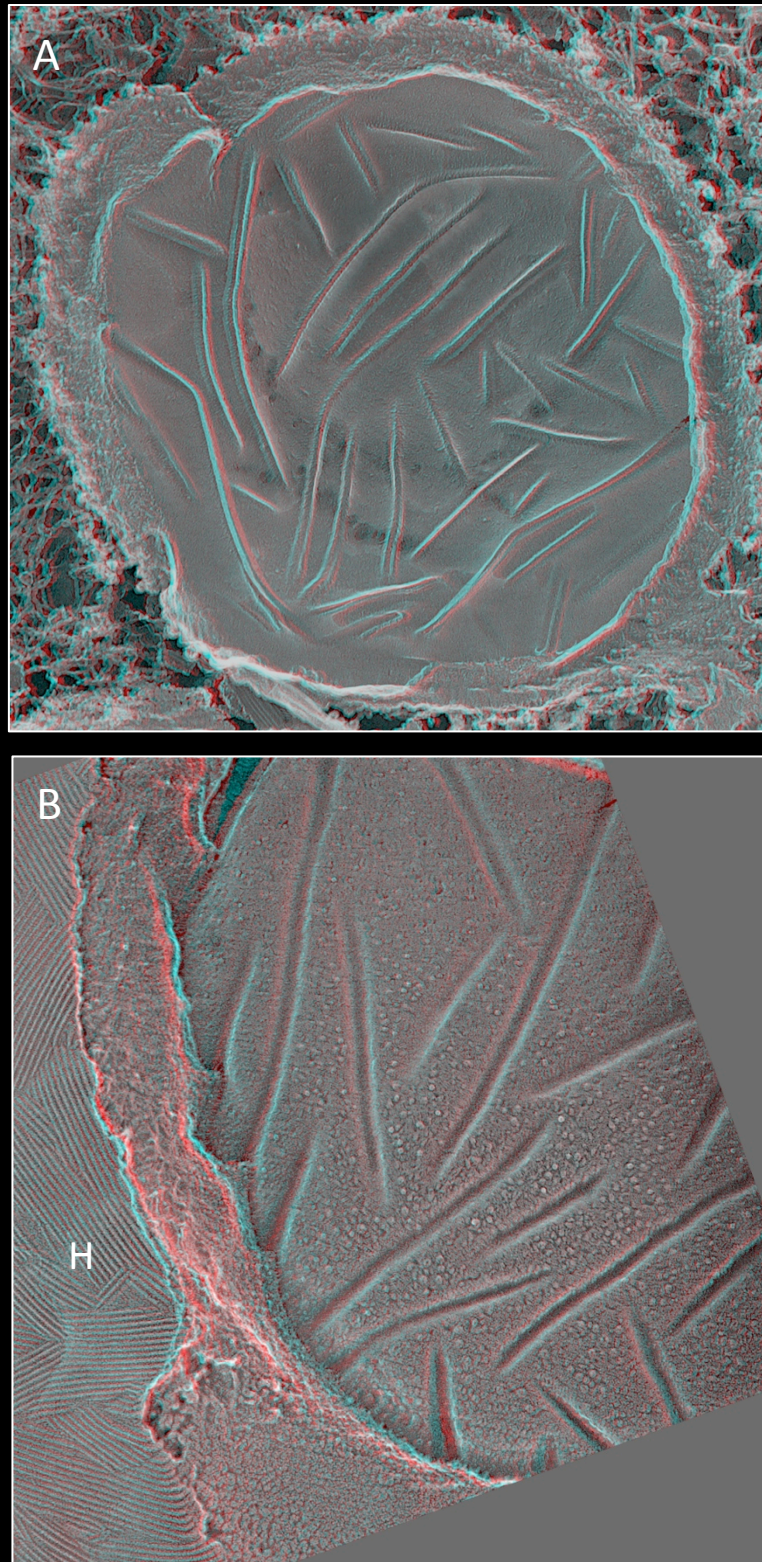


FIG S1 Convex (A) and concave (B) eisosome faces of *Penicillium* sp. isolate K17. Stereo images (use red/green glasses)



H, hydrophobin layer of cell wall

FIG S2 *Candelaria concolor* mycobiont:
(A) and (B) convex faces, (C) concave face

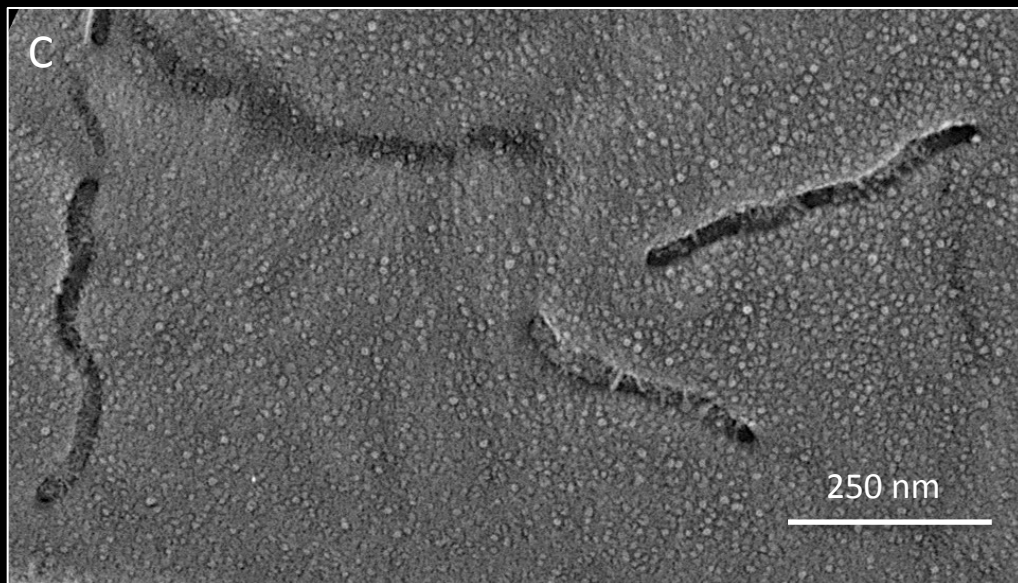
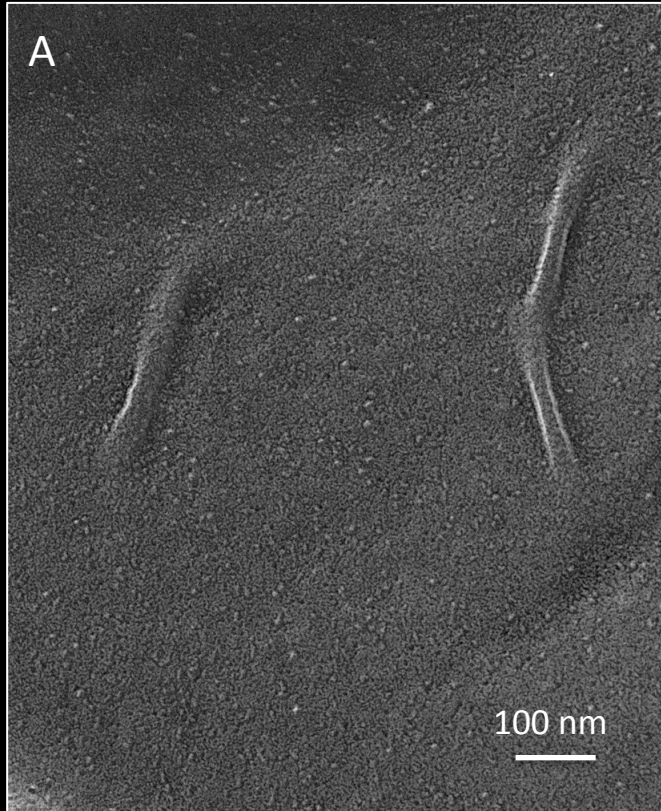


FIG S3 *Cladonia grayi* mycobiont: convex face

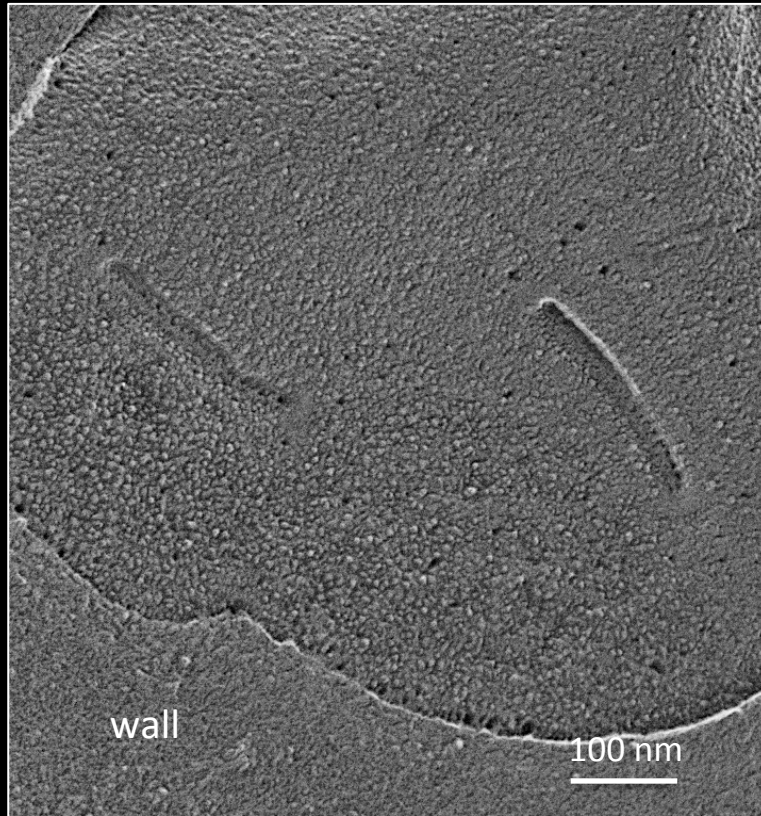


FIG S4 *Cladonia grayi* on agar:
(A) convex face
(B) concave face

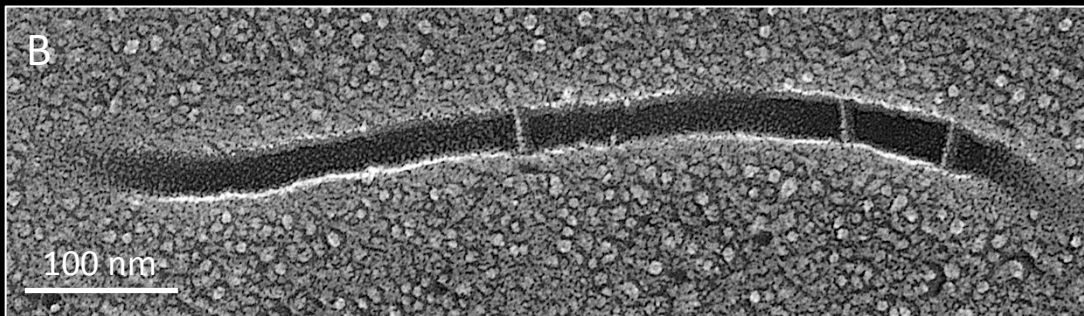
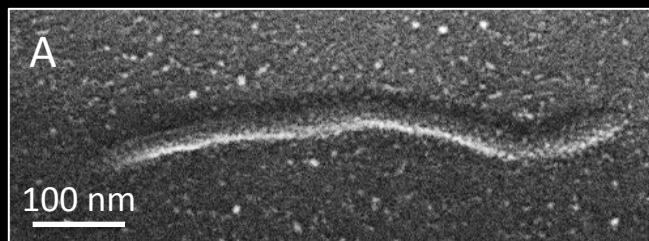


FIG S5 (A) *Cyanidioschyzon merolae*
(B) *Cyanidioschyzon* YNP 1A

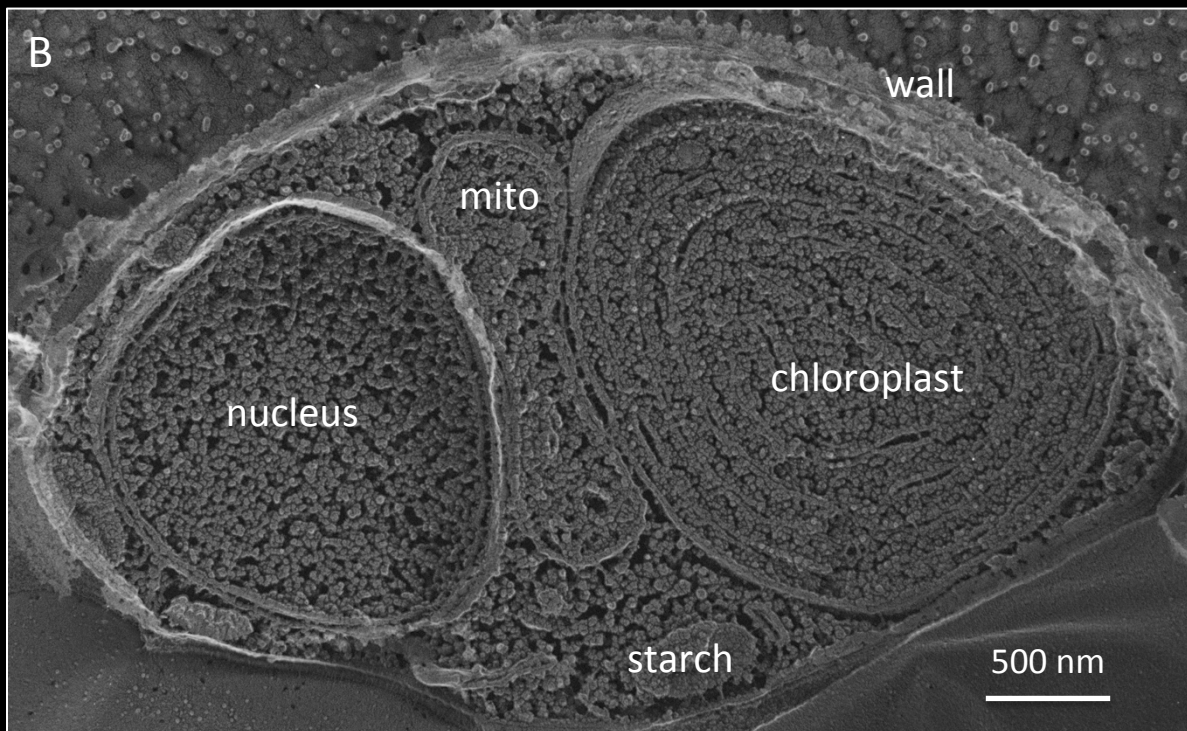
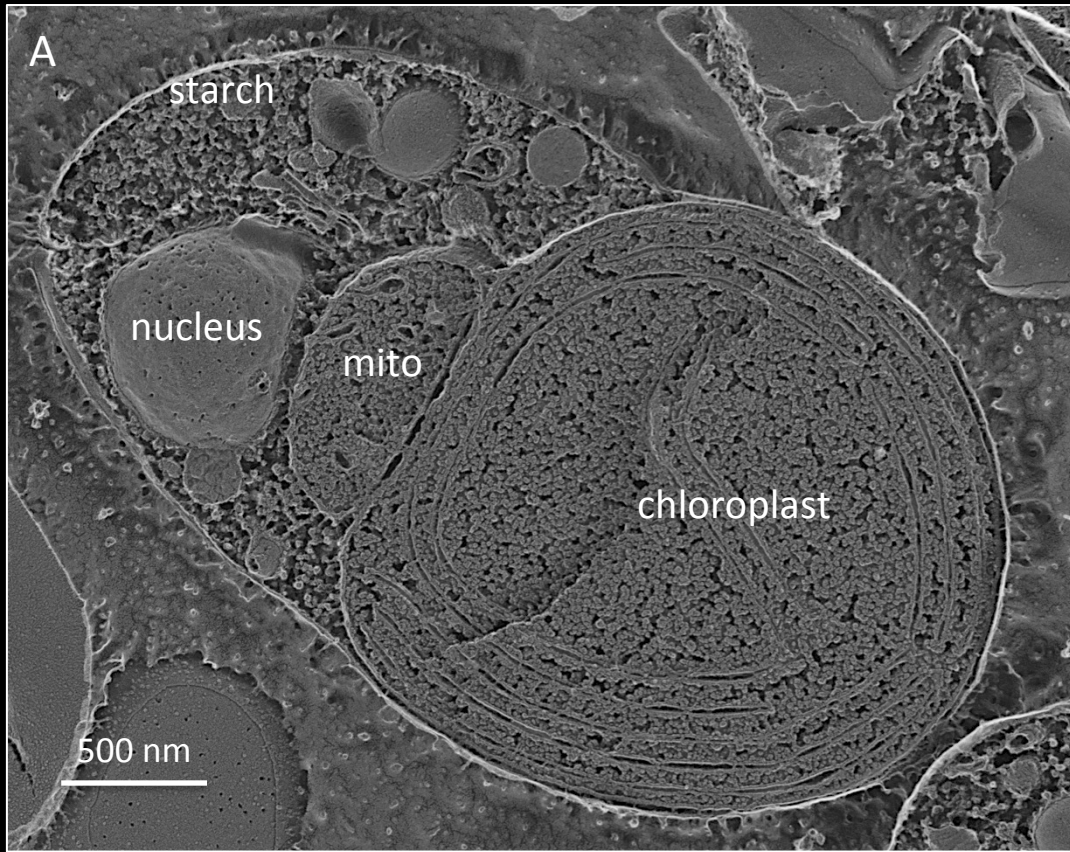
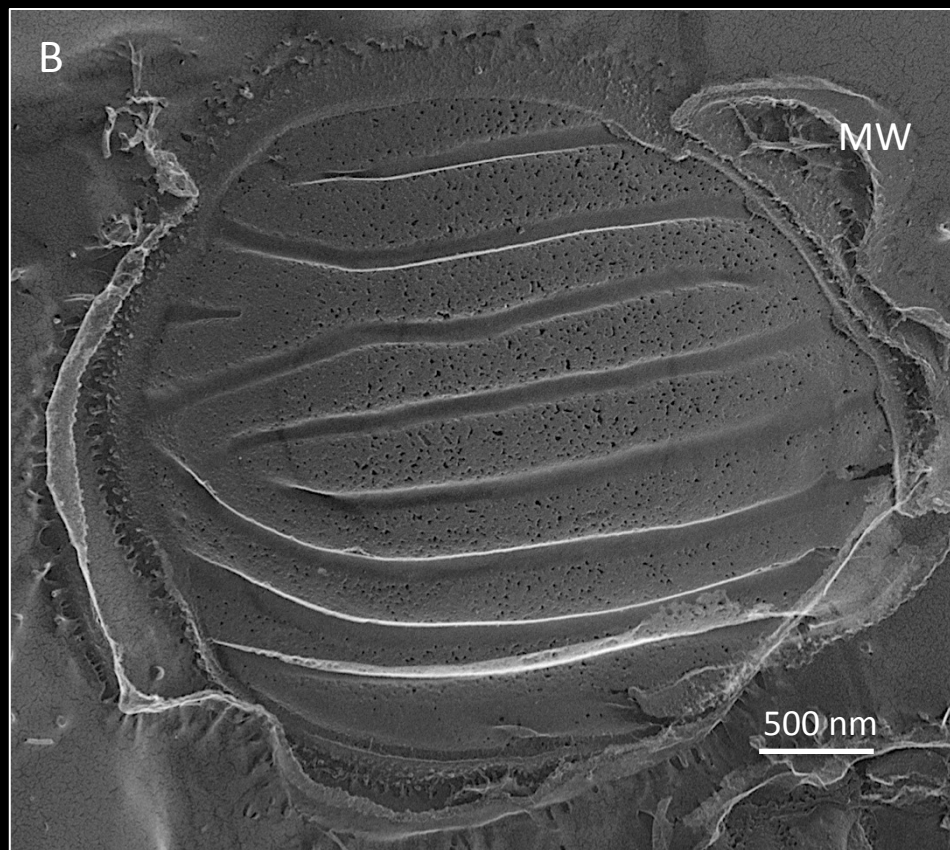
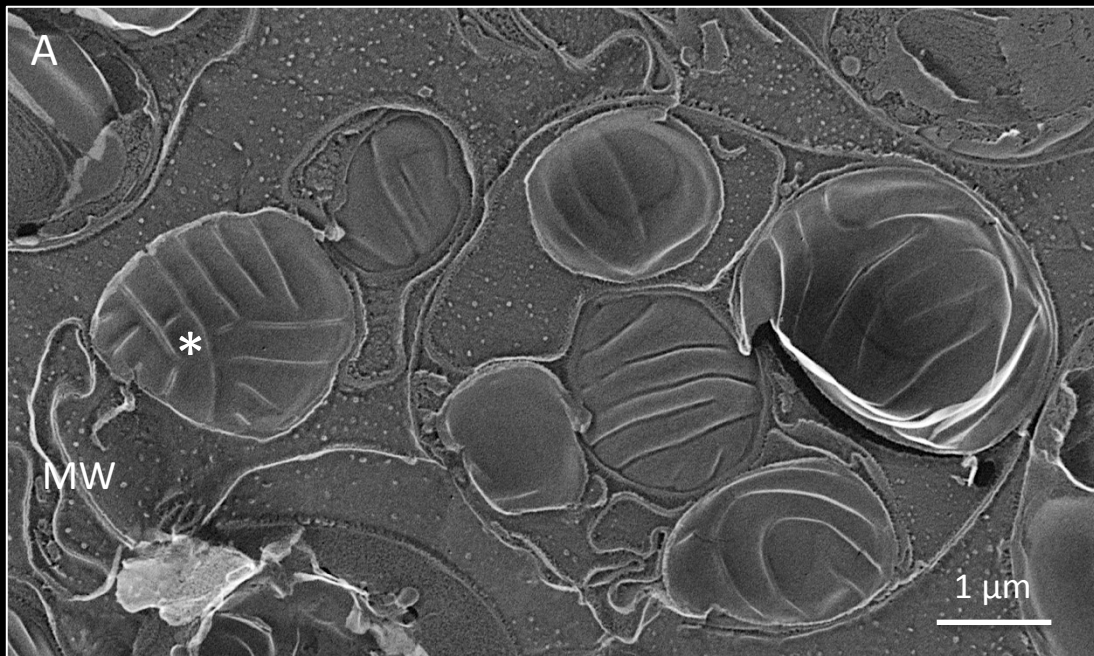


