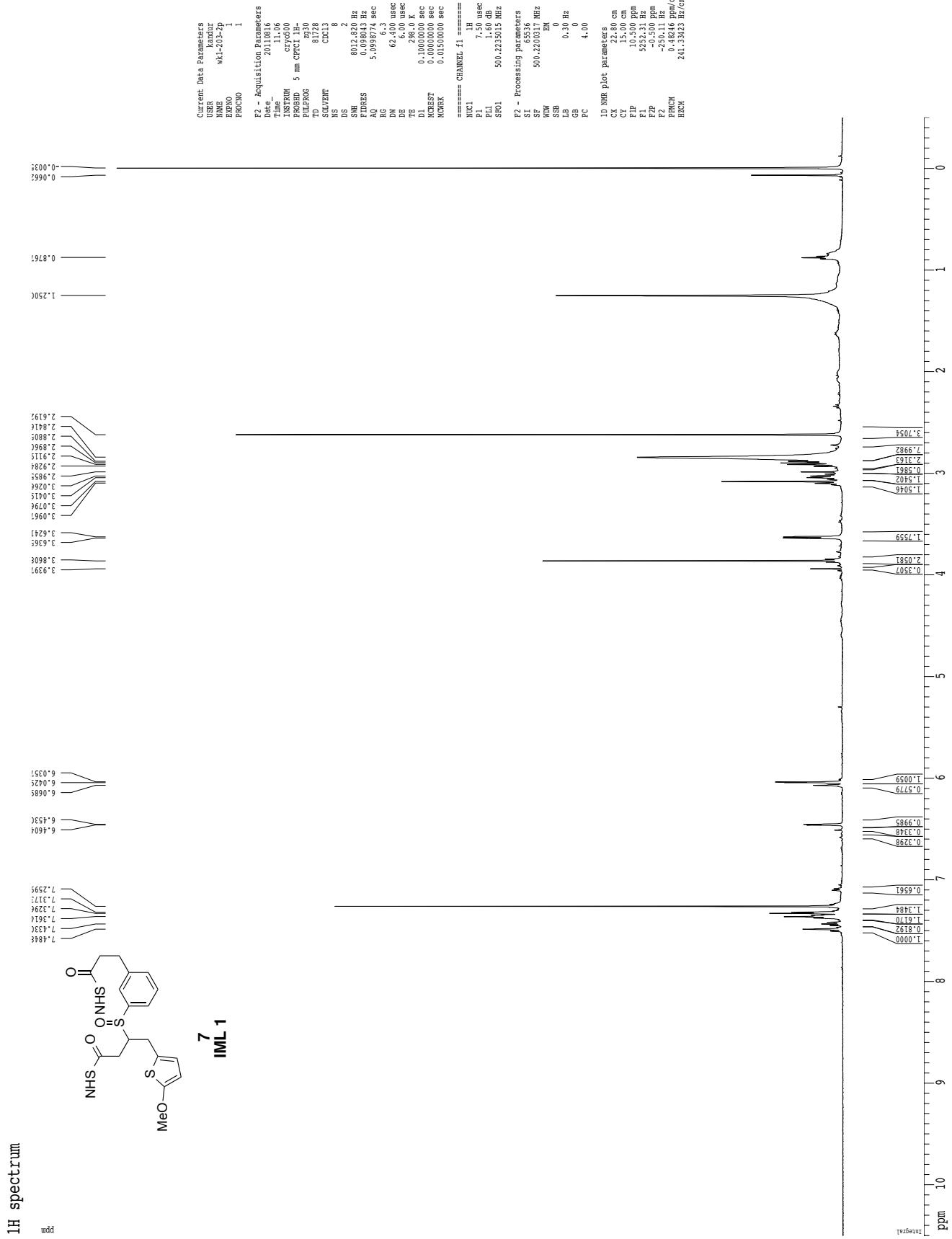
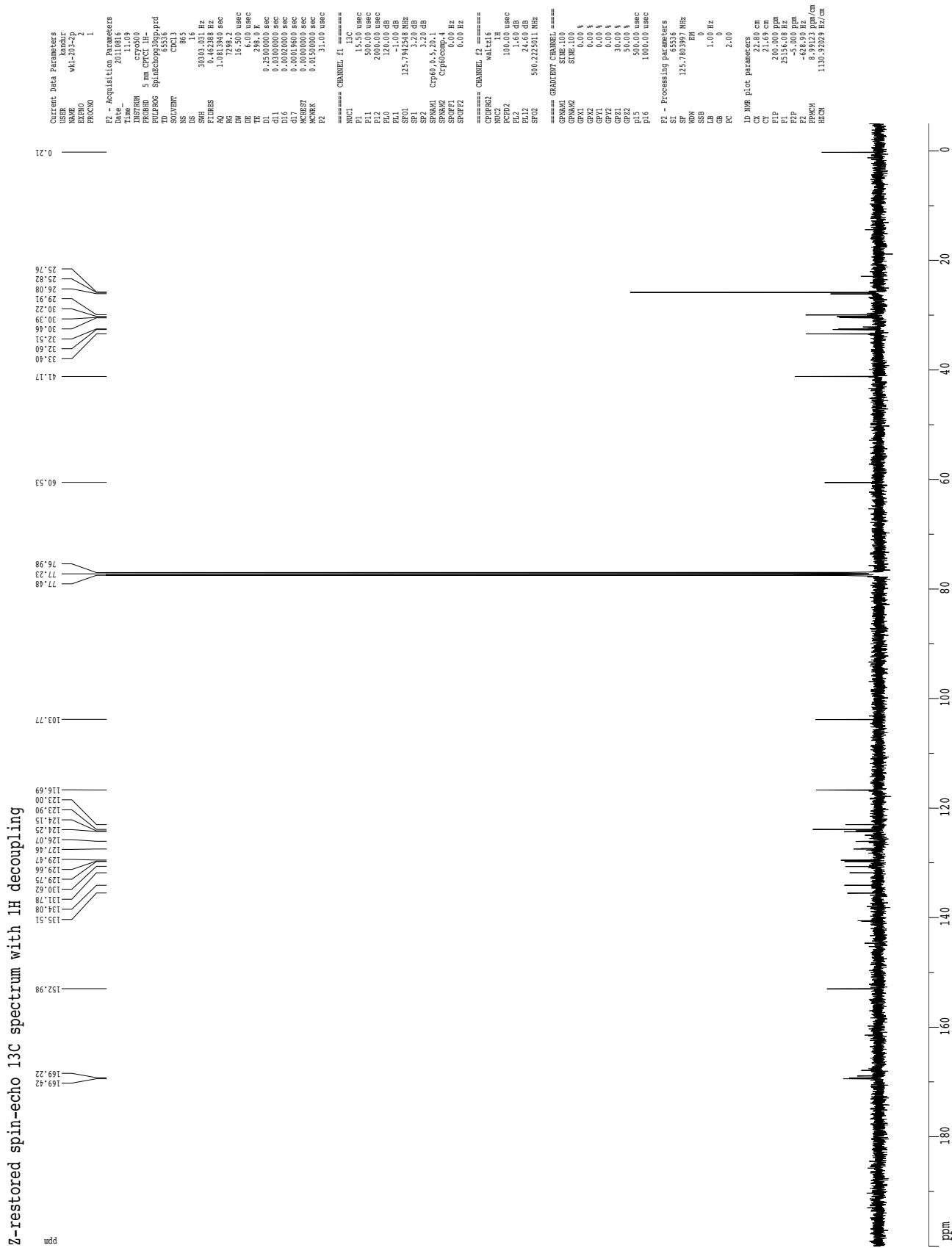


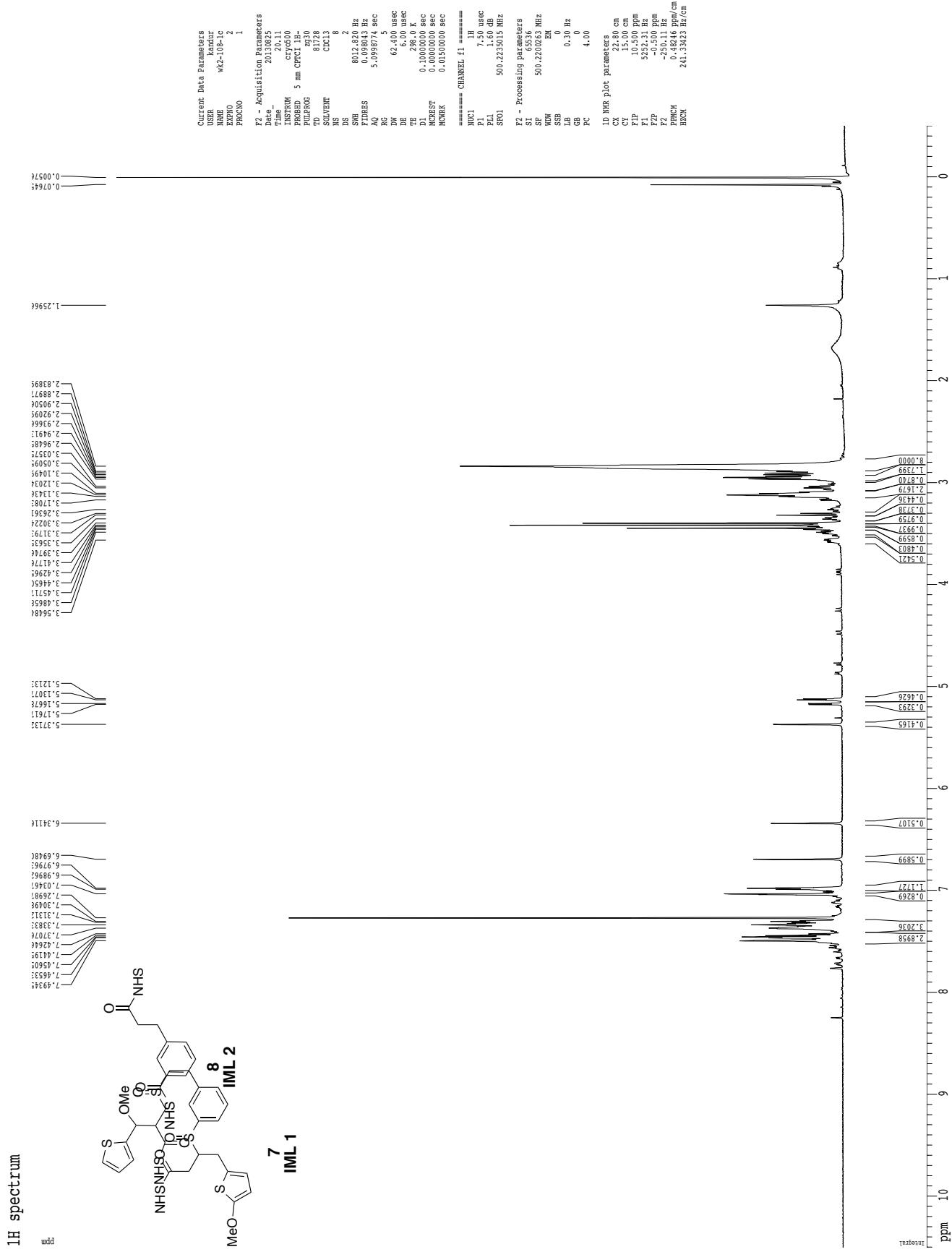
Figure 2. IML 2 Methoxy-loss in MS

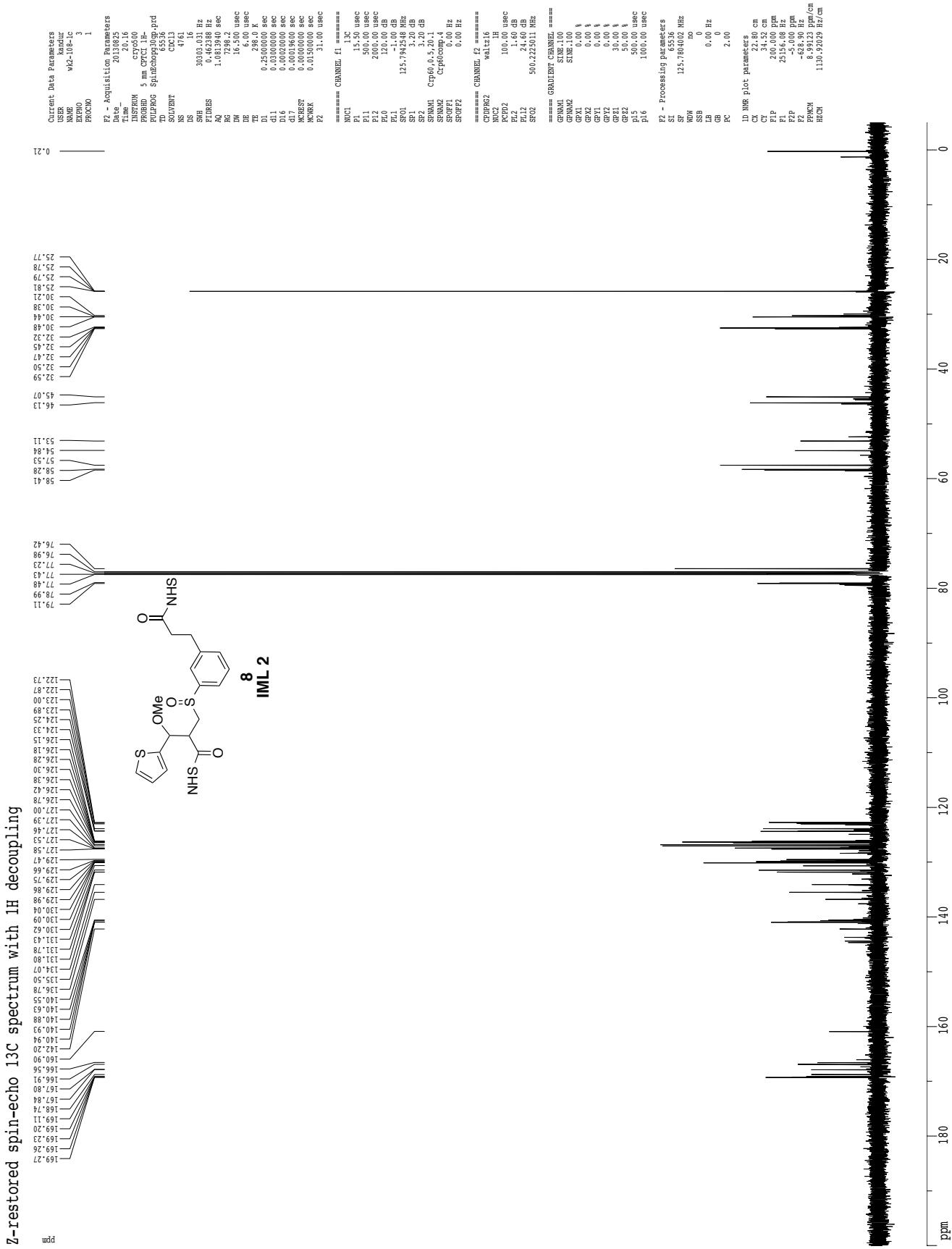
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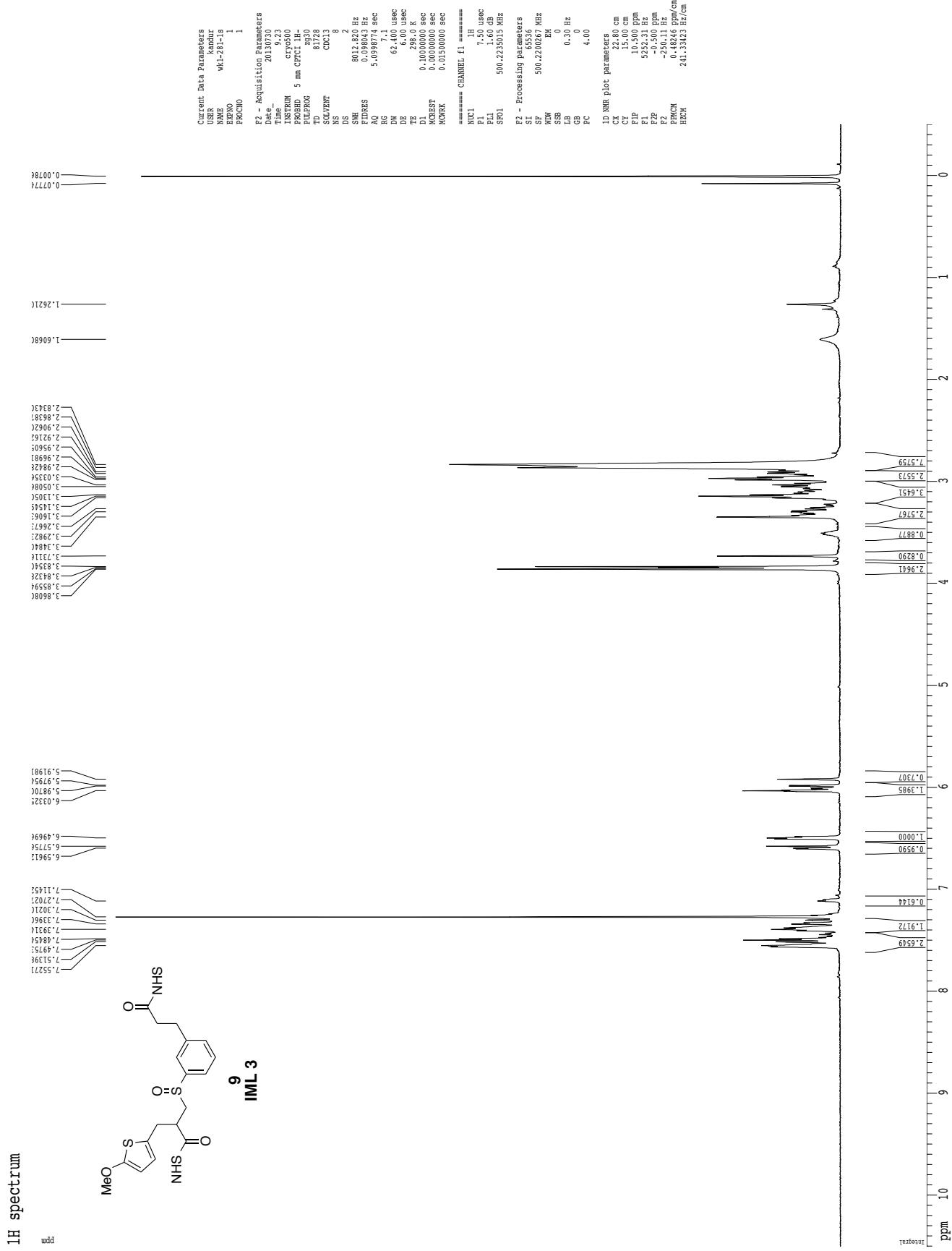
1. A. B. Pangborn, M. A. Giardello, R. H. Grubbs, R. K. Rosen, and F. J. Timmers, *Organometallics*, 1996, **15**, 1518–1520.
2. S. Gao, C. Tseng, C. H. Tsai, and C.-F. Yao, *Tetrahedron*, 2008, **64**, 1955–1961.
3. N. M. Leonard and J. Brunckova, *J. Org. Chem.*, 2011, **76**, 9169–9174.

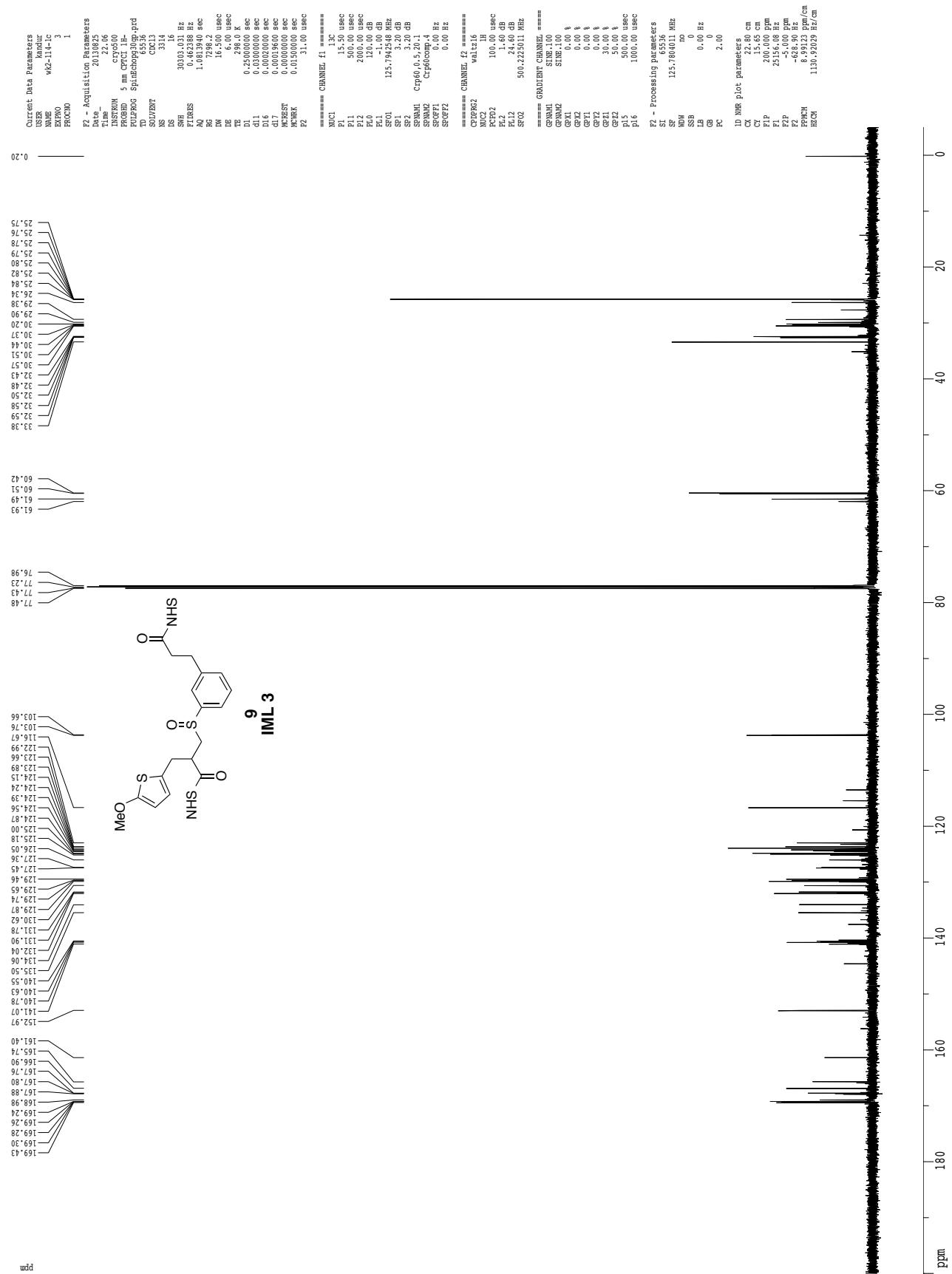


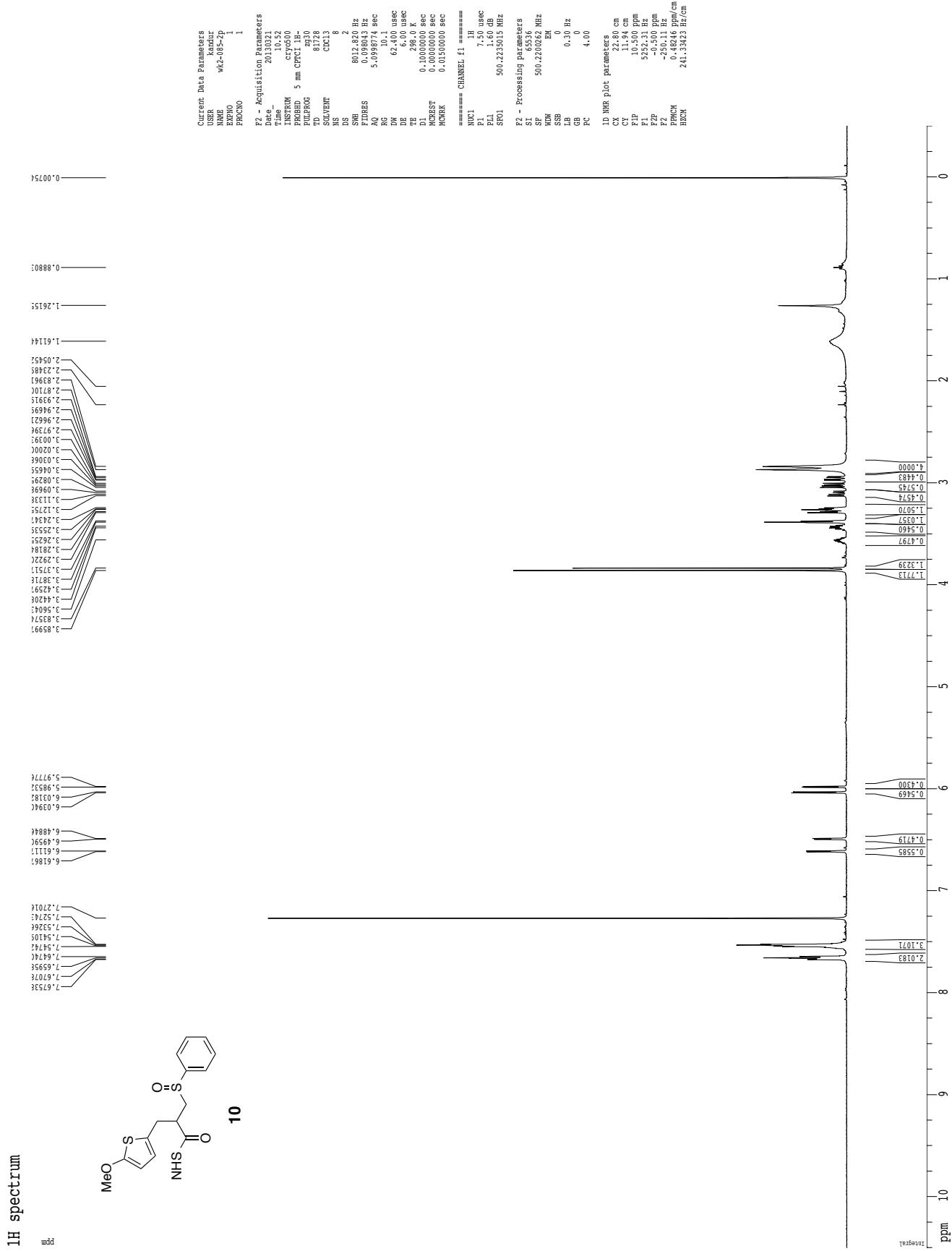


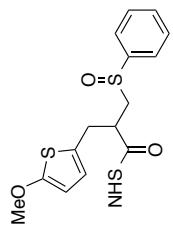
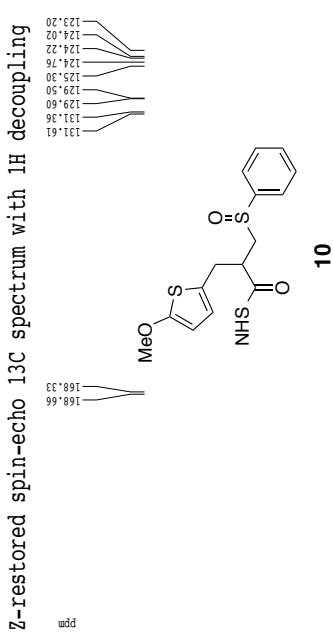




