

**Online Supporting Information for**  
**“Facile Discovery of Cell-surface Protein Targets of Cancer Cell Aptamers”**  
by  
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**Table S1.** Specificity of Sgc-3b and Sgc-4e aptamers to different cell lines.

Cell lines	Origin	Sgc-3b	Sgc-4e
RAEC	rat aortic endothelial cells	-	-
MRC-5	human fetal lung fibroblast	-	++
A549	human alveolar basal epithelial cells	-	-
Hela	human cervical cancer	-	-
Huh-7	human hepatoma	-	-
T24	human bladder carcinoma	-	++
SK-Hep-1	human hepatocellular carcinoma	-	+++
MCF-7	human breast adenocarcinoma	-	-
MCF-7R	human breast adenocarcinoma	-	+++
SKOV-3	human ovarian carcinoma	-	+
K562	human myelogenous leukemia	-	-
MDA-MB-231	human breast cancer	-	-
A431	human epithelial carcinoma	-	-
Jurkat E6-1	human T-cell leukemia	+++	++++
HCT-8	human ileocecal adenocarcinoma	-	-
HEK-293	human embryonic kidney	-	++
PC-3	human prostate cancer	-	-
Hut-78	human T cell lymphoma	-	-
LoVo	human colon cancer	-	-

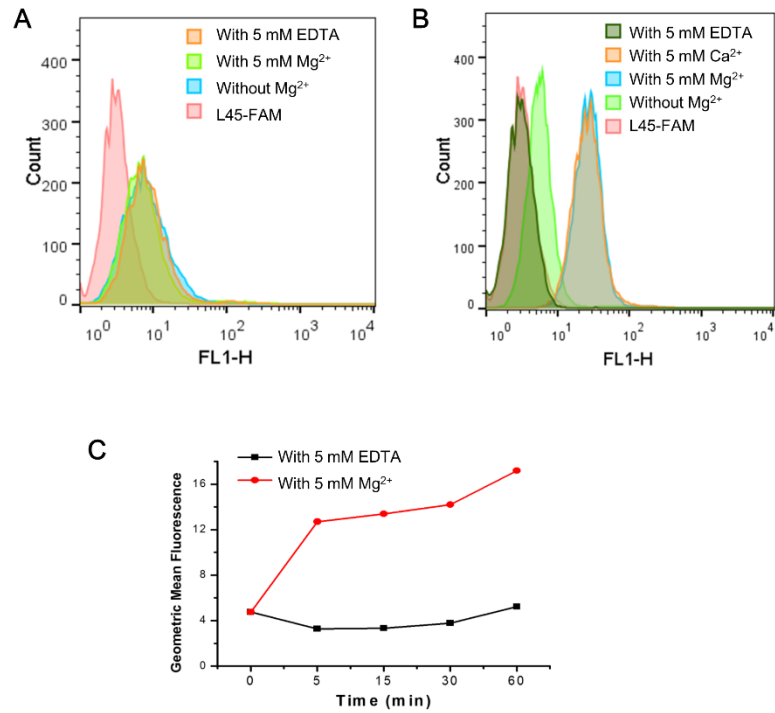
The binding capacity of the aptamer to the cells: -, <10%; +, 10-35%; ++, 35-60%; +++, 60-85%; +++++, >85%.

**Table S2.** The identification of endogenous biotinylated proteins with or without formaldehyde cross-linking using. The data represent the means and standard deviations of results obtained from three independent SILAC experiments.