FIG S6 *Cyanidioschyzon* YNP 1A:
Eisosomes in mature and daughter cells



The eisosomes are generally circumferential, but exceptions occur (asterisk). MW, discarded mother walls.

FIG S7 *Cyanidioschyzon* YNP 1A:
Eisosomes in mature and daughter cells

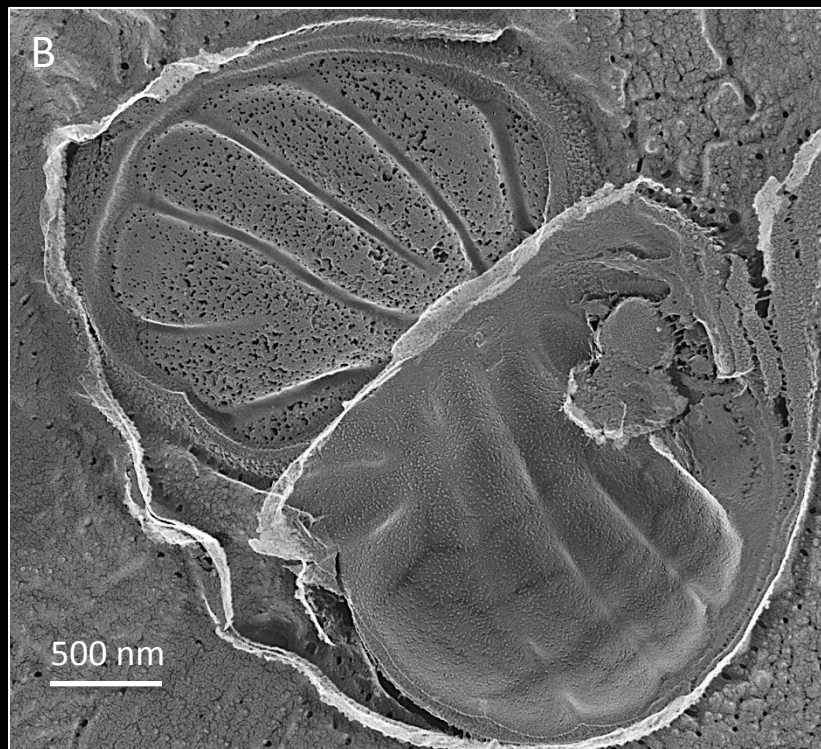
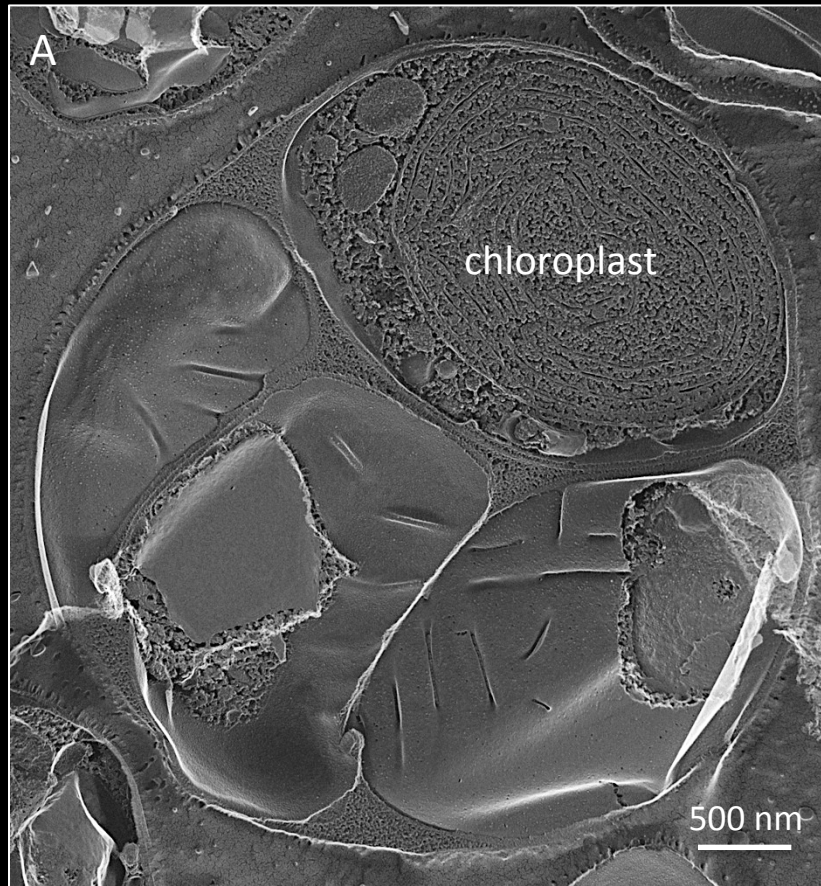