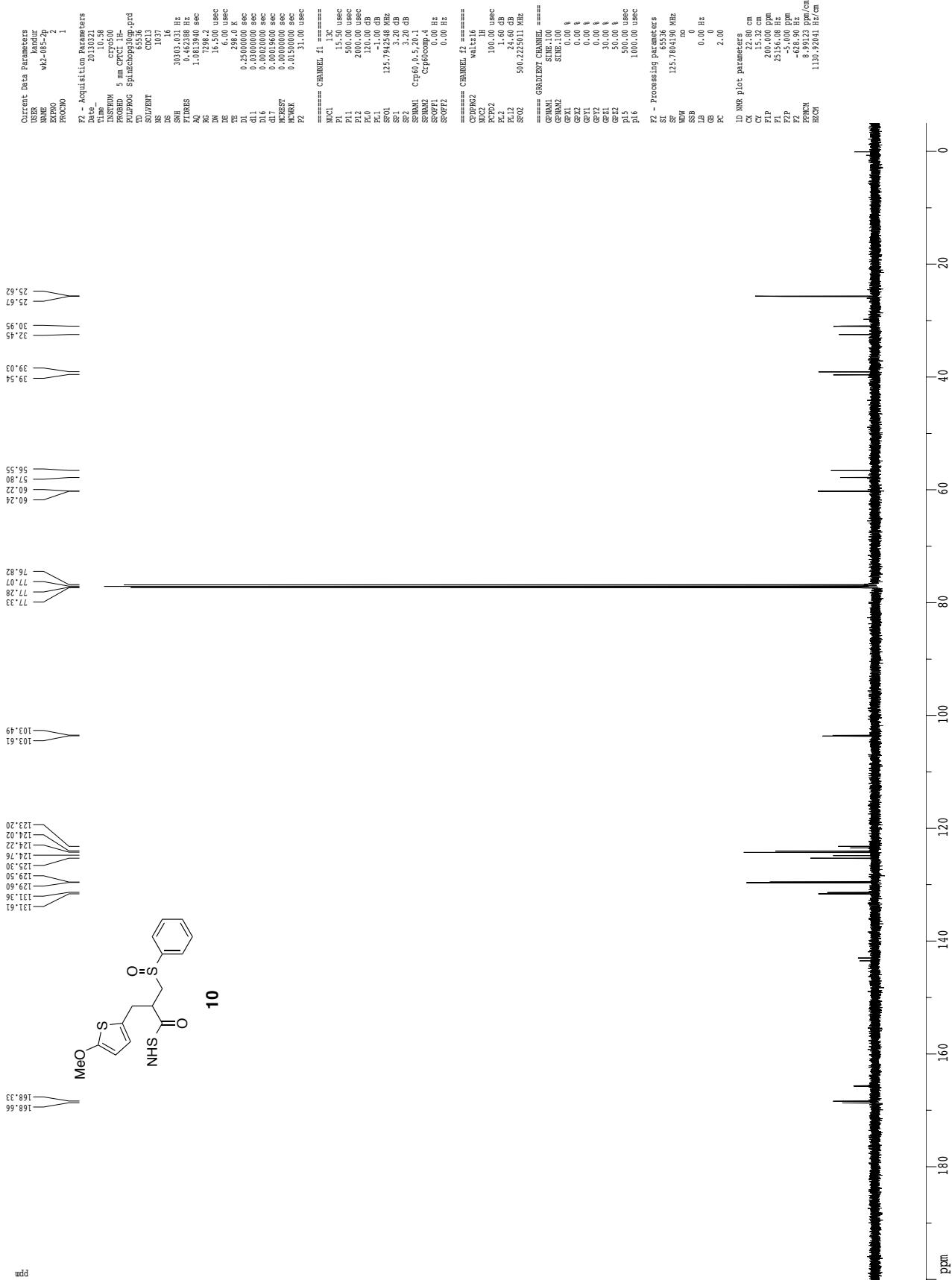


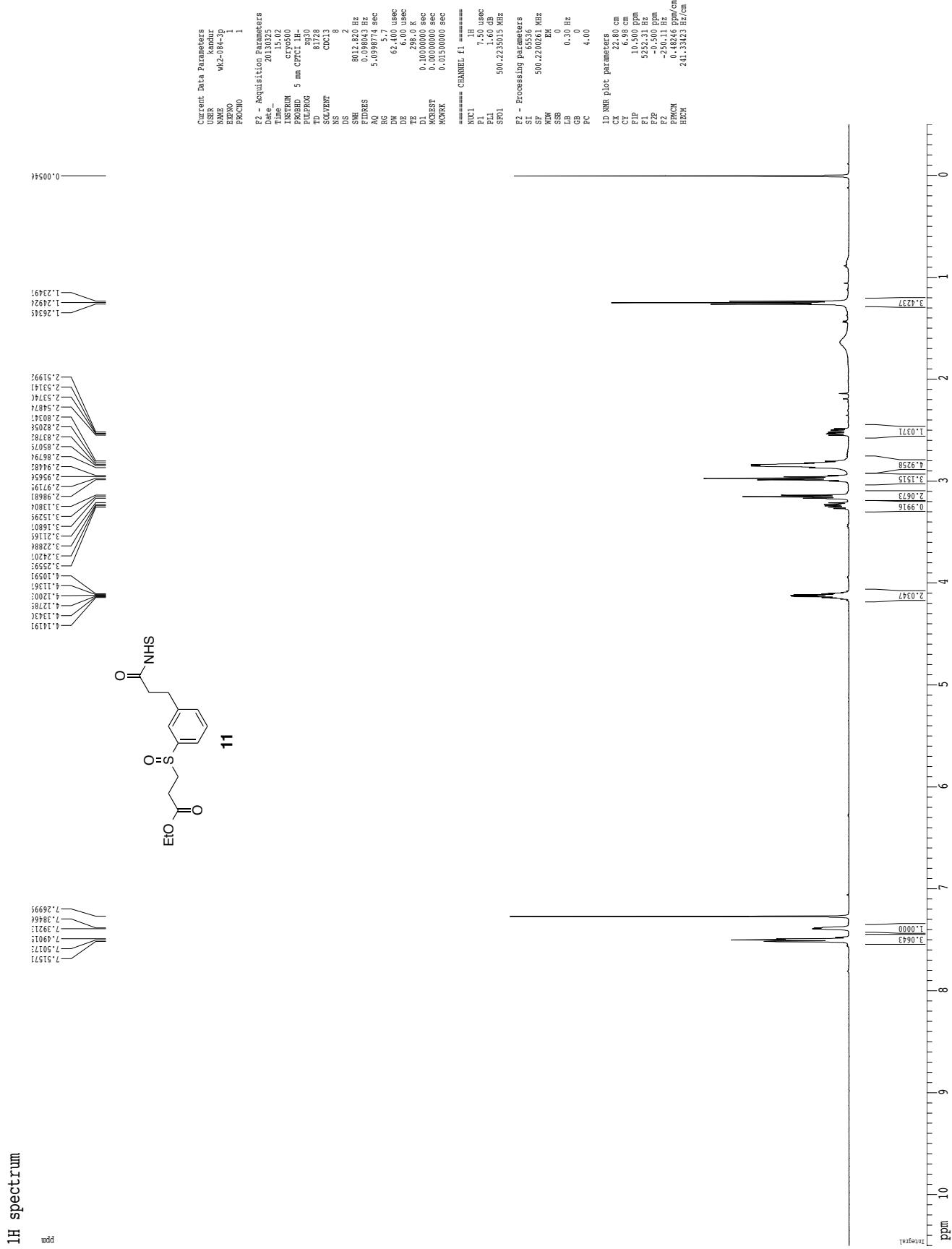
Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

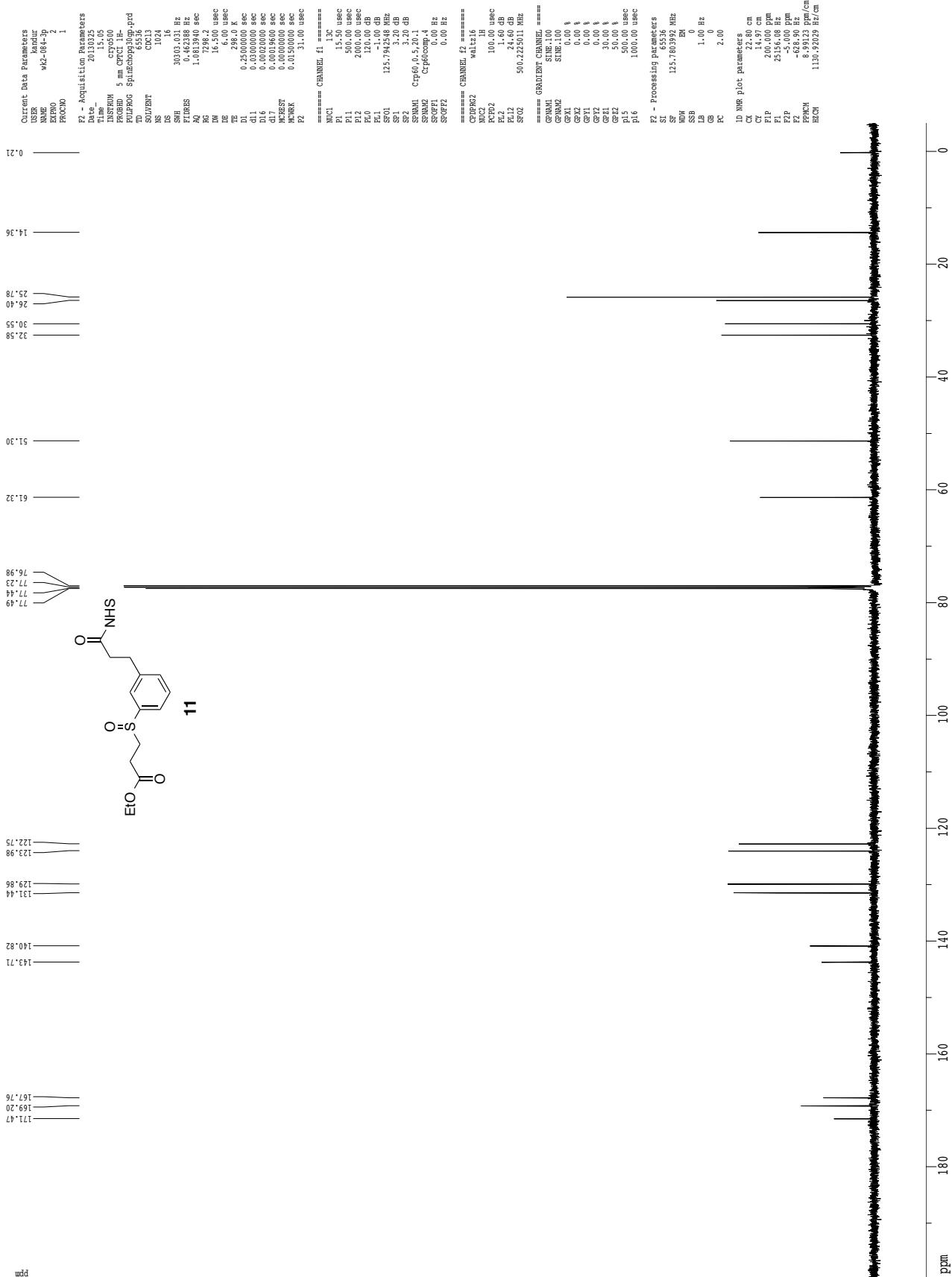
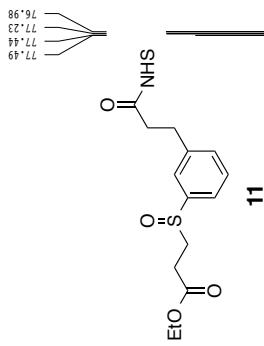


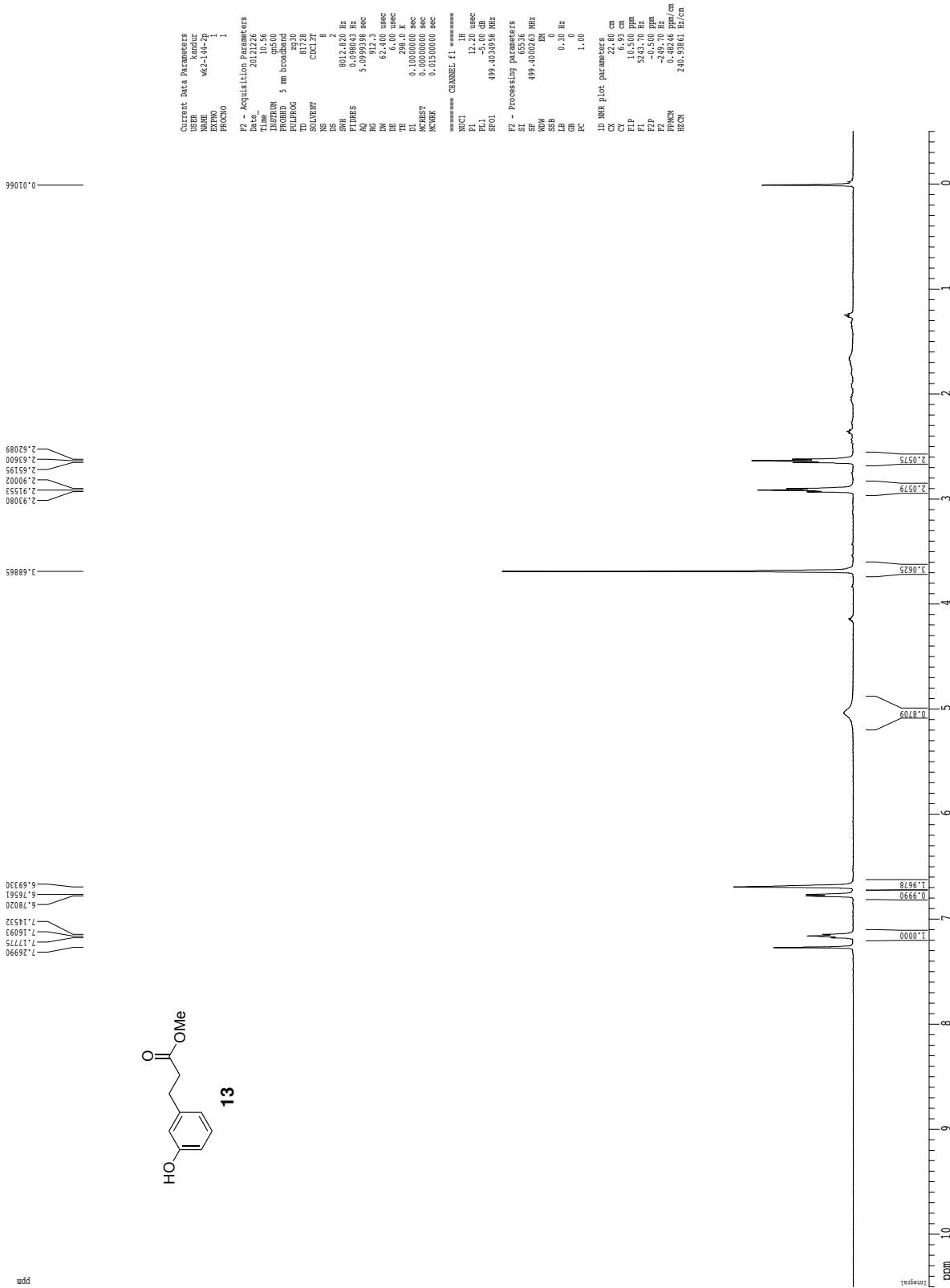


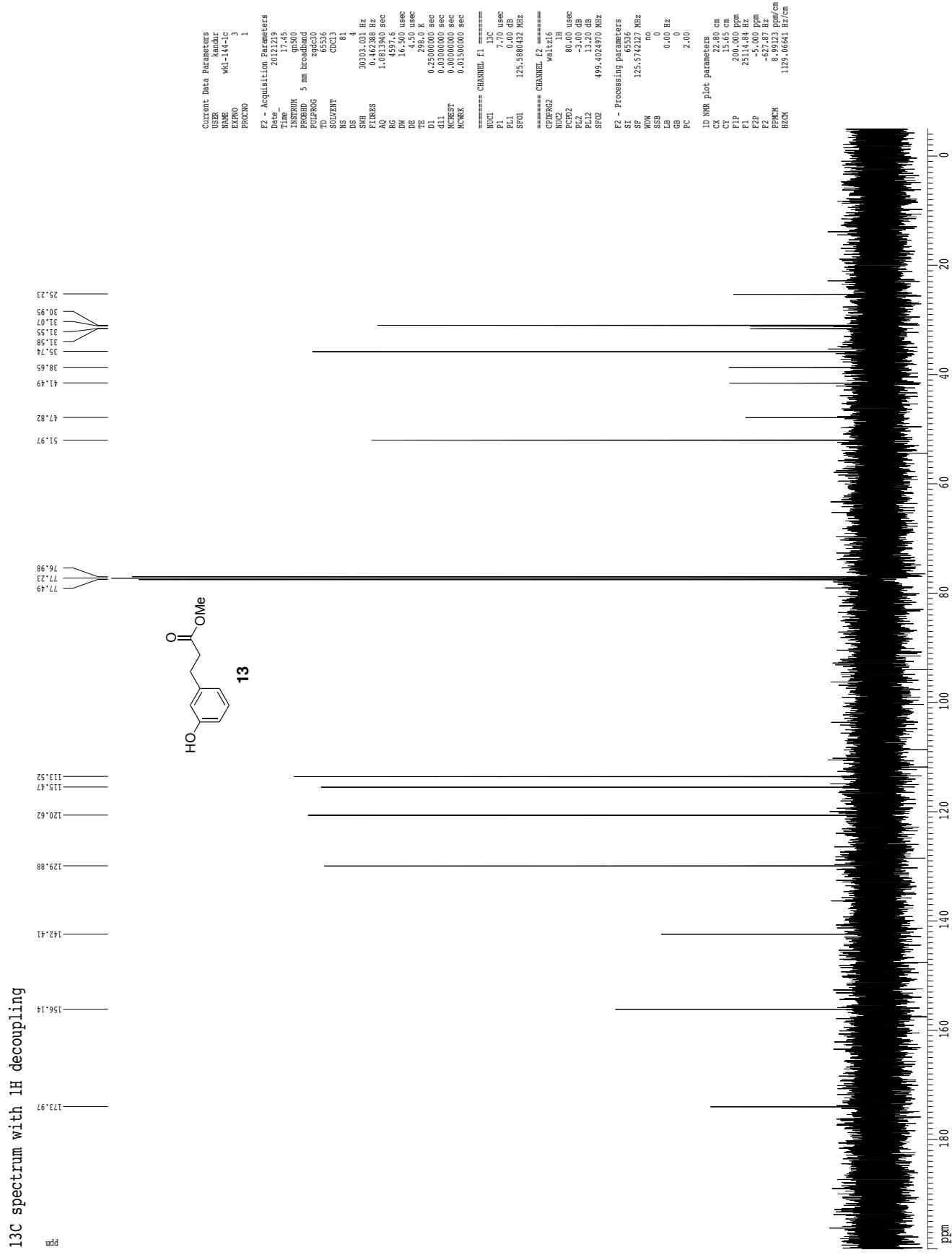
$\pi/2$ -restored spin-echo ^{13}C spectrum with 1H decoupling

