

Supplementary Material

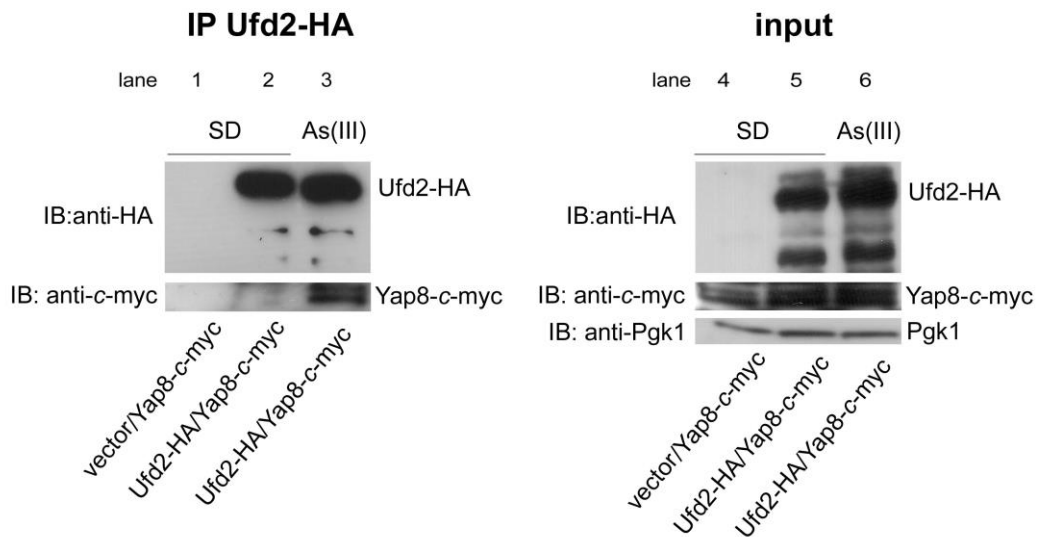


Fig. S1. Reciprocal co-immunoprecipitation assays showing arsenic-mediated interaction between Yap8 and Ufd2. BY4742 cells co-transformed with vector/*YAP8-c-myc* (lanes 1 and 4) or *UFD2-HA/YAP8-c-myc* (lanes 2, 3, 5 and 6) were exposed or not to 1.5 mM As(III) for 90 min and Ufd2-HA was immunoprecipitated with anti-HA antibody. Immunoblotting was performed using anti-HA, anti-*c-myc* and anti-Pgk1 antibodies. A representative experiment is shown. IP – immunoprecipitation; IB – immunoblotting; SD – Synthetic Defined medium, control condition.

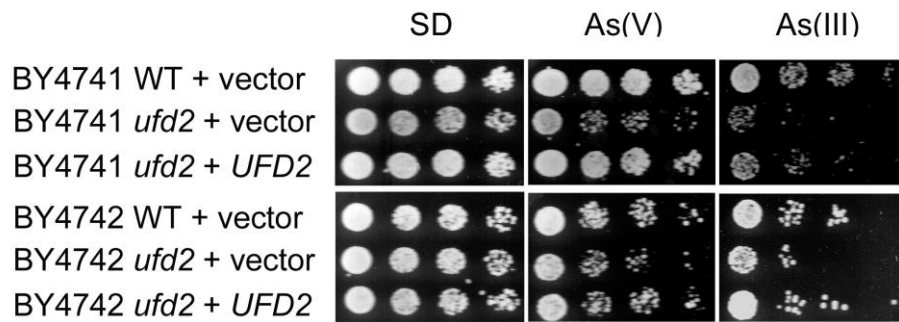


Fig. S2. As(V)- and As(III)-sensitivity phenotypes of *ufd2* are restored by expressing an episomal copy of *UFD2*. Exponential phase BY4741 and BY4742 wild type (WT) and *ufd2* cells expressing an episomal copy of *UFD2* or the respective control vector were serially diluted and spotted onto SD media supplemented or not with 2 mM As(V) or 1.5 mM As(III). Growth was recorded after 2 days incubation at 30°C. A representative experiment is shown.