FIG S8 *Cyanidioschyzon* YNP 1A:
(A) convex face, (B) concave face

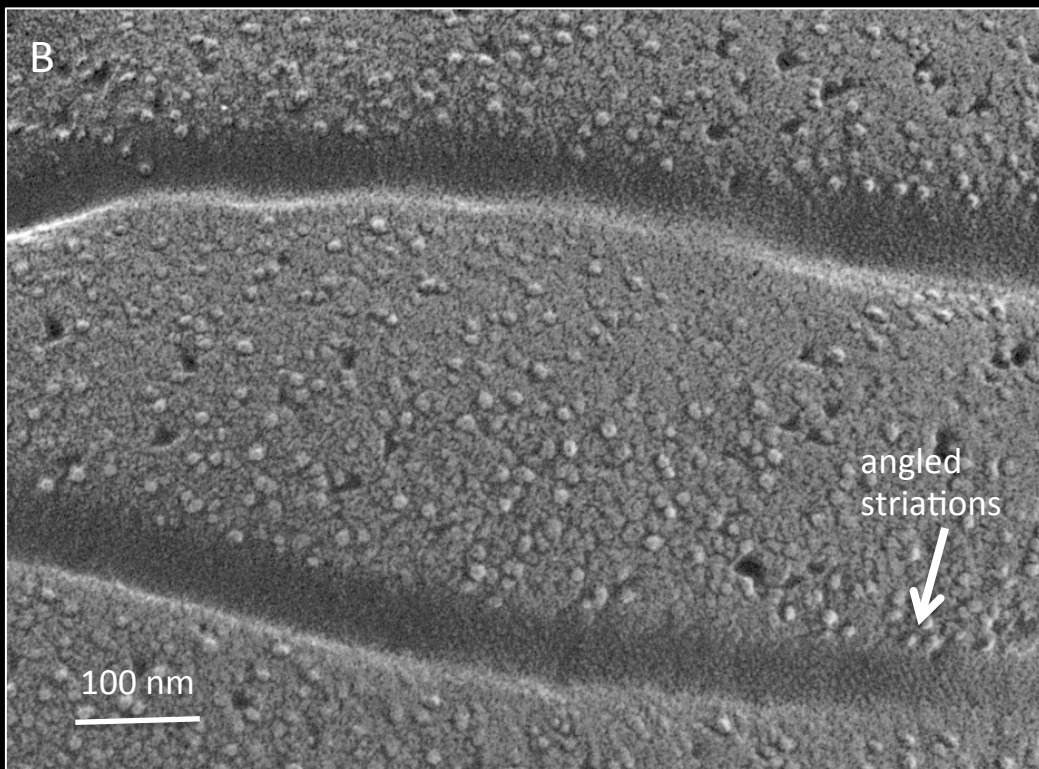
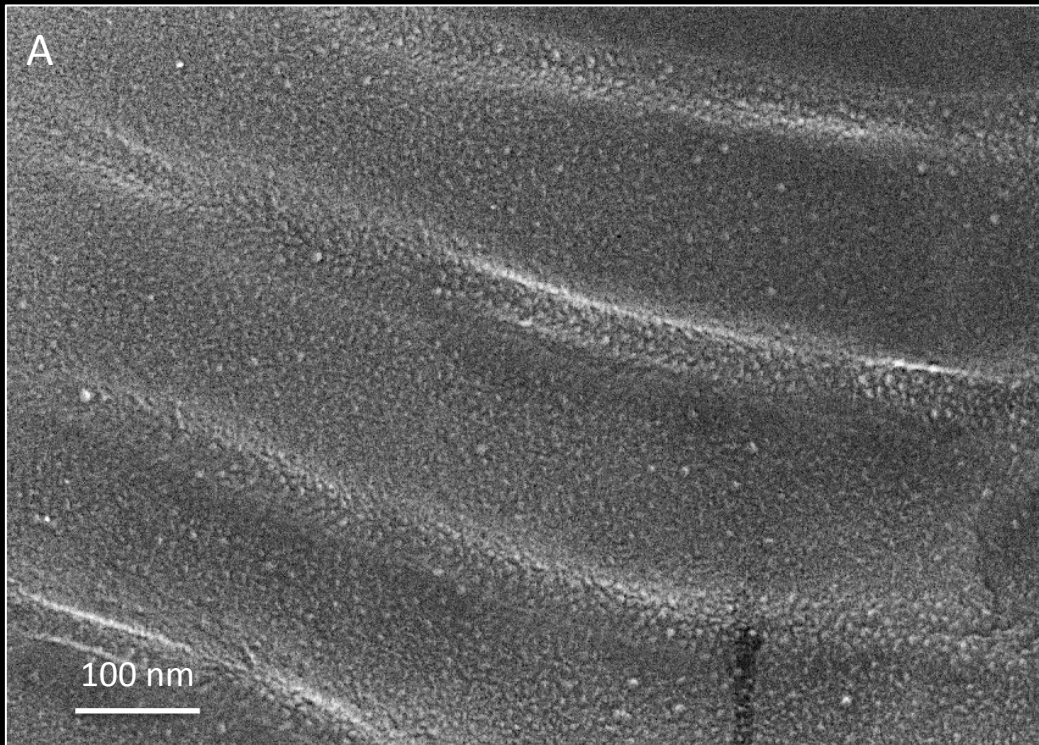
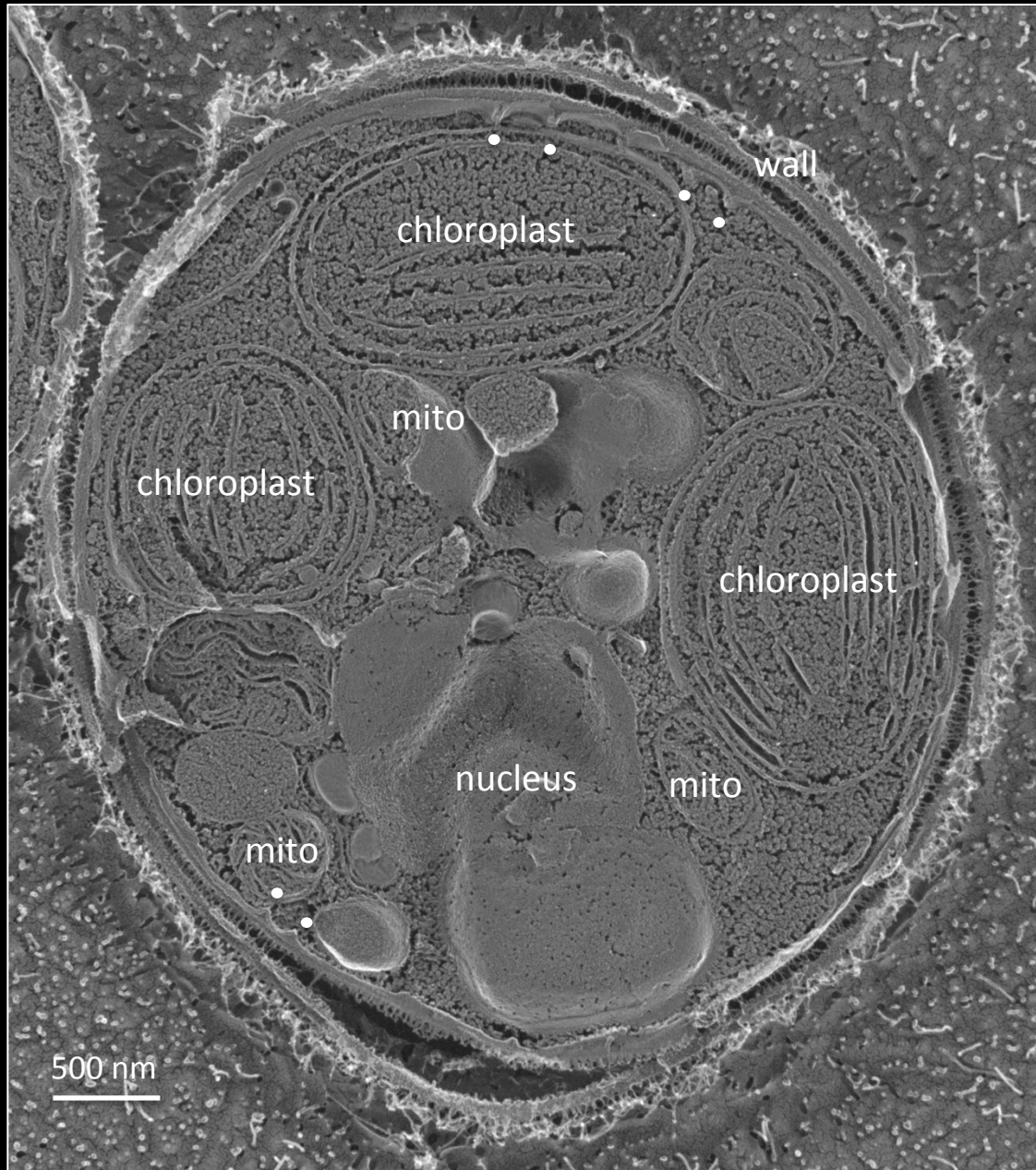
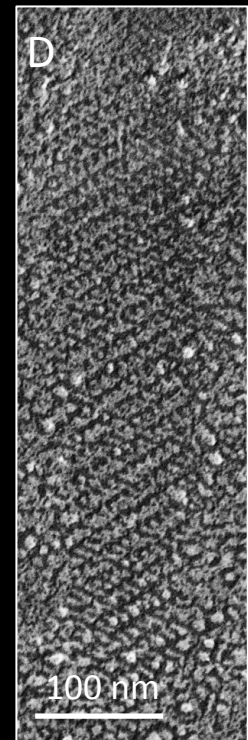
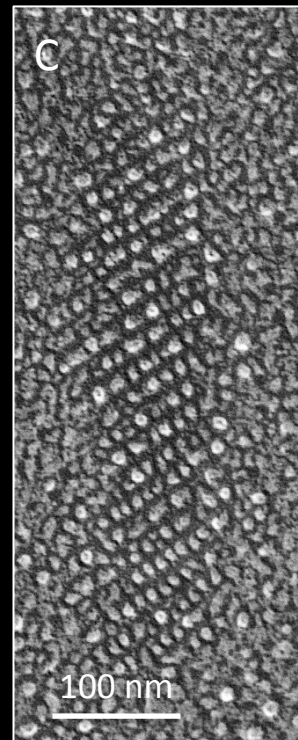
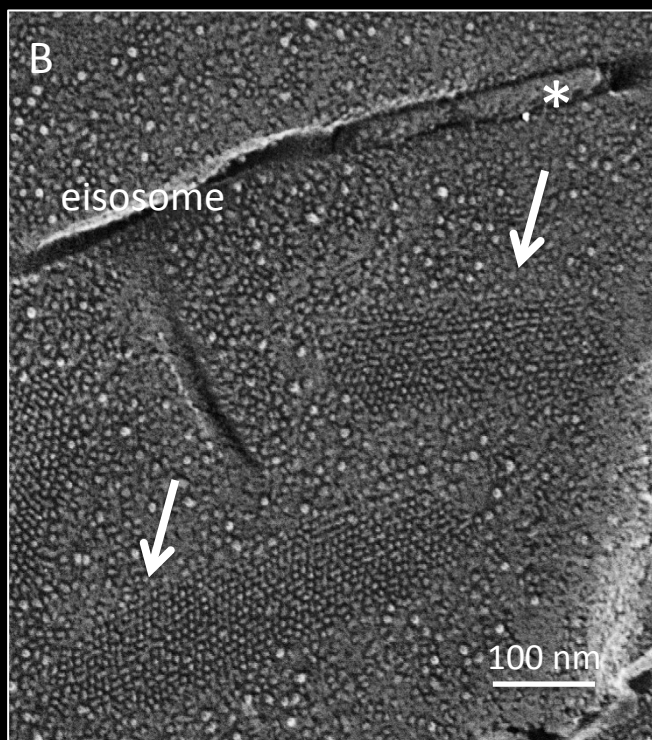
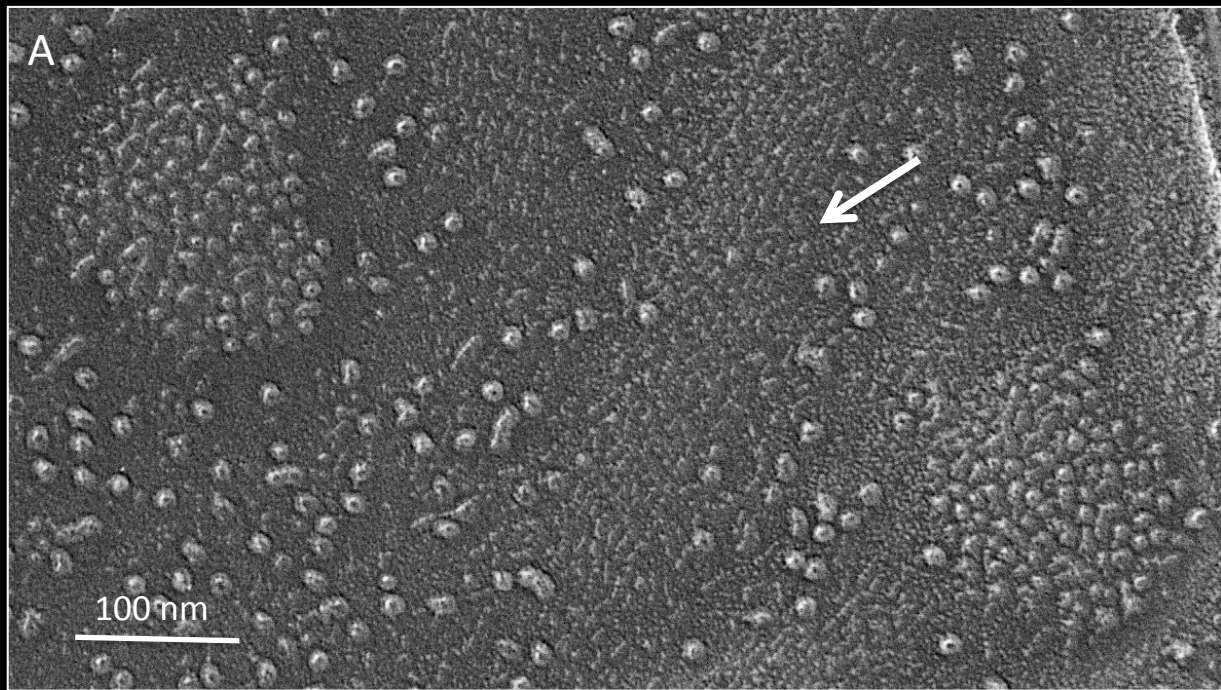


FIG S9 *Galdieria sulphuraria*



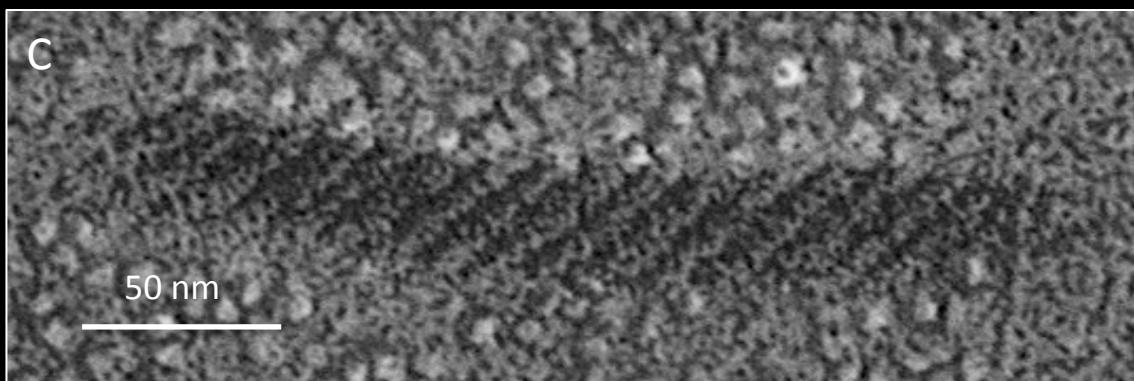
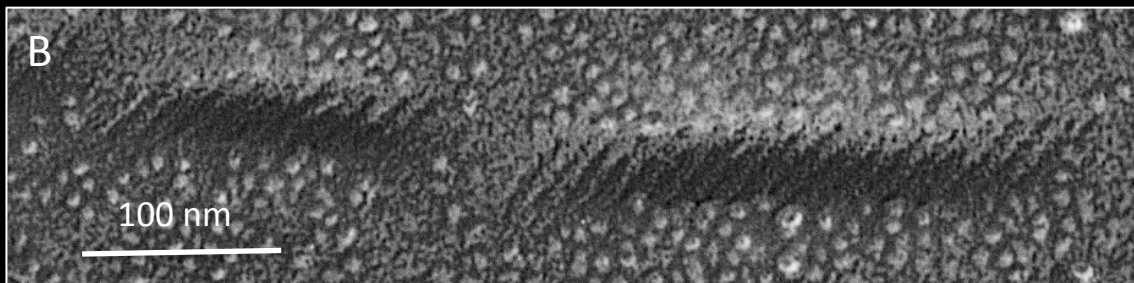
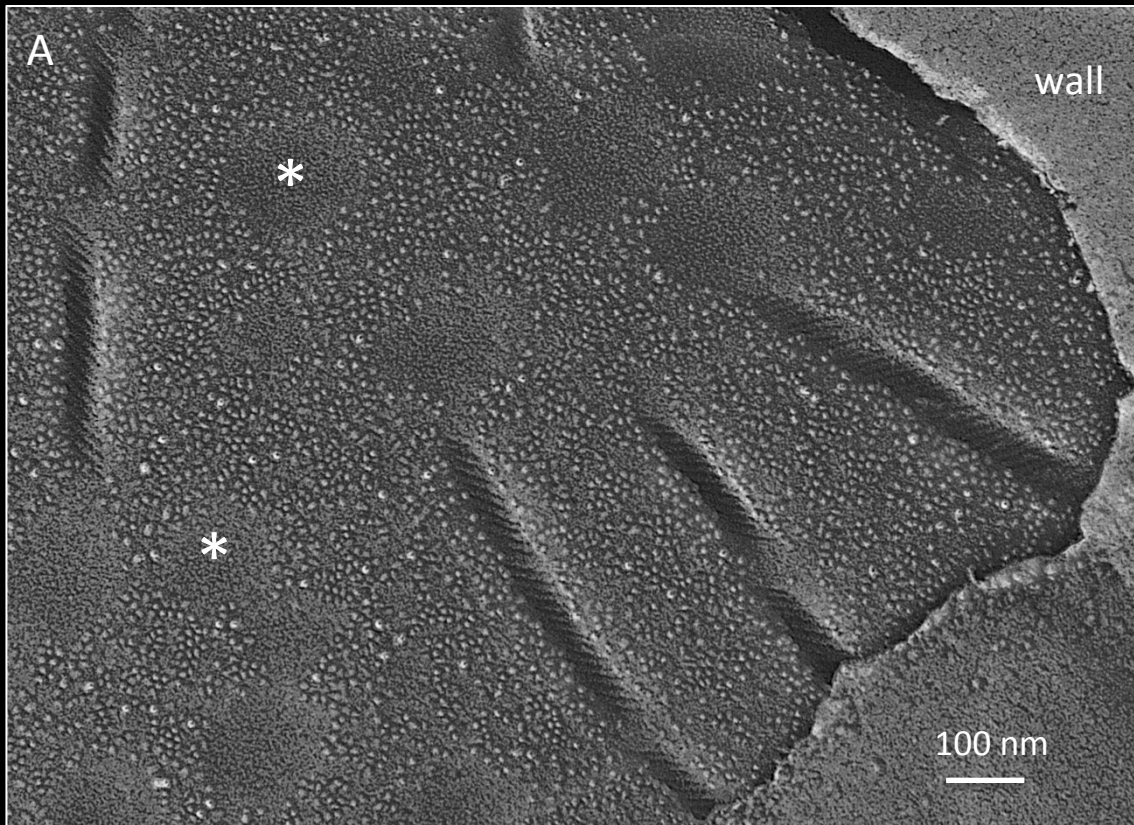
G. sulphuraria cell with several chloroplast profiles, several mitochondria, and no cytoplasmic starch. Eisosomes at white dots.

FIG S10 *Galdieria sulphuraria*:
Non-etched plasma membrane differentiations



(A) Circular IMP “islands” and parallel rectangular arrays (arrow).
(B) Parallel rectangular arrays (arrows) and eisosome with cell-wall inclusion (asterisk). (C and D) Rectangular arrays of angled IMPs.

FIG S11 *Galdieria sulphuraria*
Non-etched concave eisosomes



Asterisks: P-faces of circular IMP "islands" (c.f. Fig. S10)

FIG S12 *Galdieria sulphuraria*: non-etched convex faces.
Stereo image (use red/green glasses)