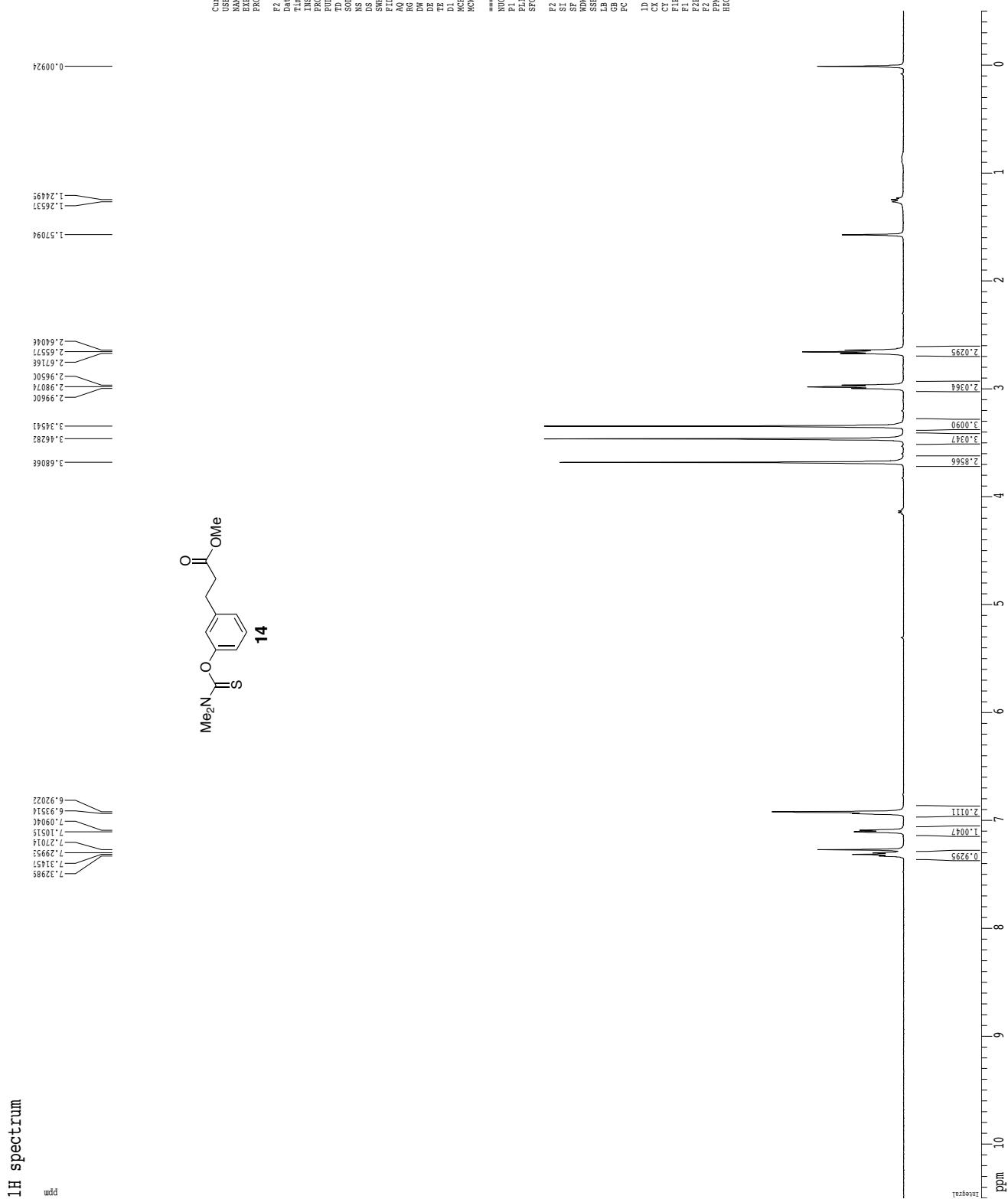


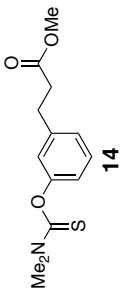




¹H spectrum







$\pi/2$ -restored spin-echo ^{13}C spectrum with 1H decoupling



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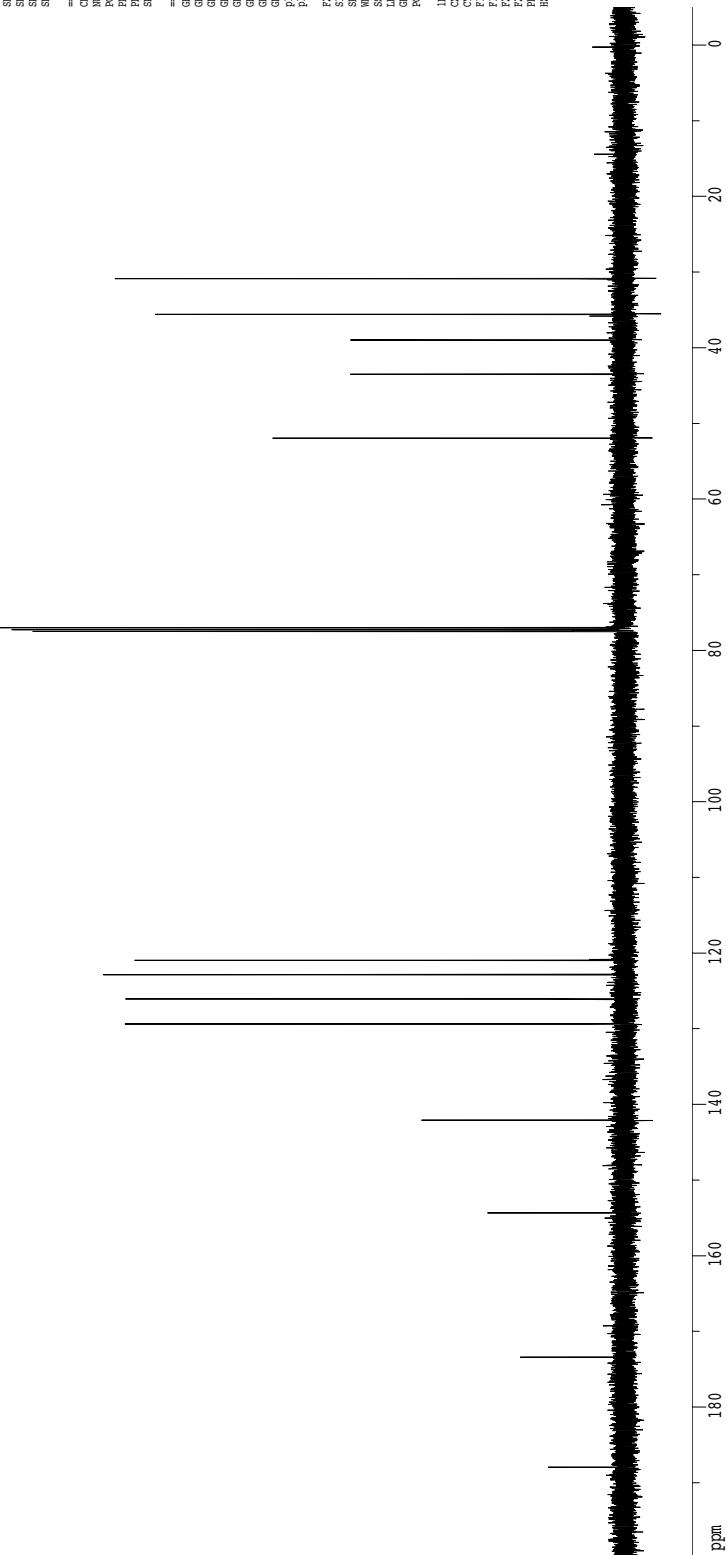
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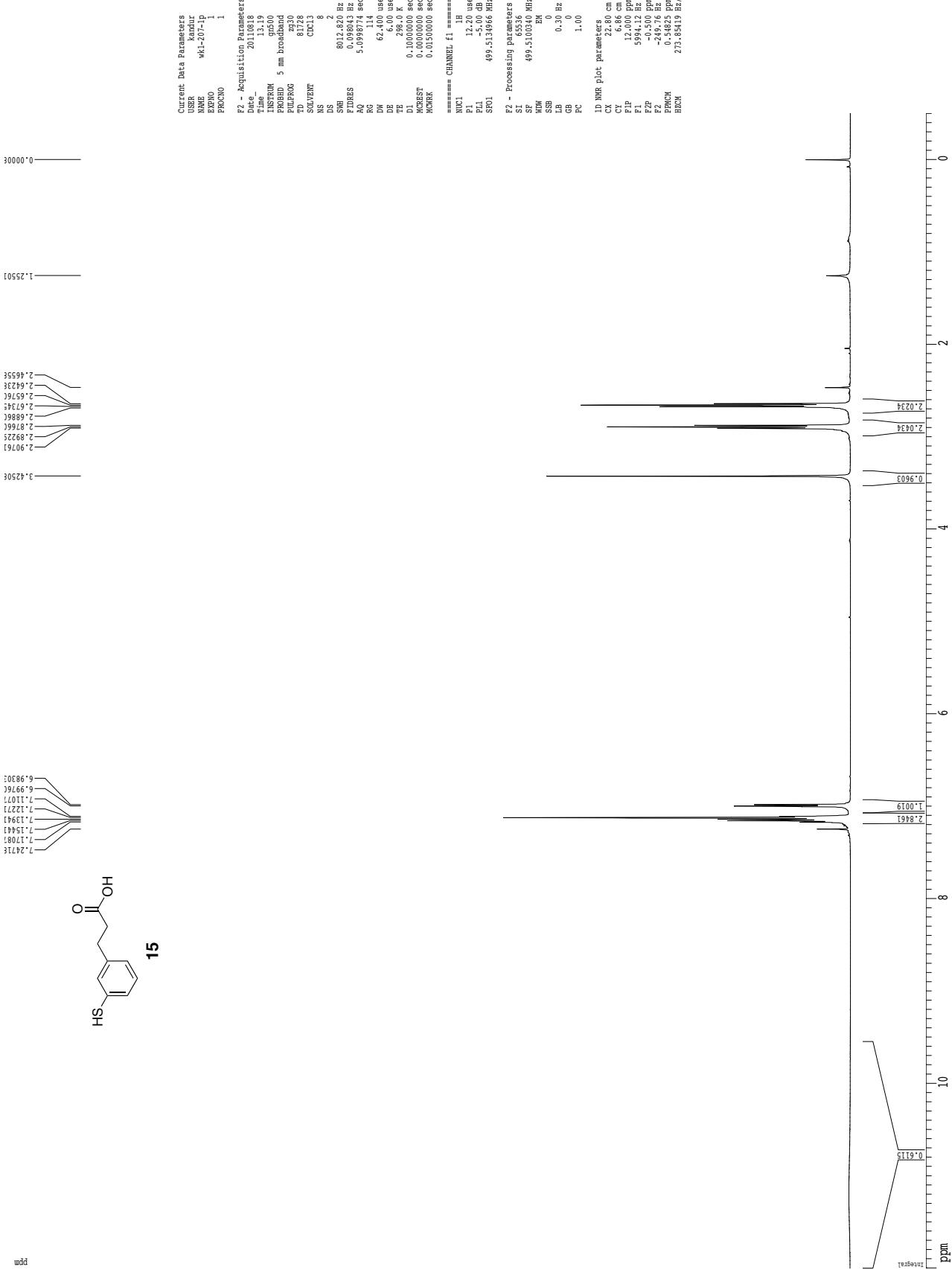
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		MICROST				

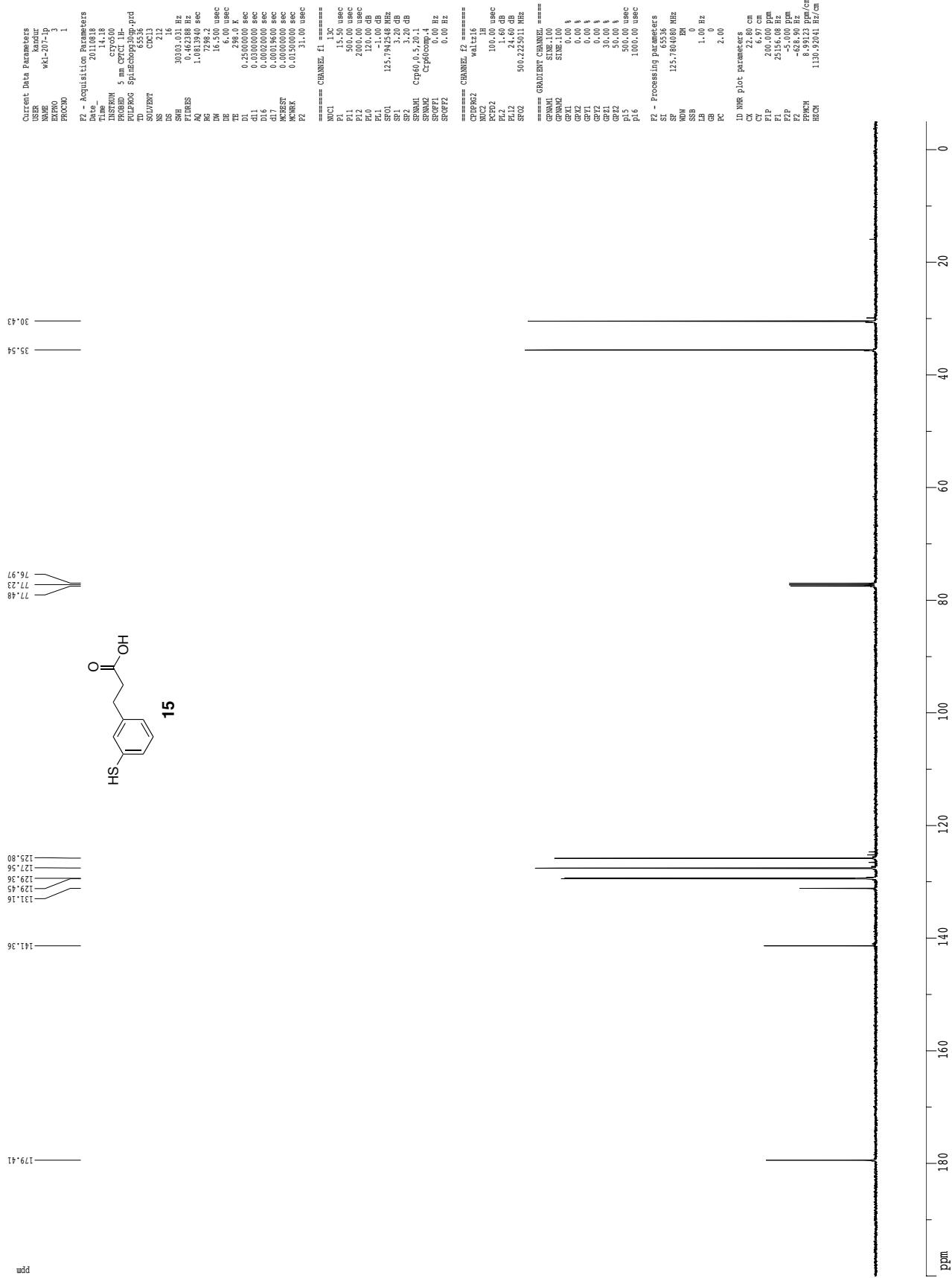
```
===== CHANNEL f1 =====
      NUC1          1.3C
      P1           15.50 usec
      P1.1         500.00 usec
      P1.2        1200.00 usec
      P1.0          11.00 db
      S1           125.194548 MHz
      S2            3.20 dB
      S2.1          3.20 dB
      S2.2          20.11
      SFRM1        CTFD0.0, 5.4
      SFRM2        CTFD0.0, 4.4
      SFOP1         0.00 dB
      SFOP2         0.00 dB
      SFOP3         0.00 Hz

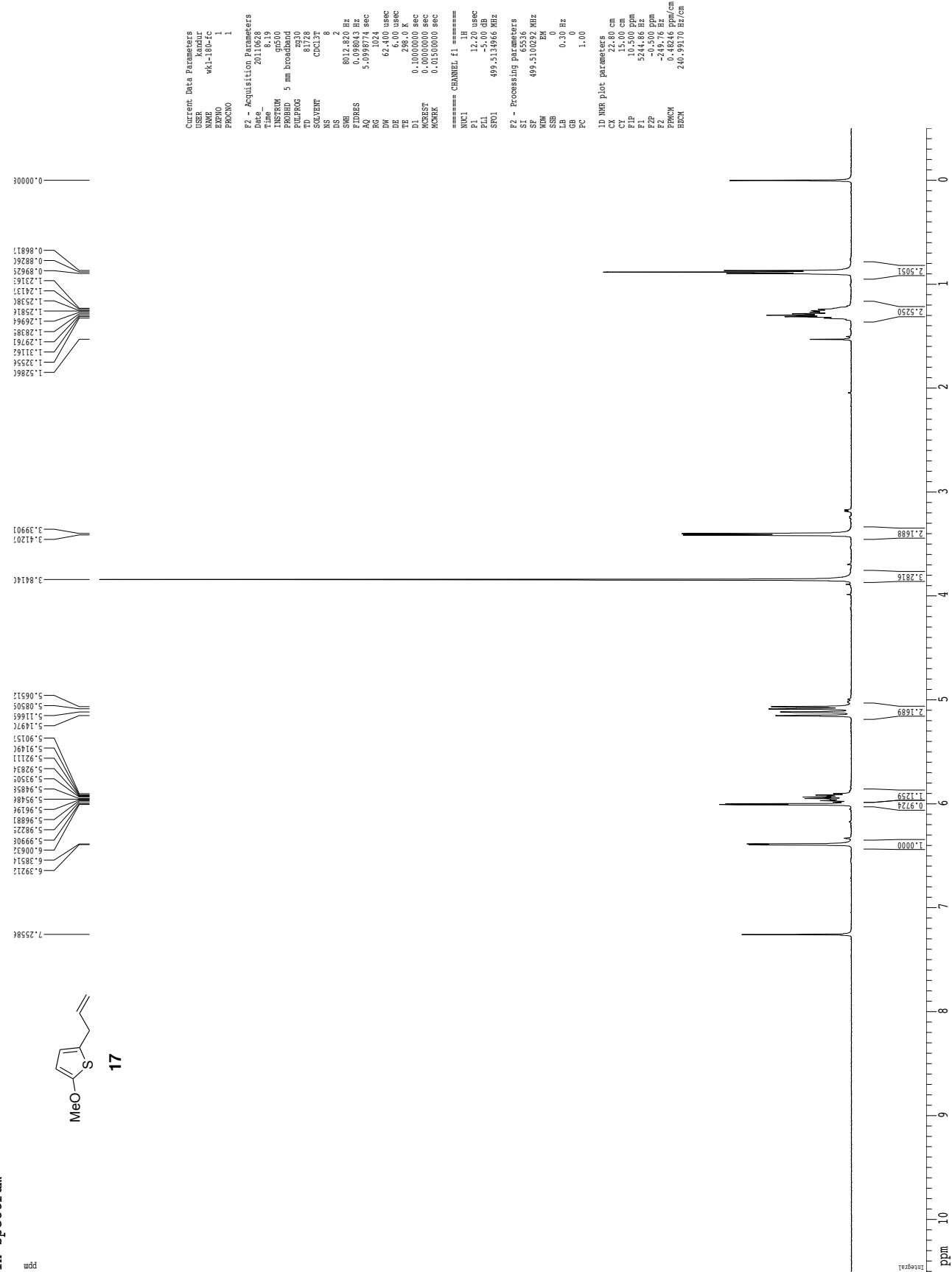
===== CHANNEL f2 =====
      GDFP62
      waltz16
      NUC2          1.18
      L1
```

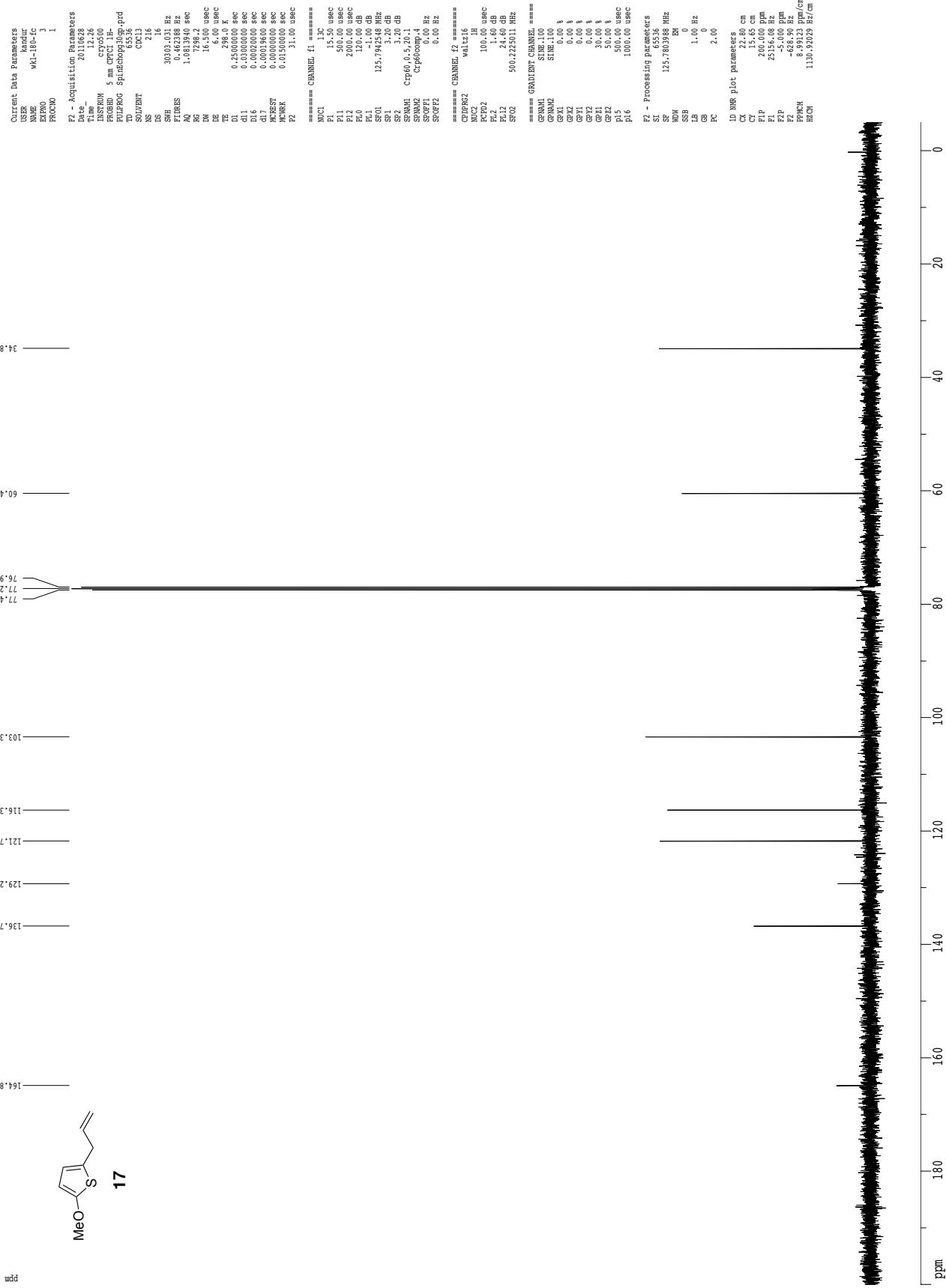
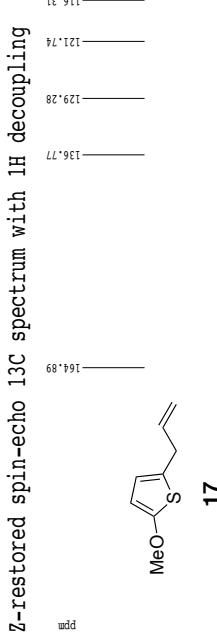
PL2 1.60 dB
PL12 24.60 dB
SF02 500.2225011 MHz

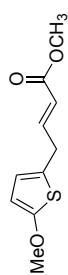


¹H spectrum

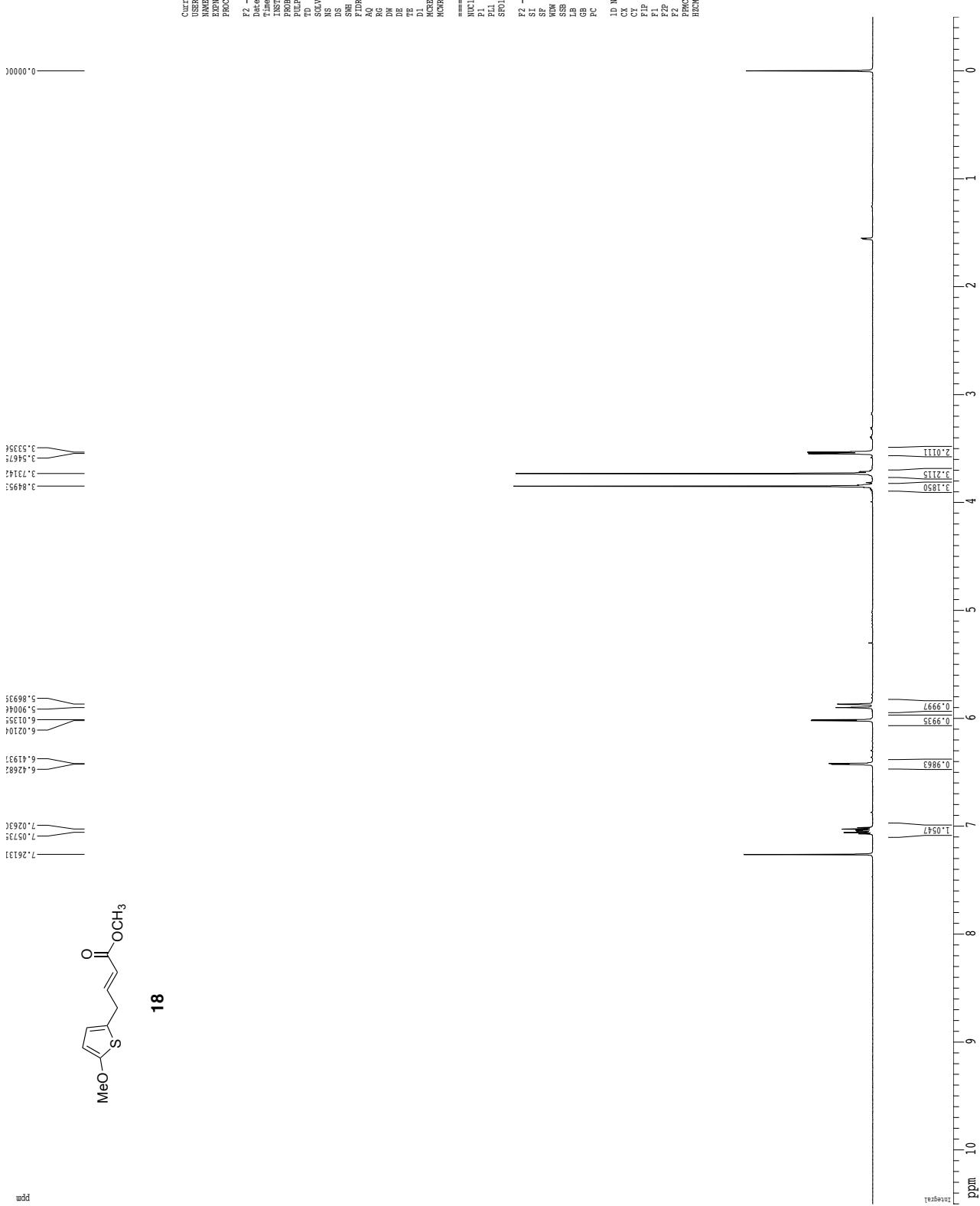
Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

