

Supplementary Figure 1

Electropherograms of all identified mutations in *ATF6*.

Mutant sequence (top) compared to wild-type sequence (bottom). Nucleotide and protein sequence (one-letter code) are presented beneath the electropherogram. Exonic sequence is given in uppercase letters, and intronic sequence is given in lowercase letters. Arrows indicate the mutation.

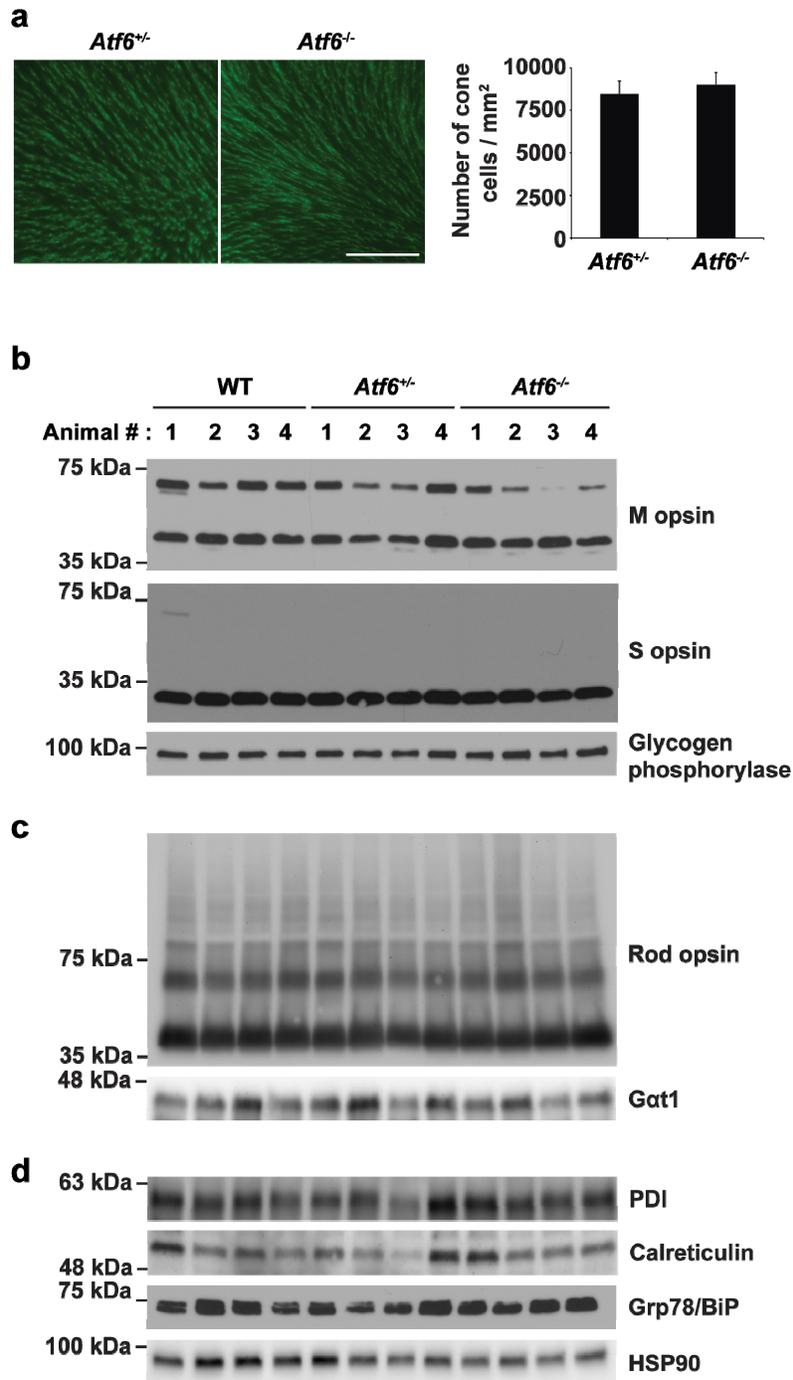
| p.R324C | | | | | |
|-----------------|-----|---------------|---|-------------|-----|
| H.sapiens | 311 | QRMIKNRESACQS | R | KKKKEYMLGLE | 335 |
| P.troglodytes | 311 | QRMIKNRESACQS | R | KKKKEYMLGLE | 335 |
| C.lupus | 366 | QRMIKNRESACQS | R | KKKKEYMLGLE | 390 |
| M.musculus | 298 | QRMIKNRESACQS | R | KKKKEYMLGLE | 322 |
| R.norvegicus | 385 | QRMIKNRESACQS | R | KKKKEYMLGLE | 409 |
| G.gallus | 270 | QRMIKNRESAFQS | R | KKKKEYMLGLE | 294 |
| D.rerio | 297 | QRMIKNRESASLS | R | KKKKEYLMTLE | 321 |
| A.gambiae | 203 | QRMIKNRQSALES | R | QKKKEYVTSLE | 227 |
| H.sapiens ATF6B | 339 | QRMIKNRESACQS | R | RKKKEYLQGLE | 354 |