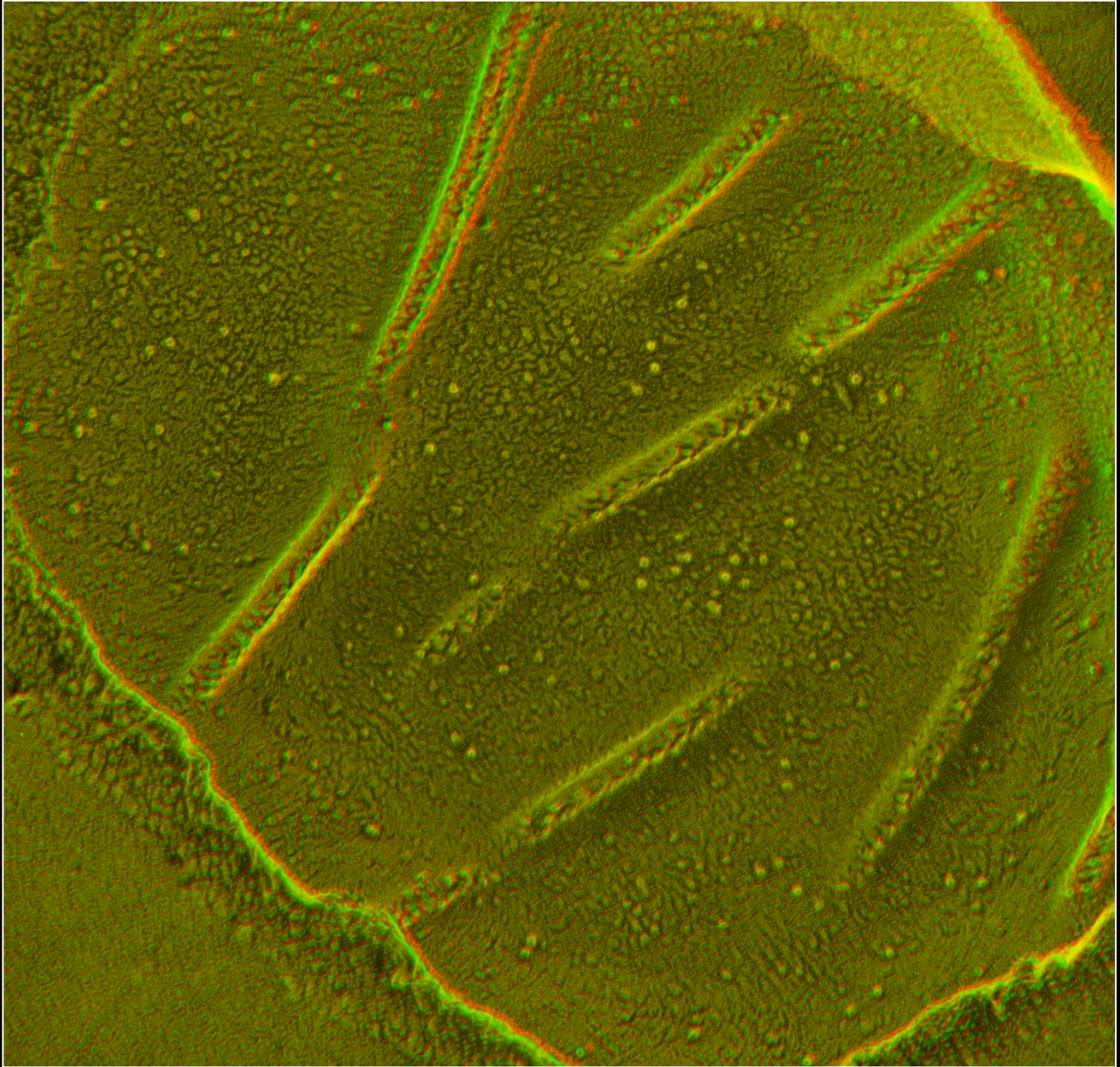
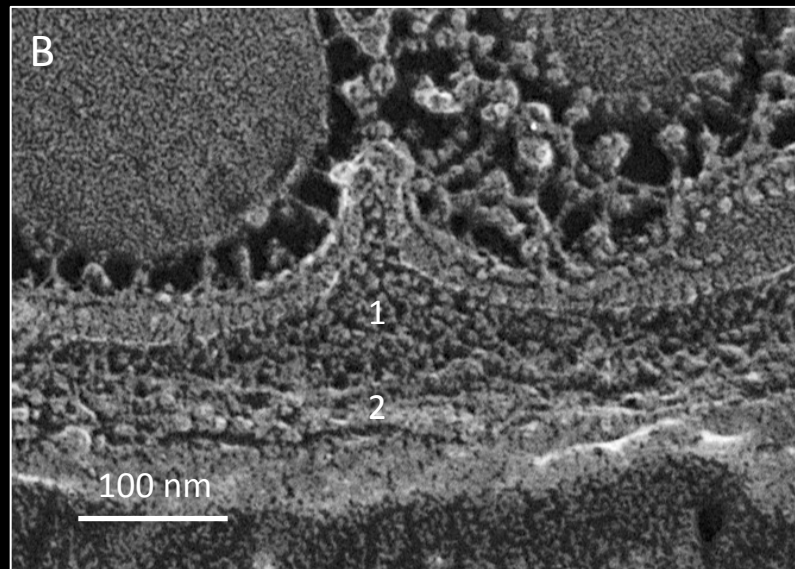
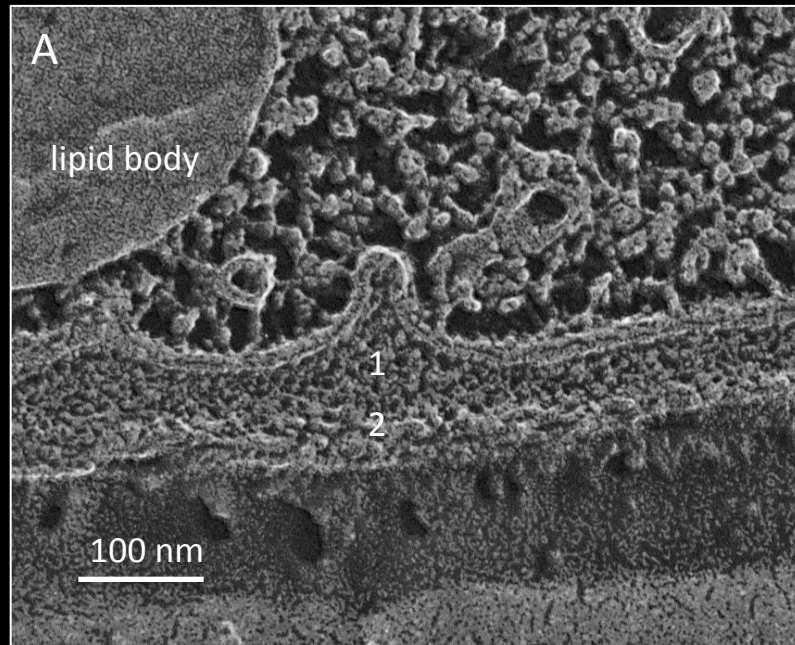


FIG S13 *Galdieria sulphuraria*: non-etched concave faces (arrows)
Stereo image (use red/green glasses)

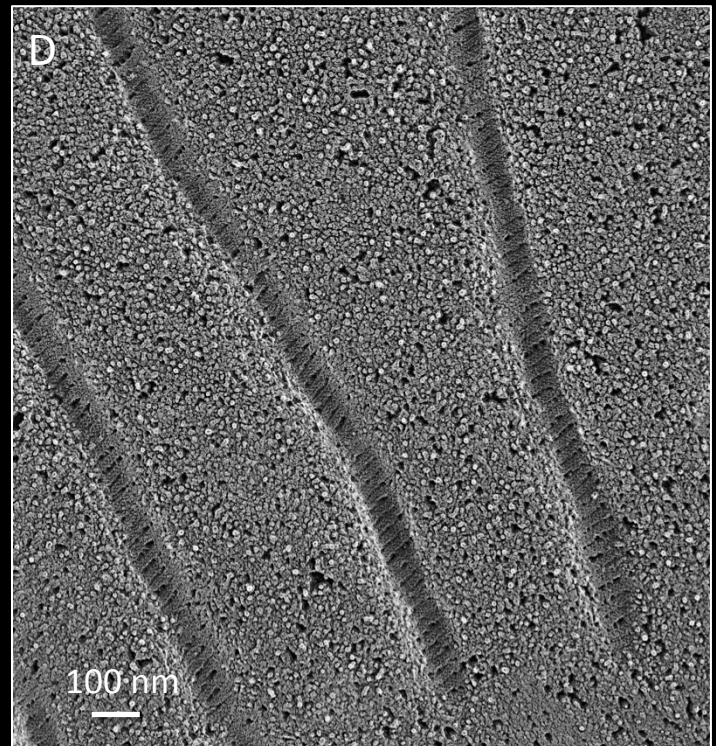
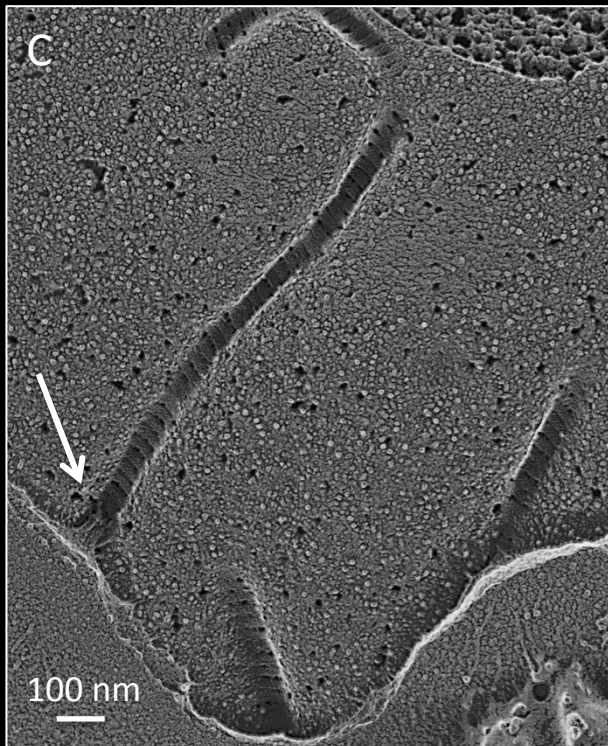
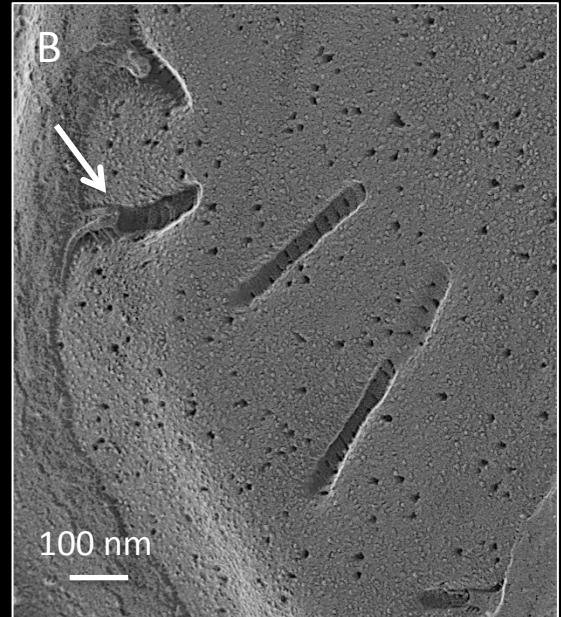
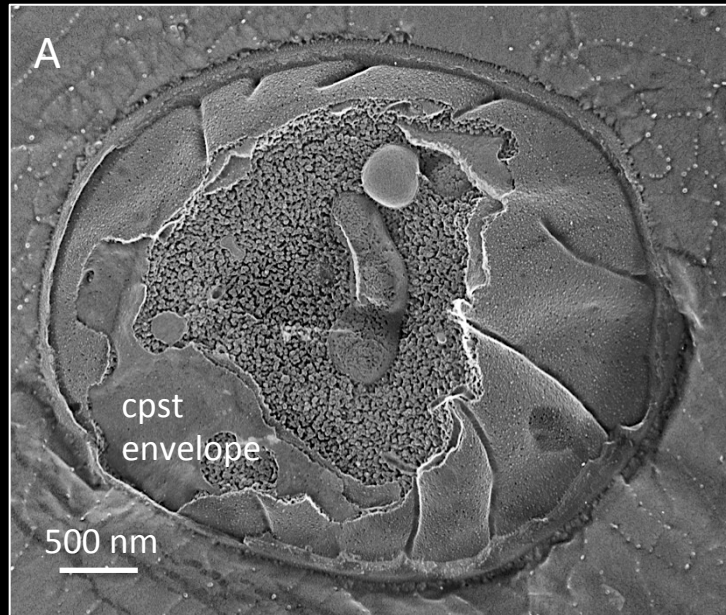


FIG S14 *Auxenochlorella protothecoides*:
eisosomes and walls



The inner granular layer 1 penetrates the eisosome furrow; the outer denser layer 2 does not.

FIG S15 *Auxenochlorella protothecoides* eisosomes:
Concave faces (most elongated)



Arrows, wall entering eisosomes; cpst, chloroplast

FIG S16 *Auxenochlorella protothecoides* eisosomes:
Convex faces (punctate)

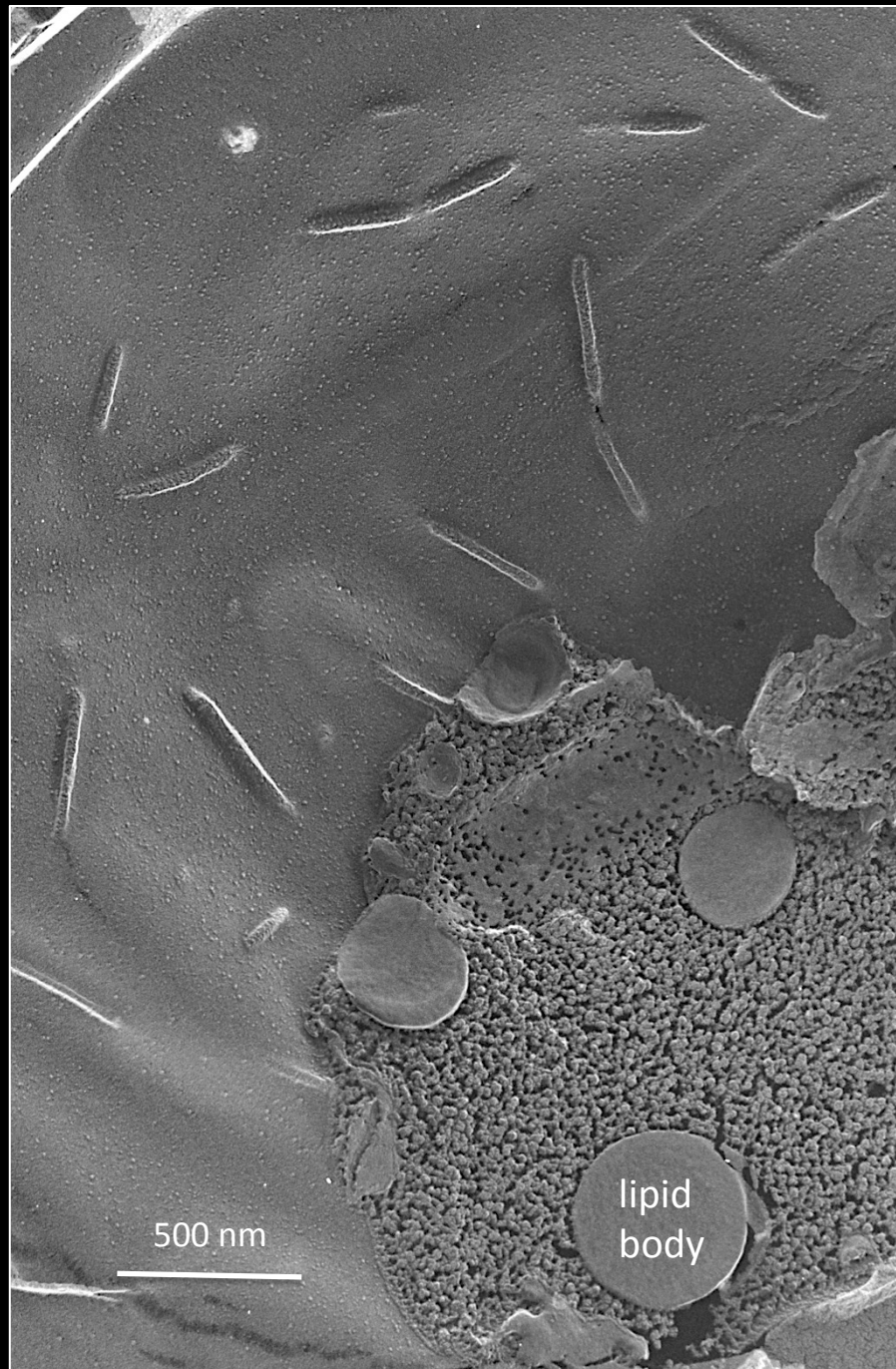


FIG S17 *Auxenochlorella protothecoides* eisosomes:
Periodicity of angled striations

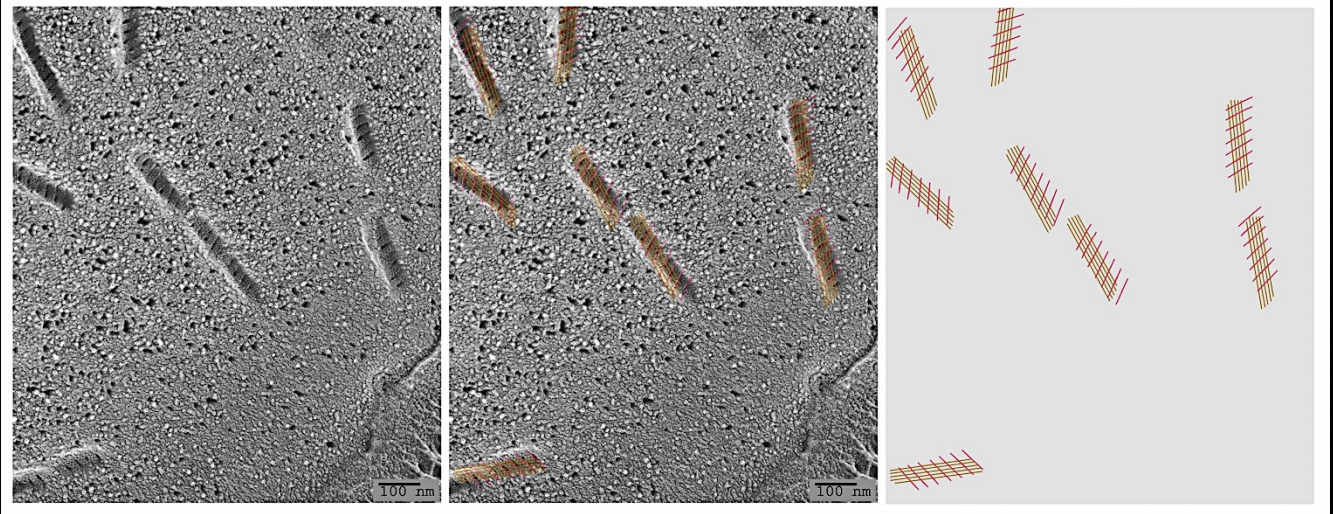


FIG S18 *Chlamydomonas monoica*:
Convex faces with irregularly shaped and distributed particles