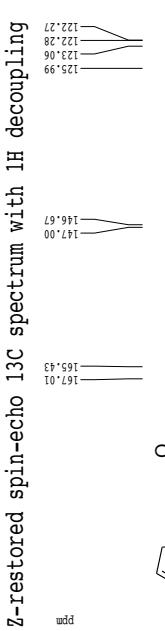




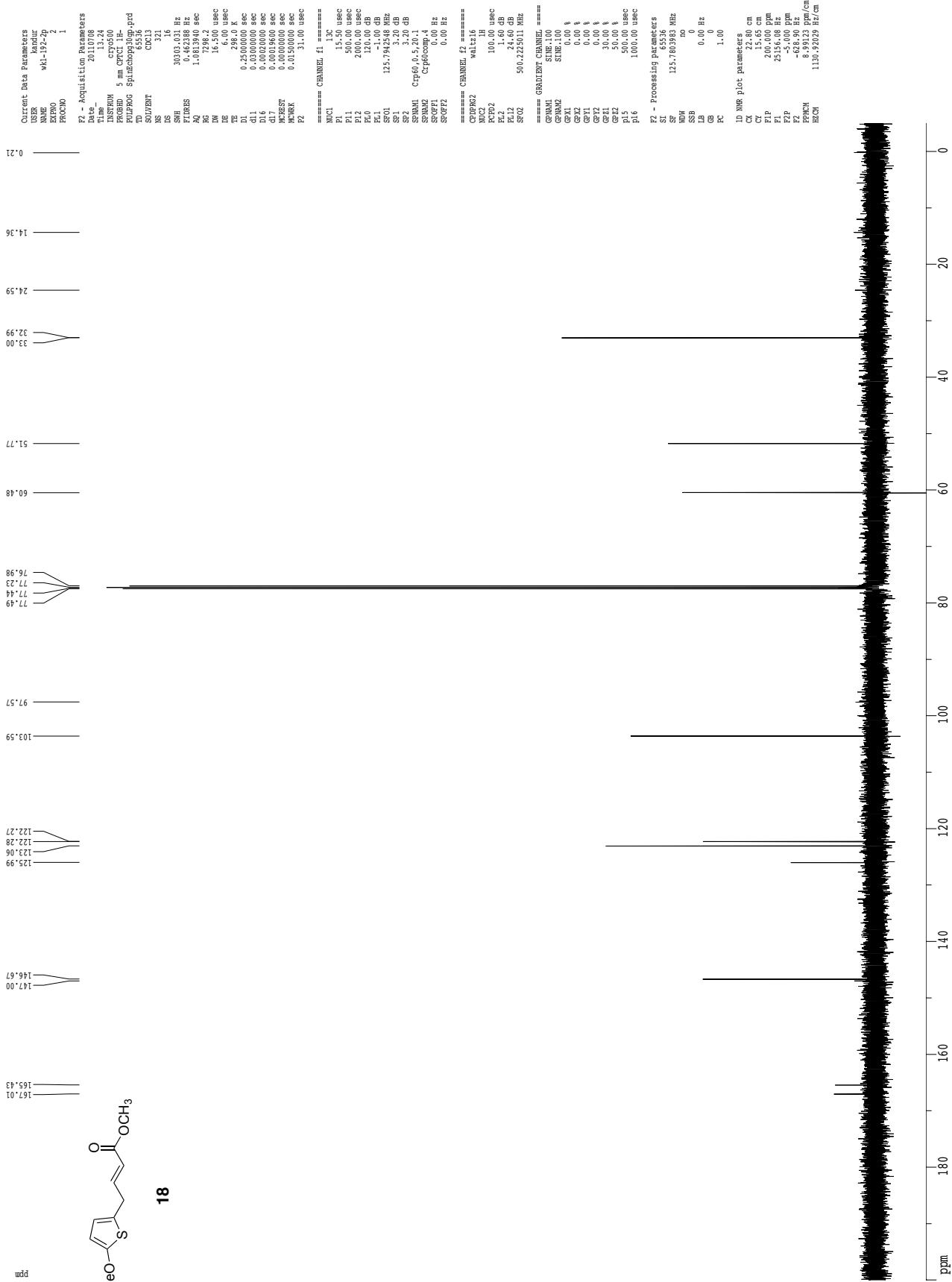


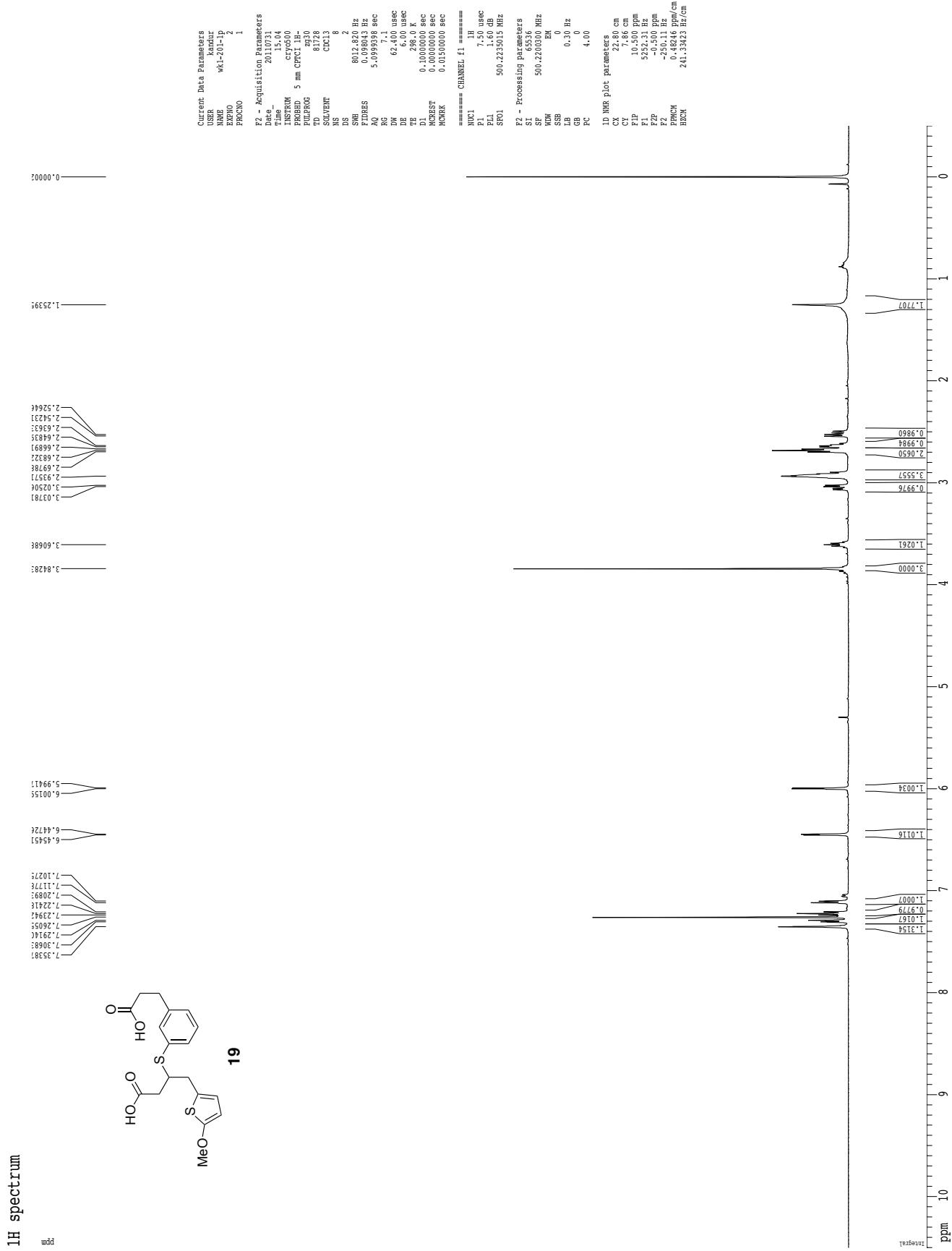
1H spectrum

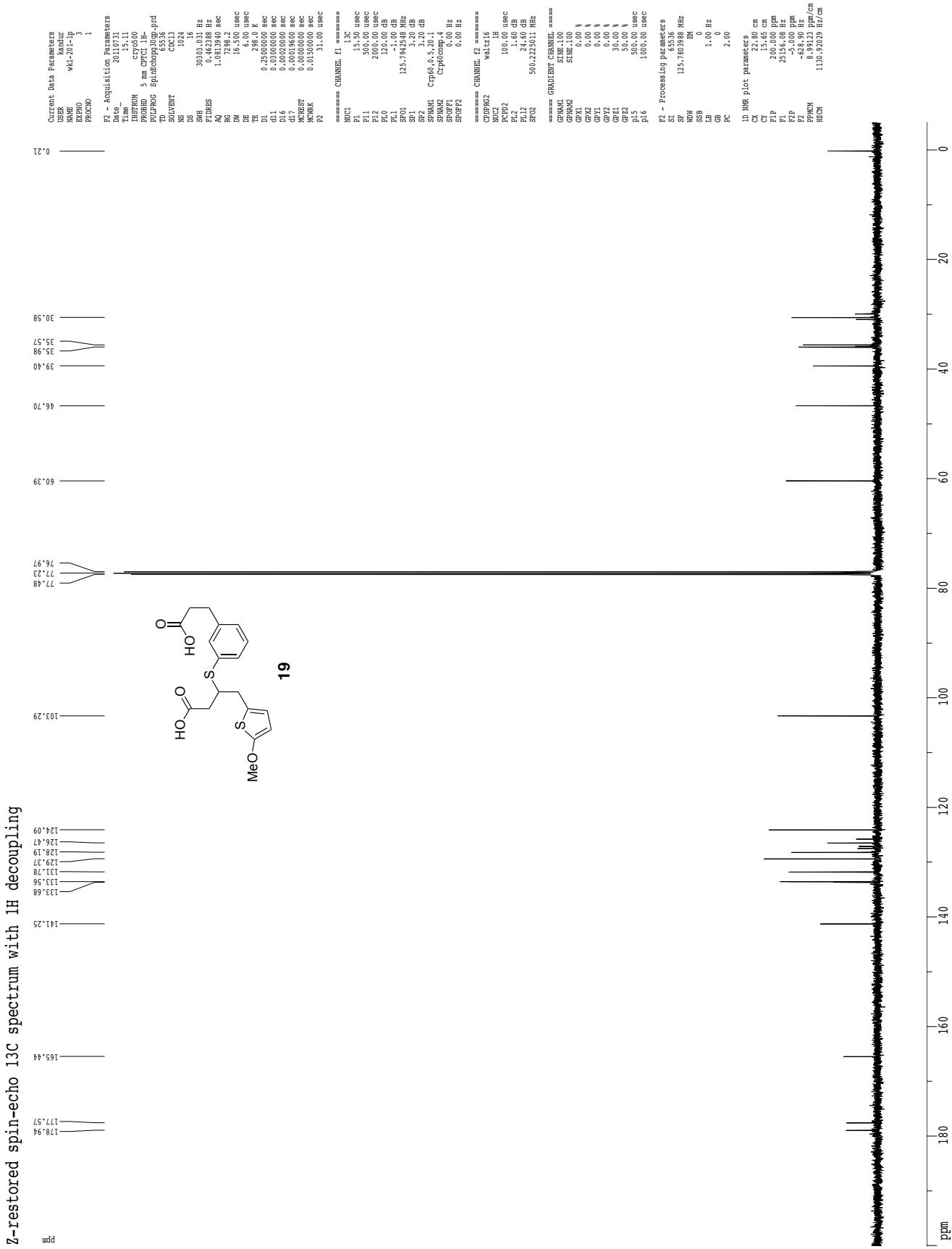


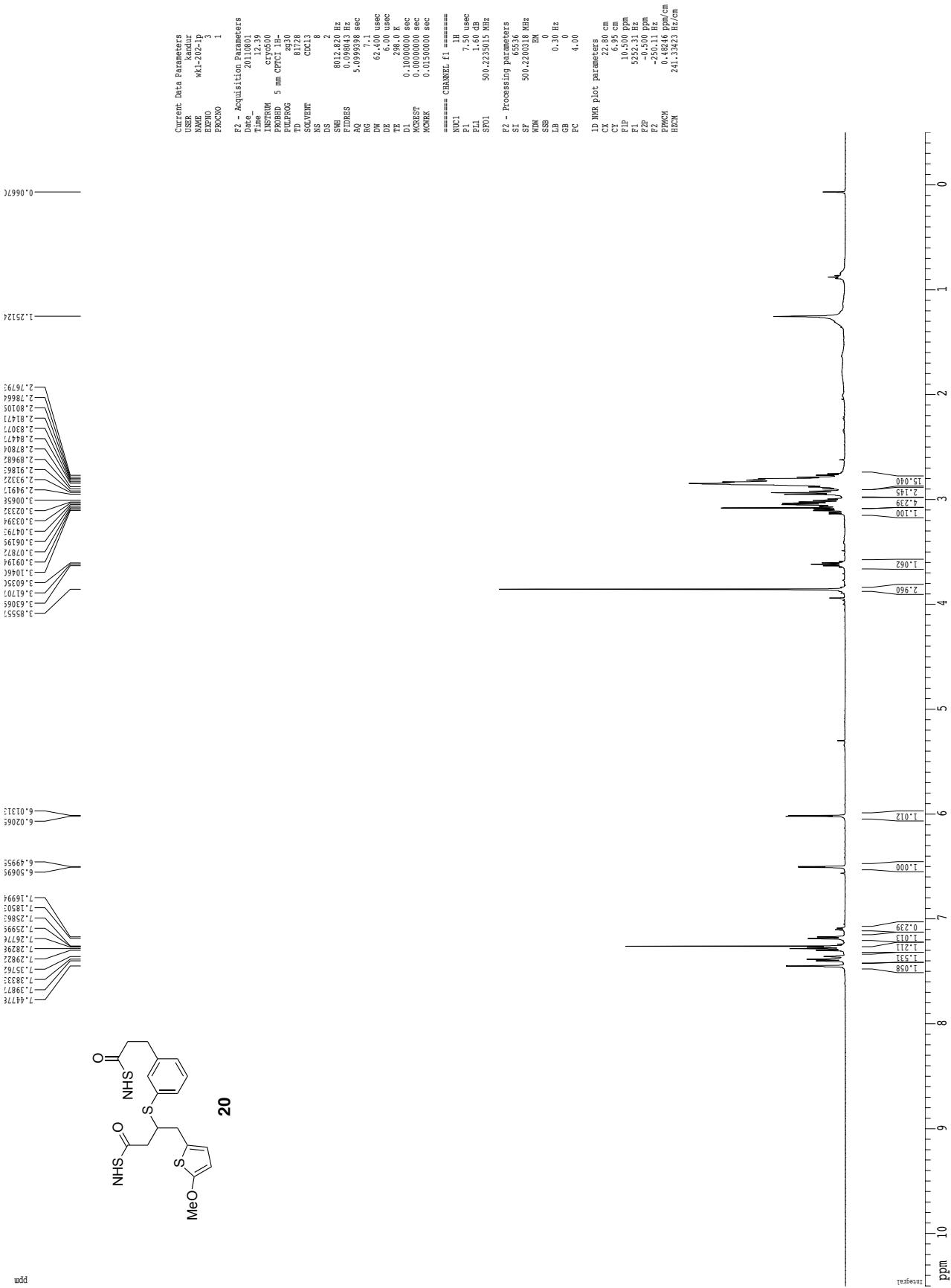


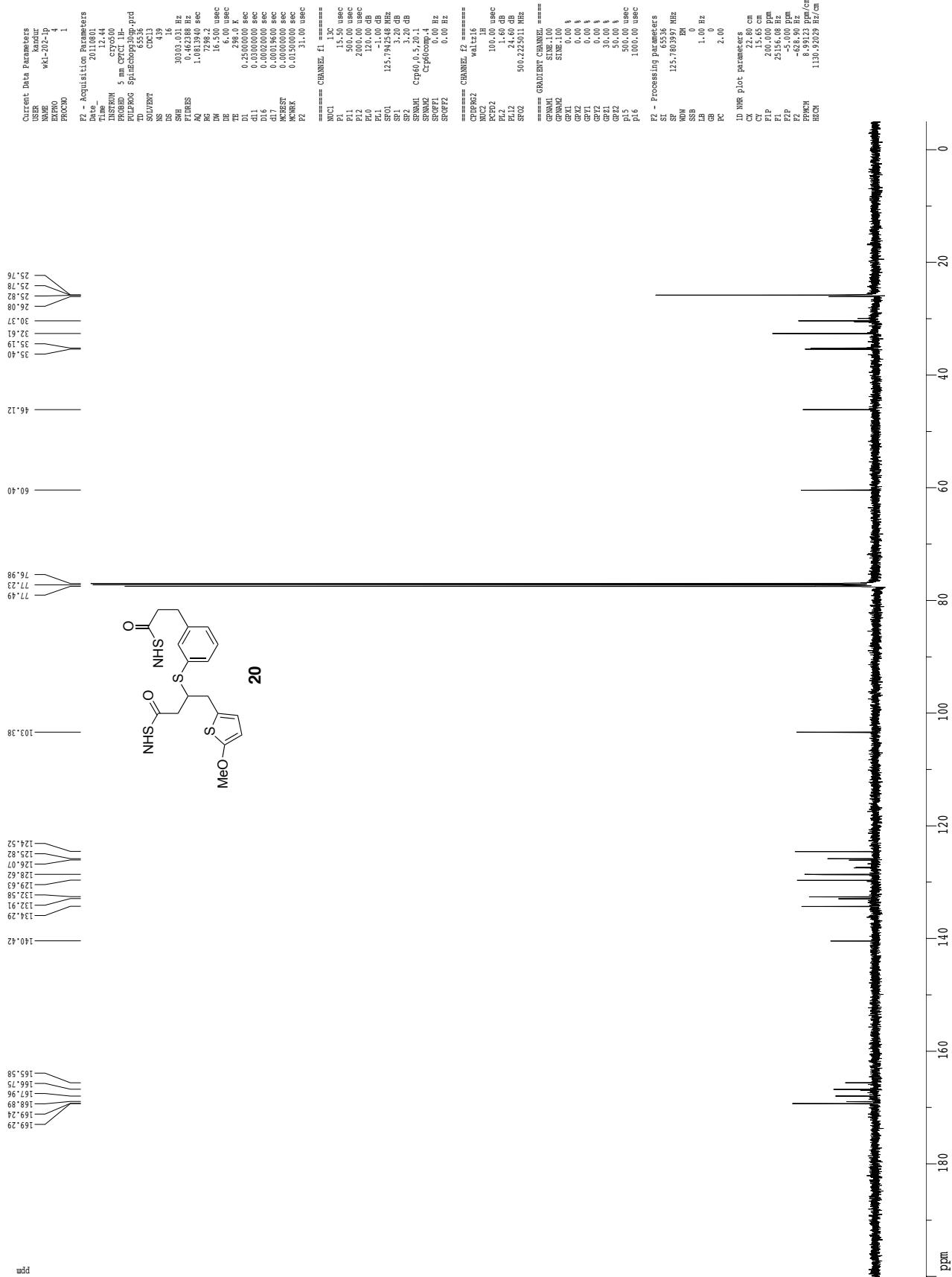
τ -restored spin-echo ^{13}C spectrum with 1H decoupling

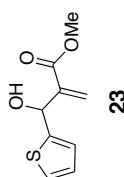




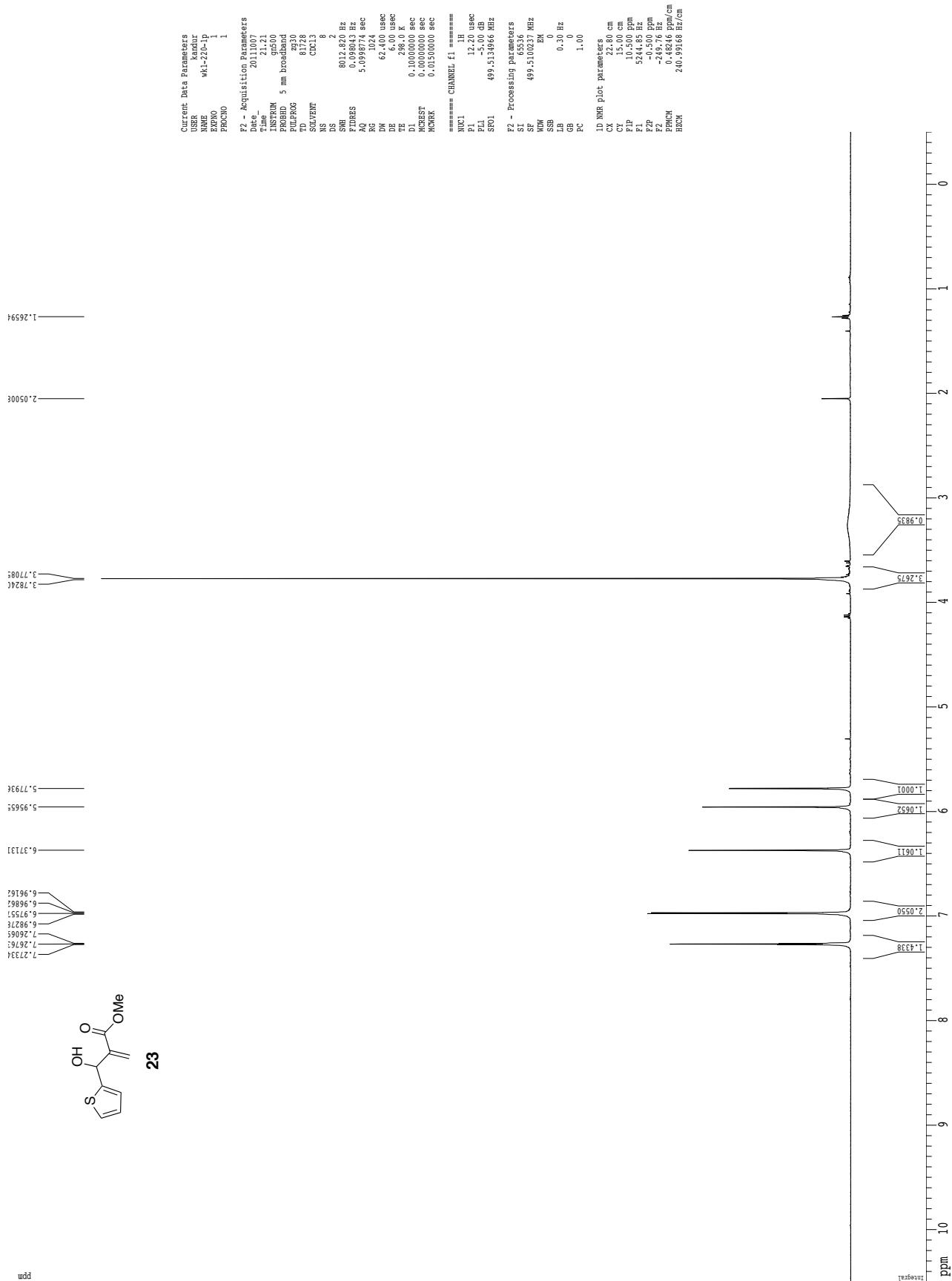


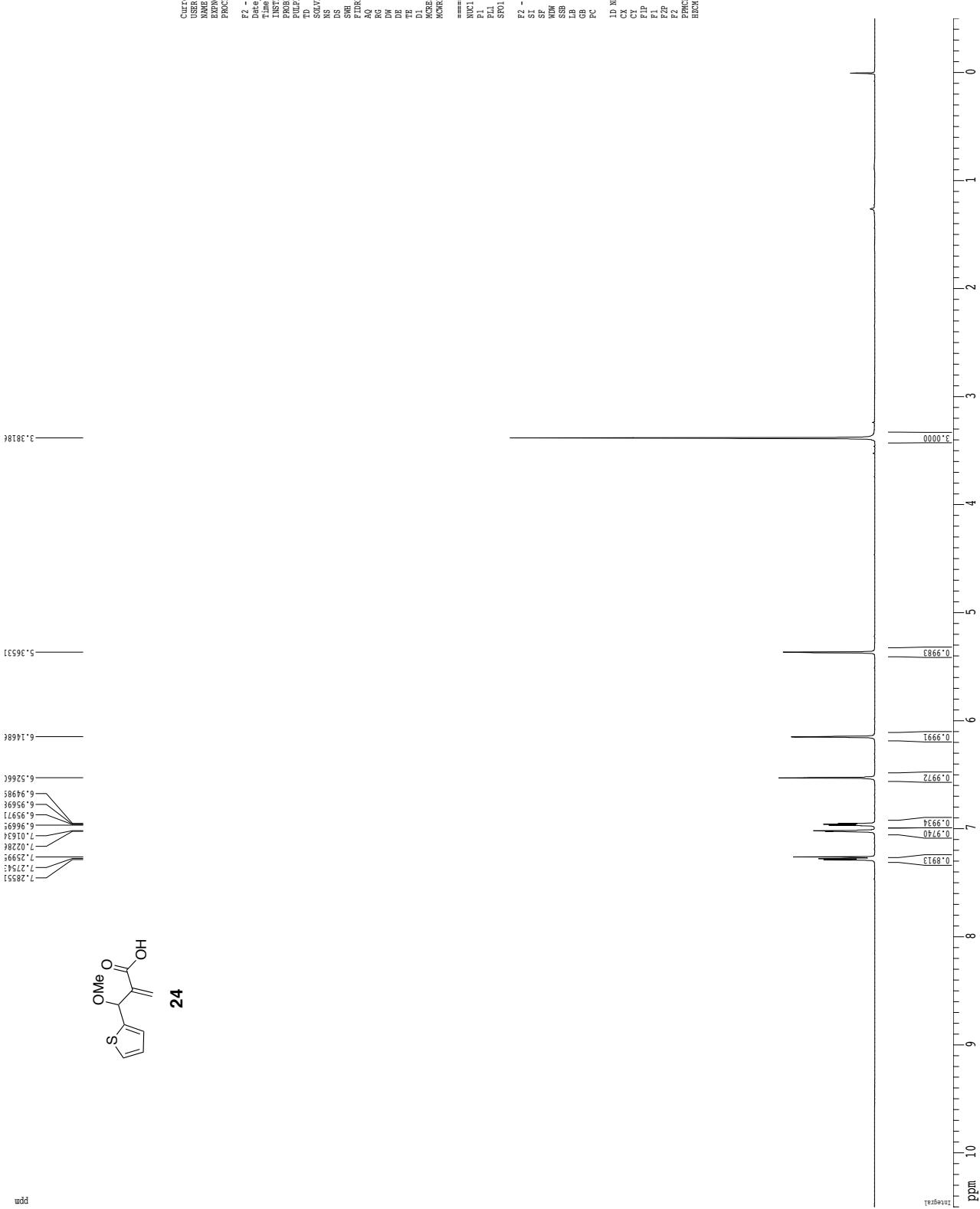
¹H Spectrum

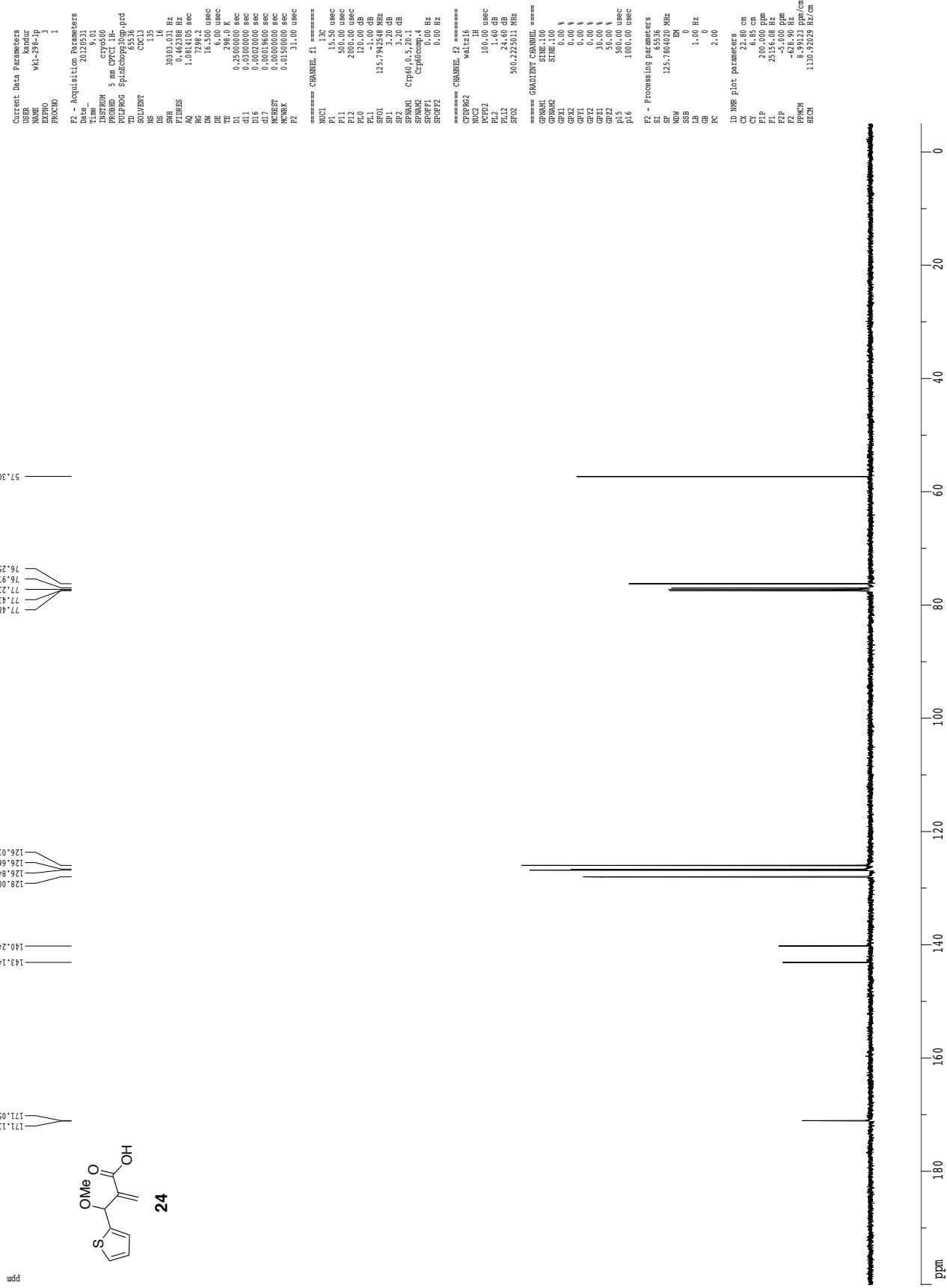
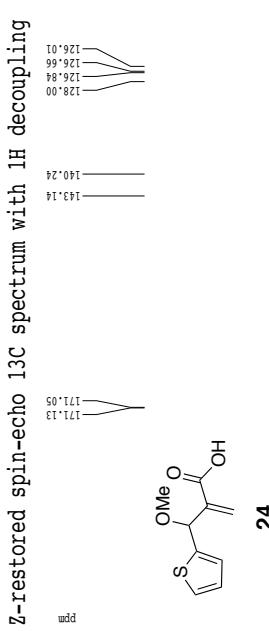
Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