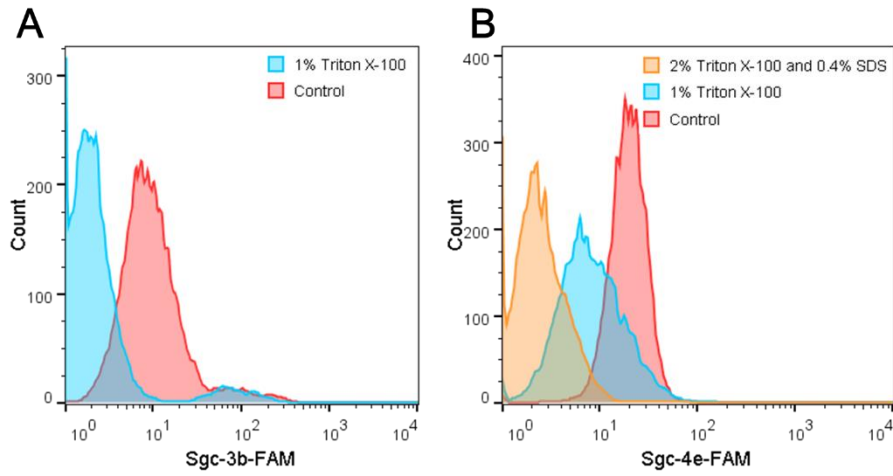
Refseq Protein Accession	Gene Symbol	Protein Name	Unique Peptides	Sequence coverage [%]	PEP <sup>[a]</sup>	Protein Abundance Ratio (streptavidin beads/ biotin-saturated streptavidin beads)	
						With Cross-linking	No Cross-linking
NP_000911	PC	Pyruvate carboxylase	11	13.1	0	17.5±1.2	13.7±3.4
NP_942131	ACACA	Isoform 4 of Acetyl-CoA carboxylase 1	22	11.7	0	>20	17±2.4
NP_000273	PCCA	propionyl-coenzyme A carboxylase, $\alpha$ polypeptide isoform a precursor	4	6.9	2.48E-85	>20	18.7±2.6
NP_000523	PCCB	Propionyl coenzyme A carboxylase, $\beta$ polypeptide, isoform CRA_c	9	22.4	2.9E-263	>20	>20
NP_064551	MCCC1	Methylcrotonoyl-CoA carboxylase subunit $\alpha$	7	14.1	9.1E-178	>20	14±4.1

<sup>[a]</sup>PEP, the posterior error probability. This value essentially operates as a p-value, where smaller is better.

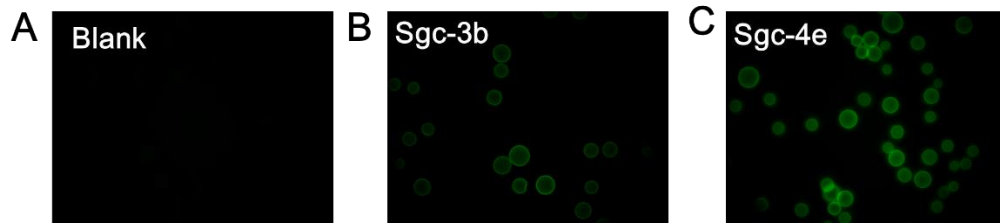
**Fig. S1.** A) Binding of Sgc-3b to Jurkat E6-1 cells in the absence or presence of  $Mg^{2+}$  (5 mM),  $Ca^{2+}$  (5 mM) or EDTA (5 mM). B) Binding of Sgc-4e to Jurkat E6-1 cells in the absence or presence of  $Mg^{2+}$  (5 mM),  $Ca^{2+}$  (5 mM) or EDTA (5 mM). C) Fluorescence of Jurkat E6-1 cells cross-linked with Sgc-4e in the presence of  $Mg^{2+}$  or EDTA in the buffer for different periods of time.



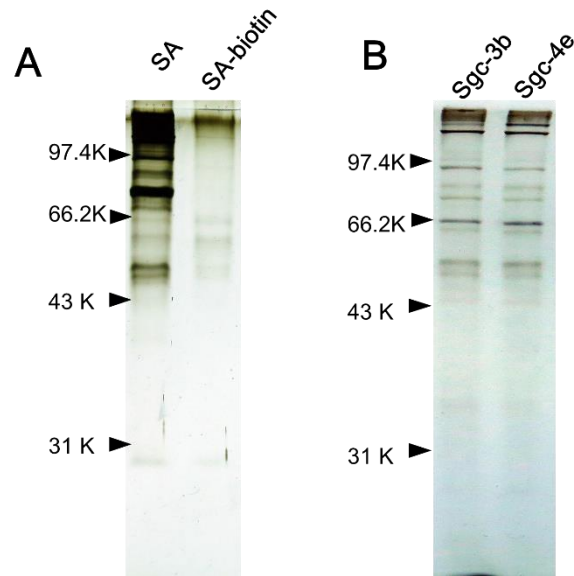
**Fig. S2.** A) Binding of Sgc-3b to Jurkat E6-1 cells after cross-linking (Control) and elution of aptamer-target complex using 1% Triton X-100. B) Binding of Sgc-4e to Jurkat E6-1 cells following cross-linking and elution of aptamer-target complex using 1% Triton X-100 or 2% Triton X-100 and 0.4% SDS.