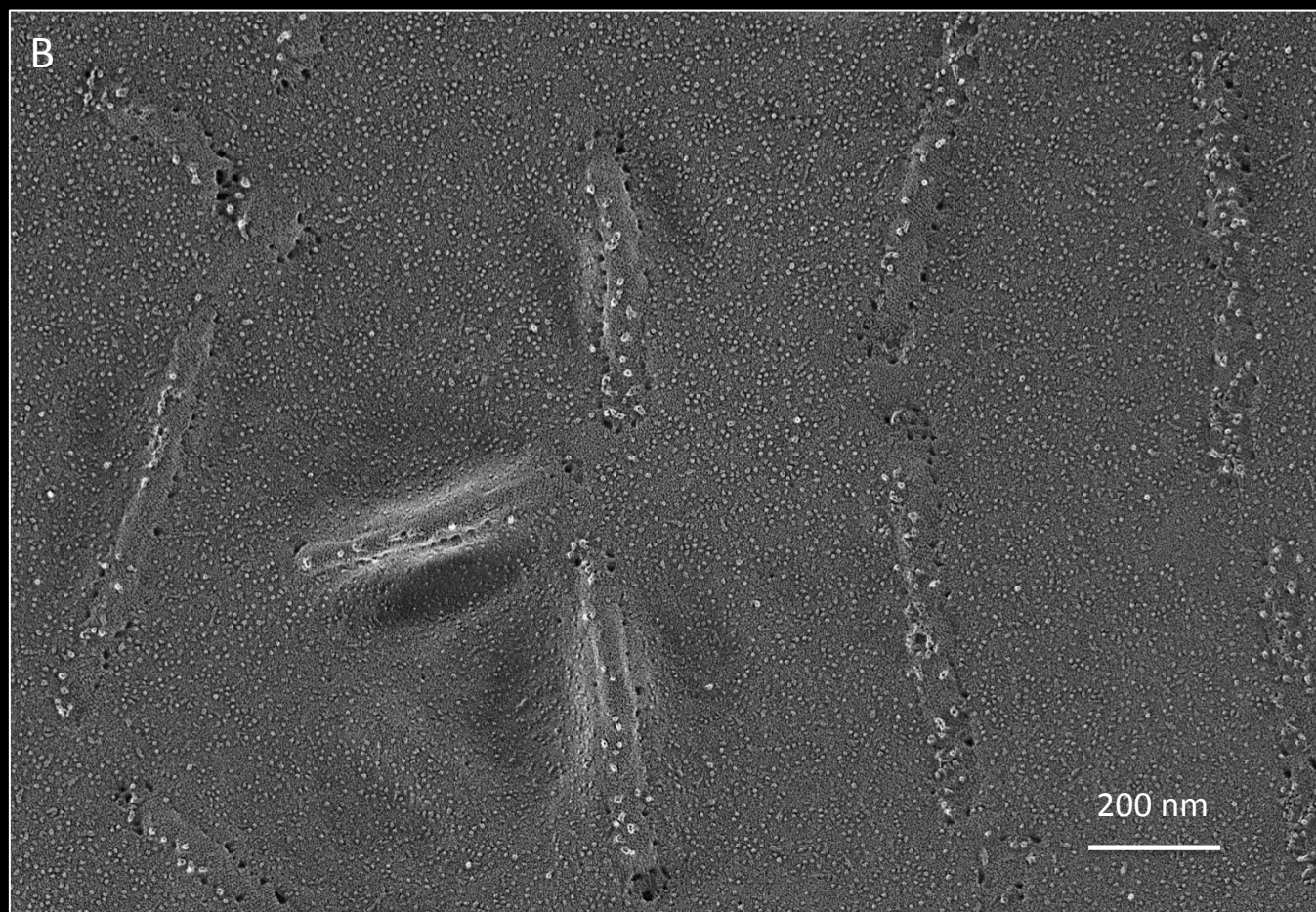
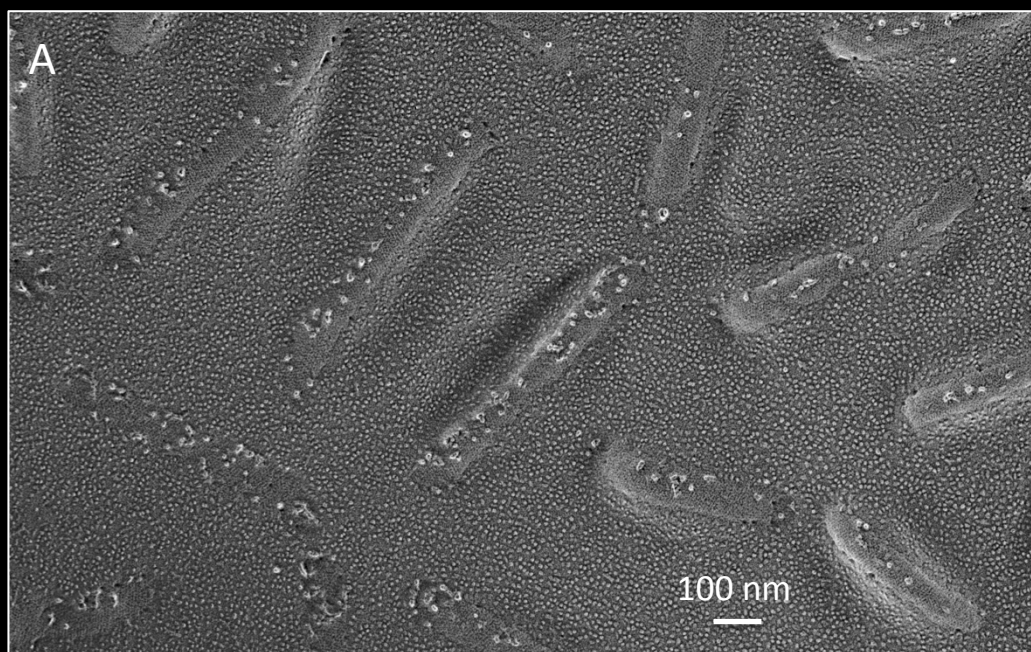


FIG S19 *Chlamydomonas monoica* concave faces.
Stereo image (use red/green glasses)

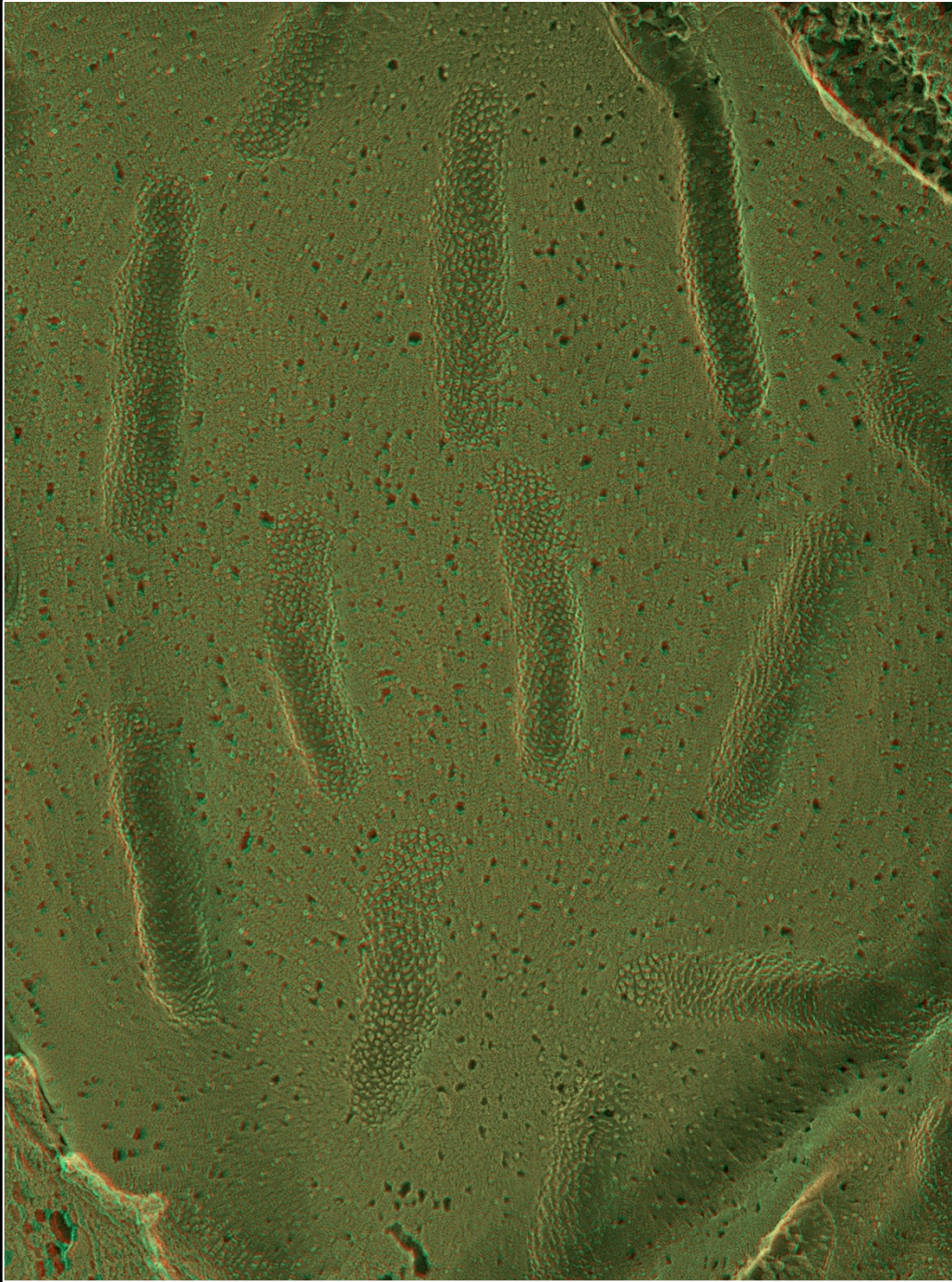


FIG S20 *Chlamydomonas monoica* convex faces
Stereo image (use red/green glasses)

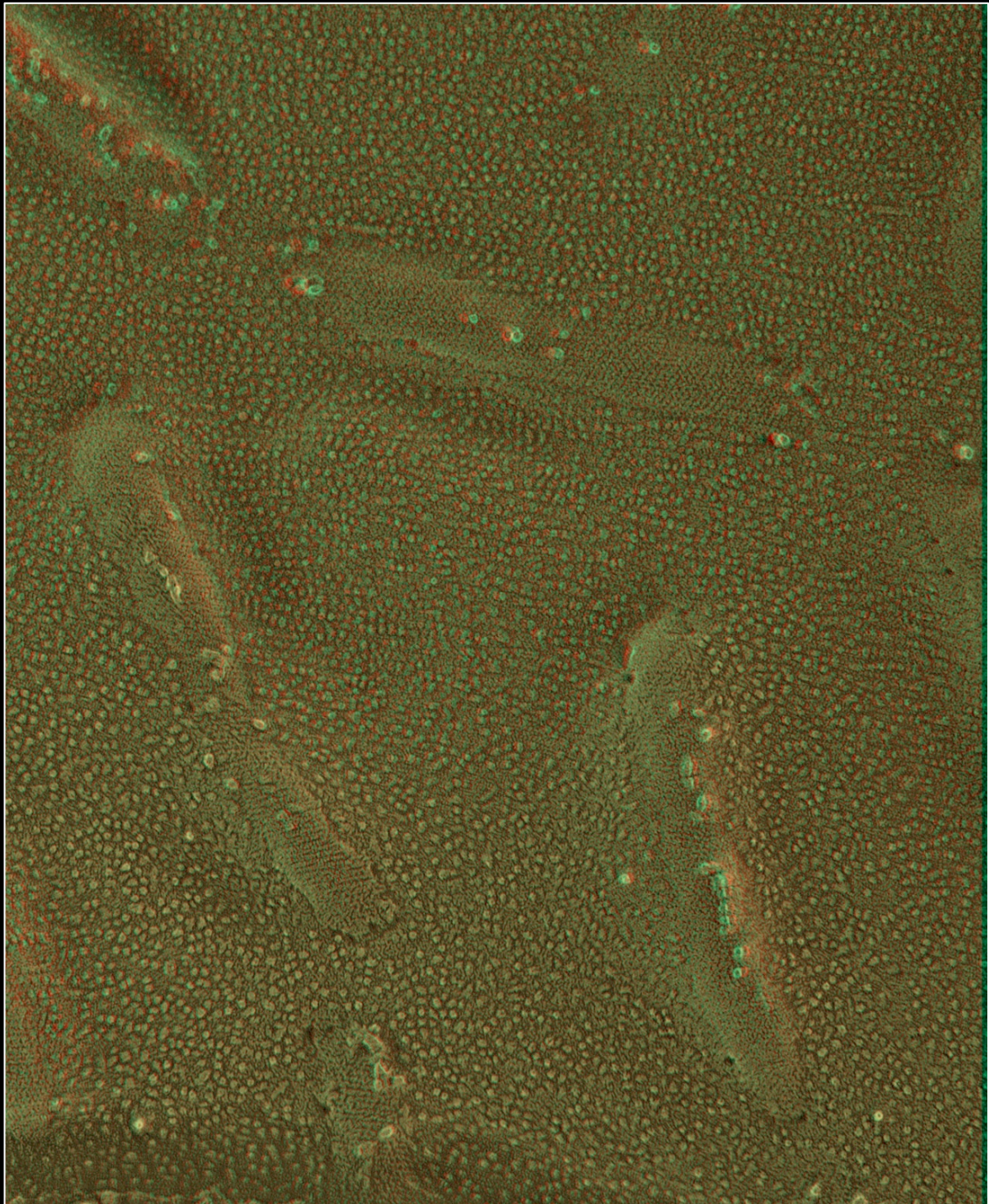


FIG S21 *Chlamydomonas monoica* concave faces.
Stereo image (use red/green glasses) (see Fig. S22 for labels)

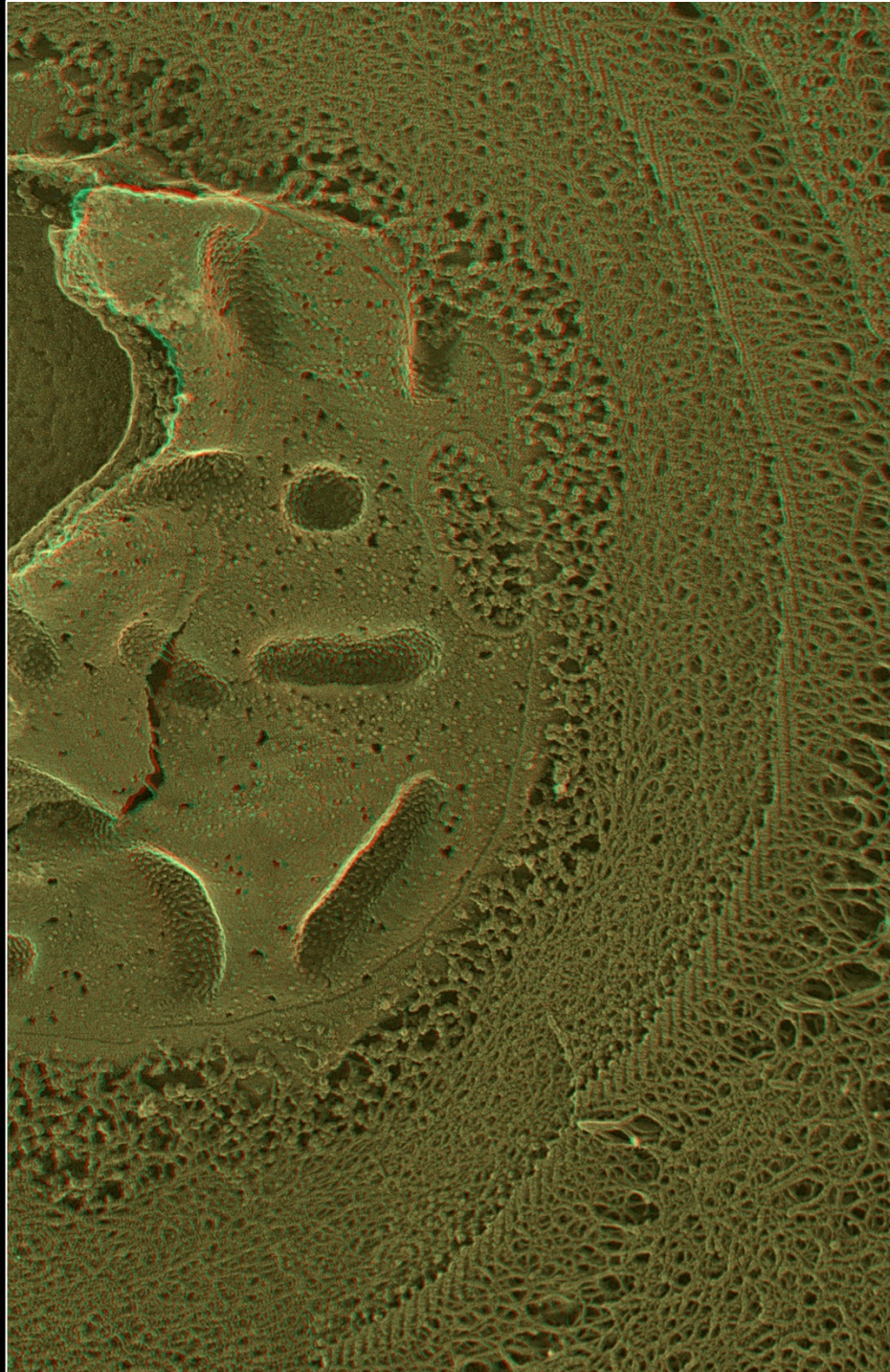
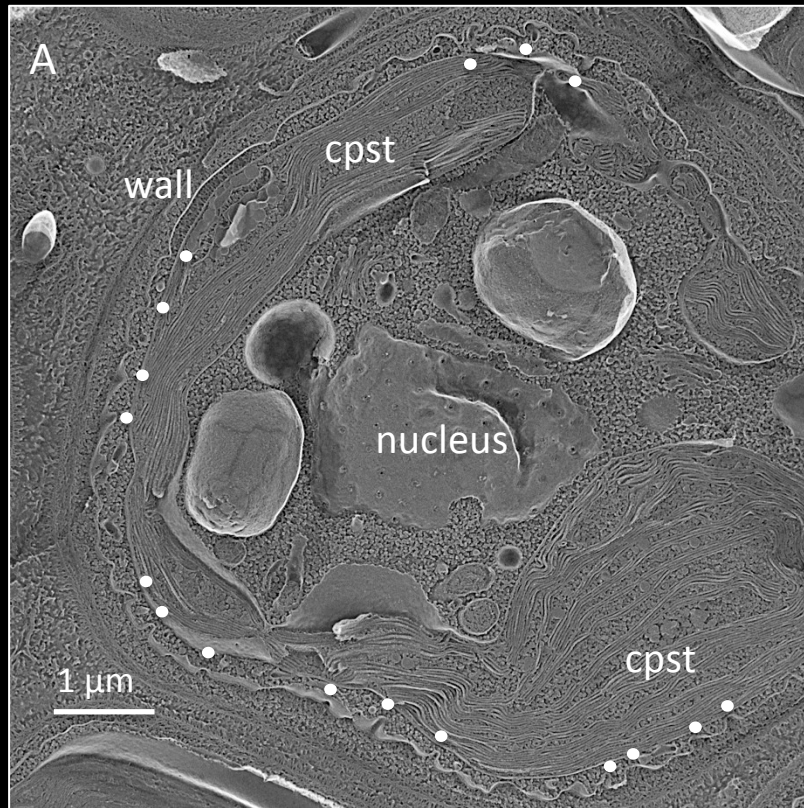