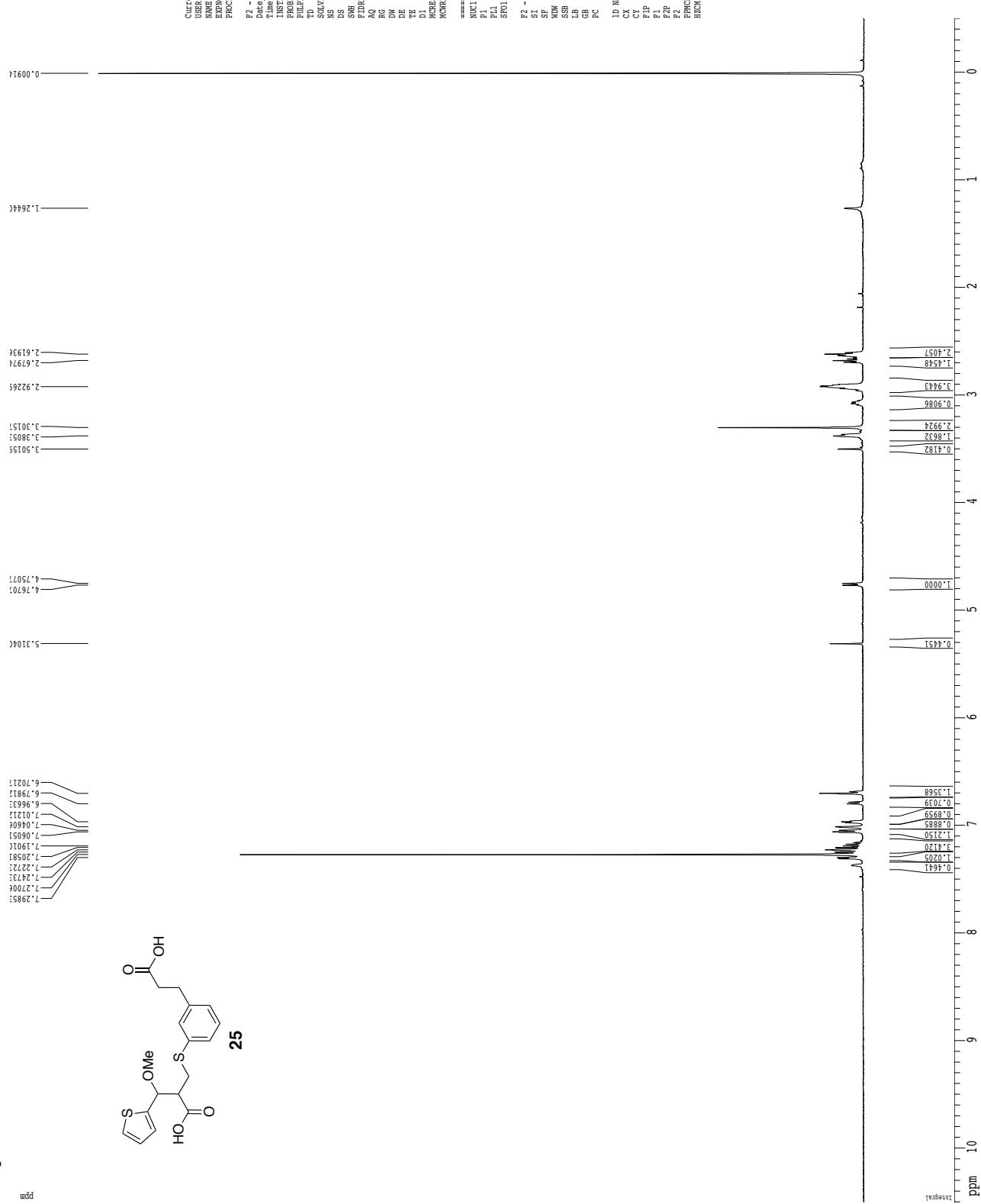


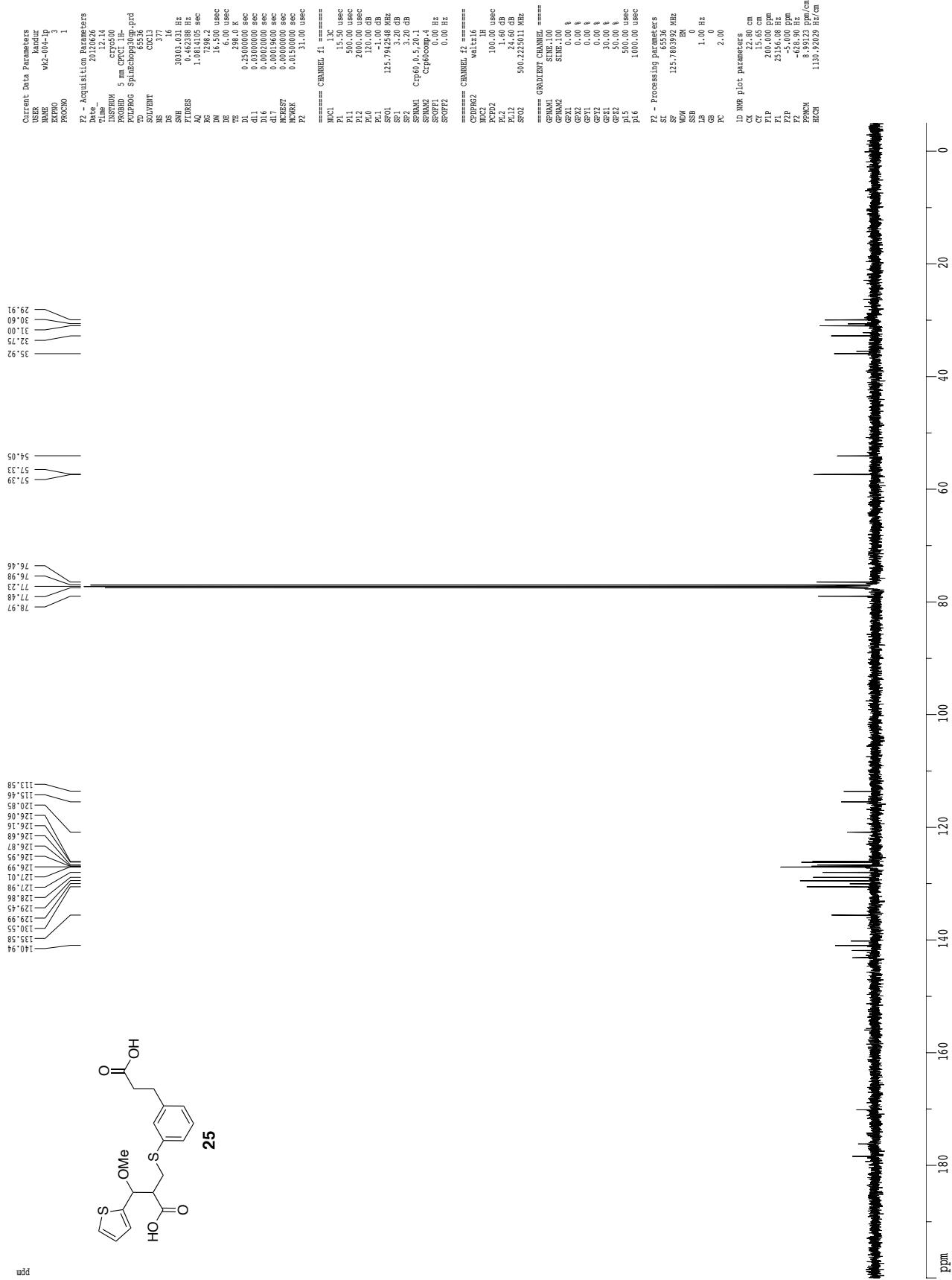
1H spectrum

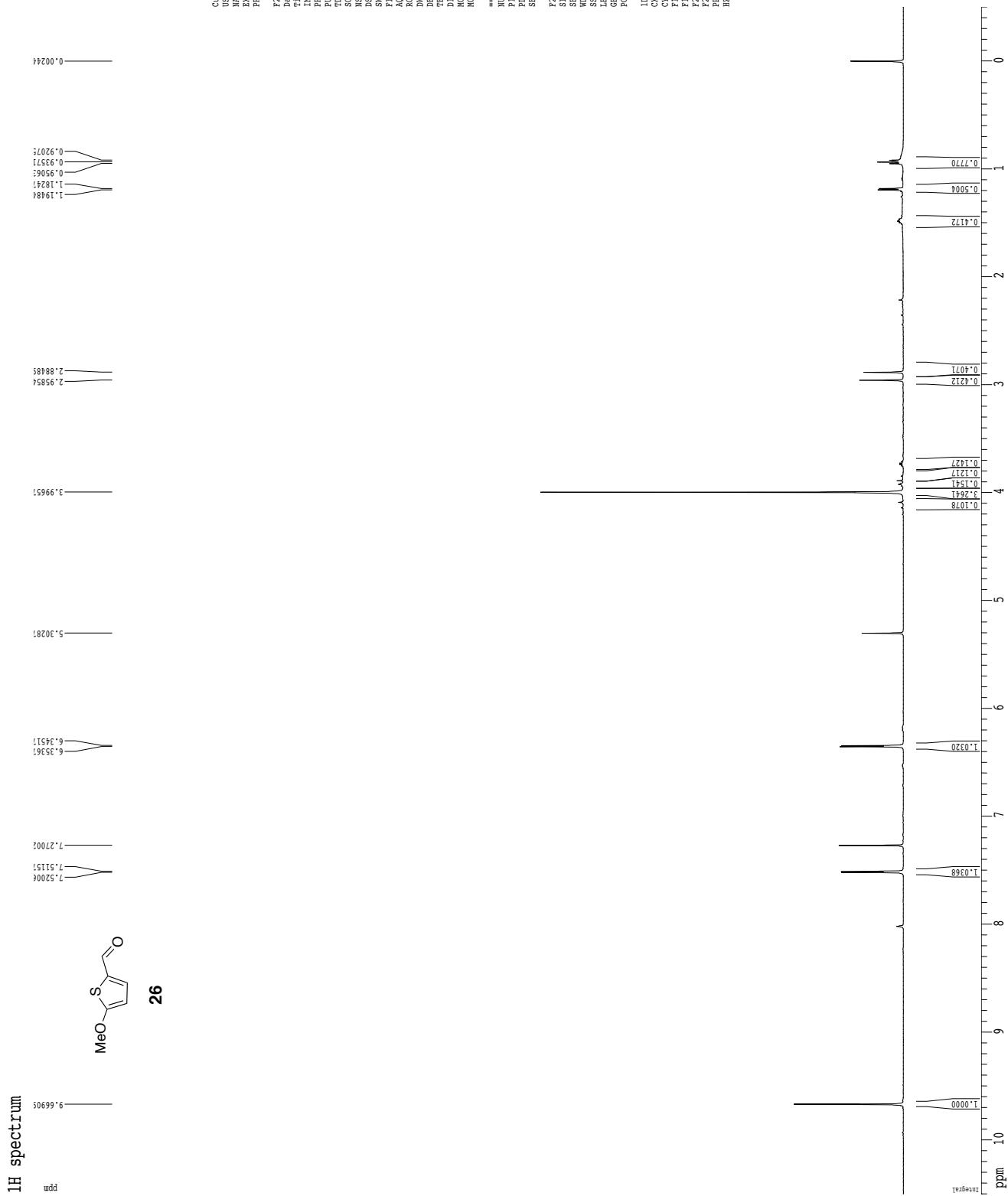


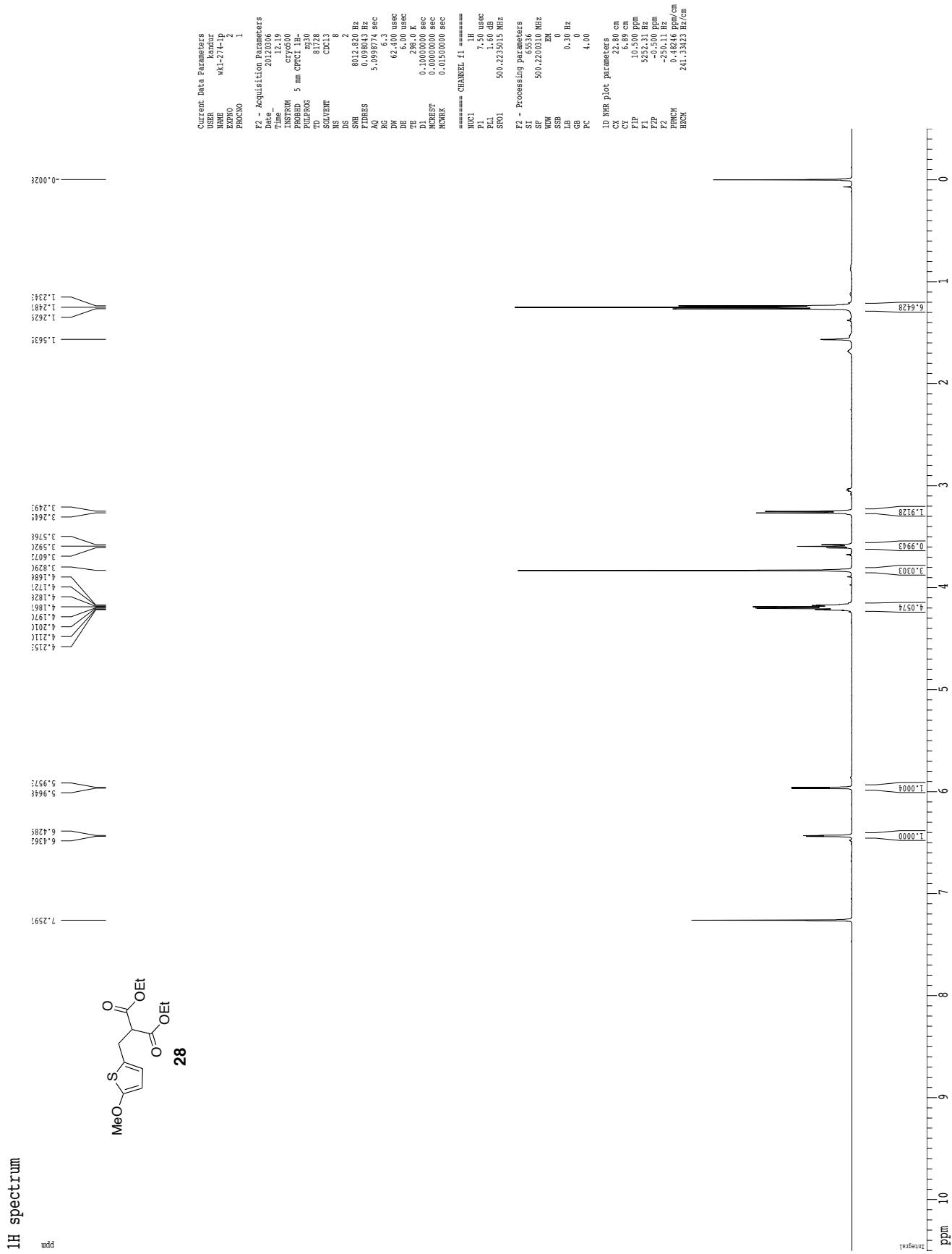
¹H spectrum

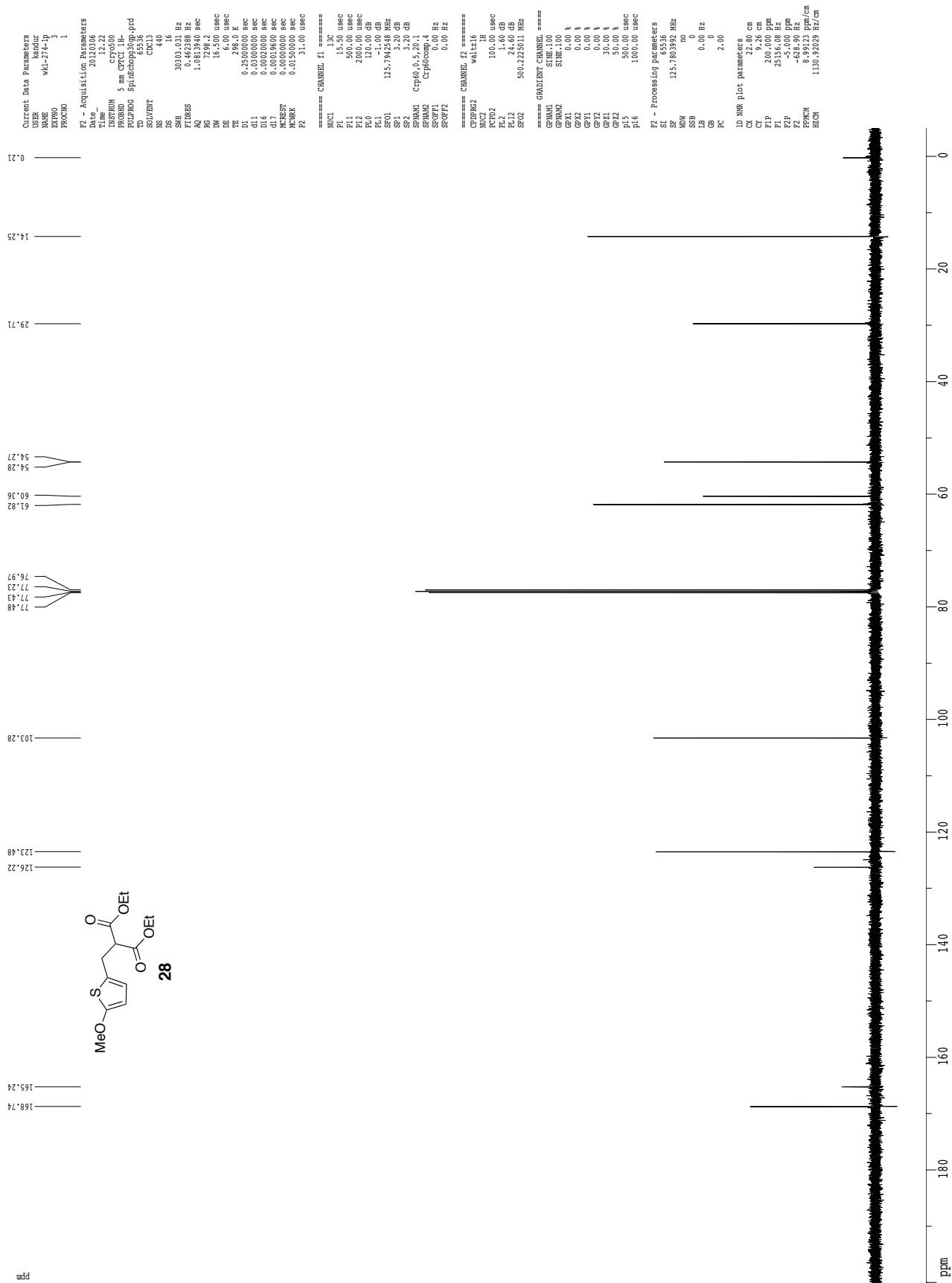


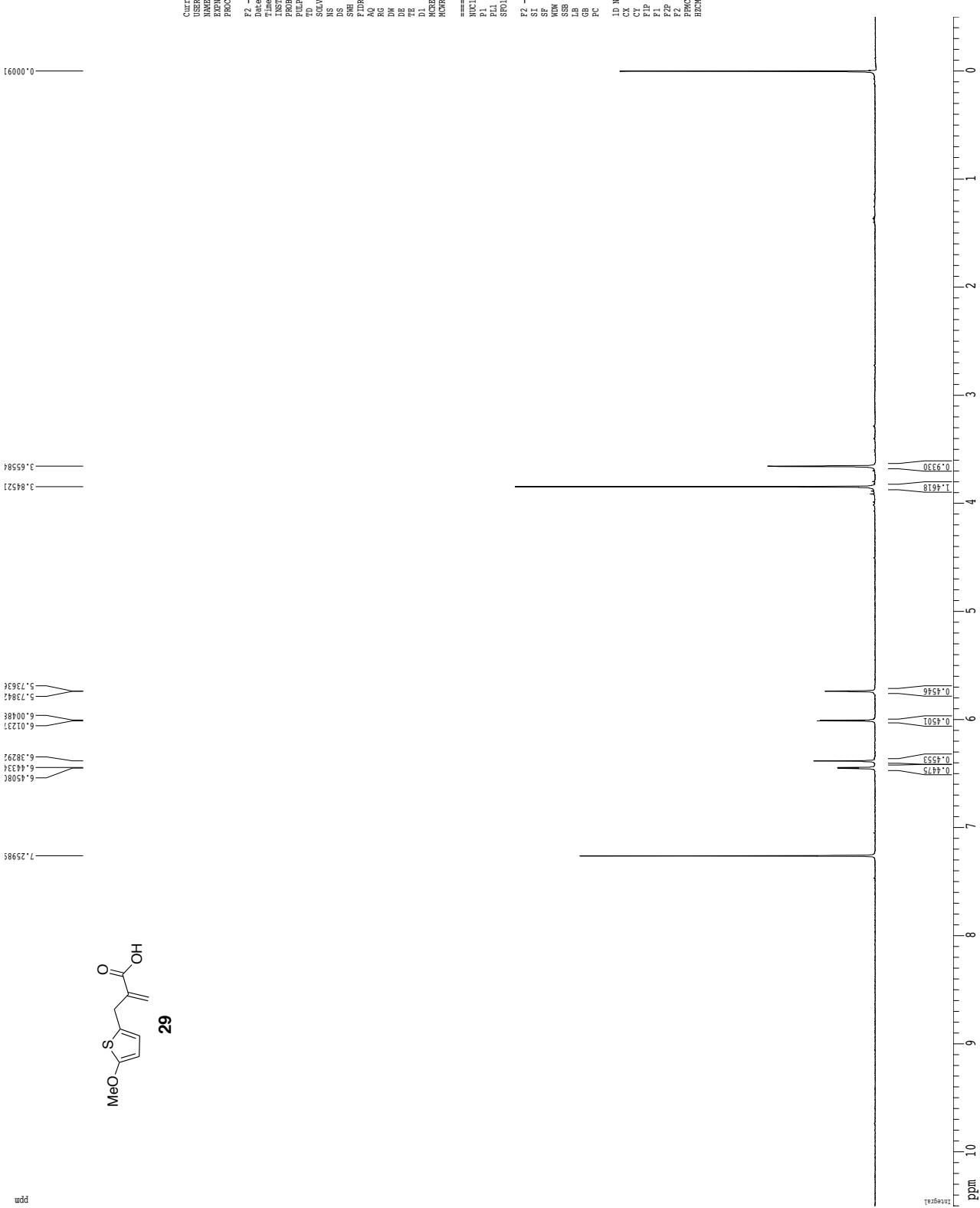
¹H spectrum

Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

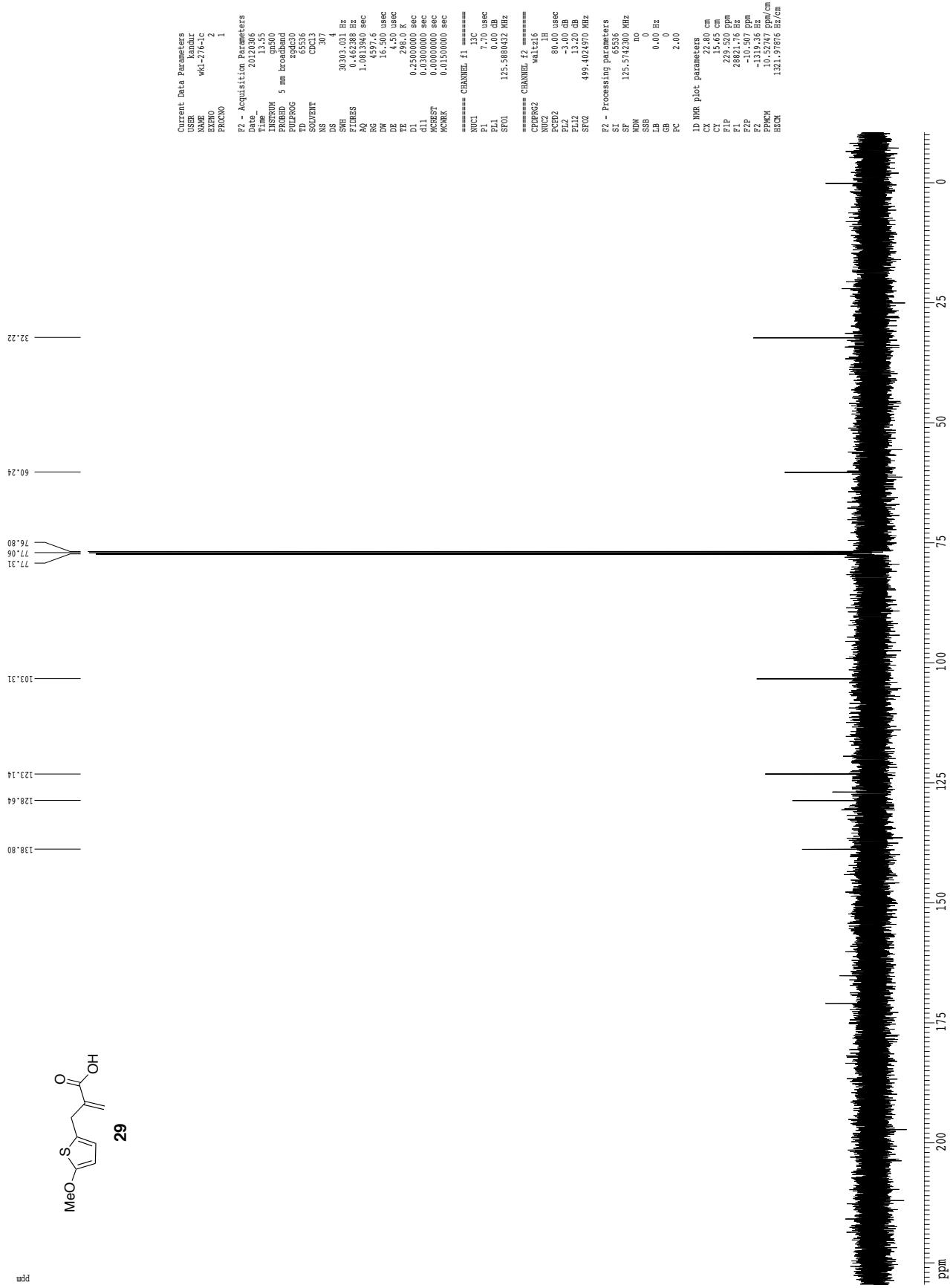