| p.Y567N | | | | | |
|-----------------|-----|---------------|---|-------------|-----|
| H.sapiens | 554 | DFFEAIRRRGDTF | Y | VVSFRRDHLLL | 578 |
| P.troglodytes | 554 | DFFEAIRRRGDTF | Y | VVSFRRDHLLL | 578 |
| M.mulatta | 536 | DFFEAIRRRGDTF | Y | VVSFRRDHLLL | 560 |
| C.lupus | 542 | GFFDAIRRRGDTF | Y | VVSFRRDHLLL | 566 |
| B.taurus | 604 | DFFEAIRRRGDTF | Y | VVSFRRDHLLL | 628 |
| M.musculus | 540 | GFFDAIRRRGDTF | Y | VVSFRRDHLLL | 564 |
| R.norvegicus | 540 | DFFEAIRRRGDTF | Y | VVSFRRDHLLL | 564 |
| G.gallus | 555 | DFFEAIHRKEDTF | Y | VVSFRRDHLLL | 579 |
| A.gambiae | 498 | EFFEEIGRRDDTF | Y | LVSFSEEHLLL | 522 |
| D.rerio | 539 | DFFDELNRRGDTF | Y | VISFRRDHLLL | 563 |
| H.sapiens ATF6B | 590 | AFLDAIDRREDTF | Y | VVSFRRDHLLL | 514 |

Supplementary Figure 2

Comparative sequence analysis showing conservation of ATF6A at the protein level.

Comparative sequence analysis showing conservation of ATF6A at the protein level according to HomoloGene. In addition, alignment with human ATF6B is shown on the last line for comparison. Top, protein alignment for the p.Arg324Cys missense mutation. Bottom, protein alignment for the p.Tyr567Asn mutation.



Supplementary Figure 3

Atf6^{-/-} mouse whole-mount and biochemistry data.

(a) Whole-mount retina preparations from 1-year-old *Atf6*^{+/-} and *Atf6*^{-/-} mice were stained with FITC-PNA (left immunofluorescent images), and the numbers of FITC-PNA-positive cone cells were quantified with Keyence BZ image analysis software for eight different eyes. Scale bar, 500 μ m. (b–d) Whole retinas were collected and lysed from 90-d-old wild-type, *Atf6*^{+/-} and *Atf6*^{-/-} mice ($n = 4$ per genotype). (b) Cone-specific proteins (M opsin, S opsin and glycogen phosphorylase), (c) rod-specific proteins (rhodopsin and Gat1 / rod transducin) and (d) ER stress-induced proteins (PDI, calreticulin and BiP (*Grp78*)) were detected by immunoblotting. HSP90 served as a protein loading control. WT, wild type; FITC, fluorescein isothiocyanate; PNA, peanut agglutinine.