FIG S22 *Chlamydomonas monoica*:
Wall/eisosome relationships



White dots,
eisosomes;
cpst, chloroplast

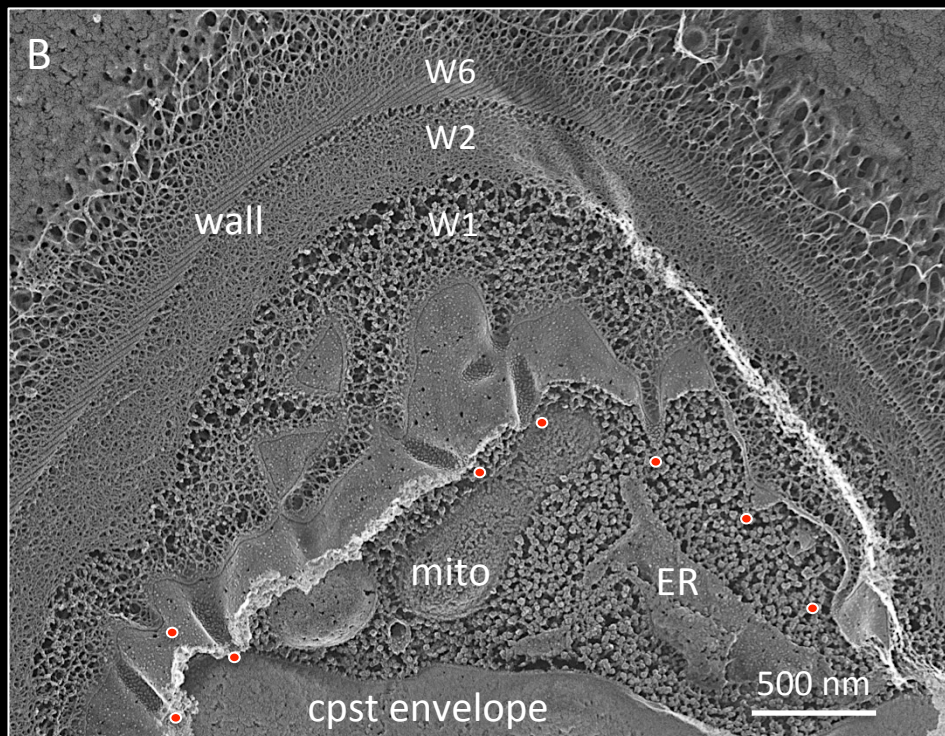
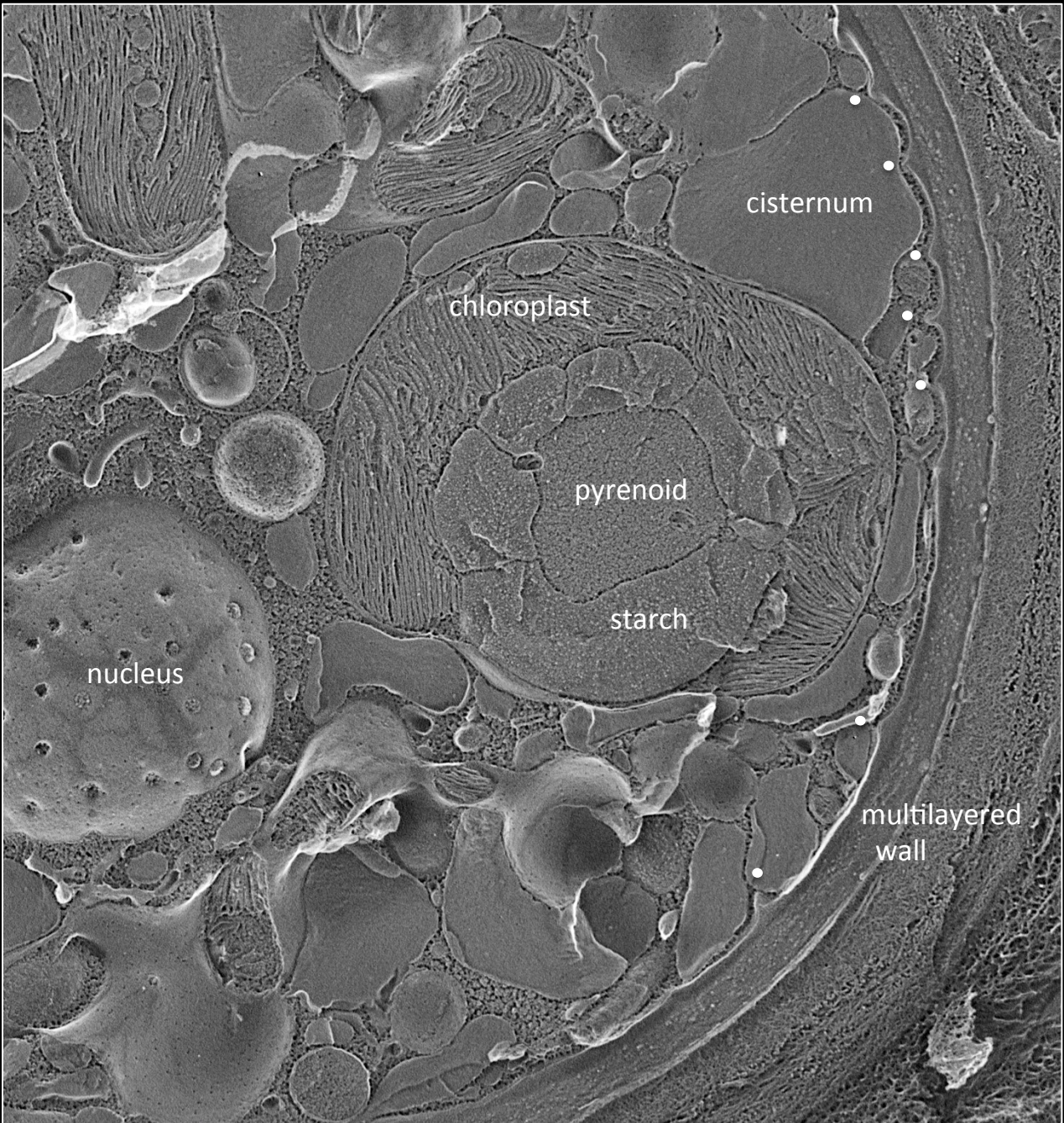


FIG S23 *Chlamydomonas reinhardtii* zygote



White dots, eisosomes

FIG S24 *Chlamydomonas reinhardtii* zygotes:
(A) concave faces (B) convex faces

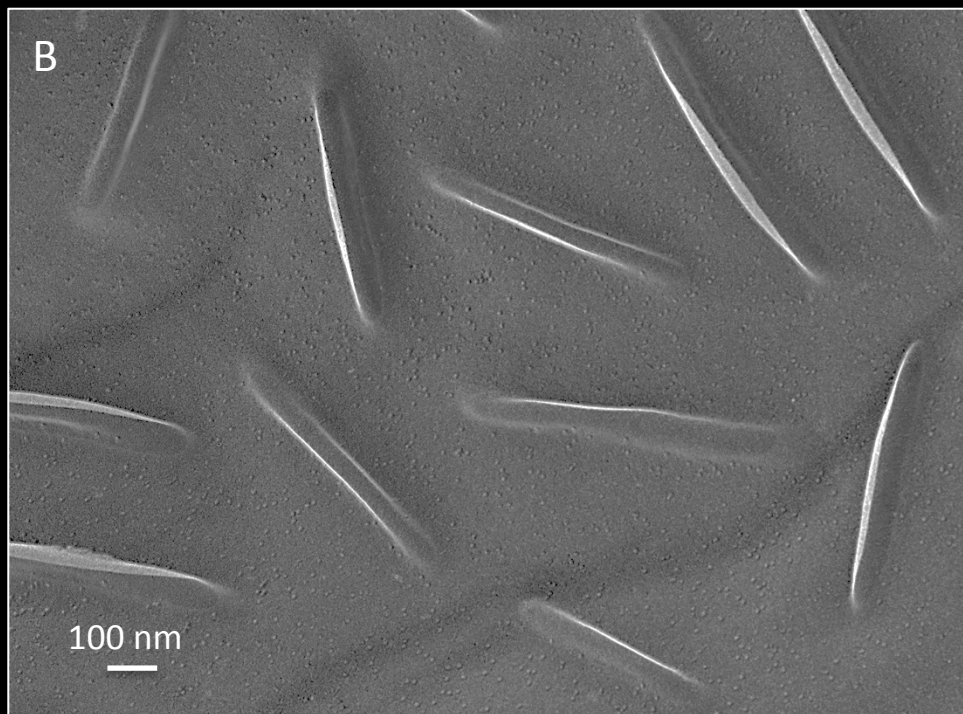
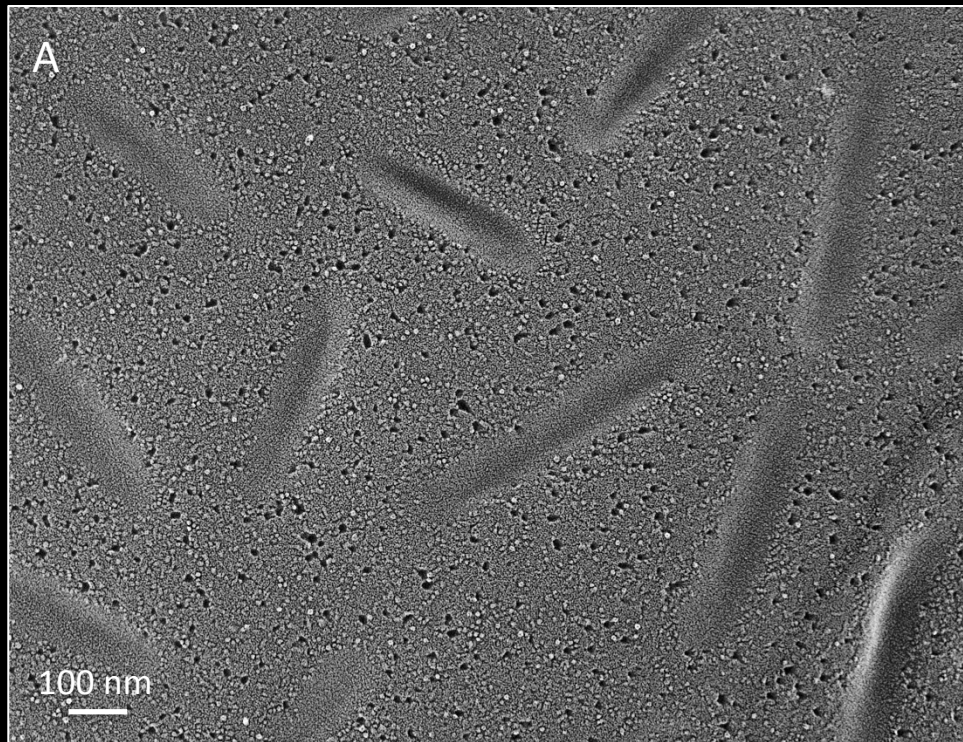
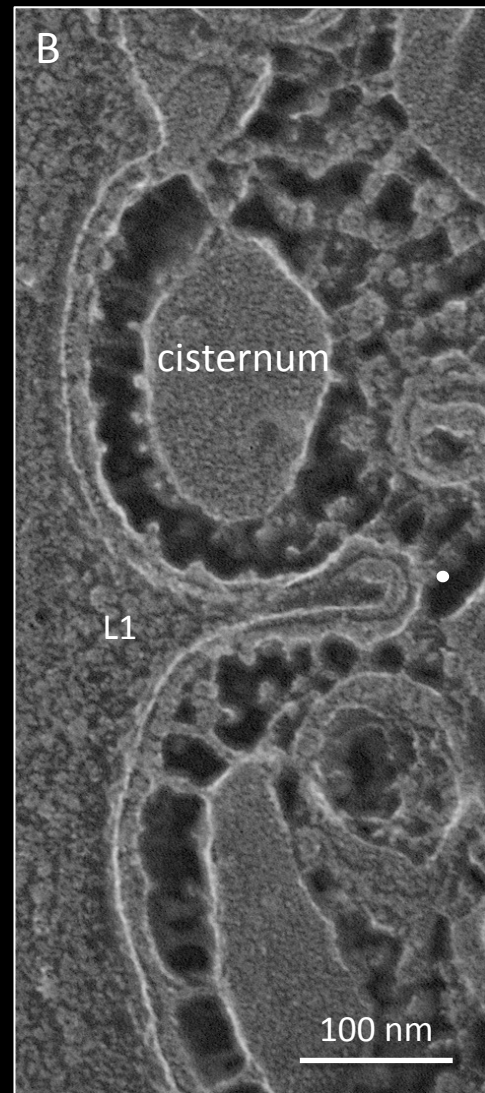
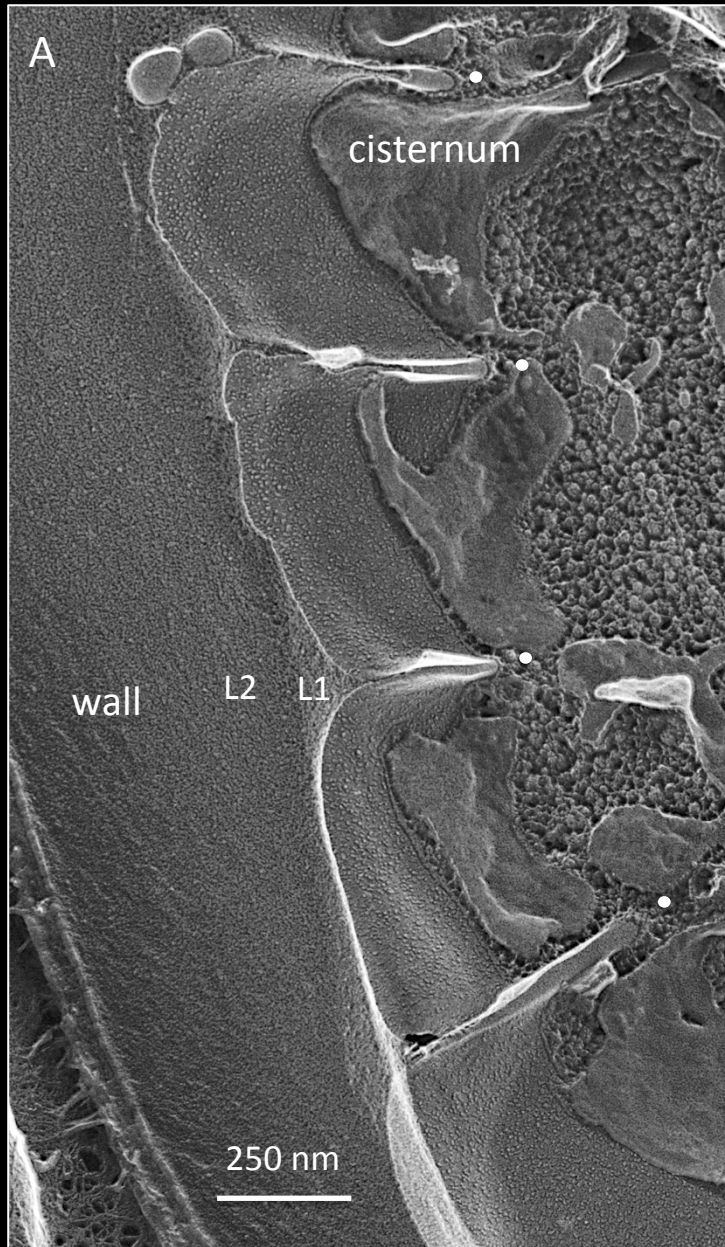
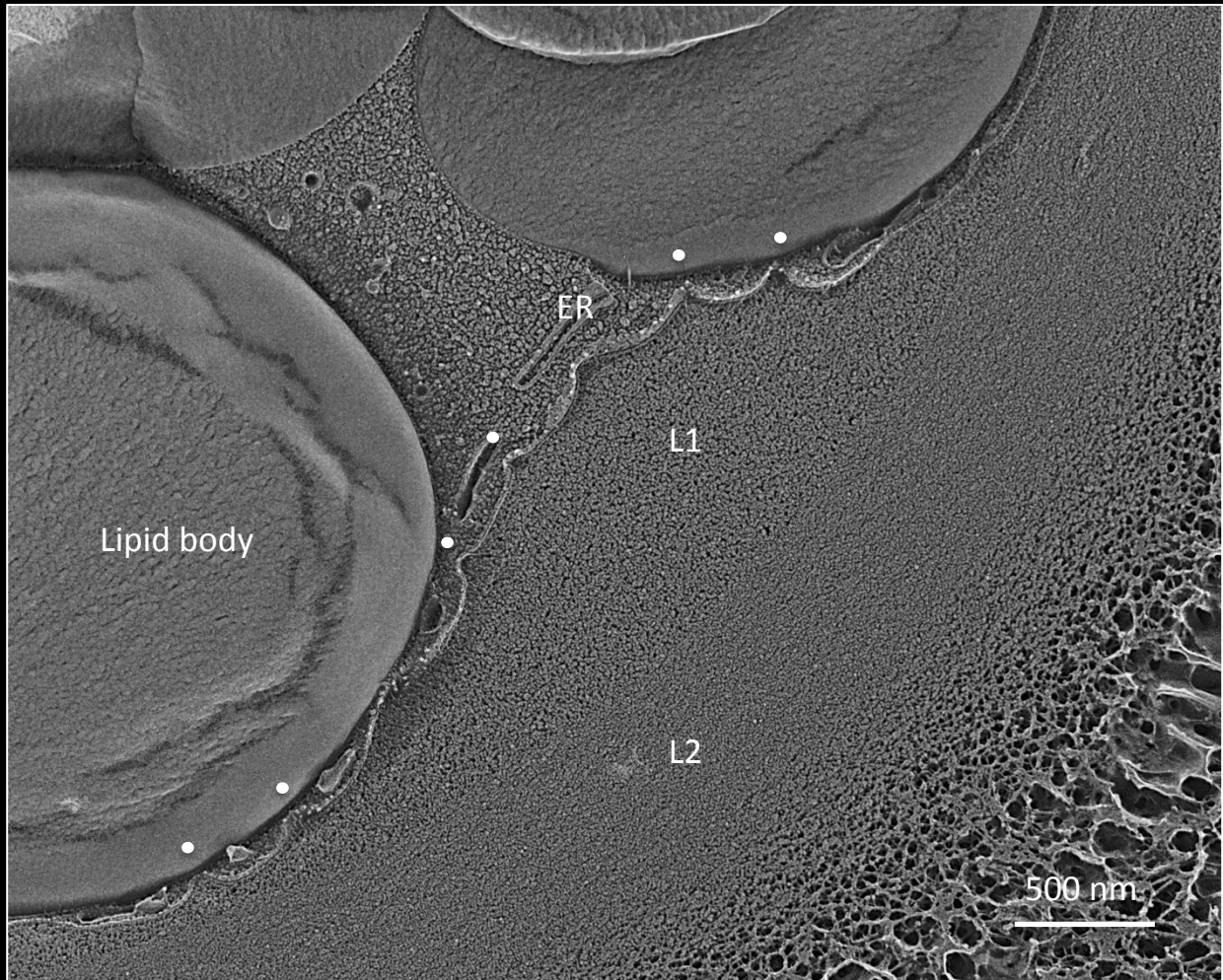