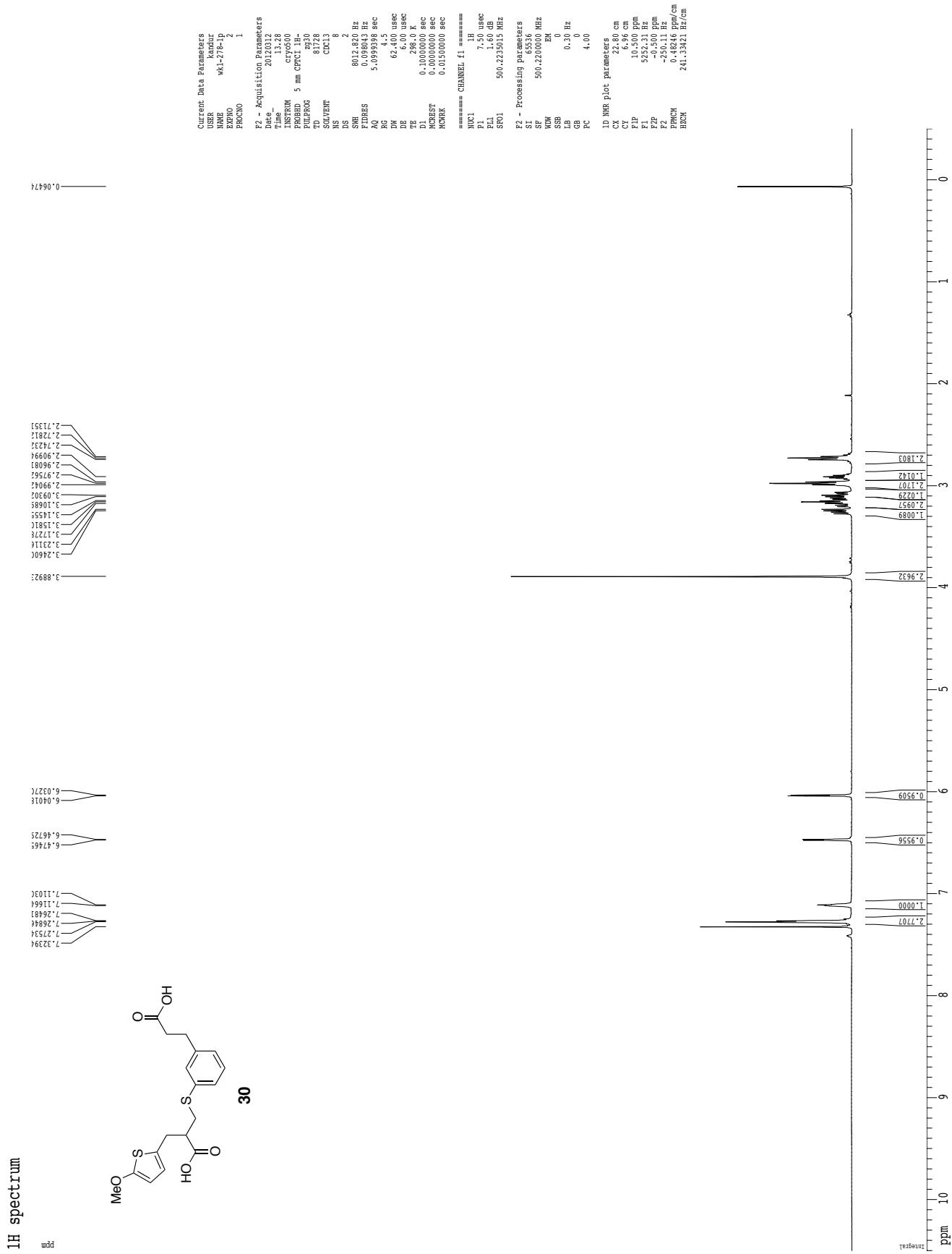


Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

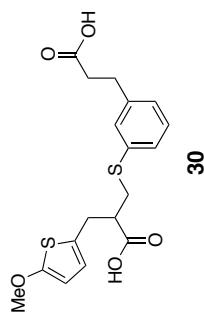
¹H spectrum

13C spectrum with 1H decoupling

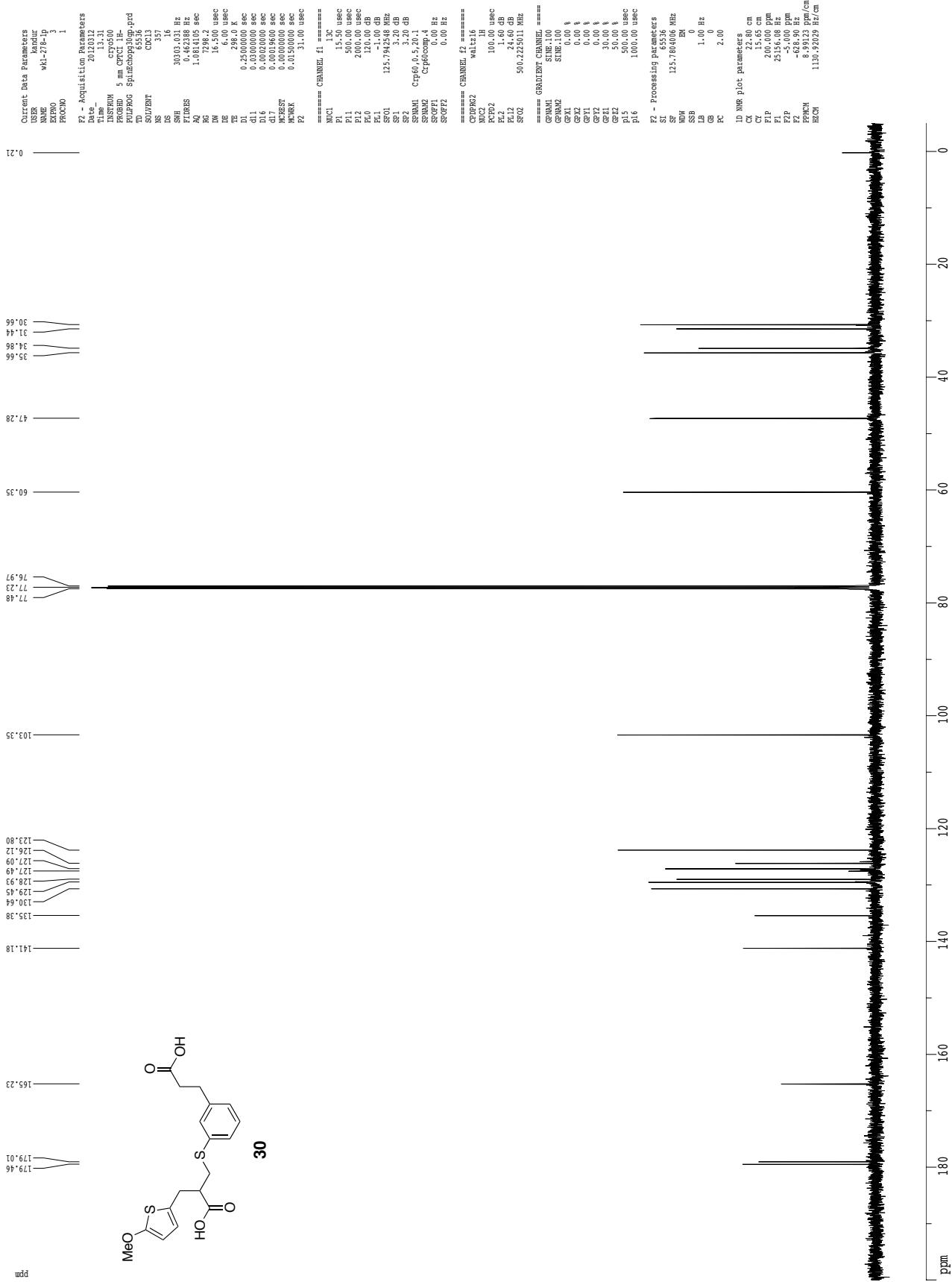


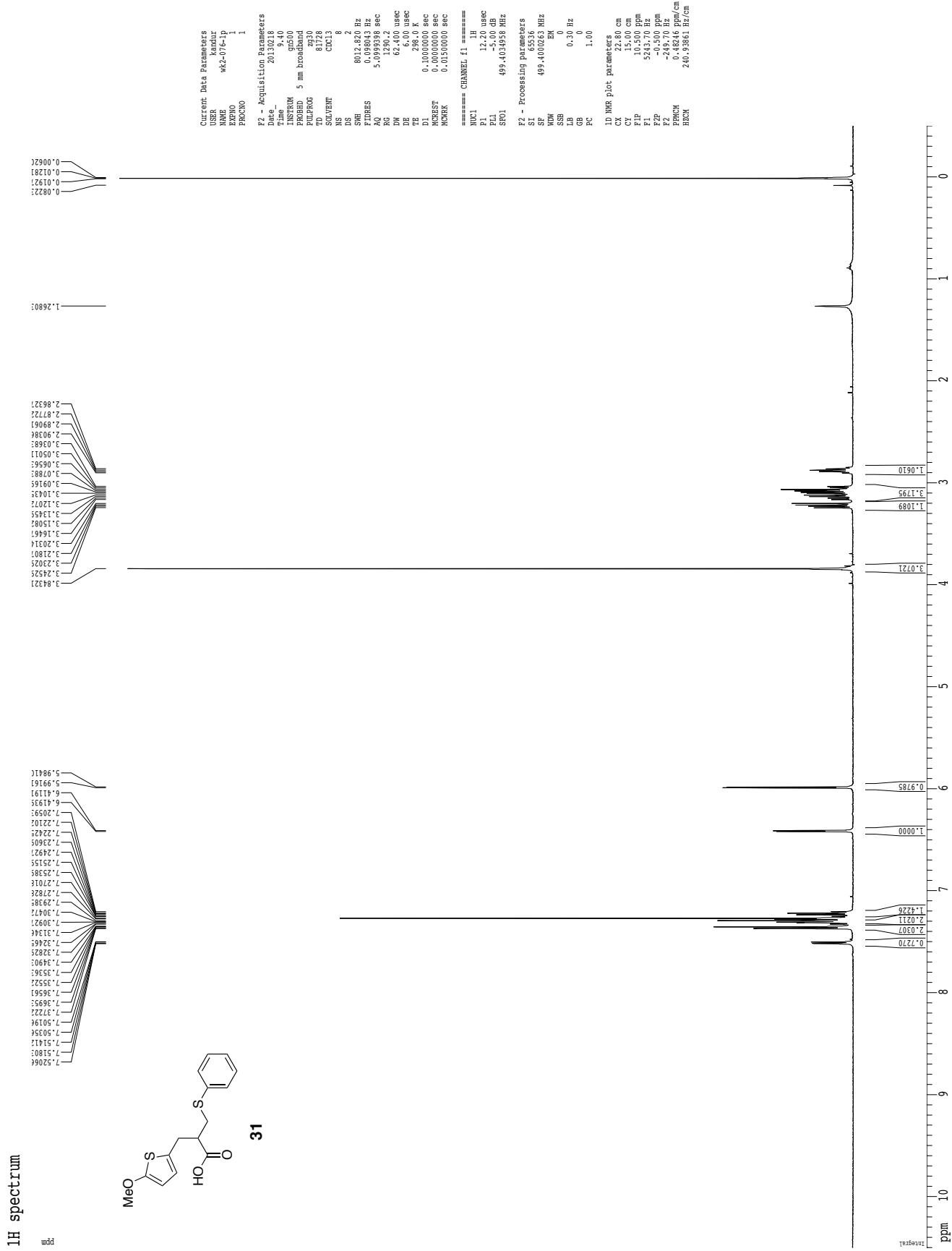


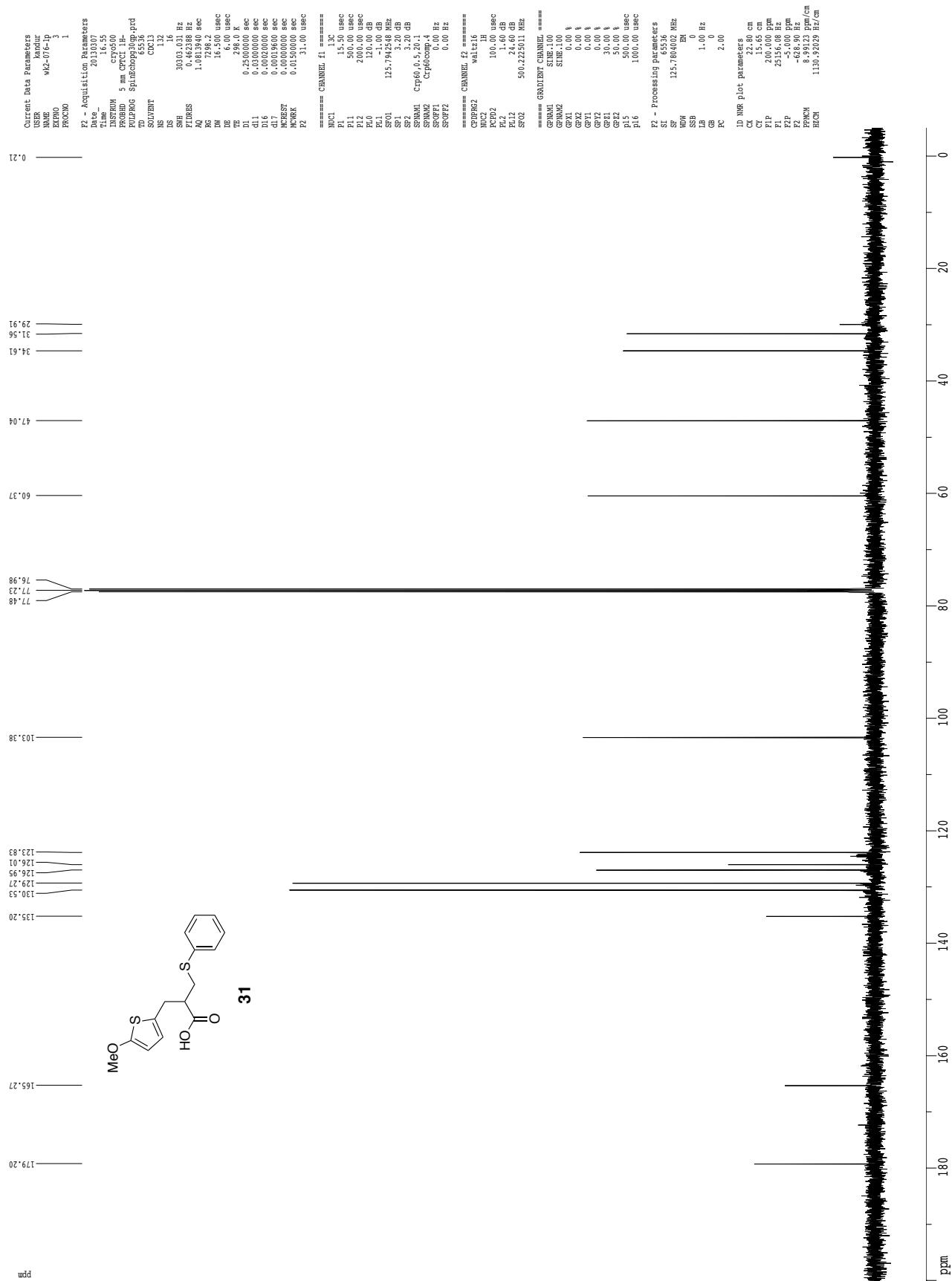
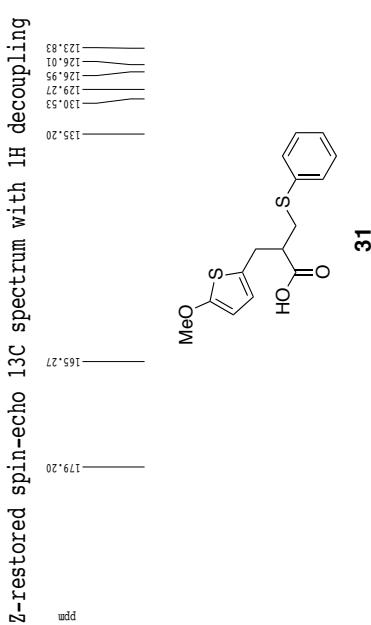
Z-restored spin-echo ^{13}C spectrum with 1H decoupling

