

1 **Legends to Supplementary Figures**

2 **Supplementary Figure S1**

3 Ctf18-RFC co-purifies with DNA polymerase epsilon. **(A)** Ctf18-TAP was
4 purified from 4 litres of cell culture, and the final material was resolved in one
5 lane of a 4-12% gradient gel, which was then cut into 60 bands, before
6 analysis of the protein content of each band by mass spectrometry. The
7 histograms show the number of detected peptides per band of the gel, for
8 each of the indicated proteins, with Mascot scores indicated in brackets and
9 bold. **(B)** Summary of mass spectrometry data after two-step purifications
10 (IgG-Sepharose and release of bound material with TEV protease, then
11 Calmodulin-affinity resin) of cell extracts from *DPB2-TAP* and control strains.
12 For each protein detected in the mass spectrometry analysis, the number of
13 spectral counts is shown for each strain, indicating that Ctf18-Dcc1-Ctf8 were
14 specifically enriched upon purification of DNA polymerase epsilon.

15

16 **Supplementary Figure S2**

17 Ctf18 does not interact with the Dpb3 or Dpb4 subunits of Pol ϵ in the yeast
18 two-hybrid assay. Two-hybrid analysis was performed as in Figure 2,
19 indicating that Ctf18-RFC did not interact in this assay with Dpb3 or Dpb4 **(A)**.
20 As a control, we confirmed that Dpb3 and Dpb4 were able to interact with
21 each other in the same assay **(B)**.

22

1 **Supplementary Figure S3**

2 Pol2CT interacts with itself in the yeast two-hybrid assay. The indicated
3 fragments were tested in the yeast two-hybrid assay, as above in Figure 2A.

4

5 **Supplementary Figure S4**

6 The Pol ϵ binding module of Ctf18-RFC comprises a conserved motif at the
7 end of Ctf18 that interacts with Dcc1-Ctf8 and Pol2NT. **(A)** Alignment of the
8 carboxy terminal half of Ctf18 from the indicated budding yeast species,
9 generated as in Figure 3B. Asterisks denote 10 conserved hydrophobic
10 residues at the end of Ctf18. **(B)** An equivalent alignment of the carboxy
11 terminal half of Ctf18 from the indicated species (X.l. = *Xenopus laevis*; H.s =
12 *Homo sapiens*; D.m. = *Drosophila melanogaster*; S.p. =
13 *Schizosaccharomyces pombe*; O.s. = *Oryza sativa*; S.c. = *Saccharomyces*
14 *cerevisiae*).

15

16 **Supplementary Figure S5**

17 Residues at the extreme carboxyl terminus of Ctf18 are important for
18 interaction with Pol2NT and Dcc1. The indicated Ctf18 fragments were tested
19 for their ability to interact with Pol2NT **(A)** and Dcc1 **(B)** in the two-hybrid
20 assay, as in Figures 2 and 3.

21

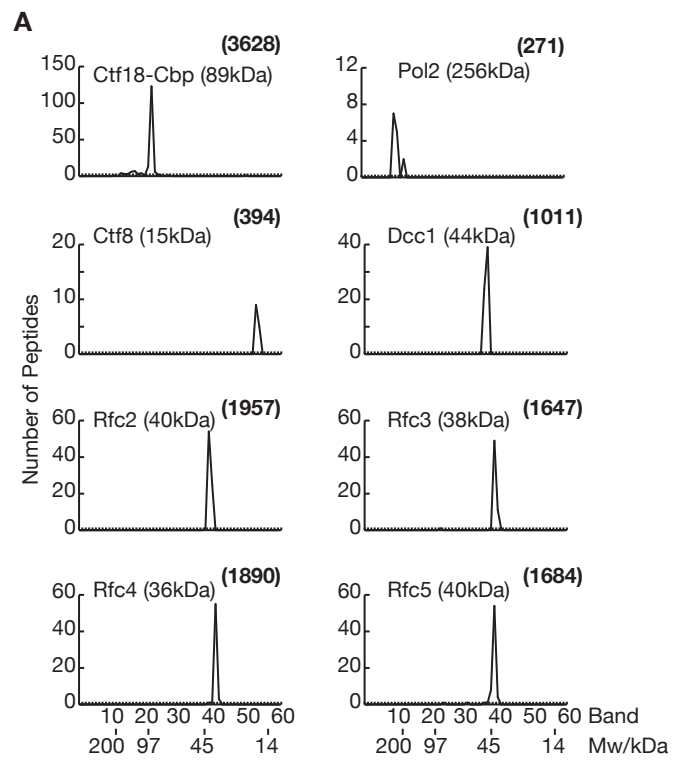
22 **Supplementary Figure S6**

23 Comparison of the hydroxyurea sensitivity caused by various mutations in
24 Ctf18-RFC. Cells were processed as for Figure 4A.