FIG S25 *Polytomella parva* cysts



White dots, eisosomes; L1, granular wall layer; L2 denser wall layer.

FIG S26 *Borodinellopsis texensis* cyst



White dots, eisosomes; L1, granular wall layer; L2 denser wall layer.

FIG S27 *Euplotes* sp. cysts
(A) Freeze-substituted thin section (B) convex faces
White dots: eisosomes

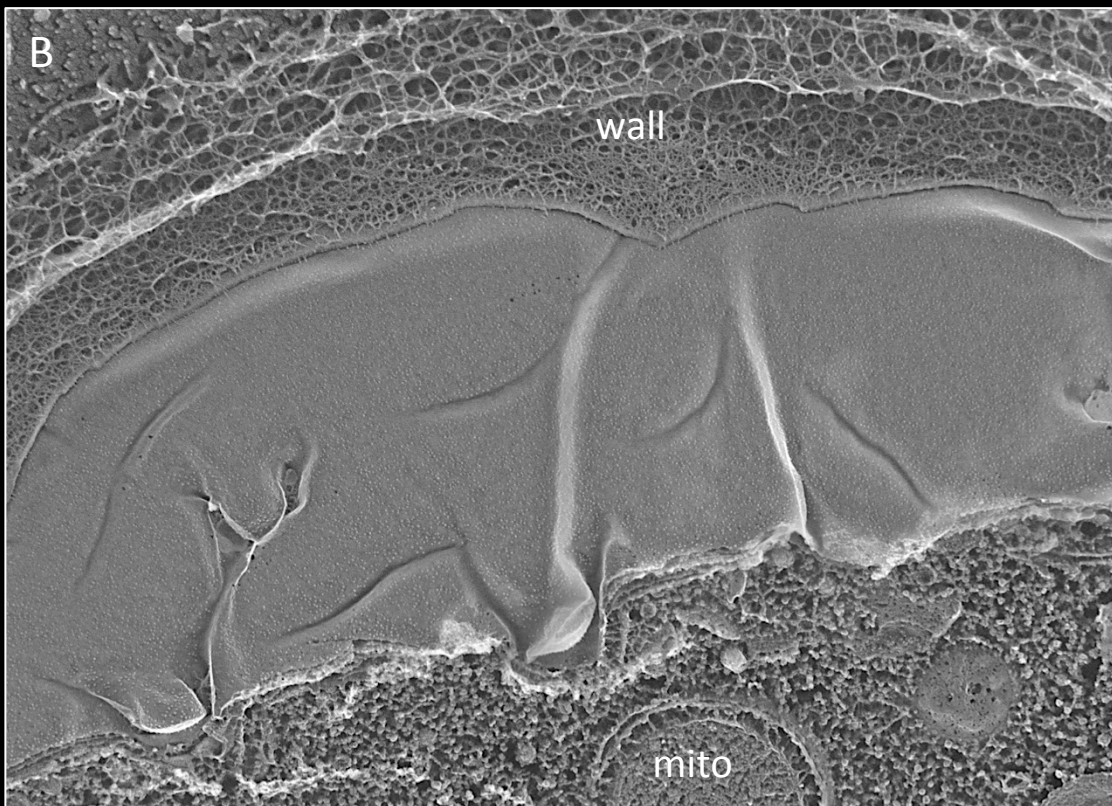
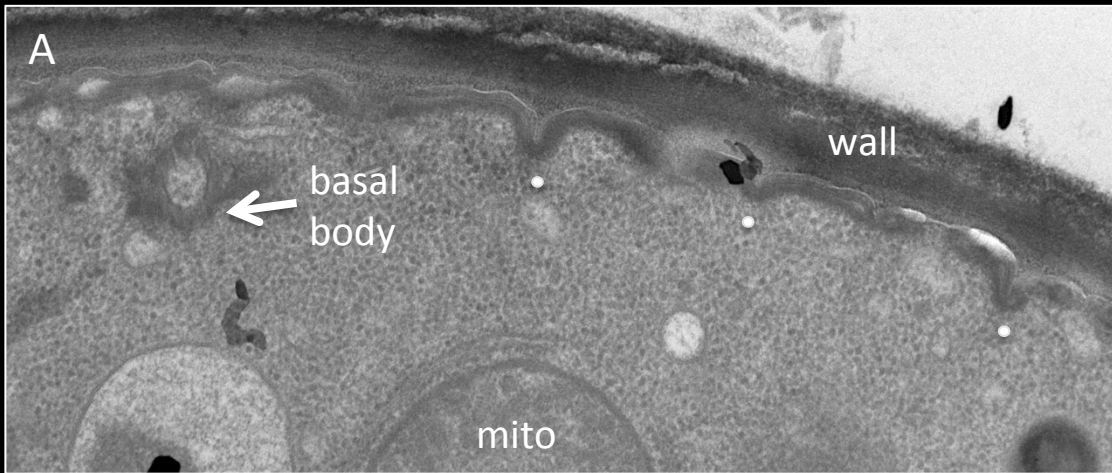
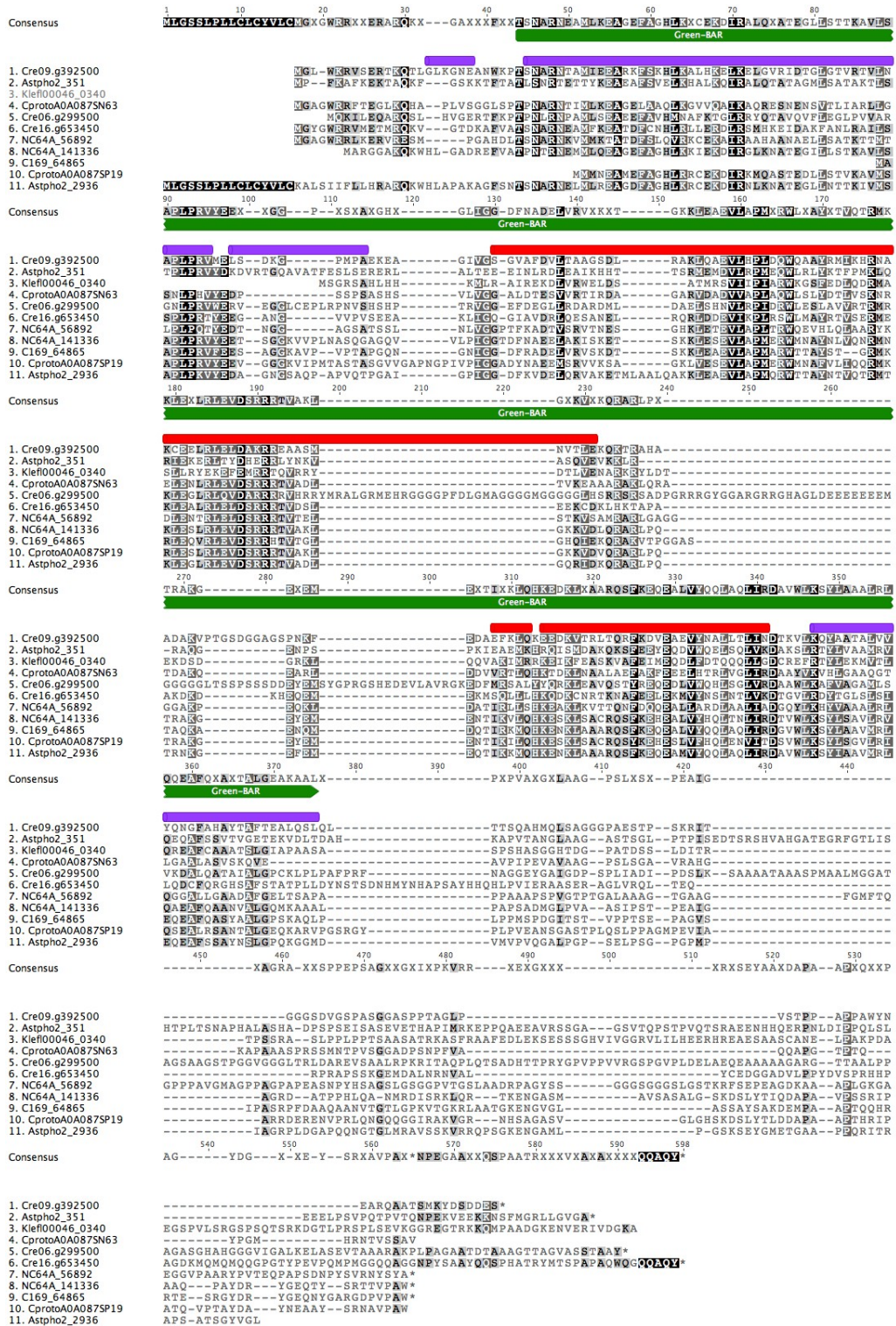


FIG S28 Alignment of Green-BAR proteins.

Red, predicted α -helix; purple, predicted amphipathic α -helix.



Cre, *Chlamydomonas reinhardtii*; Astpho2, *Astrochloris* sp.; Klefl, *Klebsormidium flaccidum*; Cproto, *Chlorella protothecoides*; NC64A, *Chlorella* sp.; C169, *Coccomyxa subellipsoidea*.

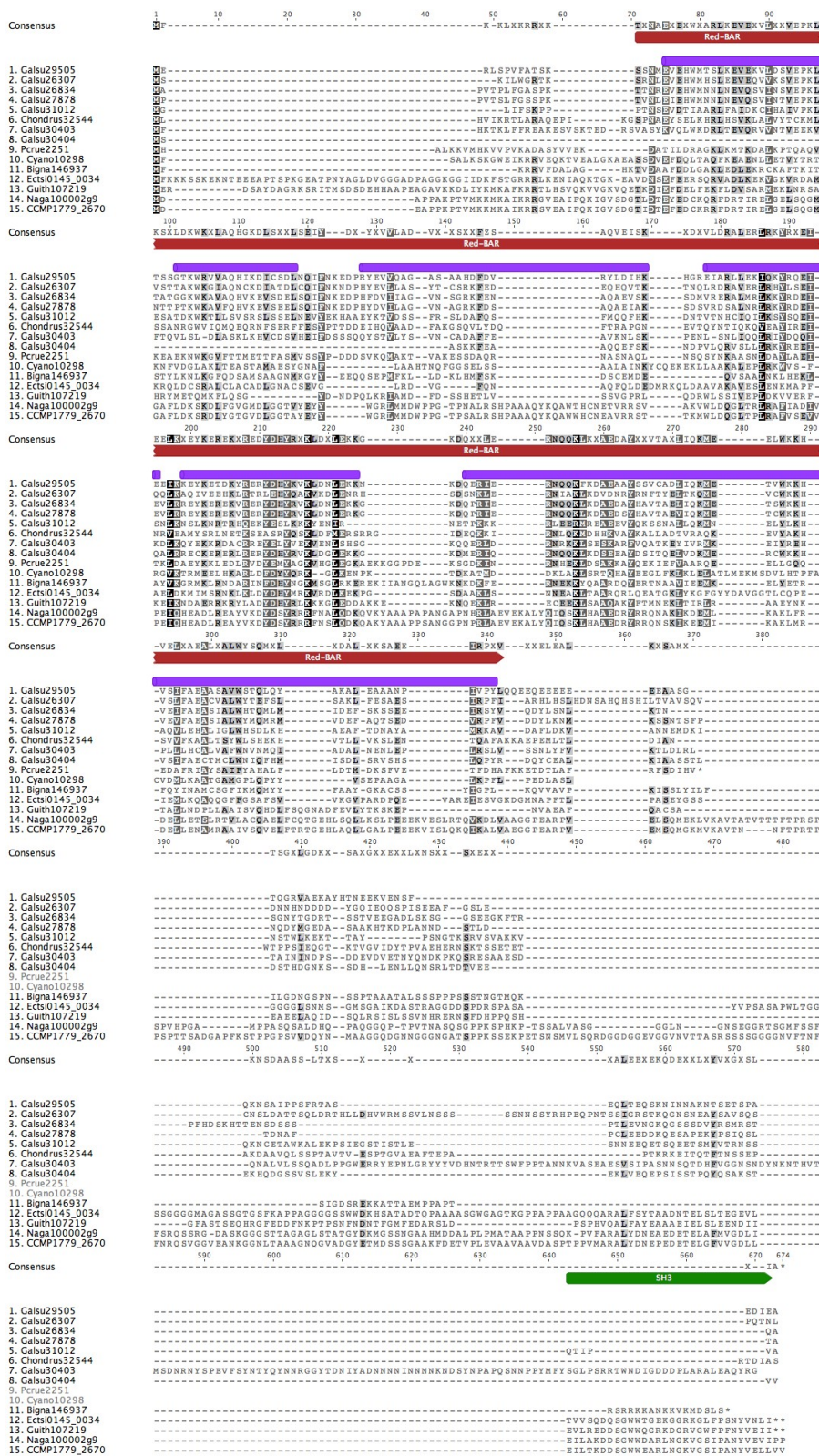
FIG S29 Alignment of Prasino-BAR proteins.
Red, predicted α -helix; purple, predicted amphipathic α -helix



Bathy, *Bathycoccus prasinos*; RCC299, *Micromonas pusilla*; Ostta, *Ostreococcus tauri*

FIG S30 Alignment of Red-BAR proteins.