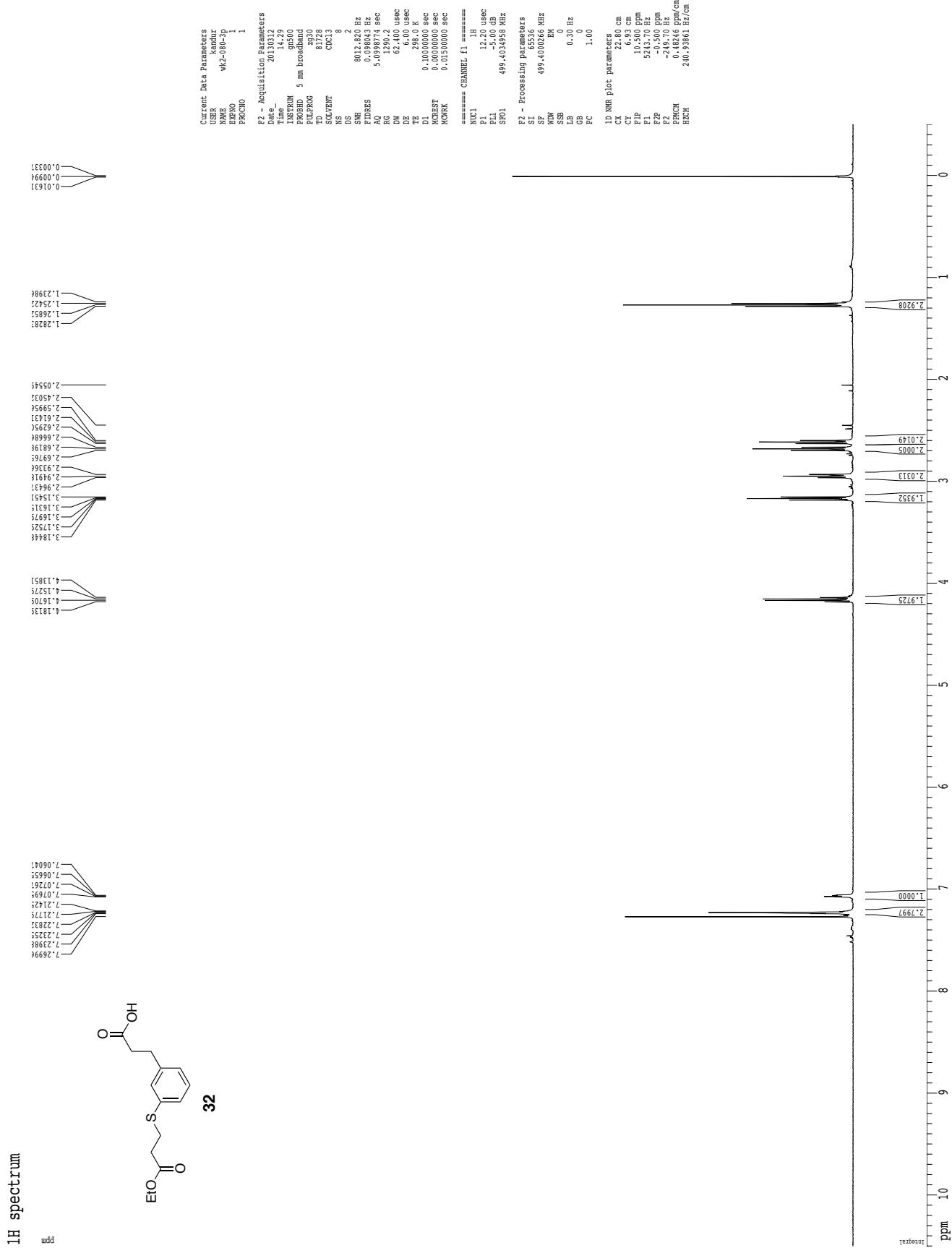


30

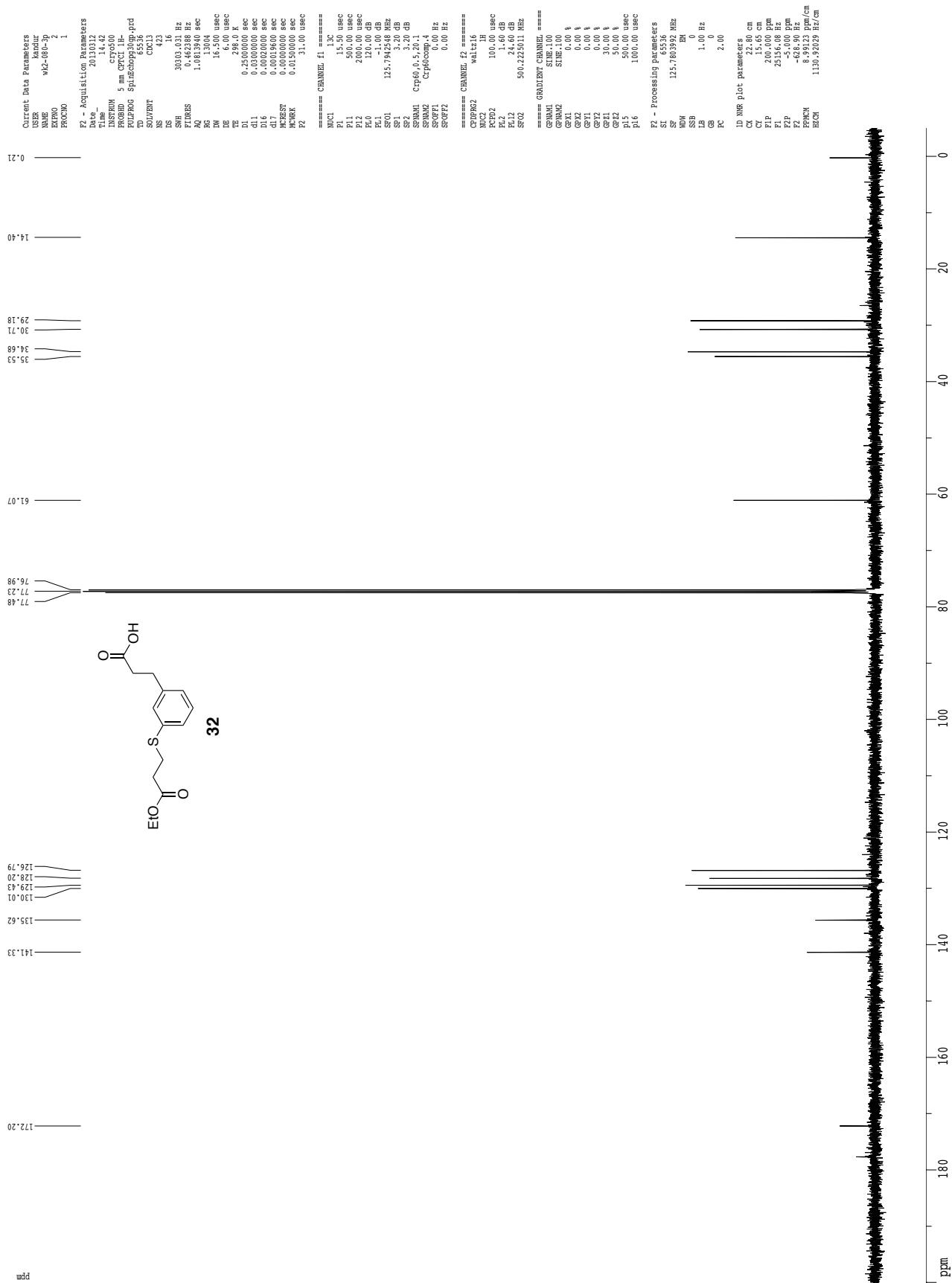


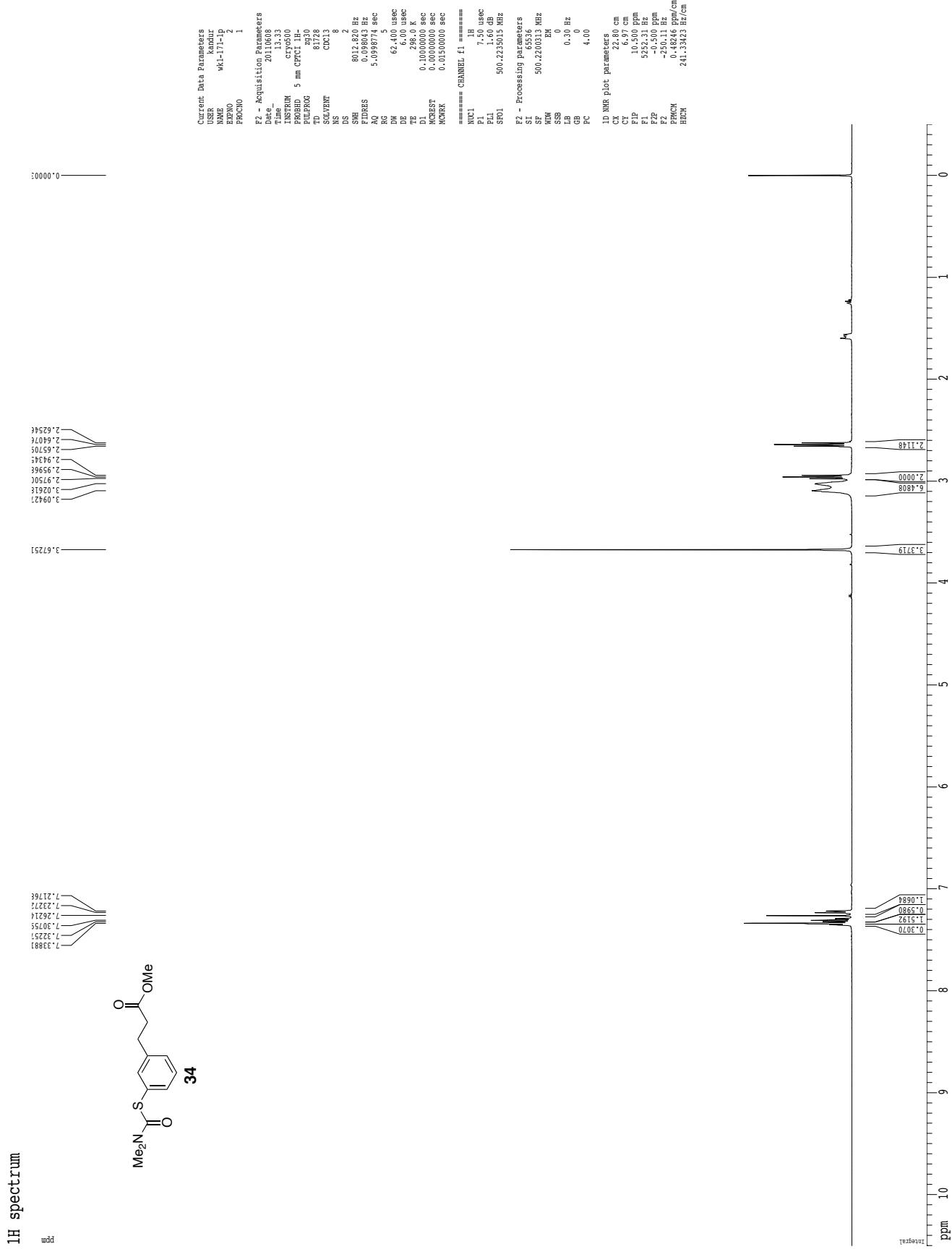


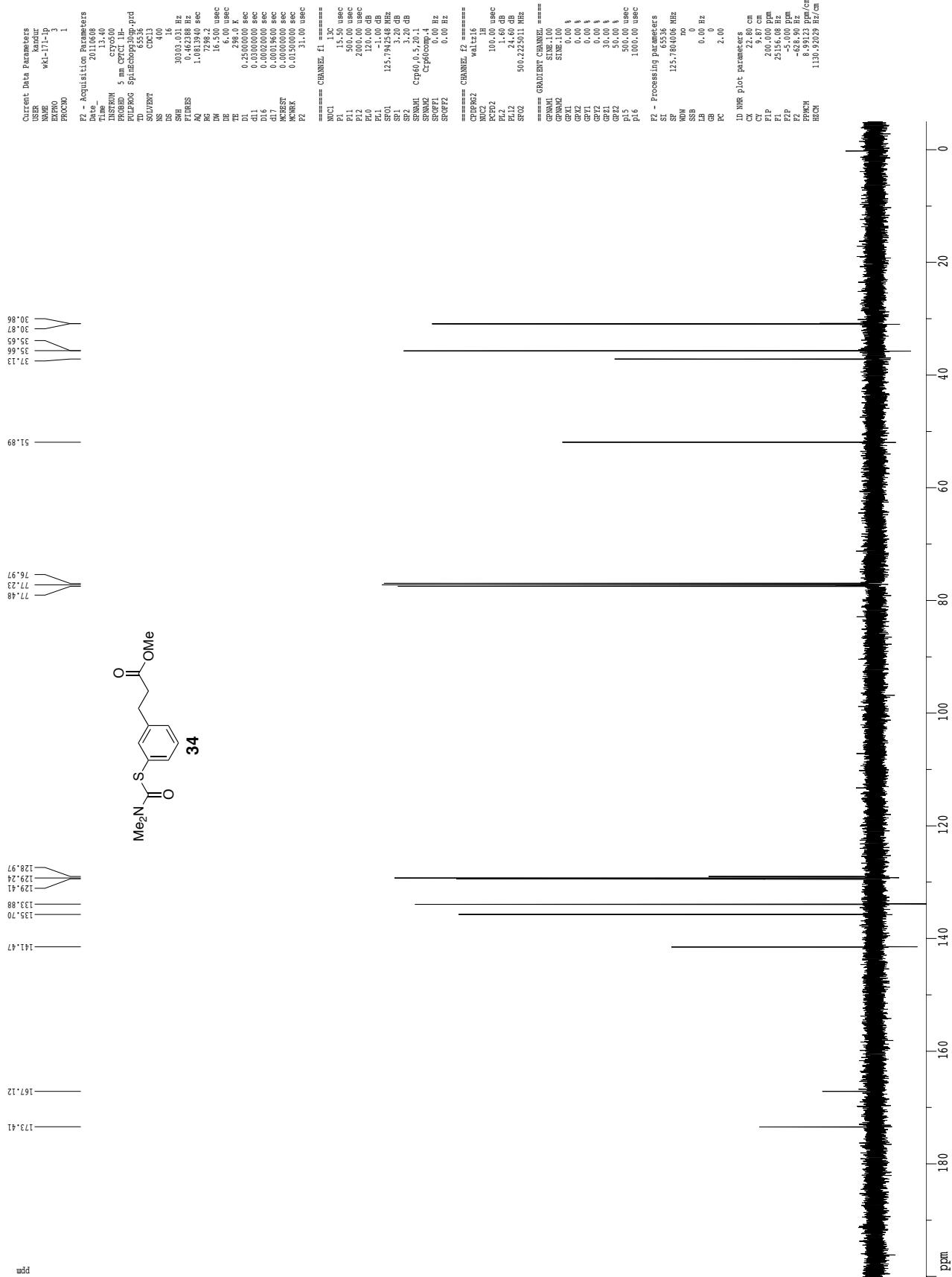


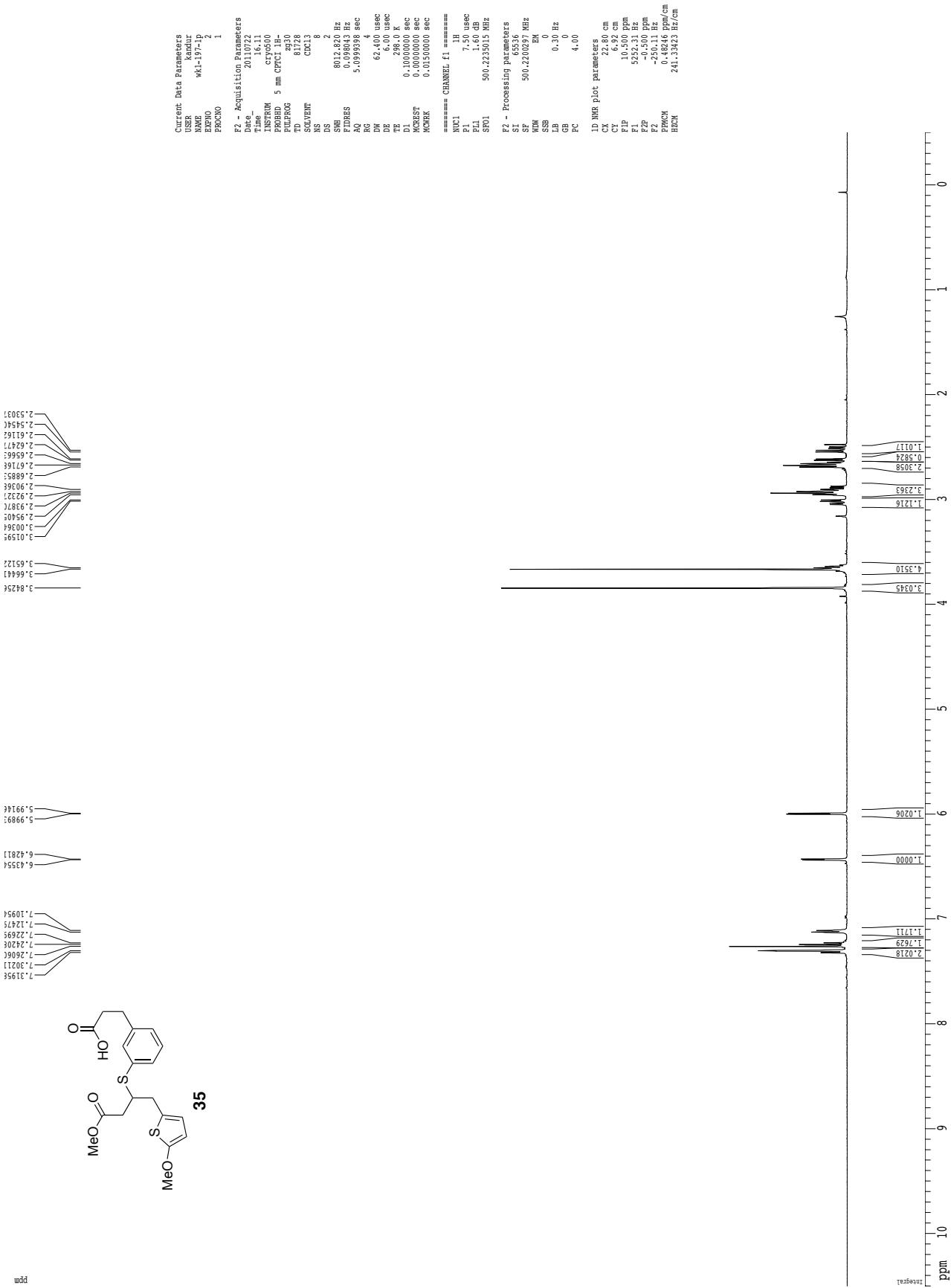


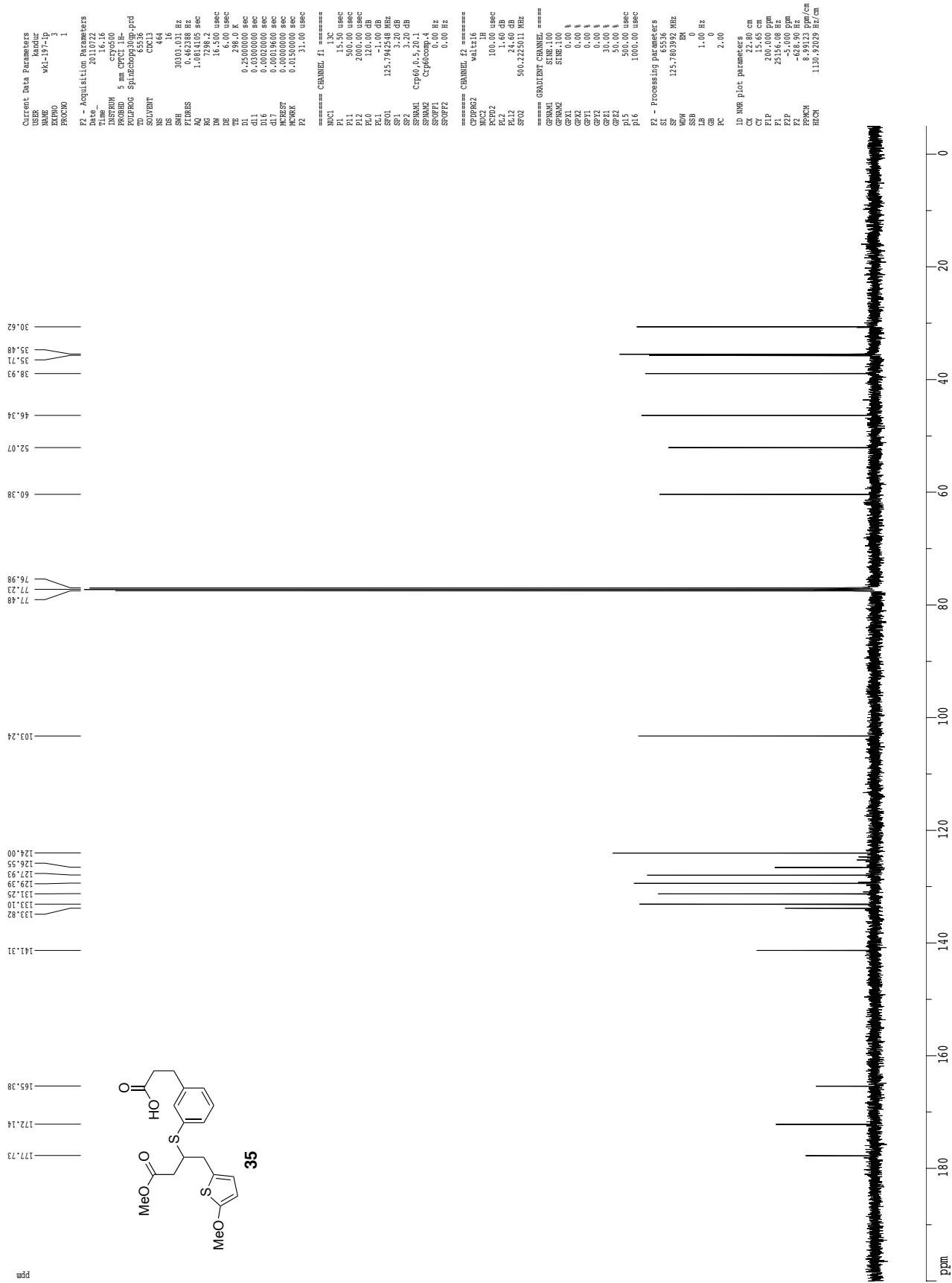
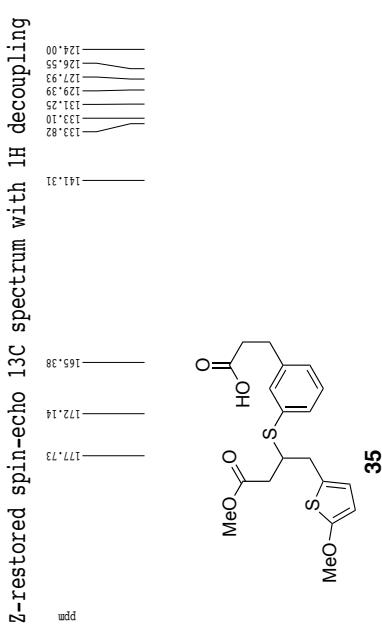
$\pi/2$ -restored spin-echo ^{13}C spectrum with 1H decoupling

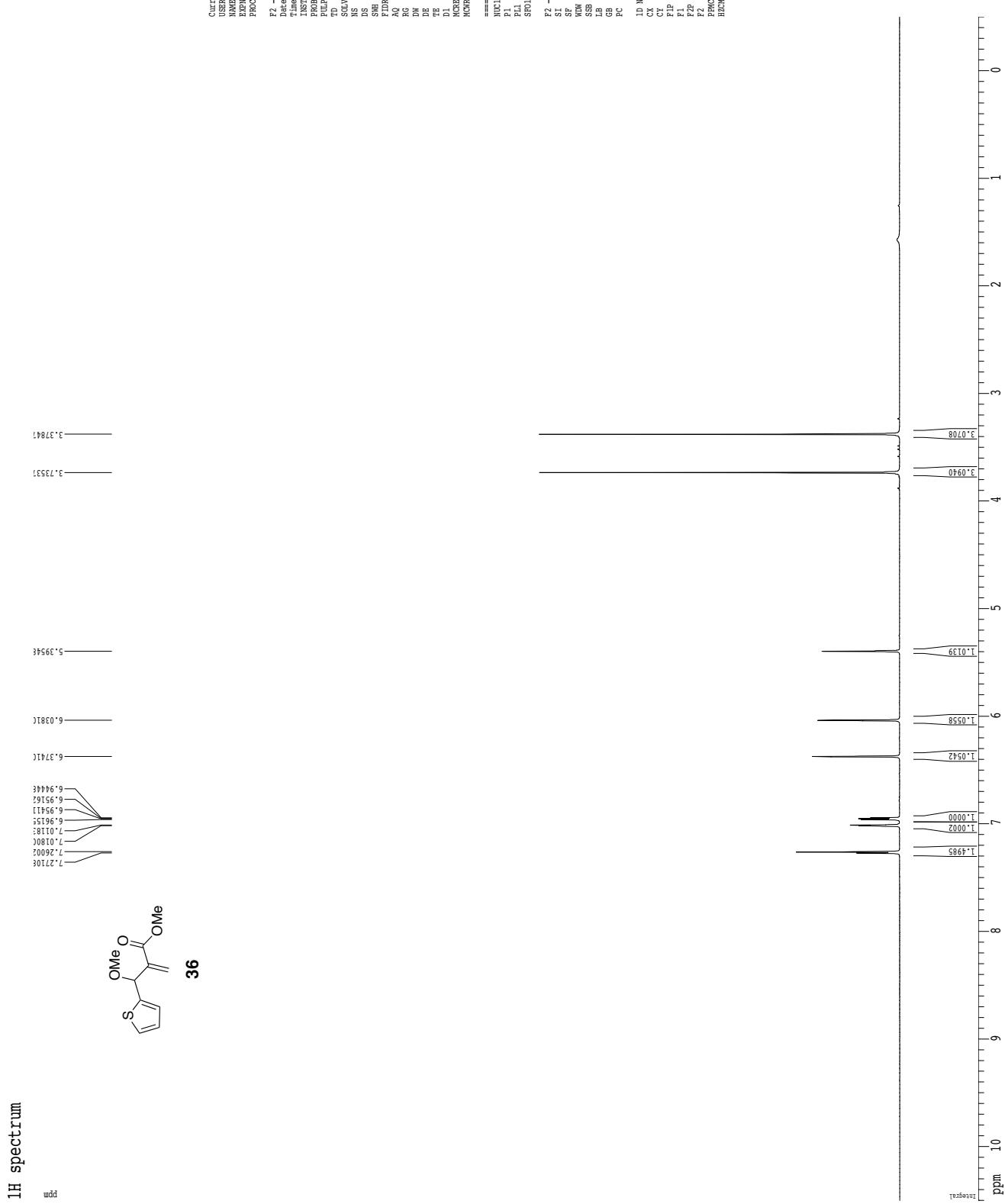


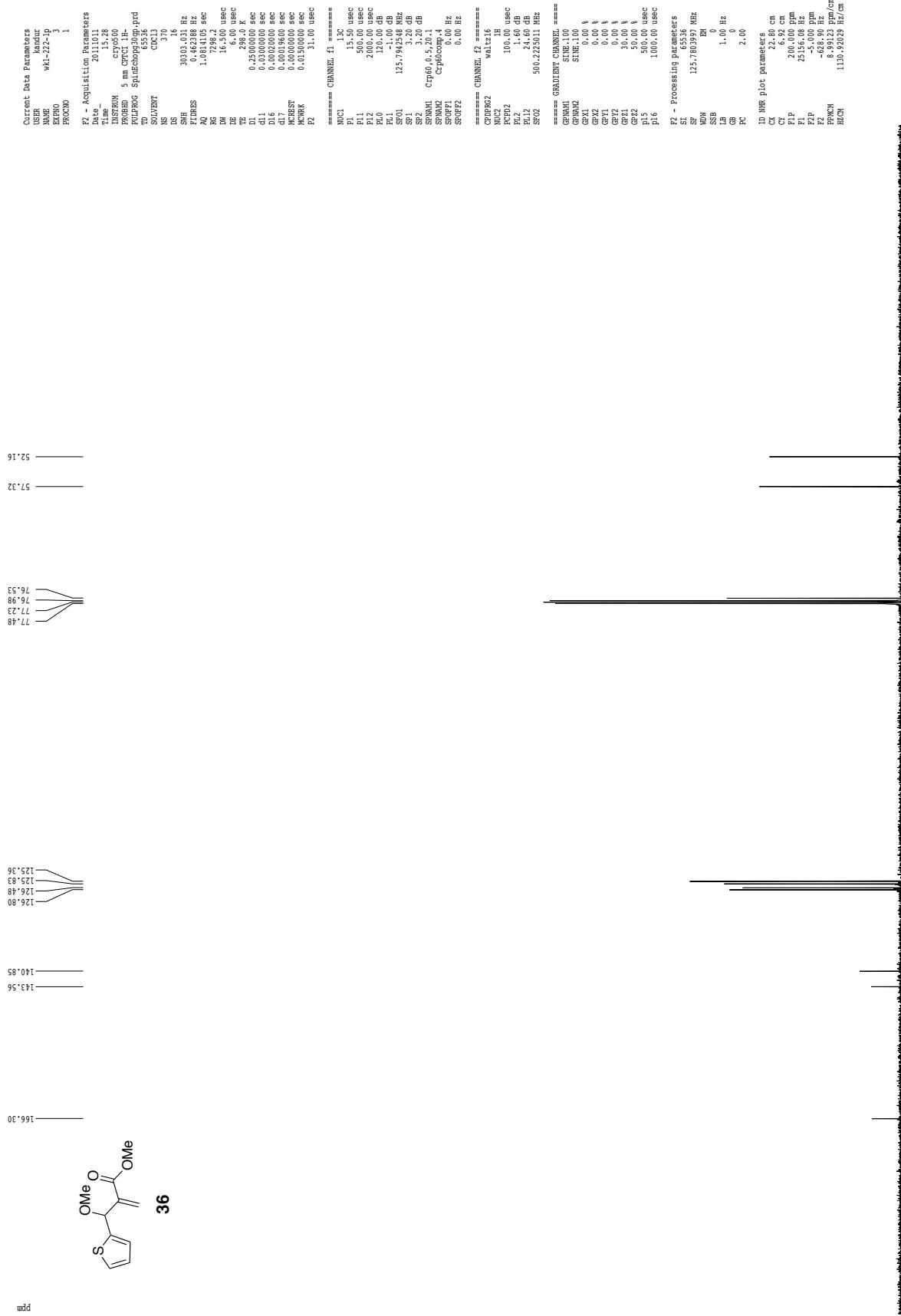
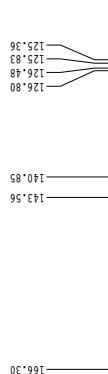
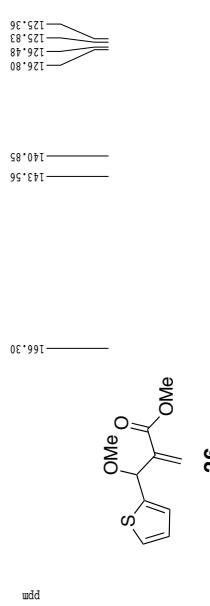


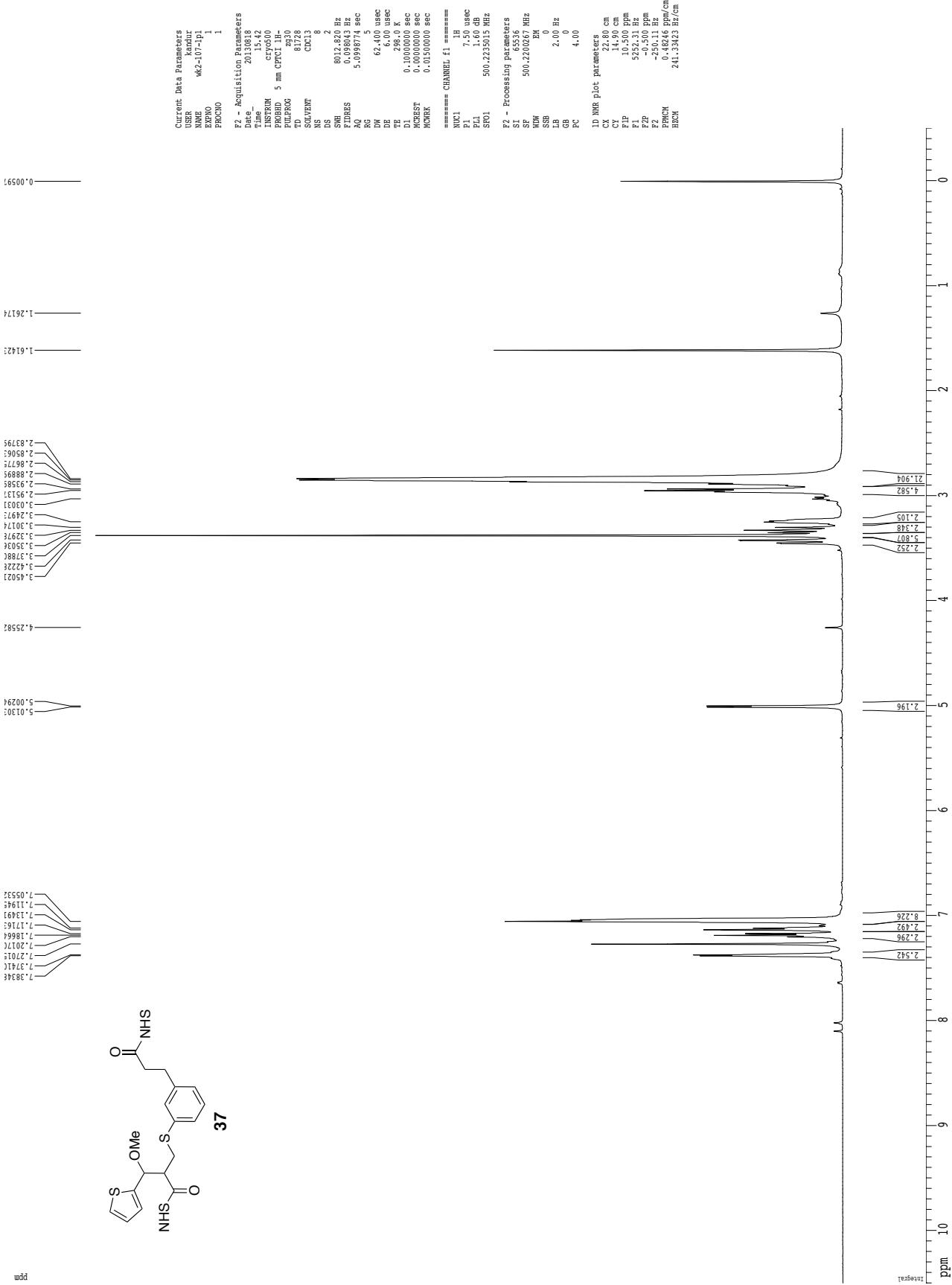
Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

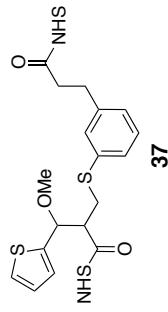
¹H spectrum





Z-restored spin-echo ^{13}C spectrum with ^1H decoupling

¹H spectrum



ppm

Current Data Parameters

USER	kardar
NAME	wk-107-1pi
EXPTNO	2
FACUNO	1

P2 - Acquisition Parameters

Date-	2010810
Time-	15.40
INSTRUM	cryogen
PROBID	5 mm EPRCI 1H- PULPROG: spinEPRDg.prd
TD	6536
SOLVENT	CCL3
NS	274
DS	16
TE	298.0 K
SWH	3039.031 Hz
FLAMES	0.46388 sec
AQ	1.00000 sec
TS	723.2 sec
RG	16.500 sec
DE	6.00 usec
TEC	0.2500000 sec
D1	0.0300000 sec
D11	1.50 usec
D16	0.0002000 sec
D17	0.0001600 sec
MEREST	0.0000000 sec
MERKK	0.0150000 sec
T2	31.00 usec

===== CHANNEL f1 =====

NUC1	1H
PL1	500.00 usec
PL11	200.00 usec
PL12	120.00 dB
PL13	-1.00 dB
SP01	125.794548 MHz
SP1	3.20 dB
SP2	3.20 dB
SP3	0.50 dB
SP4	0.00 dB
SP401	0.00 Hz
SP402	0.00 Hz
SP4T12	0.00 Hz

===== CHANNEL f2 =====

NUC2	1H
PCP12	100.00 usec
PL2	1.60 dB
PL12	24.60 dB
SP02	500.232311 MHz

===== GRADIENT CHANNEL =====

GRAD1	SINE B 100
GP11	0.00 %
GP12	0.00 %
GP21	0.00 %
GP22	30.00 %
p15	50.00 %
p16	100.00 usec

