

## Supplementary Data

### Supplementary Methods

#### Immunofluorescence staining

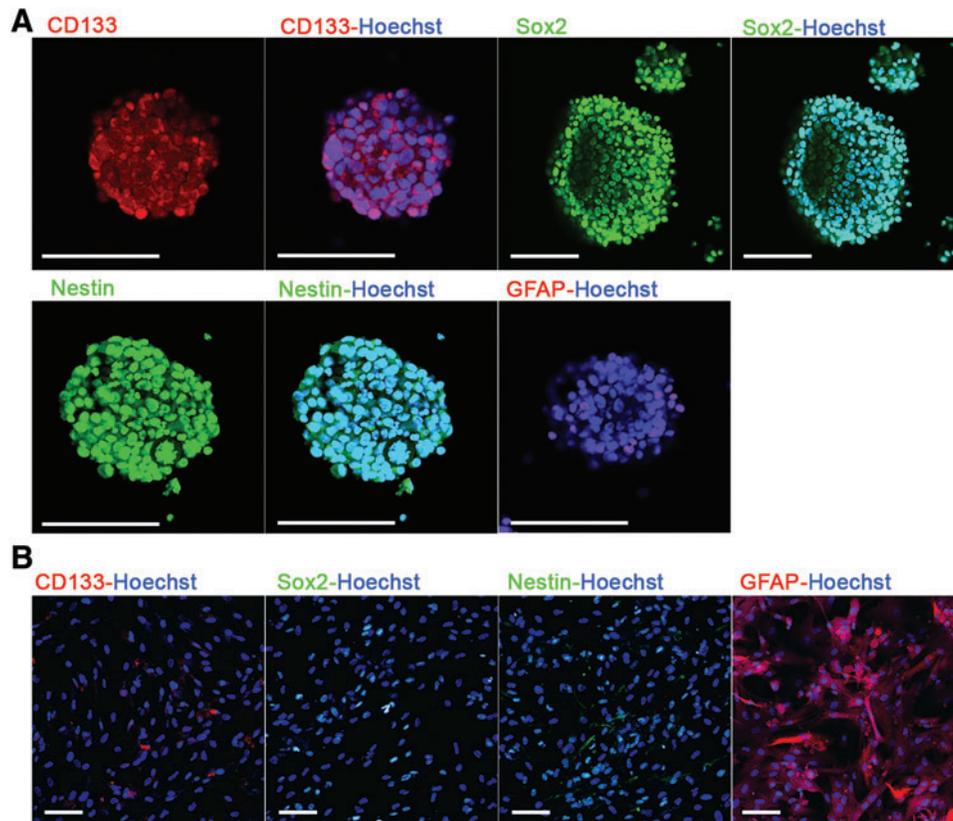
To corroborate the expression of stem markers, GBM1 cells were cultured in liquid media and collected after 4 days. Human astrocytes (ScienCell, Carlsbad, CA) were used as negative control. Cells were washed and fixed with formaldehyde 4%, permeabilized with Triton X-100 0.1%, and incubated with Sox2 Alexa Fluor-488<sup>®</sup>, Nestin Alexa Fluor-488<sup>®</sup> (eBioscience, Santa Clara, CA), human CD133-APC (Miltenyi Biotech, Inc., San Diego, CA), and GFAP-Cy3<sup>®</sup> (Sigma-Aldrich, St. Louis, MO) at dilutions of 1:100, 1:100, 1:10, and 1:100, respectively. Counterstaining for

nuclei was performed with Hoechst 33342 (Life Technologies, Carlsbad, CA).

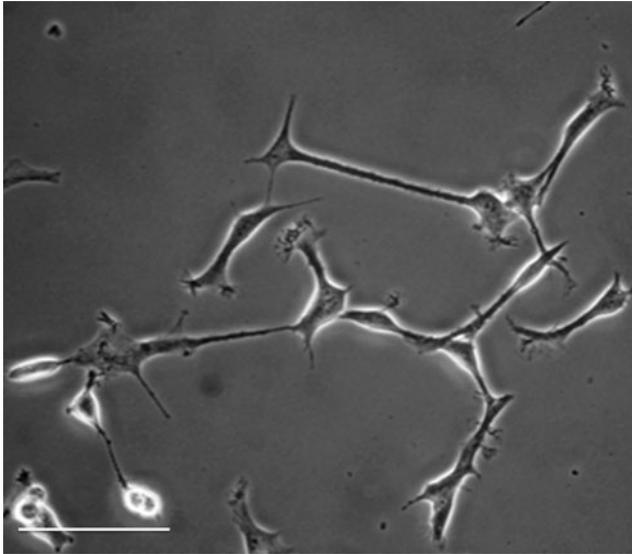
To analyze glioblastoma stem cell (GSC) collective protrusions in Matrigel 50%, neurospheres were fixed after 24 h of culture with formaldehyde 4%, permeabilized with Triton X-100 0.1%, and incubated with Alexa Fluor 488 Phalloidin (Life Technologies) (dilution 1:100) at 4°C for 6 h.