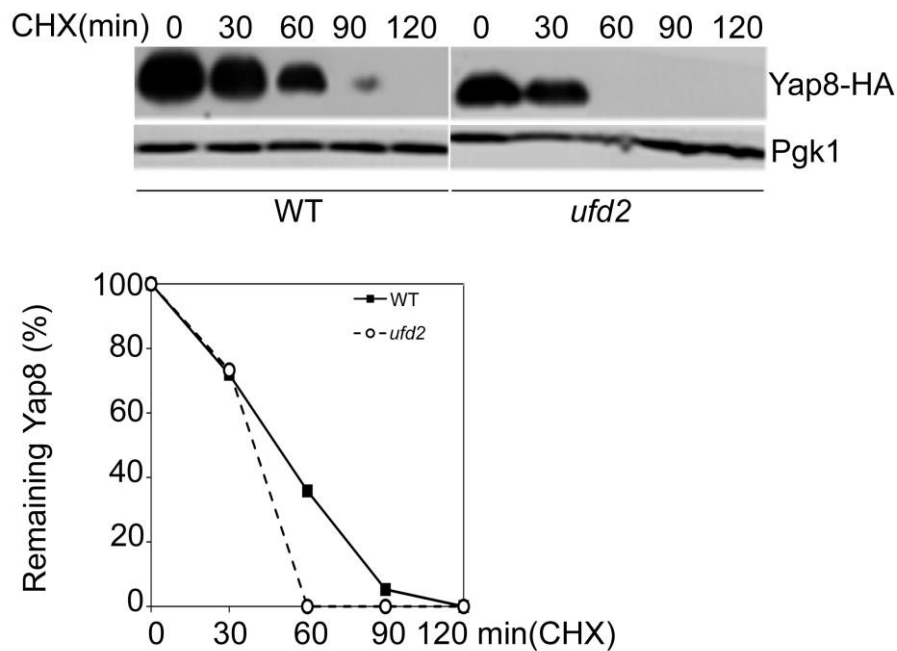


Fig. S3. Ufd2 mediates Yap8 stabilization under As(V) stress conditions. BY4742 wild type (WT) and *ufd2* mutant strains expressing Yap8-HA were pre-treated with 2 mM As(V) for 60 min, washed and subsequently treated with 0.1 mg/mL cycloheximide (CHX) up to 120 min prior to immunoblotting using anti-HA and anti-Pgk1 antibodies. The graph represents the percentage of remaining Yap8 protein after CHX addition. A representative experiment is shown.

Table S1. *Saccharomyces cerevisiae* strains used in this study

Strain	Genotype	Reference or source
BY4742	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0</i>	EUROSCARF*
<i>ufd2</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YDL190c::<i>kanMX4</i></i>	EUROSCARF
<i>yap8</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YPR199c::<i>kanMX4</i></i>	EUROSCARF
<i>yap8ufd2</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YPR199c::<i>kanMX4</i> YDL190c::<i>HIS3MX4</i></i>	This study
<i>ubc4</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YBR082c::<i>kanMX4</i></i>	ATCC**
<i>rad23</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YEL037c::<i>kanMX4</i></i>	EUROSCARF
<i>dsk2</i>	<i>MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YMR276w::<i>kanMX4</i></i>	EUROSCARF
Y187	<i>MATα ura3-52 his3-200 ade2-101 trp1-901 leu2-3 112 gal4Δ met-gal80Δ URA3::<i>GALIUAS-GALITATA-lacZ MEL1</i></i>	Clontech Laboratories, Inc.
BY4741	<i>MATα his3Δ1 leu2Δ0 met15Δ0 ura3Δ0</i>	EUROSCARF
<i>ufd2</i>	<i>MATα his3Δ1 leu2Δ0 met15Δ0 ura3Δ0 YDL190C::<i>kanMX4</i></i>	EUROSCARF
<i>Ufd2^{U-boxΔ}</i>	<i>MATα his3Δ1 leu2Δ0 met15Δ0 ura3Δ0 YDL190C^{2568-2886 bp}::<i>HIS3</i></i>	Chang Liu <i>et al.</i> (2011)