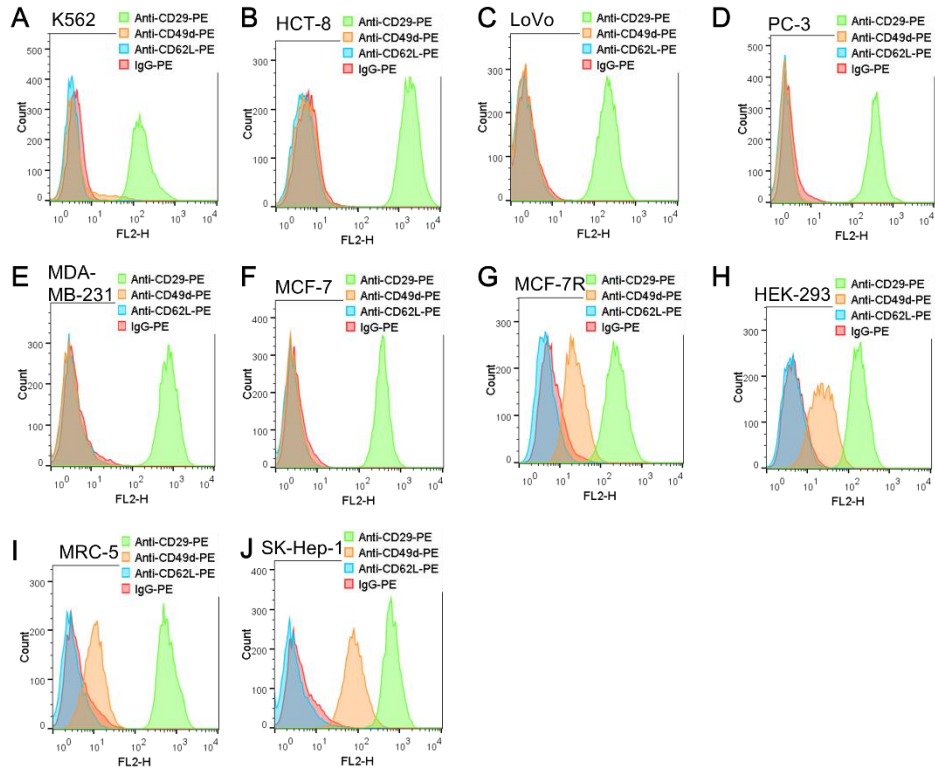
**Fig. S3.** Fluorescence images of streptavidin-coated beads (GE company) pulling down the aptamer-target complex from Jurkat E6-1 cells: A) Blank; B) aptamer Sgc-3b; C) aptamer Sgc-4e.



**Fig. S4.** A) Silver-stained SDS-PAGE (10%) of proteins that were pulled down using streptavidin-coated beads (SA) or biotin-saturated streptavidin-coated beads (SA-biotin). B) Silver-stained SDS-PAGE (10%) of proteins that were pulled down using streptavidin beads coated with Sgc-3b or Sgc-4e.

