

Phenotype of murine effector CD8<sup>+</sup> T cells activated ex vivo and in vivo. (a) Flow cytometric analysis of CD8 and CD44 expression on murine T cells post-activation ex vivo; data are representative of transferred populations used in homing assays in 3 independent experiments ( $n \ge 2$  mice per group). (b) Chemokine receptor expression was compared between CD8<sup>+</sup> CD44<sup>hi</sup> T cells activated either ex vivo (WT OT-I) or in vivo (i.e., T cell populations in tumor draining lymph nodes [TdLN] from mice 7 days after implantation of B16-OVA tumors or in dLN 7 days post DC vaccination in hind footpad). Data shown in bar graph (mean  $\pm$  s.e.m.) are from n = 3 mice per group with representative histograms shown. Gray-filled histograms represent isotype control Ab staining.

b



**Murine effector CD8+ T cell trafficking to tumors under homeostatic and elevated trafficking conditions.** (a) Schematic and quantification for competitive homing studies between WT cells pretreated with PTX and *Cxcr3+* cells in the same recipient mice. Data represent ratio of adoptively transferred *Cxcr3+*:PTX treated T cells determined by flow cytometry in tumor and spleen. Data showing that the 1:1

## Supplementary Figure 2 (continued)

ratio in cells recovered from tumors was equivalent to the 1:1 input ratio indicate that *Cxcr3<sup>-/-</sup>* cells and PTX-treated cells homed to the same extent. Data (mean  $\pm$  s.e.m.) are from 3 independent experiments (n ≥ 2 mice per group). (**b**) Absolute number of WT OT-I and *Cxcr3<sup>-/-</sup>* OT-I T cells homed to B16-OVA tumors under baseline normothermic (NT) conditions or in mice administered preconditioning systemic thermal therapy (STT). Data (mean  $\pm$  s.e.m.) are from n ≥ 2 mice per group and are representative of 3 independent experiments. (**c**) Schematic for comparison of endogenous CD8<sup>+</sup> T cell infiltration in WT and *Cxcr3<sup>-/-</sup>* mice at 2 weeks after implantation of B16-OVA tumors (tumor volume was 475  $\pm$  266 and 638  $\pm$  306 mm<sup>3</sup> for WT and *Cxcr3<sup>-/-</sup>* recipients, respectively). Data represent number of endogenous CD8<sup>+</sup> T cells determined by flow cytometry per 10<sup>6</sup> total cells in tumor or spleen. Data (mean  $\pm$  s.e.m.) are from n ≥ 3 mice per group. (**b**, **c**) \* *P* < 0.05, unpaired two-tailed Student's *t*-test.



Effector T cells require  $G_{\alpha i}$  signaling in LPS-inflamed vessels. (a) Phenotype of WT OT-I and Cxcr3<sup>-/-</sup>OT-I T cells for prototypical trafficking molecules post-activation ex vivo; this phenotype is representative of populations used for intravital imaging studies in tumor vessels and normal skin. Gray-filled histograms represent isotype control Ab staining. (b) Representative photomicrographs depict stable interactions of fluorescently-tagged WT OT-I cells or PTX-treated WT OT-I in normal skin vessels in untreated mice or mice pre-treated with LPS. Data (mean ± s.e.m.) for rolling fractions and sticking fractions are from  $\geq$  3 independent experiments (n  $\geq$  2 mice per group). \* P < 0.002, unpaired two-tailed Student's t-test. Scale bar, 100µM.

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Phenotype of inflammatory monocytes used for trafficking studies. Representative flow cytometric analysis of adoptively transferred inflammatory monocytes after CD115<sup>+</sup> selection by AutoMacs. Purity of transferred WT or *Ccr2<sup>-/-</sup>* monocytes (left) and expression of inflammatory markers (right). Gray-filled histograms represent isotype control Ab staining; numbers denote percent positively stained cells. Data are representative of  $\geq$  3 independent experiments.



WT and *Cxcr3<sup>-/-</sup>* OT-I exhibit equivalent cytotoxicity in vitro. (a) Intracellular staining of WT effector OT-I and *Cxcr3<sup>-/-</sup>* OT-I for IFN-γ and granzyme B (GzmB). Gray-filled histograms represent isotype control Ab staining. (b) In vitro cytotoxicity of SIINFEKL-pulsed target splenocytes, (c) B16-OVA, and (d) EG7 target cells. The proportion of live fluorescent OVA-specific and non-specific tumor targets was evaluated in the absence of T cells (-), or after incubation with effector WT OT-I or with *Cxcr3<sup>-/-</sup>* OT-I T cells for 24 hrs. Left, representative flow plots of target cells. Right, percent specific lysis of OVA-expressing targets. (b-d) Data (mean  $\pm$  s.e.m.) are from 2 independent experiments performed in triplicate. \* *P* < 0.05; ns, not significant; unpaired two-tailed Student's *t*-test.