*EUROpean Saccharomyces Cerevisiae ARchive for Functional analysis

** American Type Culture Collection

Table S2. Plasmids used in this study

Name	Features	Reference or source
pRS416	<i>CEN, URA3</i>	Agilent Technologies
pRS416- <i>YAP8-HA</i>		Our unpublished work
pRS416- <i>YAP8-c-myc</i>		Amaral et al. (2013)
pRS416- <i>UFD2-HA</i>		This study
pGADT7-Rec	2 μ , <i>LEU2, HA</i>	Clontech
pGADT7- <i>T-antigen</i>		Clontech
pGADT7- <i>UFD2</i>		This study
pGBKT7	2 μ , <i>TRP1, c-myc</i>	Clontech
pGBKT7- <i>LamC</i>		Clontech
pGBKT7- <i>p53</i>		Clontech
pGBKT7- <i>YAP8</i>		Our unpublished work
YCplac111	<i>CEN, LEU2</i>	Agilent Technologies
YCplac111- <i>UFD2-HA</i>		This study
YEplac181	2 μ , <i>LEU2</i>	Agilent Technologies
YEplac181- <i>YAP8-HA</i>		Our unpublished work
pGal- <i>MPS1-c-myc</i>	<i>Integrative, URA3, GAL1^{promoter}</i>	Chang Liu et al. (2011)

Table S3. Oligonucleotides used in this study

#	Name	Sequence (5' – 3' UTR)
1	UFD2-HIS-Fw ^D	GGGAAAAGTTAACTTTGAAAGTAGAACCCCTCATTCCATAGATCGTAC GCTGCAGG
2	UFD2-HIS-Rv ^D	TTGATTAGGGTCAATTTTGCAATTTATTCTATCACTTATTTTAGGGAG ACCGGCAGAT
3	A1-UFD2 ^D	GGCTTGCTGGTACAATATGG
4	A4-UFD2 ^D	CCAGAGCTTTGAGAAGAG
5	UFD2 400-Fw*	GAAAGGTAAAGTTGAC
6	UFD2 1300-Fw*	CTTAAACTCAAGGAC
7	UFD2 2200-Fw*	GGTAAATTAGTGACG
8	UFD2-pGADT7-Fw ^C	ACCGCCATAGAAGATATTTTAC
9	UFD2-pGADT7-Rv ^C	TCACTCGCTTGCTTTATG
10	pRS416- <i>SmaI</i> -UFD2-P1000-Fw ^C	GAATTCCTGCAGCCCAATATTCTGTTATTG
11	HA-UFD2-Rv ^C	AGCGTAATCTGGAACATCGTATGGGTACATTCACTTATTCATCA
12	HA-UFD2-Term500-Fw ^C	ATGTACCCATACGATGTTCCAGATTACGCTTAGAATAAAATTGCAA
13	UFD2-Term500-pRS416- <i>SmaI</i> -Rv ^C	ACTAGTGGATCCCCCAGTTGGCTGAATTGA
14	UFD2-Fw●	GACTTCCTGTGGATGAAG
15	UFD2-Rv●	CTCTCCTCTCTTGTGGTG
16	ACR2-Fw●	AGGCAACTCAAGGCCTAAT
17	ACR2-Rv●	GAACATGCCAAGCGTTTGTA
18	ACR3-Fw●	AAGAGGGTCTGGGGAAGAAA
19	ACR3-Rv●	GCAATTGCCAGGGATAGTTC
20	YAP8-Fw●	AACCGCCACATGTAACACT
21	YAP8-Rv●	TCCAACACACACTGAGAGCAG
22	ACT1-Fw●	CTATTGGTAACGAAAGATTCA
23	ACT1-Rv●	CCTTACGGACATCGACATCA

Fw - forward; Rv - reverse

^DGene disruption and mutant confirmation, *Sequencing, ^C Cloning, ●qRT-PCR