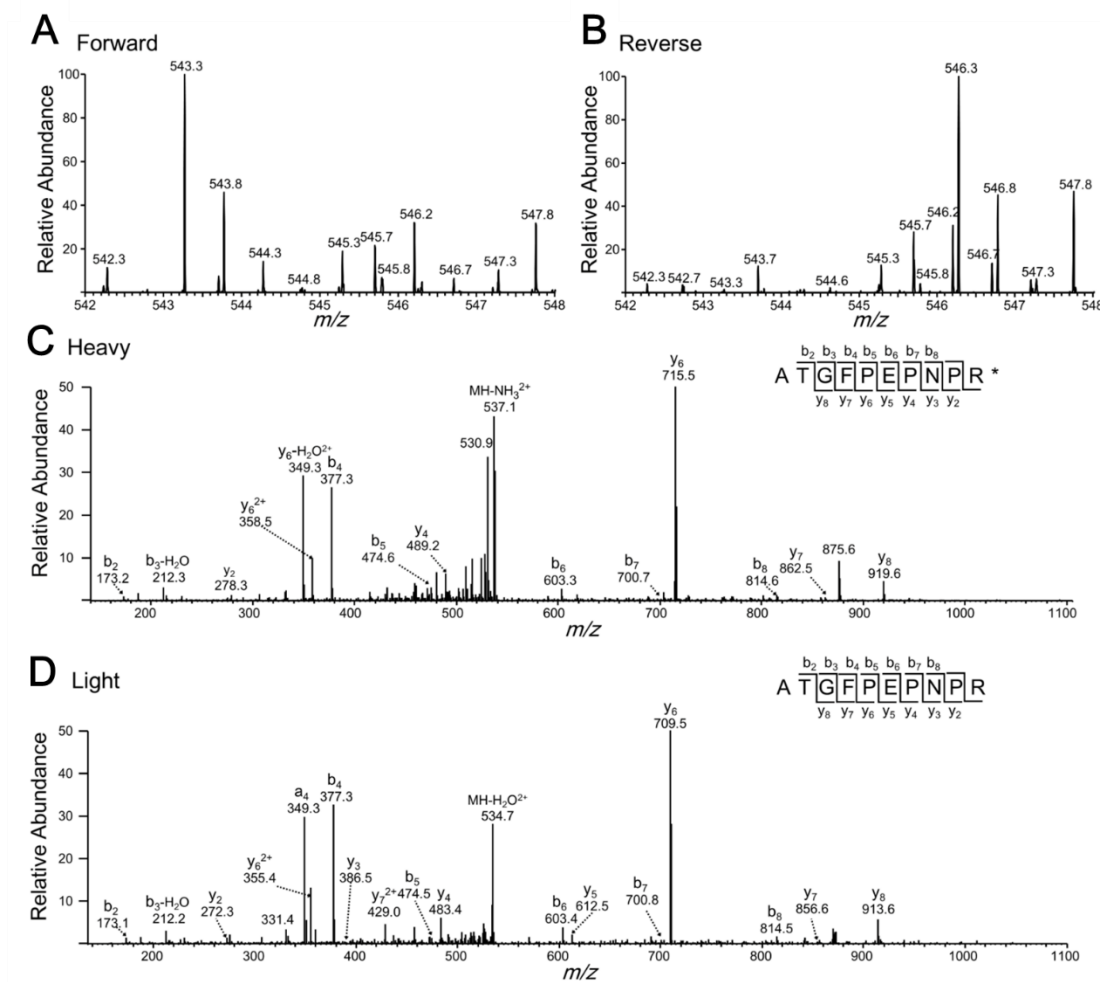


**Fig. S5.** Flow cytometry assay results showed the expression of CD62L, CD49d or CD29 on K562(A), HCT-8(B), LoVo(C), PC-3(D), MD-MB-231(E), MCF-7(F), MCF-7R(G), HEK-293(H), MRC-5(I) and SK-Hep-1(J) cell lines. IgG-PE, which do not bind to cells, was used as negative control.