Supplementary Tables:

Supplementary Table 1: Summary of patients' demographics, clinical data and *ATF6* mutations and genotypes

| Patient ID | <i>ATF6</i> mutation Nucleotide level | <i>ATF6</i> mutation Protein level | Ethnic descent / Age / Gender | VA (Refraction) OD, OS | Fundus | OCT | Color vision | Visual field | Scotopic / photopic ERG | Glare | Nystagmus | Progression |
|---------------|---------------------------------------|------------------------------------|-------------------------------|---|--|--|---------------------|--------------------------|-------------------------|-------|---|-------------|
| CHRO282-II:1 | c.82+5G>T homozygous | Splice defect | South Tyrolean / 42 / M | 20/100 (+0.5 +2.0 90°), 20/2000 (+1.0 +2.0 90°) | Macular changes | n.a. | Achromat | n.a. | n.a. | Yes | Nystagmus at age 5 months | No |
| CHRO628-II:2 | c.970C>T homozygous | p.Arg324Cys | Irish / 19 / M | 20/63 (+2.0 -1.5 162°), 20/100 (+2.25 -0.75 5°) | Small RPE-defect in the fovea, pathological reflexes | Foveal hypoplasia, missing foveal pit, IS/OS and RPE disruption | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Nystagmus at birth, currently no nystagmus | No |
| CHRO628-II:4 | c.970C>T homozygous | p.Arg324Cys | Irish / 16 / F | 20/100 (0.0 -1.0 180°), 20/63 (-1.0) | Small RPE-defect in the fovea, pathological reflexes | Foveal hypoplasia, missing foveal pit, IS/OS and RPE disruption | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Nystagmus at birth, currently no nystagmus | No |
| CHRO628-II:6 | c.970C>T homozygous | p.Arg324Cys | Irish / 9 / F | 20/200 (+3.0), 20/200 (+3.0) | Small RPE-defect in the fovea, pathological reflexes | Foveal hypoplasia, missing foveal pit, IS/OS and RPE disruption | n.a. | n.a. | n.a. | Yes | Nystagmus at birth, currently no nystagmus | No |
| CHRO91-II:1 | c.970C>T homozygous | p.Arg324Cys | British / 47 / F | 20/448 (-4.0 -4.25 10°), 20/252 (-4.5 -4.00 2°) | Mild peripapillary atrophy, small amount of foveal atrophy | Foveal hypoplasia,, IS/OS absence | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Pendular nystagmus at birth, now mild nystagmus | No |
| CHRO91-II:2 | c.970C>T homozygous | p.Arg324Cys | British / 45 / F | 20/209 (+6.5 -2.0 5°), 20/115 (+6.0 -1.5 15°) | Mild peripapillary atrophy, small amount of foveal atrophy | Foveal hypoplasia, IS/OS absence | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Pendular nystagmus at birth, now mild nystagmus | No |
| CHRO91-II:3 | c.970C>T homozygous | p.Arg324Cys | British / 43 / M | 20/110 (-6.5), 20/152 (-9.0) | Marked peripapillary atrophy, marked foveal atrophy | Foveal hypoplasia, marked loss of outer retina and RPE | Achromat | Relative central scotoma | n.a. | Yes | Pendular nystagmus at birth, now mild nystagmus | No |
| CHRO709-II:1 | c.1187+5G>C homozygous | Splice defect | Asian-Indian / 27 / F | 20/132 (0.0 -2.5 15°), 20/145 (-1.5 -2.25 10°) | Mild pigmentary changes at macula | Foveal hypoplasia, IS/OS absence | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Pendular nystagmus at birth, now mild nystagmus | No |
| CHRO709-II:2 | c.1187+5G>C homozygous | Splice defect | Asian-Indian / 23 / F | 20/110 (+ 2.5 -2.0 180°), 20/166 (+2.0 -1.0 50°) | Mild pigmentary changes at macula | Foveal hypoplasia, IS/OS absence | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Pendular nystagmus at birth, now mild nystagmus | No |
| CHRO593-IV:1 | c.1533+1G>C homozygous | Splice defect | French-Canadian / 17 / M | 20/160 (+4.25 +0.75 92°), 20/160 (+2.0 +1.25 92°) | Bull's eye maculopathy | n.a. | Incomplete achromat | n.a. | Normal / Reduced | Yes | Currently convergence controlled nystagmus | n.a. |
| CHRO593-II:3 | c.1533+1G>C homozygous | Splice defect | French-Canadian / 94 / M | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. |
| MOGL411-III:4 | c.1533+1G>C homozygous | Splice defect | French-Canadian / 59 / F | 20/100 (+6.0 + 1.0 90°), 20/100 (+6.0 +1.0 90°) | Foveal atrophy | Foveal hypoplasia with no umbo formation, cone loss in the central fovea | Achromat | Normal | Normal / Reduced | Yes | Yes | n.a. |