Purple, predicted amphipathic α -helix



Galsu, *Galdieria sulfuraria*; Chondrus, *Chondrus crispus*; Pcrue, *Porphyridium cruensis*; Cyano, *Cyanophora paradoxa*; Bigna, *Bigelowiella natans*; Ectsi, *Ectocarpus siliculosus*; Guith, *Guillardia theta*; Naga, *Nannochloropsis gaditana*; CCMP, *Nannochloropsis oceanica*.

FIG S31 Alignment of yeast Lsp1/Pil1 eisosome proteins
 Red, predicted α -helix; purple, predicted amphipathic α -helix



FIG S32 Alignment of animal amphyphysin proteins Red, predicted α -helix; purple, predicted amphipathic α -helix

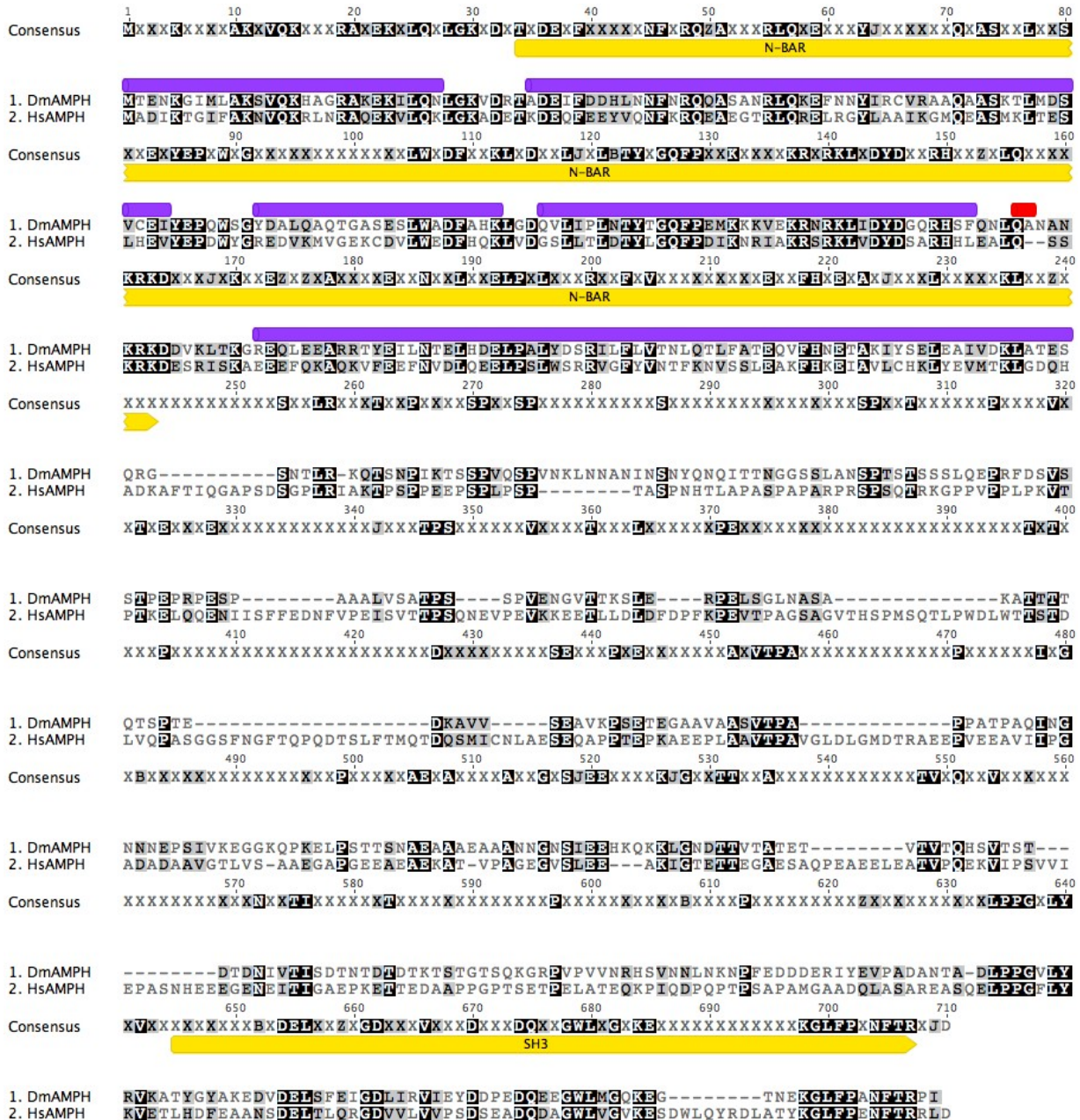


FIG S33 3D structure of the two faces of BAR-domain monomers of yeast Lsp1p (3plt), *Drosophila* amphiphysin (1uru), *Chlamydomonas* Green-Bar (Gbar) and *Galdieria* Red-BAR (3caz).

Surface charge potential: Blue, positive; Red, negative; Yellow, polar; White, hydrophobic
Arrows, N-terminal amphipathic α -helices

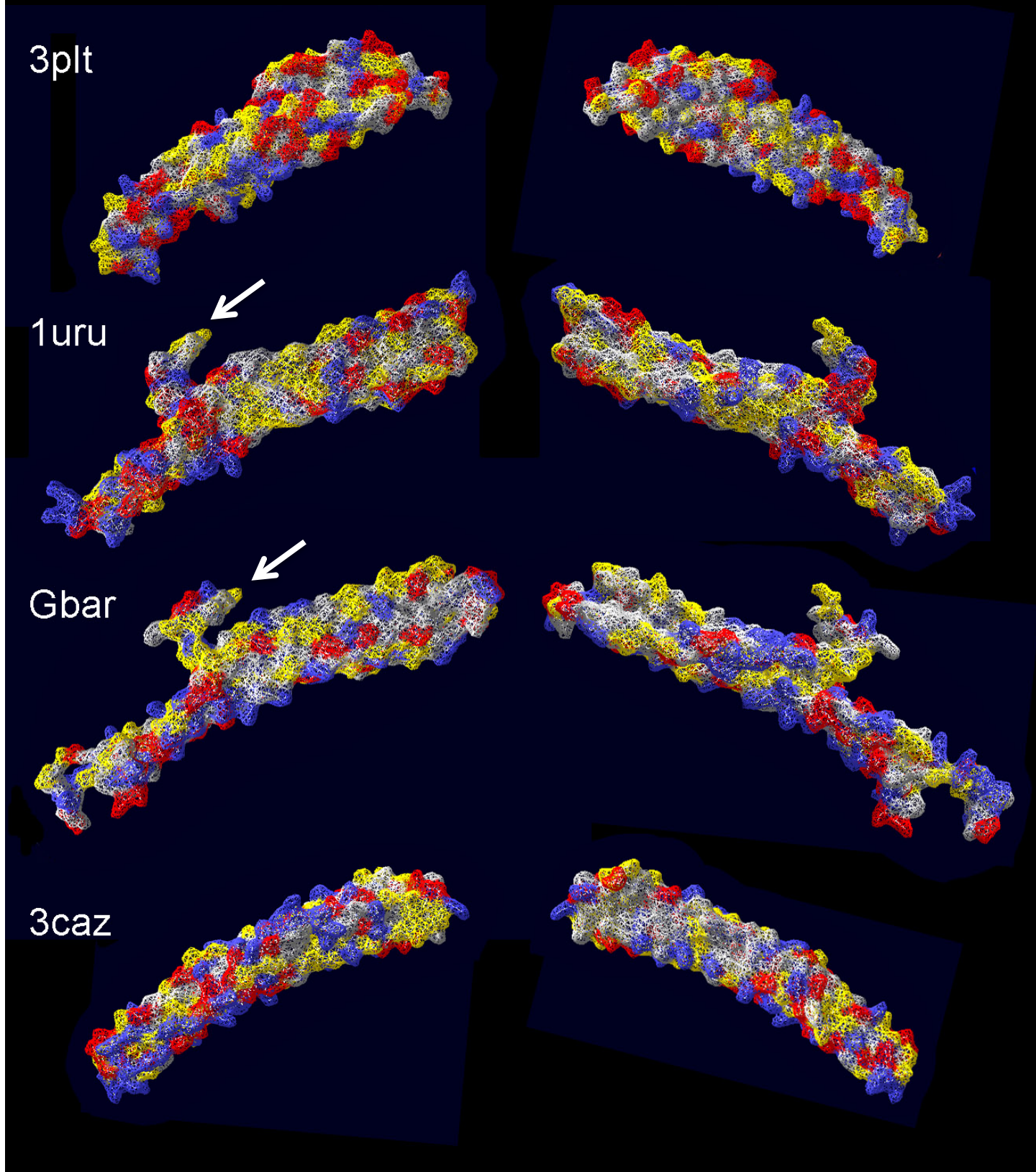


Table S1 Microalgae that do **not** produce eisosomes during vegetative growth.

<u>Class</u>	<u>Genus and species</u>	<u>Cell wall?</u>
Chlorophyceae	<i>Dunaliella tertiolecta</i>	No
	<i>Dunaliella salina</i>	No
	<i>Polytomella parva</i>	No
	<i>Chlamydomonas reinhardtii</i>	Yes
	<i>Volvox carteri</i>	Yes
	<i>Haematococcus</i> sp.	Yes
	<i>Borodinellopsis texensis</i>	Yes
Trebouxiophyceae	<i>Botryococcus braunii</i> Race A	Yes
	<i>Botryococcus braunii</i> Race B	Yes
	<i>Botryococcus braunii</i> Race C	Yes
Prasinophyceae	<i>Micromonas</i> CCMP 1545	No
	<i>Bathycoccus</i> RC 216	No (scales)
Cyanidiophyceae	<i>Cyanidioschyzon merolae</i>	No
Eustigmatophyceae	<i>Nannochloropsis salina</i>	Yes
	<i>Nannochloropsis oceanica</i>	Yes
	<i>Nannochloropsis gaditana</i>	Yes
	<i>Nannochloropsis oculata</i>	Yes
	<i>Eustigmatos vischeri</i>	Yes
Bacillariophyceae	<i>Phaeodactylum tricornutum</i>	Yes
	<i>Thalassiosira pseudonanna</i>	Yes
	<i>Cyclotella cryptica</i>	Yes
Chrysophyceae	<i>Ochromonas danica</i>	No
Phaeophyceae	<i>Pelagomonas</i> sp. CCMP 1756	Yes
Dinophyceae	<i>Symbiodinium</i> sp	Yes

Table S2. BAR-domain containing genes described in this report. Abbreviations: AGD, Arf-GAP; Ank, Ankyrin; PH, Pleckstrin-homology; PX, Phosphoinositide-binding structural domain; RNI, RNase-inhibitor; SNX, Sorting nexin; Vps5, Vacuolar protein sorting 5; ext, additional C-terminal sequences with no identified sorting domains.

Species	ID	Domain Structure (protein length)	Family
<i>Chlamydomonas reinhardtii</i>			
	Cre16.g653450	BAR + ext (398 aa)	Green-BAR
	Cre06.g299500	BAR + ext (530 aa)	Green-BAR
	Cre09.g392500	BAR + ext (340 aa)	Green-BAR
	Cre03.g154150	BAR + PH + ArfGAP + Ank	AGD homolog
	Cre06.g281050	PX + Vps5	AtSNX1 homolog
	Cre07.g326450	PX + Vps5	AtSNX2b homolog
	Cre10.g462300	PX+divergent Vps5	AtSNX2b homolog
<i>Chlorella sp. NC64A</i>			
	jgi ChINC64A_1 141336	BAR + ext (359 aa)	Green-BAR
	jgi ChINC64A_1 56892	Divergent BAR + ext (752 aa)	Green-BAR
	jgi ChINC64A_1 142705	PX + Vps5	SNX homolog
<i>Cocomixa subellipsoidea C-169</i>			
	jgi Coc_C169_1 64865	Divergent BAR + ext (296 aa)	Green-BAR
	jgi Coc_C169_1 66447	PX + Vps5	SNX homolog
	jgi Coc_C169_1 11161	PX + Vps5	SNX homolog
<i>Asterochloris sp.</i>			
	Astpho2 2936	BAR + ext (897 aa)	Green-BAR
	Astpho2 351	Divergent BAR + ext (465 aa)	Green-BAR
	Astpho2 8374	BAR + PH + ArfGAP + Ank	AGD homolog
	Astpho2 2830	PX + Vps5	SNX homolog
<i>Chlorella protothecoides</i>			
	Cproto_AOA087SP19	BAR + ext (347 aa)	Green-BAR
	Cproto_AOA087SN63	BAR + ext (315 aa)	Green-BAR
	Cproto_AOA087SN62	RabGAP-TBC + BAR	Others
<i>Klebsormidium flaccidum</i>			
	Klefl00046_0340	BAR + ext (323 aa)	Green-BAR
	Klefl00290_0020	BAR + PH + ArfGAP + Ank	AGD homolog
	Klefl00109_0220	PX + Vps5	SNX homolog
	Klefl00036_0250	PX + Vps5	SNX homolog
	Klefl00014_0220	BAR + ext (882 aa)	P-rich associated
<i>Micromonas RCC299</i>			
	jgi MicpuN3 62600	Ext + BAR (347 aa)	Prasino-BAR
	jgi MicpuN3 89004	BAR + PH + ArfGAP + Ank	AGD homolog
	jgi MicpuN3 55855	BAR + PH + ArfGAP	AGD homolog
	jgi MicpuN3 62692	PX + Vps5	SNX homolog

Table S2 (cont.)

<i>Ostreococcus tauri</i>		
jgi Ostta4 32396	Ext + BAR (428 aa)	Prasino-BAR
jgi Ostta4 16066	PX + Vps5	SNX homolog
jgi Ostta4 35163	PX + Vps5+P-rich	SNX homolog
jgi Ostta4 36908	PX + Vps5	SNX homolog
<i>Bathycoccus prasinus</i>		
Bathy08g02610	Ext + BAR (577 aa)	Prasino-BAR
<i>Galdieria sulfuraria</i>		
gi 452824006 gb EME31012.1	BAR + ext (303 aa)	Red-BAR
gi 452820841 gb EME27878.1	BAR + ext (286 aa)	Red-BAR
gi 452822486 gb EME29505.1	BAR + ext (292 aa)	Red-BAR
gi 452819782 gb EME26834.1	BAR + ext (297 aa)	Red-BAR
gi 452819243 gb EME26307.1	BAR + ext (307 aa)	Red-BAR
gi 452823393 gb EME30404.1	BAR + ext (218 aa)	Red-BAR
gi 452824006 gb EME31012.1	BAR + ext (303 aa)	Red-BAR
gi 452823392 gb EME30403.1	BAR + WW	Red-BAR
gi 452825554 gb EME32550.1	BAR + F-box + RNI	Others
<i>Cyanidioschyzon merolae</i>		
gnl CMER CMR296C	BAR only (237 aa)	Others
gnl CMER CMS157C	BAR only (243 aa)	Others
<i>Cyanophora paradoxa</i>		
ConsensusfromContig39931	BAR + ext (342 aa)	Others
ConsensusfromContig10298	BAR only (242 aa)	Red-BAR
ConsensusfromContig53923	BAR + ext (431 aa)	Others
ConsensusfromContig9140	Divergent BAR (138 aa)	Others
ConsensusfromContig9428	PX+BAR	SNX homolog
ConsensusfromContig48286	Vps5 only	SNX homolog
ConsensusfromContig55813	Vps5 only	SNX homolog
<i>Chondrus crispus</i>		
gi 507108597 emb CDF32544.1	BAR + ext (327 aa)	Red-BAR
<i>Porphyridium cruensis</i>		
evm.model.contig_2251.3	BAR + ext (250 aa)	Red-BAR
evm.model.contig_2493.1	BAR + ext (481 aa)	Others
evm.model.contig_3425.9	BAR + ext (517 aa)	Others
<i>Bigelowiella natans</i>		
jgi Bigna1 146937	BAR + ext (309 aa)	Red-BAR
<i>Guillardia theta</i>		
jgi Guith1 107219	BAR + SH3	Red-BAR
<i>Ectocarpus siliculosus</i>		
Esi0145_0034	BAR + SH3	Red-BAR
<i>Nannochloropsis gaditana</i>		
Naga100002g9	BAR + SH3	Red-BAR
<i>Nannochloropsis oceania</i>		
CCMP1779_2670	BAR + SH3	Red-BAR

Table S3 Genomes of algae and protists lacking Green-BAR and Red-BAR homologues.

Aureococcus anophagefferens (Pelagophyte)

Fragilariopsis cylindrus (Bacillariophyte)

Phaeodactylum triconutum (Bacillariophyte)

Thalassiosira pseudonana (Bacillariophyte)

Emiliana huxleyi (Bacillariophyte)

Pytophthora sojae (Oomycete)

Oxytricha trifallax (Ciliate)

Ichthyophthirius multifiliis (Ciliate)

Styolnychia lemnae (Ciliate)

Paramecium tetraurelia (Ciliate)

Tetrahymena thermophile (Ciliate)