1

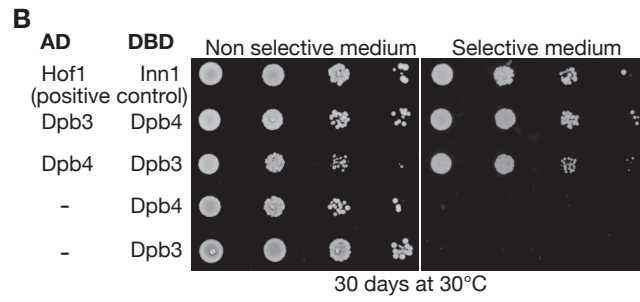
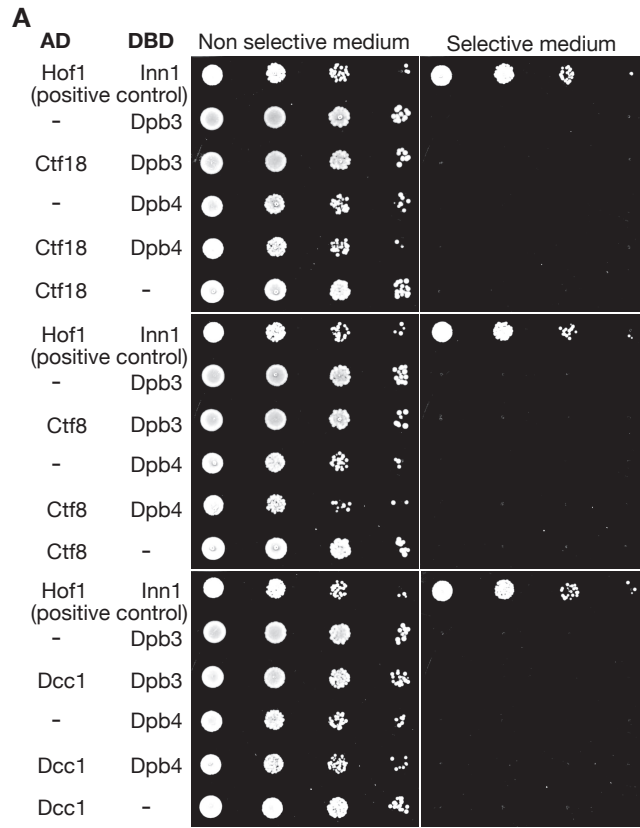
2 **Supplementary Figure S7**

3 Increased phosphorylation of Serine 129 of histone H2A indicates DNA
4 damage in ctf18-2A upon release into S-phase in the presence of
5 hydroxyurea. One of the replicates from Figure 5A was used to monitor DNA
6 content by flow cytometry (upper panels) and γ -H2A by immunoblotting (lower
7 panels). Ponceau-stained total protein provided a loading control.