#### Differentiation of GSCs

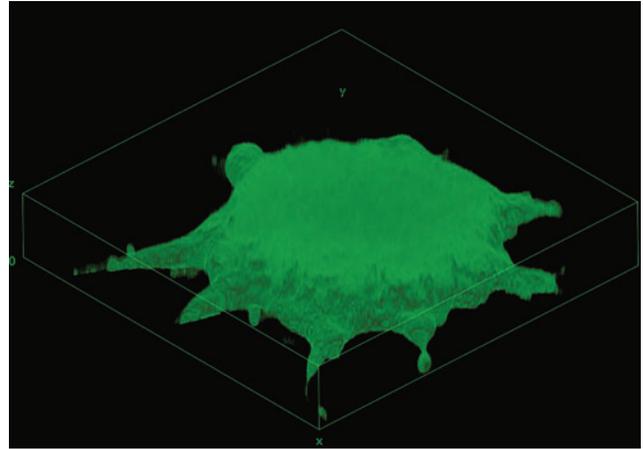
Differentiation media included DMEM/F12 and 10% FBS (Life Technologies); cells were fed with media and FBS every other day for at least 12 days.



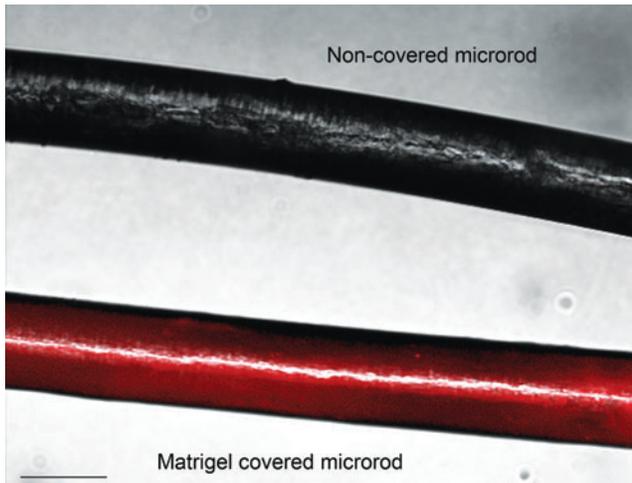
**SUPPLEMENTARY FIG. S1.** (A) GBM1 neurospheres (passages 17–23) exhibit markers of glioblastoma cancer stem-like cells (CD133, Sox2, and Nestin) after 5 days of liquid culture. GFAP marker for differentiated astrocytes is not expressed. (B) Human primary astrocytes do not express stem-like markers (CD133, Sox2, and Nestin), but are positive for GFAP expression. Scale bars 100  $\mu$ m.