| Patient ID | ATF6 mutation Nucleotide level | ATF6 mutation Protein level | Ethnic descent / Age / Gender | VA (Refraction) OD, OS | Fundus | OCT | Color vision | Visual field | Scotopic / photopic ERG | Glare | Nystagmus | Progression |
|----------------------|--|-----------------------------|-------------------------------|--|---|--|---------------------|---|-------------------------|-------|--|-------------|
| MOGL411-MOGL467-IV:1 | c.1533+1G>C homozygous | Splice defect | French-Canadian / 25 / M | 20/100 (-6.5 + 3.5 90°), 20/100 (-9.5 + 5.0 90°) | Foveal atrophy | Foveal hypoplasia with no umbo formation, cone loss in the central fovea | Achromat | Normal | Reduced / Reduced | Yes | No | n.a. |
| MOGL5414-II:1 | c.1533+1G>C homozygous | Splice defect | French-Canadian / 32 / M | 20/70 (-5.5 + 1.75 90°), 20/80 (-5.75 + 1.25 90°) | Foveal atrophy | Foveal hypoplasia with no umbo formation, cone loss in the central fovea | Achromat | Normal | Normal / Non-detectable | Yes | No | n.a. |
| CHRO649-II:1 | c.1699T>A homozygous | p.Tyr567Asn | Iranian / 26 / F | 20/200 (-1.5+3.5 85°), 20/100 (-1.5+2.75 105°) | Dull foveal reflex with RPE changes at the fovea OU, fine RPE mottling left eye | Partial foveal hypoplasia; presence of optical gap (hyporefectivity) at the fovea in both eyes | Incomplete achromat | Relative central scotoma | Normal / Non-detectable | Yes | Nystagmus since infancy; now milder | Yes |
| ZD179-II:1 | c.353delC homozygous | p.Pro118Leufs*31 | Turkish / 41 / F | 20/200 (0.0 -4.0 180°), 20/200 (0.0 -4.5 170°) | Dull foveal reflex with RPE changes at the fovea | No foveal impression, no subfoveal inner outer segment border | Achromat | n.a. | Normal / Non-detectable | Yes | Yes | n.a. |
| CHRO436-II:1 | c.797dupC / c.1110dupA compound-heterozygous | p.Asn267* / p.Val371Serfs*3 | German / 22 / M | 20/100 (+7.25 -2.25 5°), 20/200 (+7.25 -2.25 5°) | Small RPE-defect in the fovea, pathological reflexes, small drusen | Foveal hypoplasia, missing foveal pit, IS/OS and RPE disruption | Achromat | Relative central scotoma | Normal / Non-detectable | Yes | Nystagmus at birth, currently no nystagmus | No |
| CHRO436-II:2 | c.797dupC / c.1110dupA compound-heterozygous | p.Asn267* / p.Val371Serfs*3 | German / 17 / F | 20/200 (+7.25 -1.25 100°), 20/200 (+8.25 -1.25 20°) | Small RPE-defects in the fovea | Foveal hypoplasia, missing foveal pit, IS/OS and RPE disruption | Achromat | Slightly narrowed outer boundaries due to glare | Normal / Non-detectable | Yes | Yes | No |

Abbreviations: OD: right eye; OS: left eye; OU: both eyes; n.a.: not available; M: male; F: female; RPE: retinal pigment epithelium; IS: inner segment; OS: outer segment.