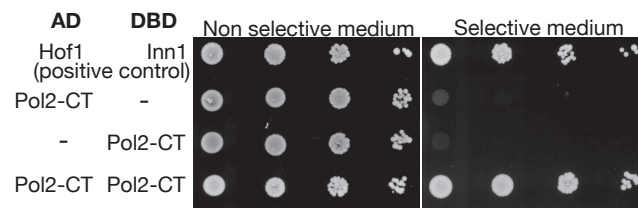


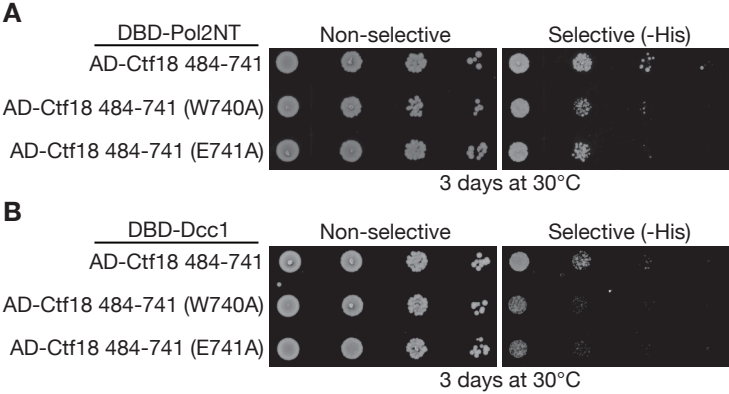
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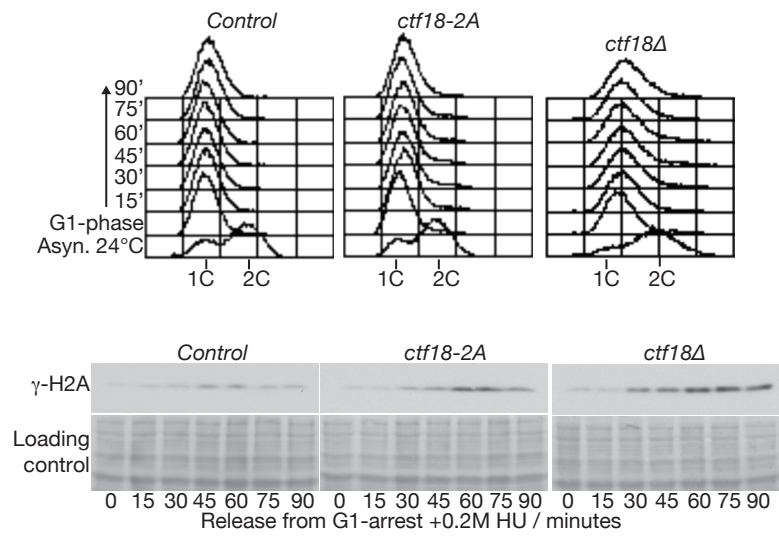
Protein	Mw (kDa)	Spectral counts (<i>DPB2-TAP</i>)	Spectral counts (<i>Control</i>)
Dpb2-TAP	99	1193	11
Dpb3	23	157	2
Dpb4	22	218	0
Pol2	256	4286	146
Ctf18	84	91	6
Dcc1	44	19	2
Ctf8	15	25	0