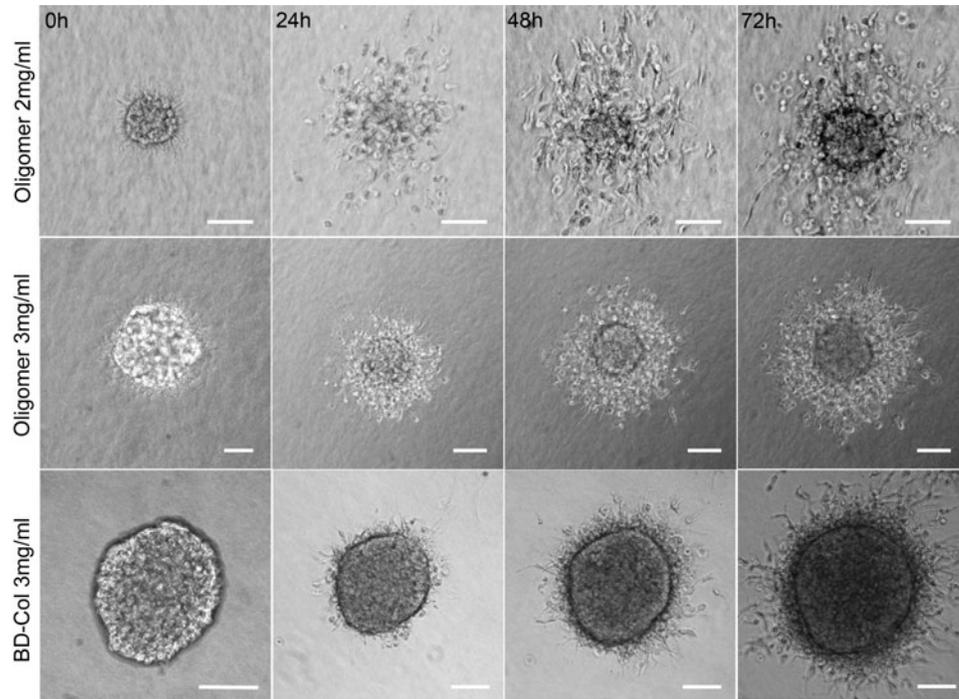
**SUPPLEMENTARY FIG. S2.** Glioblastoma stem cells (GSCs) do not form neurospheres and differentiate into astrocytes after 15 days of culture in the differentiation medium. Scale bars 100  $\mu\text{m}$ .



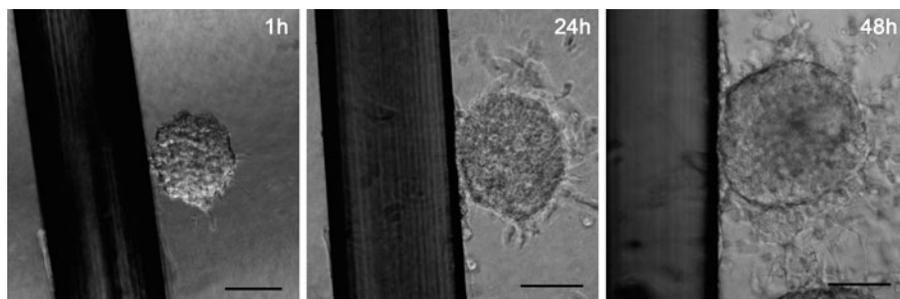
**SUPPLEMENTARY FIG. S4.** Z-reconstruction of GSC migrating in Matrigel 50% after 24 h shows that collective migrating extensions are located in the same focal plane.



**SUPPLEMENTARY FIG. S3.** Microrods covered and noncovered with Matrigel. Immunostaining with anti-Collagen type IV eFluor<sup>®</sup> 660 (red) was used to corroborate the presence of Matrigel deposited on the surface of the microrods. Scale bars 100  $\mu\text{m}$ .



**SUPPLEMENTARY FIG. S5.** GSC (GBAM1) neurosphere migratory behavior over time when cultured in matrices of Oligomer 3 mg/mL, Oligomer 2 mg/mL, and BD-Col 3 mg/mL. The neurospheres show expansive growth accompanied by cell detachment and mesenchymal-like single migration. Scale bars 100  $\mu$ m.



**SUPPLEMENTARY FIG. S6.** GSCs (GBAM1) present multiple migration modes in a HA:10-Col:2mg/mL matrix with embedded noncoated rods, single-cell migration toward the matrix, and collective strand migration along the microrods. Scale bars 100  $\mu$ m.

SUPPLEMENTARY TABLE S1. VISCOELASTIC PROPERTIES  
 OF THE COMPOSITE MATRICES OF OLIGOMER  
 COLLAGEN AND HYALURONAN ( $N=3$ , MEAN  $\pm$  SEM)

<i>Matrix</i>	<i>Viscoelastic properties</i>		
	$G'$ (Pa)	$G''$ (Pa)	$\delta$ (degrees)
HA:2-Col:2 mg/mL	126.2 $\pm$ 14.6	15.8 $\pm$ 0.4	7.2 $\pm$ 0.1
HA:5-Col:2 mg/mL	107.9 $\pm$ 19.6	13.4 $\pm$ 2.2	7.1 $\pm$ 0.1
HA:10-Col:2 mg/mL	36.1 $\pm$ 0.6	3.7 $\pm$ 0.1	5.9 $\pm$ 0.0

HA, hyaluronan.