### Mikucki et al. Supplementary Figure 6



#### **Supplementary Figure 6**

WT and *Cxcr3*<sup>-/-</sup> OT-I accumulation and control of tumor growth in vivo. (a) No T cells (-), effector WT OT-I, or *Cxcr3*<sup>-/-</sup> OT-I T cells were adoptively transferred (ACT) into mice bearing B16-OVA tumors; time of ACT administration is denoted by the arrow. Tumor growth (left) and survival (right) were monitored over time. Data (mean ± s.e.m.) are representative of 3 independent experiments (n = 5 mice per group). \* *P* < 0.02, \*\* *P* < 0.005, WT OT-I compared to no adoptive transfer or transfer of *Cxcr3*<sup>-/-</sup> OT-I; statistical significance of tumor growth, two-way ANOVA for repeated measures; survival, Kaplan-Meier log rank tests. (b) Tumor-infiltrating CD45.1<sup>+</sup> WT or *Cxcr3*<sup>-/-</sup> CD45.1<sup>+</sup> OT-I T cells were tracked 3 weeks after ACT in CD45.2<sup>+</sup> congenically mismatched mice bearing B16-OVA tumors. Data (mean ± s.e.m.) are from 1 experiment (n ≥ 5 mice per group). \* *P* < 0.05; unpaired two-tailed Student's *t*-test.

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**Ex vivo activated human T cells express functional CXCR3, CCR5, and CCR2 and respond to both human and murine chemokines in vitro.** (**a**) Chemokine receptor phenotype on CD4<sup>+</sup> population in ex vivo activated human T cells. (**b**) Chemotaxis assay demonstrating migration of human T cells (donor 1) to both human and murine recombinant chemokines in vitro; similar results were detected in independent experiments for activated T cells from donor 2. (**c**) Flow cytometric analysis of CXCR3 surface expression on activated human PBL after desensitization with human CXCL10 (left). Chemotaxis assays for CXCR3-dependent migration after CXCR3 desensitization (CXCR3 dsn) or antibody blockade (right). Gray-filled histograms represent isotype control Ab staining. (**b, c**) Data (mean ± s.e.m.) are representative of 2 independent experiments. \* *P* < 0.05, unpaired two-tailed Student's *t*-test. α-CXCR3, CXCR3 blocking Ab.



CXCR3 does not alterhuman effector T cell localization in the spleens of SCID mice bearing human melanoma xenografts. Short-term (1 h) competitive homing studies using adoptive cell transfer (ACT) of untreated (Untx) human effector T cells from donor 1 or donor 2 that were comixed at a 1:1 ratio with untreated cells, PTX-pretreated cells,  $\alpha$ -CXCR3 Ab-pretreated cells, or cells where CXCR3 was desensitized by exposure to recombinant CXCL10 prior to transfer into mice (CXCR3 dsn). Ratio of adoptively transferred T cells relative to untreated cells in spleens of M537 or M888 tumor-bearing SCID mice following short-term competitive homing assays is shown. PTX, pertussis toxin;  $\alpha$ -CXCR3, CXCR3 blocking Ab. Data (mean  $\pm$  s.e.m.) are from  $\geq$  2 independent experiments (n = 2 mice per group); ns, not significant; unpaired two-tailed Student's *t*-test.



Murine effector T cells require CXCR3 during trafficking to human melanoma xenografts. (a) Murine WT effector cells were comixed at a 1:1 ratio with WT cells, PTX-pretreated WT cells, or  $Cxcr3^{-/-}$  cells (all T cells on a C57BL/6 background). Ratio of adoptively transferred T cells relative to WT in M537 or M888 melanoma tumor-bearing SCID mice following short-term (1 h) competitive homing assays is shown. \* P < 0.001, compared to WT; ns, not significant; unpaired two-tailed Student's *t*-test. (b) Schematic and quantification for competitive homing studies between PTX-pretreated WT and  $Cxcr3^{-/-}$  murine cells following adoptive T cell transfer (ACT) in M537 and M888 human melanoma xenografts. (a, b) Data (mean  $\pm$  s.e.m.) are from  $\geq$  3 independent experiments (n  $\geq$  2 mice per group).

b