Garcia-Rodriguez Fig S3







Strain	Genotype	Source
W303-1a	<i>MATa ade2-1 ura3-1 his3-1 trp1-1 leu2-3, 112 can1-100</i>	R. Rothstein
YAC53	<i>MATa sml1Δ::HIS3 rad53Δ::ADE2</i>	This study
YASD375	<i>MATa TAP-SLD5 (KanMX) pep4Δ::URA3 ADE2</i>	This study
YCS394	<i>MATa MCM4-5-FLAG (hphNT) leu2-3,112:: pRS305-GAL-DPB2-9myc (LEU2) GAL TIR1-9myc (klTRP1) dpb2-IAA17 (kanMX) pep4Δ::ADE2</i>	This study
YCS396	<i>MATa MCM4-5-FLAG (hphNT) leu2-3,112:: pRS305-GAL-dpb2-NT (1-168)-9myc GAL TIR1-9myc (klTRP1) dpb2-IAA17 (kanMX) pep4Δ::ADE2</i>	This study
YCS1180	<i>MATa MCM4-5FLAG (hphNT) pep4Δ::ADE2</i>	This study
YGDP1970	<i>MATa CTF18-TAP (HIS3MX) pep4Δ::URA3 ADE2</i>	This study
YGDP1971	<i>MATa CTF18-TAP (HIS3MX) dcc1Δ::hphNT pep4Δ::URA3 ADE2</i>	This study
YGDP1972	<i>MATa DCC1-TAP (HIS3MX) pep4Δ::URA3 ADE2</i>	This study
YGDP1973	<i>MATa DCC1-TAP (HIS3MX) ctf18Δ::K.I. TRP1 pep4Δ::URA3 ADE2</i>	This study
YGDP993	<i>MATa MCM4-5FLAG (hphNT) mrc1Δ::K.I. TRP1 pep4Δ::ADE2</i>	This study
YLG33	<i>MATa TAP-SLD5 (KanMX) ctf18Δ::K.I. TRP1 pep4Δ::URA3 ADE2</i>	This study
YLG60	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4Δ gal80Δ LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ ctf8Δ::URA3</i>	This study
YLG63	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4Δ gal80Δ LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ dcc1Δ::URA3</i>	This study
YLG98	<i>MATa MRC1-18MYC (K.I. TRP1) pep4Δ::URA3 ADE2</i>	This study
YLG100	<i>MATa MRC1-18MYC (K.I. TRP1) CTF18-TAP (HIS3MX) pep4Δ::URA3 ADE2</i>	This study
YLG263	<i>MATΔ / MATα ade2-1 / ade2-1 ura3- 1 / ura3-1 his3-1 / his3-1 trp1-1 / trp1-1 leu2-3, 112 / leu2-3,112 can1-100 / can1-100 ctf18-2A (K.I. TRP1) / CTF18 ctf4Δ::KanMX / CTF4 pep4Δ::ADE2 / PEP4</i>	This study
YLG249	<i>MATa ctf18-2A (K.I. TRP1) pep4Δ::ADE2</i>	This study
YLG284	<i>MATa / MATα ade2-1 / ade2-1 ura3- 1 / ura3-1 his3-1 / his3-1 trp1-1 / trp1-1 leu2-3, 112 / leu2-3,112 can1-100 / can1-100 CTF18 (K.I. TRP1) / CTF18 ctf4Δ::KanMX / CTF4 pep4Δ::ADE2 / PEP4</i>	This study
YLG292	<i>MATa / MATα ade2-1 / ade2-1 ura3- 1 / ura3-1 his3-1 / his3-1 trp1-1 / trp1-1 leu2-3, 112 / leu2-3,112 can1-100 / can1-100 CTF18 (K.I. TRP1) / CTF18 mrc1Δ::hphNT / MRC1 pep4Δ::ADE2 / PEP4</i>	This study