Supplementary Table 2: PCR and Sequencing Primers for *ATF6* used in this study.

| Exon | Forward Primer (5'-3') | Reverse Primer (5'-3') |
|----------------------|----------------------------|----------------------------|
| <i>ATF6</i> -Exon 1 | CTGAAAAC TCCAAAAGGGAAA | GCGGAAAGTAGGGAGGAGGAA |
| <i>ATF6</i> -Exon 2 | GGTGACATAGGGACACAGTGC | AGCCTGAACCTGTTGTCCTG |
| <i>ATF6</i> -Exon 3 | TGCCAAATTGTGTCTCACAG | GCCCAATAACCCCACCTAAT |
| <i>ATF6</i> -Exon 4 | TTTTGGTGACCTTAGCTTCCA | CCCACAAGGCTCTTTCTTGA |
| <i>ATF6</i> -Exon 5 | TCCTTTGAAGTTACCCTGAAGTG | AAAACAGCAAGCCAGCCTAA |
| <i>ATF6</i> -Exon 6 | TGTGTAGAGAAAGGTGTGTGATAGAA | GCACGTGCTCAGAAATTATAACC |
| <i>ATF6</i> -Exon 7 | TCCAATCCCTTGGTGTGAT | TGTGCAATTCAGCACACTCTT |
| <i>ATF6</i> -Exon 8 | AGTTTGAGCAGTGTGTTTTCAA | ACCCTCCTTTTCTTAGCACT |
| <i>ATF6</i> -Exon 9 | AAGCGTTGCTTTTTCTGAAATC | TGACAAAAGCTAAAGAATAATGAGAA |
| <i>ATF6</i> -Exon 10 | AGCTGCATGTAGCAGGCATA | CACTTAGGCCAAAAGATAGATGC |
| <i>ATF6</i> -Exon 11 | TTTCCATGAAAAGTTAACACTAATGA | CCAACTCAGATGTTCTGCAA |
| <i>ATF6</i> -Exon 12 | GTTTGAATGCATGGTTGACAGG | CAGGAGACAGTGGGAAGAGG |
| <i>ATF6</i> -Exon 13 | TGTCAGAAATCTTAGCAATGGAA | TTTAAGGTGAAAGGGAAGTGTGA |
| <i>ATF6</i> -Exon 14 | GAATGGGAAGTATTTTGGGAAA | GTGTACGGCAAGCACTCAAA |
| <i>ATF6</i> -Exon 15 | TTTTTTCATCATCCCAACGAA | TCCTGGTTAAACAAAGAAATCTCA |
| <i>ATF6</i> -Exon 16 | CCTCTACACCCCTTGAAGT | TTCTCTGCCTGCCACCAAG |

PCR was performed with standard chemistry on ABI Veriti Cycler applying the following PCR program: 95°C 3min – 95°C 15sec, 55°C 15sec, 72°C 30sec, 35 cycles– 72°C 7min, cool down to 8°C (except for exons 2 and 13 where annealing temperatures of 65°C and 60°C, respectively, were used).

Supplementary Table 3: Primer sequences used for qPCR in this study.

| Gene | Forward primer (5'-3') | Reverse primer (5'-3') |
|----------------------------------|------------------------|------------------------|
| <i>GAPDH</i> (human) | GTCGGAGTCAACGGATT | AAGCTTCCCGTTCTCAG |
| <i>RPL19</i> (human) | ATGTATCACAGCCTGTACCTG | TTCTTGGTCTCTTCCCTCCTTG |
| <i>HSPA5 (GRP78/BiP)</i> (human) | GCCTGTATTTCTAGACCTGCC | TTCATCTTGCCAGCCAGTTG |
| <i>HERPUD1</i> (human) | AACGGCATGTTTTGCATCTG | GGGGAAGAAAGGTTCCGAAG |
| <i>SEL1L</i> (human) | ATCTCCAAAAGGCAGCAAGC | TGGGAGAGCCTTCCCTCAGTC |
| <i>EDEM1</i> (human) | TTCCCTCCTGGTGAATTTG | AGGCCACTCTGCTTTCCAAC |

For quantitative PCR analysis, cDNA was used as template in SYBR green qPCR supermix (Bio-Rad). *GAPDH* or *Rpl19* mRNA levels served as internal normalization standards. qPCR condition was 95°C for 5 min; 95°C for 10 sec; 60°C for 10 sec; 72°C for 10 sec, with 40 cycles of amplification.