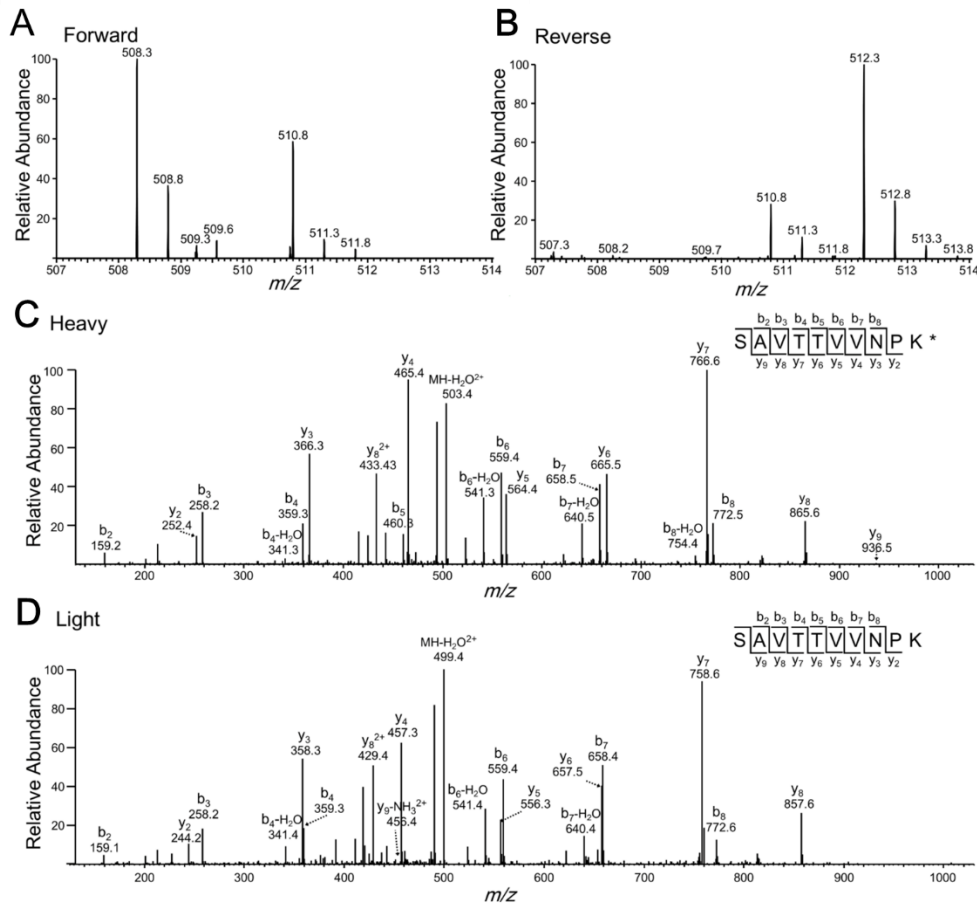




**Fig. S6.** Representative ESI-MS and MS/MS of a tryptic peptide from CD49d (the target protein for aptamer Sgc-4e). R\* designates the heavy arginine. Shown in (A) and (B) are the ESI-MS for the light ( $m/z$  543.3 for the monoisotopic peak of the  $[M+2H]^{2+}$  ion) and heavy ( $m/z$  546.3 for the monoisotopic peak of the  $[M+2H]^{2+}$  ion) arginine-containing peptide observed in forward and reverse SILAC experiments. Displayed in (C) and (D) are the MS/MS for the  $[M+2H]^{2+}$  ions of the heavy- and light-arginine-bearing peptide.



**Fig. S7.** Representative ESI-MS and MS/MS of a tryptic peptide from CD29 (the target protein for aptamer Sgc-4e). K\* designates the heavy lysine. Shown in (A) and (B) are the ESI-MS for the light ( $m/z$  508.3 for the monoisotopic peak of the  $[M+2H]^{2+}$  ion) and heavy ( $m/z$  512.3 for the monoisotopic peak of the  $[M+2H]^{2+}$  ion) lysine-containing peptide observed in forward and reverse SILAC experiments. Displayed in (C) and (D) are the MS/MS for the  $[M+2H]^{2+}$  ions of the heavy- and light-lysine-bearing peptide, respectively.



**Fig. S8.** Representative ESI-MS and MS/MS of a tryptic peptide from talin-1 (the target protein for aptamer Sgc-4e). R\* designates the heavy arginine. Shown in (A) and (B) are the ESI-MS for the light ( $m/z$  508.6 for the monoisotopic peak of the  $[M+2H]^{3+}$  ion) and heavy ( $m/z$  510.6 for the monoisotopic peak of the  $[M+2H]^{3+}$  ion) arginine-containing peptide observed in forward and reverse SILAC experiments. Displayed in (C) and (D) are the MS/MS for the  $[M+2H]^{3+}$  ions of the heavy- and light-arginine-bearing peptide.