YLG296	<i>MATa</i> / <i>MATα</i> <i>ade2-1</i> / <i>ade2-1 ura3-1</i> / <i>ura3-1 his3-1</i> / <i>his3-1 trp1-1</i> / <i>trp1-1 leu2-3, 112</i> / <i>leu2-3, 112 can1-100</i> / <i>can1-100 ctf18-2A</i> (K.I. <i>TRP1</i>) / <i>CTF18 mrc1Δ::hphNT</i> / <i>MRC1 pep4Δ::ADE2</i> / <i>PEP4</i>	This study
YLG301	<i>MATa</i> <i>DCC1-TAP</i> (<i>HIS3MX</i>) <i>CTF18</i> (K.I. <i>TRP1</i>) <i>pep4Δ::URA3 ADE2</i>	This study
YLG303	<i>MATa</i> <i>DCC1-TAP</i> (<i>HIS3MX</i>) <i>ctf18-2A</i> (K.I. <i>TRP1</i>) <i>pep4Δ::URA3 ADE2</i>	This study
YLG316	<i>MATa</i> <i>CTF18</i> (K.I. <i>TRP1</i>)	This study
YLG320	<i>MATa</i> <i>ctf18-2A</i> (K.I. <i>TRP1</i>)	This study
YLG421	<i>MATa</i> <i>MCM4-5-FLAG</i> (<i>hphNT</i>) <i>ctf18Δ::K.I.TRP1</i> <i>pep4Δ::URA3 ADE2</i>	This study
YLG423	<i>MATa</i> <i>MCM4-5-FLAG</i> (<i>hphNT</i>) <i>ctf18-2A</i> (K.I. <i>TRP1</i>) <i>pep4Δ::URA3 ADE2</i>	This study
YLG426	<i>MATa</i> <i>MCM4-5FLAG</i> (<i>hphNT</i>) <i>CTF18</i> (K.I. <i>TRP1</i>) <i>pep4Δ::URA3 ADE2</i>	This study
YLG445	<i>MATa</i> <i>CTF18</i> (K.I. <i>TRP1</i>) <i>ura3::3xURA3-tetO112</i> <i>leu2::tetR-GFP-LEU2 PDS1-mCherry</i> (<i>KanMX</i>)	This study
YLG447	<i>MATa</i> <i>ctf18-2A</i> (K.I. <i>TRP1</i>) <i>ura3::3xURA3-tetO112</i> <i>leu2::tetR-GFP-LEU2 PDS1-mCherry</i> (<i>KanMX</i>)	This study
YLG449	<i>MATa</i> <i>ctf18Δ::K.I.TRP1</i> <i>ura3::3xURA3-tetO112</i> <i>leu2::tetR-GFP-LEU2 PDS1-mCherry</i> (<i>KanMX</i>)	This study
YVM164	<i>MATa</i> <i>ctf18Δ::K.I.TRP1</i>	This study
YVM657	<i>MATa</i> <i>CTF18-TAP</i> (<i>HIS3MX</i>) <i>POL2-9MYC</i> (K.I. <i>TRP1</i>) <i>RFC4-6HA</i> (K.I. <i>TRP1</i>) <i>pep4Δ::URA3 ADE2</i>	This study
YVM738	<i>MATa</i> <i>CTF18-TAP</i> (<i>HIS3MX</i>) <i>POL2-9MYC</i> (K.I. <i>TRP1</i>) <i>RFC4-6HA</i> (K.I. <i>TRP1</i>) <i>dcc1Δ::hphNT</i> <i>pep4Δ::URA3 ADE2</i>	This study
YVM740	<i>MATa</i> <i>CTF18-TAP</i> (<i>HIS3MX</i>) <i>POL2-9MYC</i> (K.I. <i>TRP1</i>) <i>RFC4-6HA</i> (K.I. <i>TRP1</i>) <i>ctf8Δ::hphNT</i> <i>pep4Δ::URA3 ADE2</i>	This study
YVM850	<i>MATa</i> <i>DCC1-TAP</i> (<i>HIS3MX</i>) <i>ctf18Δ::K.I.TRP1</i> <i>pep4Δ::URA3 ADE2</i>	This study
PJ69-4A	<i>MATa</i> <i>trp1-901 leu2-3, 112 ura3-52 his3-200 gal4Δ gal80Δ</i> <i>LYS2::GAL1-HIS3 GAL2-ADE2</i> <i>met2::GAL7-lacZ</i>	H. Ulrich

Supplementary Table S1

Strains used in this study – all the above strains are based on the W303 background, except for the yeast two-hybrid strain PJ69-4A and its derivatives YLG60 and YLG63.

Plasmid	Description	Source
pAD27	Gal4 DNA binding domain-MYC-Inn1	This study
pAD30	Gal4 Activation domain-HA-Hof1	This study
pGADT7	Gal4 Activation domain-HA	Clontech
pGBKT7	Gal4 DNA binding domain-MYC	Clontech
pHM27	pGADT7-Mrc1	This study
pLG1	pGADT7-Ctf18	This study
pLG2	pGBKT7-Ctf18	This study
pLG4	pGBKT7-Ctf8	This study
pLG6	pGBKT7-Dcc1	This study
pLG16	pGBKT7-Pol2 1-1256	This study
pLG24	pGADT7-Ctf18 1-116	This study
pLG25	pGADT7-Ctf18 1-374	This study
pLG26	pGADT7-Ctf18 1-494	This study
pLG27	pGADT7-Ctf18 1-595	This study
pLG28	pGADT7-Ctf18 1-657	This study
pLG31	pGADT7-Ctf18 369-741	This study
pLG32	pGADT7-Ctf18 494-741	This study
pLG35	pGADT7-Ctf18 654-741	This study
pLG36	pGADT7-Ctf18 675-741	This study
pLG37	pGADT7-Ctf18 709-741	This study
pLG54	pGBKT7-Pol2 1-227	This study
pLG55	pGBKT7-Pol2 1-396	This study
pLG57	pGBKT7-Pol2 1-933	This study
pLG59	pGBKT7-Pol2 214-1265	This study
pLG60	pGBKT7-Pol2 392-1265	This study
pLG61	pGBKT7-Pol2 627-1265	This study
pLG62	pGBKT7-Pol2 930-1265	This study

Supplementary Table S2

Plasmids used in this study.