

Arsenic trioxide alters the differentiation of mouse embryonic stem cell into cardiomyocytes

Paola Rebuzzini^{a,d}, Elisa Cebal^b, Lorenzo Fassina^{c,d}, Carlo Alberto Redi^{a,d,e}, Maurizio Zuccotti^{f,*} and Silvia Garagna^{a,d,*}

^aLaboratorio di Biologia dello Sviluppo, Dipartimento di Biologia e Biotecnologie ‘Lazzaro Spallanzani’, Università degli Studi di Pavia, Pavia, Italy;

^bLaboratorio de Reproducción y Fisiopatología Materno-Embrionaria, Instituto de Fisiología, Biología Molecular y Neurociencias, Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Buenos Aires, Argentina;

^cDipartimento di Ingegneria Industriale e dell’Informazione, Università degli Studi di Pavia, Pavia, Italy;

^dCenter for Health Technologies (CHT), Via Ferrata 1, University of Pavia, Italy;

^eCentro Ricerche di Medicina Rigenerativa, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy;

^fUnità di Anatomia, Istologia ed Embriologia, Dipartimento di Scienze Biomediche, Biotecnologiche e Traslazionali, Università degli Studi di Parma, Parma, Italy.

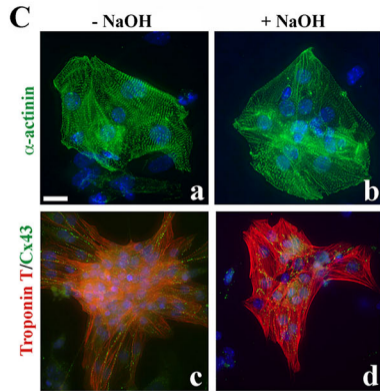
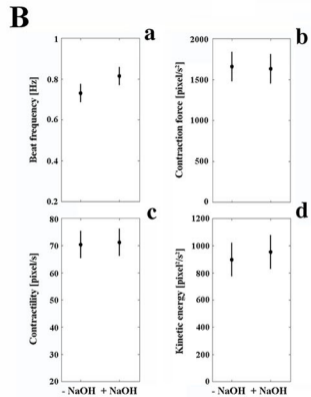
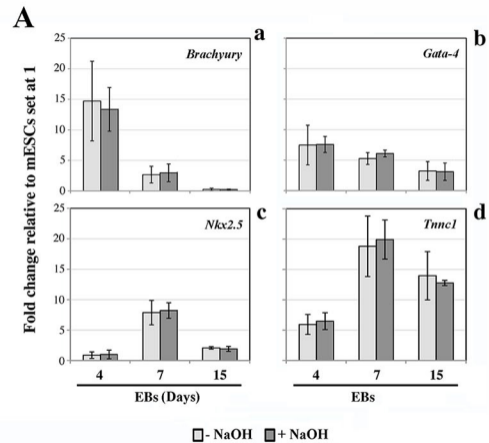
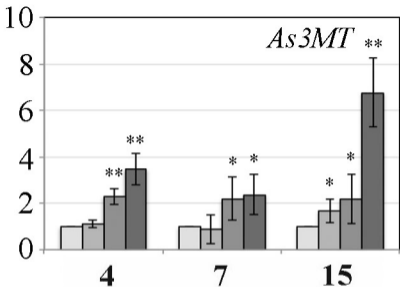


Figure 1S. Cardiomyocyte differentiation in the absence or presence of 0.01 N NaOH. **A)** Expression profile of genes that mark mesoderm (**a**, *Brachyury*), cardiac mesoderm (**b**, *Gata-4*; **c**, *Nkx2.5*) and cardiac cells (**d**, *Tnnc1*) throughout the differentiation process. Values are expressed as mean±standard deviation; **B)** Contractile properties of beating syncytia on day 15: **a)** Beat frequency [Hz]; **b)** Contraction force [pixel/s²]; **c)** Contractility [pixel/s]; **d)** Kinetic energy [pixel²/s²]. Horizontal bars represent the 95% confidence interval for the differences between means according to the Least Significant Difference statistical test. **C)** Immunofluorescence localisation of cardiac α -actinin and of cardiac troponin T (red) and Connexin 43 (green) proteins in cardiomyocytes on day 15. Bar, 20 μ m.

Three independent sets of experiments were performed.

Fold change

relative to CTR set at 1



EBs (days)

□ CTR □ 0.1 □ 0.5 □ 1.0

ATO (μM)

Figure 2S. Expression profile of *As3MT* in CTR and ATO-exposed EBs. Values are expressed as mean±standard deviation. Three independent sets of this experiment were performed. * $p < 0.05$; ** $p < 0.001$.

Methods

Disaggregation of embryoid bodies

Briefly, for each experiment, about 100 EBs were mechanically detached and centrifuged at 500 rpm for 5 min and resuspended in 1 ml of “low Ca²⁺-medium” (120 mM NaCl, 5.4 mM KCl, 5 mM sodium pyruvate, 20 mM glucose, 20 mM taurine, 10 mM HEPES) for 15 min at room temperature. Then, cells were incubated in “low Ca²⁺-medium” supplemented with 1 mg/ml collagenase and 30 mM CaCl₂ for 30 min at 37°C. After centrifugation at 500 rpm for 5 min, cells were resuspended in complete culture medium and about 50,000 cells seeded on each 0.1% gelatin-coated coverslip. After 24 h, all cells were washed twice with PBS, fixed in 4% cold paraformaldehyde/PBS. Slides were kept at 4°C until usage.

Table 1S. Oligonucleotides used for Real Time PCR amplification.

Gene	Primer forward	Primer Reverse	Amplicon length (bp)
<i>Brachyury</i>	5' CTCTAAGGAACCACCGGTCA 3'	5' AGCATGGACAGACAAGCAGA 3'	100
<i>Gata-4</i>	5' AGTTGTGCAGCTAATGCCACT 3'	5' CTGCTTTGGTAGCAGGTTTTG 3'	100
<i>Nkx2.5</i>	5'GATGGGAAAGCTCCCACTATG 3'	5'GAGACACCAGGCTACGTCAATA 3'	110
<i>Myh6</i>	5' TCACTGCGGAAACTGAAAACG 3'	5' ATGGCCATGTCCTCGATCTTG 3'	100
<i>Actn2</i>	5' AACCTGGCCATGGAAATAGCA 3'	5' TTCATCGGGTTGGGAGTGTT 3'	90
<i>Tnnc1</i>	5' CAGCAAAGGGAAGTCTGAGG 3'	5' TGCAGCATCATCTTCAGCTC 3'	102
<i>Tnnt2</i>	5' GAAGTTCGACCTGCAGGAAA 3'	5' TTCCCACGAGTTTTGGAGAC 3'	102
<i>Tnni3</i>	5' GACTTATGCCGACAGCTTAC 3'	5' GGTCAGATCTGCAATCTCAGTG 3'	105
<i>As3MT</i>	5' AAAACAAGGAGCTCGAAGGGG 3'	5' TTTCGGCTGGCTCTGTCTTAG 3'	93
<i>β2m</i>	5' GAATTCACCCCACTGAGACT 3'	5' TGCTTGATCACATGTCTCGAT 3'	103