P2 - Processing parameters

SF	6536.15 Hz
WDW	RB
SSB	0
LB	1.00 Hz
GB	0
PC	2.00

1D NMR plot parameters

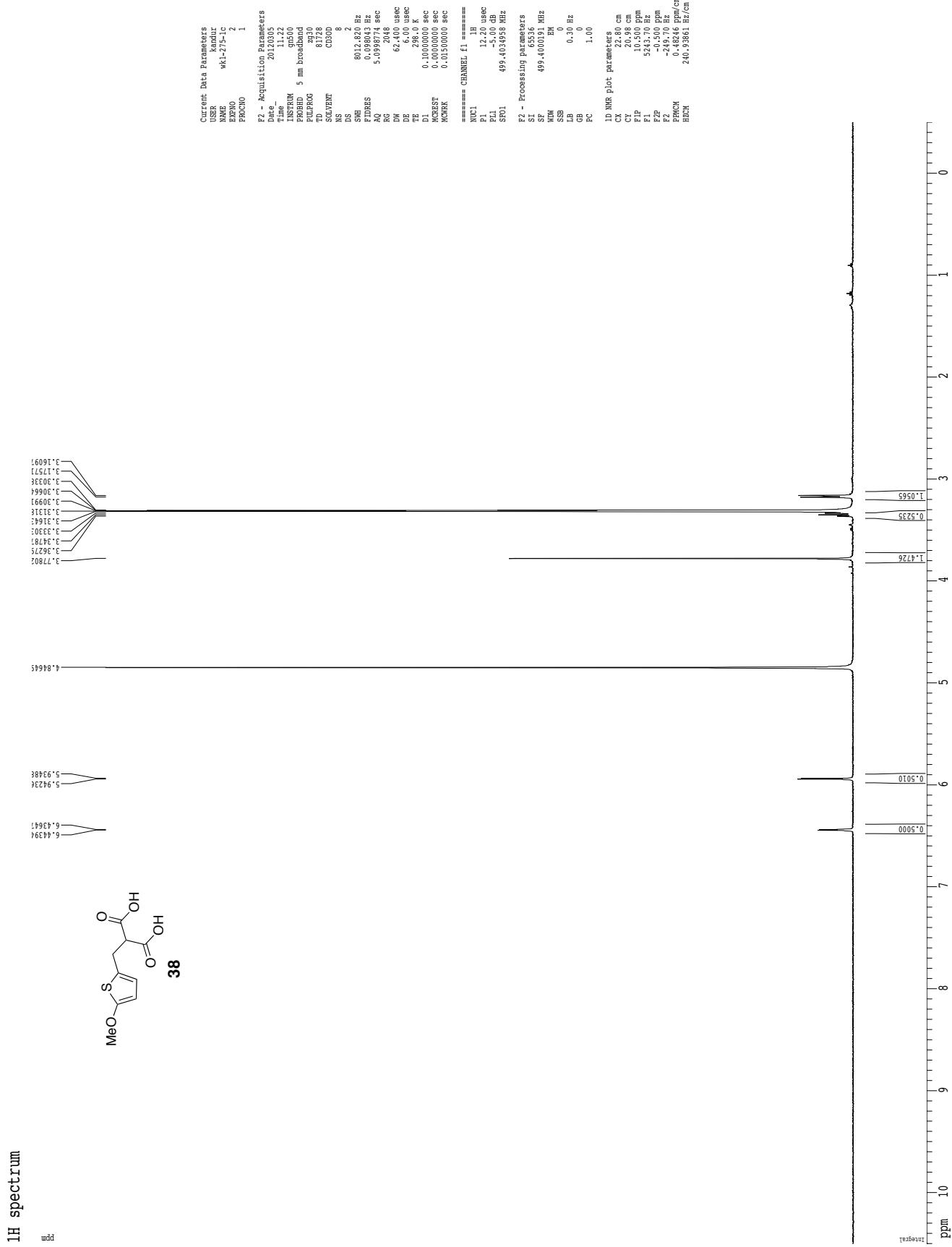
CF	22.80 cm
CT	15.65 cm
FLP	210.00 ppm
F1	2515.08 ppm
FP	5.45 ppm
F2	-0.09 ppm
FRPCM	8.99123 cm ⁻¹ /cm
HC2M	1130.9239 Hz/cm

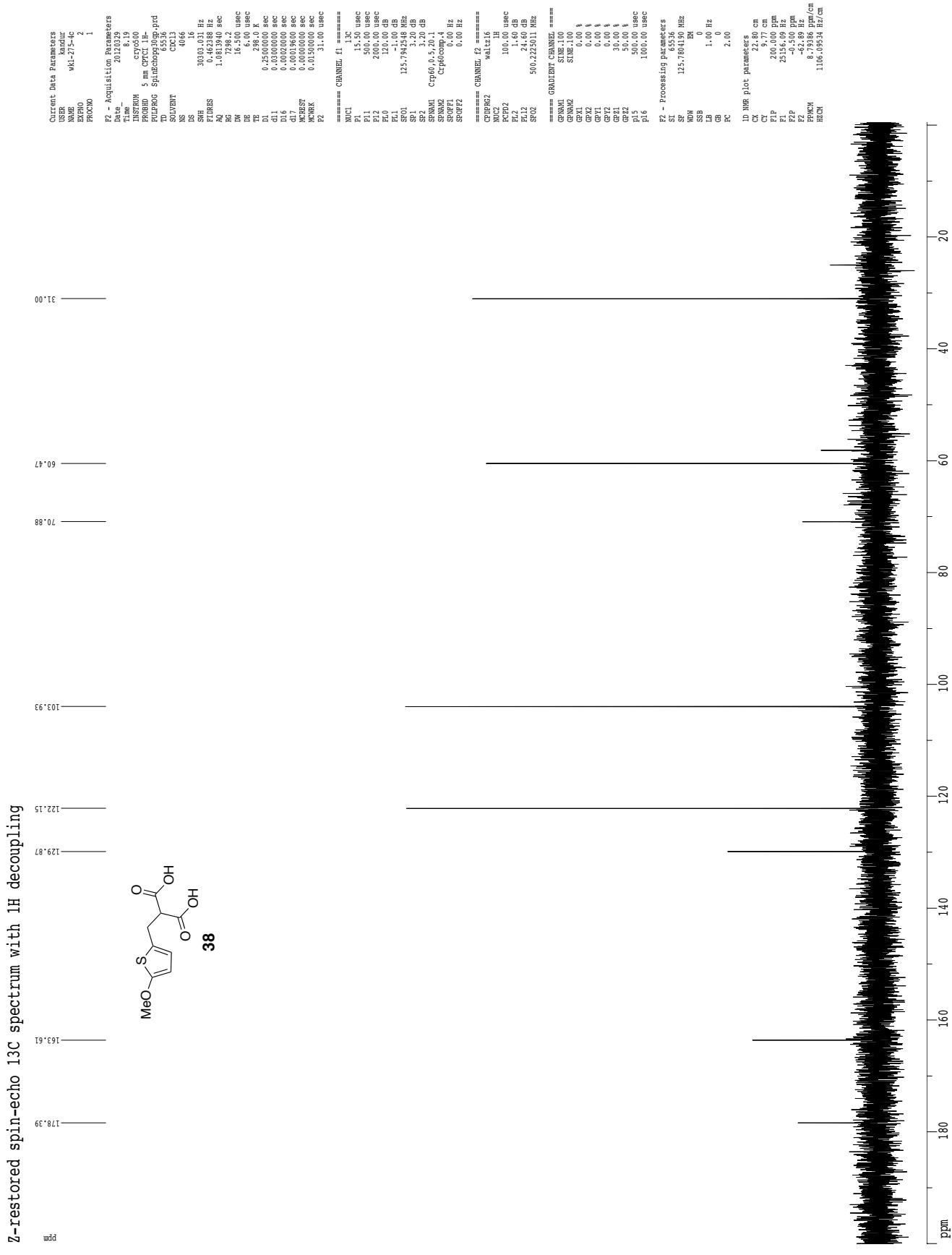
ppm

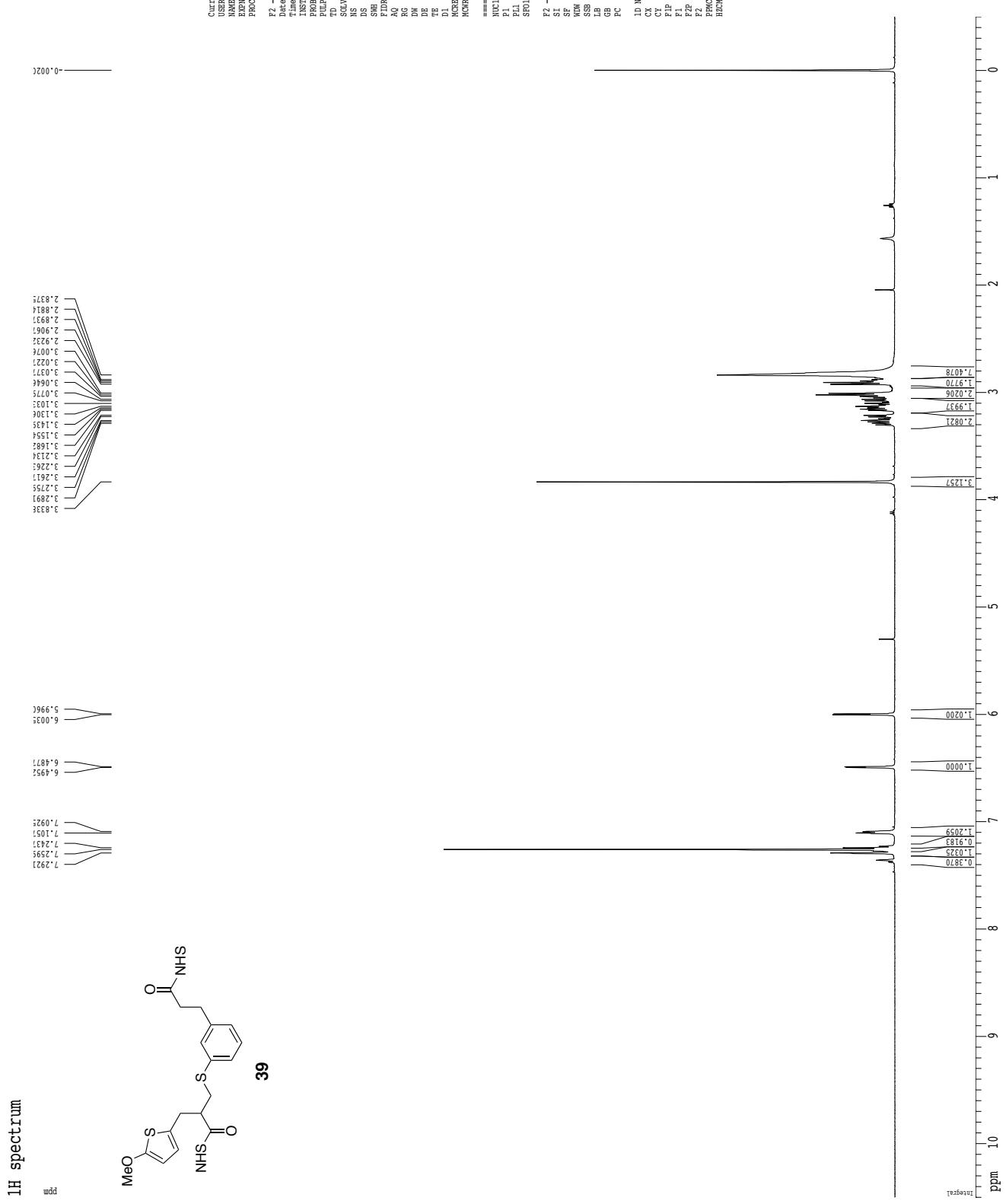
169.30
168.02
168.91
140.48
135.49
128.15
128.36
128.65
128.91
126.33
126.65
126.90
79.20
77.48
76.97
58.09
51.54
25.79
33.67
30.02
30.45
29.99
29.02
28.48
25.77
167.14
168.91
168.02
140.48
135.49
128.15
128.36
128.65
128.91
126.33
126.65
126.90
79.20
77.48
76.97
58.09
51.54
25.79
33.67
30.02
30.45
29.99
29.02
28.48
25.77

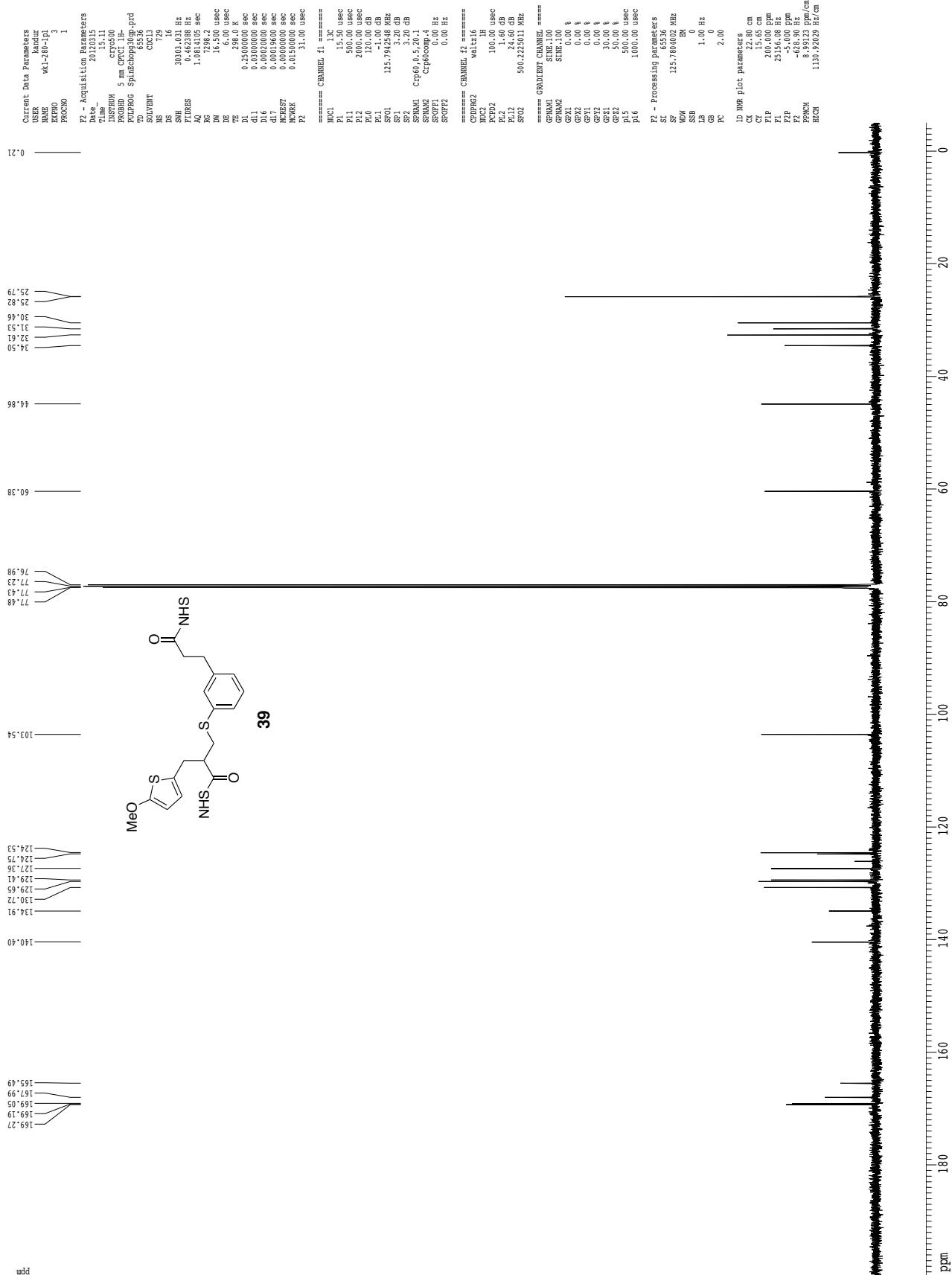
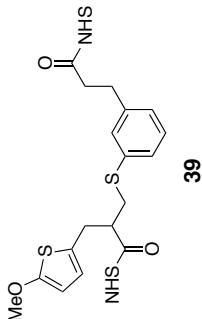
37

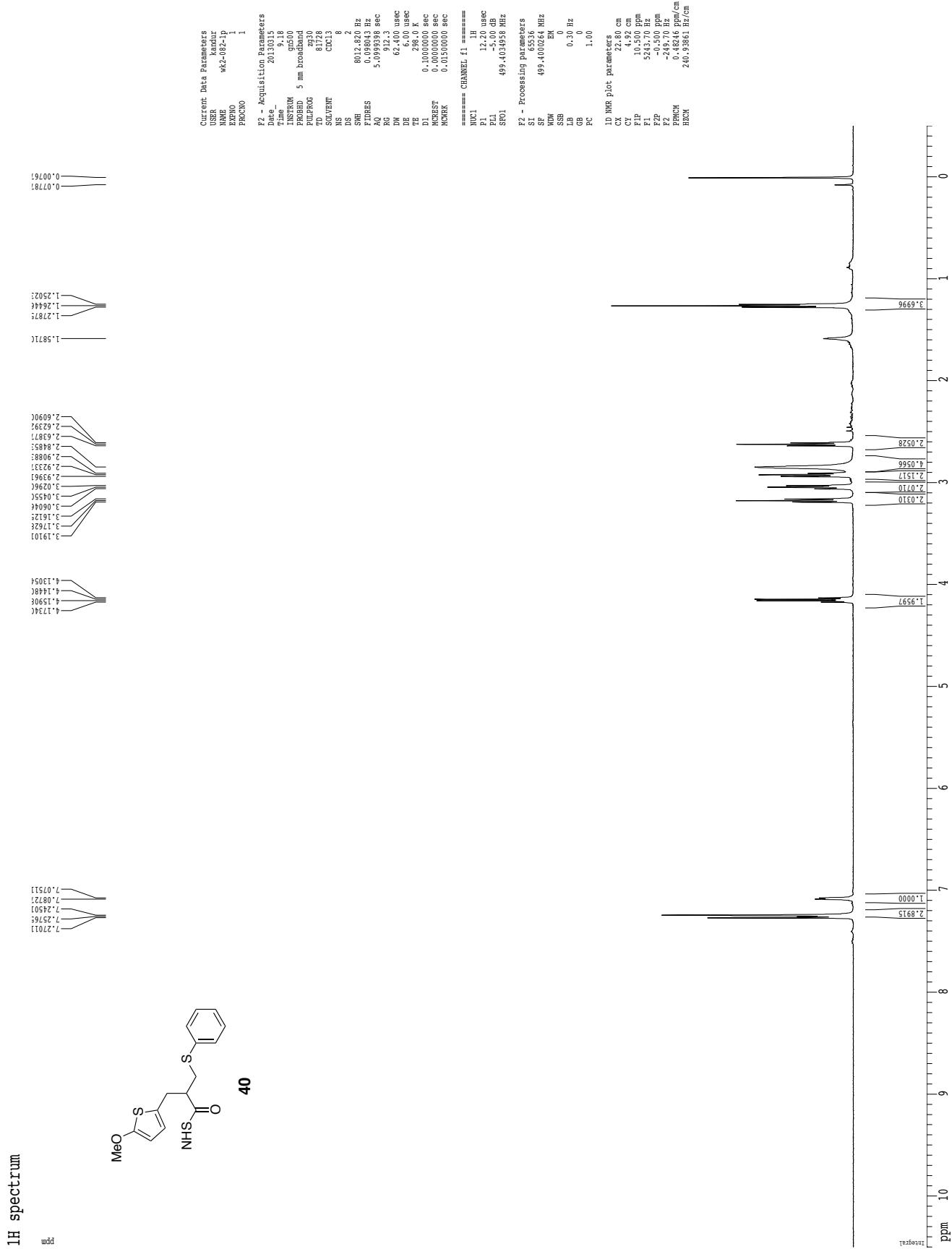
CC(C(=O)NCS)c1ccccc1Sc2ccccc2C(=O)NCS

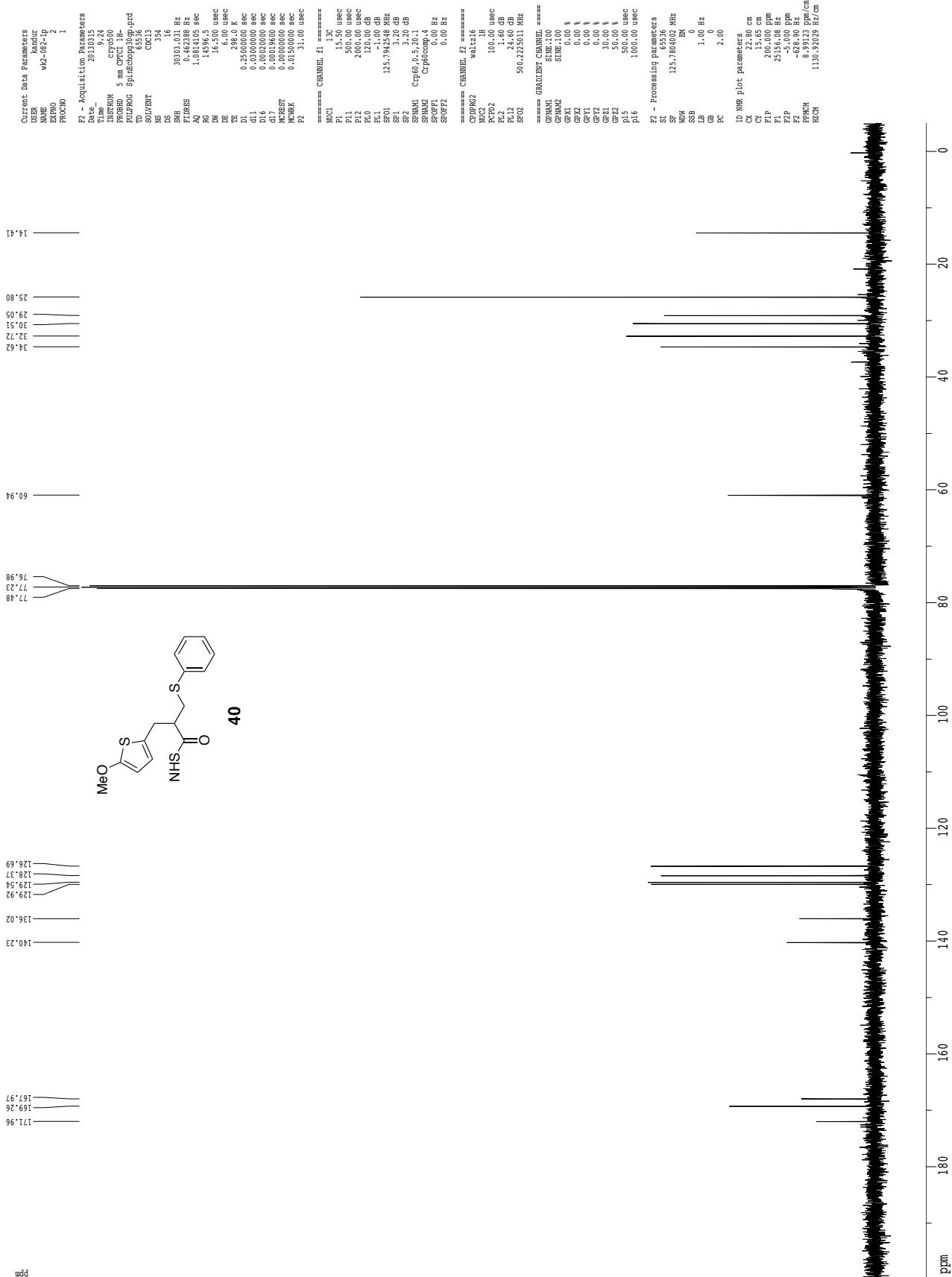
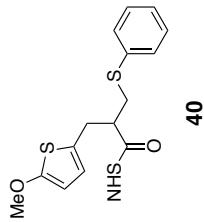


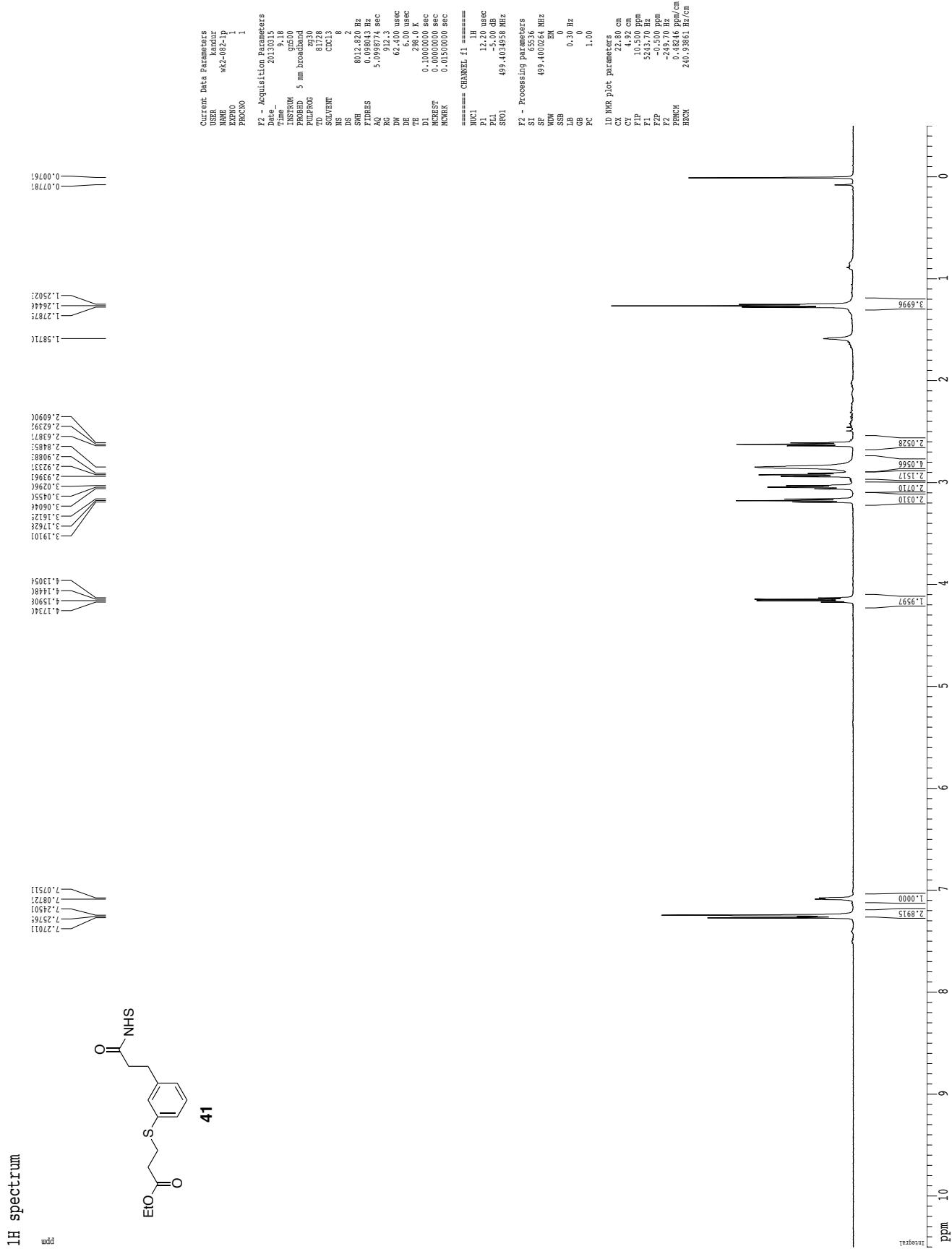












Z-restored spin-echo ^{13}C spectrum with ^1H decoupling