

## SUPPLEMENTARY MATERIAL

<b>Locus</b>	<b>Number of pairwise comparisons</b>	<b>Significant pairwise comparisons by Fisher's exact test</b>	<b>After Bonferroni correction</b>	<b>Significant pairwise comparisons by <math>\chi^2</math> test</b>	<b>After Bonferroni correction</b>
<i>Apt</i>	990	286	83	301	202
<i>Cycle</i>	300	86	15	131	18
<i>Cyp303a1</i>	1830	1119	683	1156	633
<i>Cyp305b1</i>	741	97	5	126	43
<i>Period</i>	378	40	1	48	30
<i>Phc</i>	1953	325	28	397	115
<i>SCAP</i>	780	93	0	171	0
<i>Tc</i>	946	110	0	129	0
<i>Tpi</i>	2926	375	36	549	87

**Table S1** Average  $r^2$  and number of significant pairwise comparisons before and after Bonferroni correction

Locus	Accession		Primer sequence
<i>Apt</i>	AB024903.1	forward:	CTTGGTCTACTGCCGACCTCACT
		reverse:	TAGCCATCCTCATTGTTTGGACT
<i>Cycle</i>	BAB20632.1	forward:	GCTATGGCGAGGAAGTTAGACAA
		reverse:	TCTCTCCCGTGGGCTGAGGT
<i>Cyp303a1</i>	NM_143813.2	forward:	ACGGCATTTCATGGGGCGCA
		reverse:	GTGCTTGGCTAGGCCGAACGGA
		forward, nested:	GCTCGCTGGATATTTTATACCAGA
<i>Cyp305b1</i>	NP_001106220.1	forward:	ACACGTCTGCGTTTCTCCAA
		reverse:	GGTGTAGCCAATATAACCAATCAAC
<i>down3</i>		forward:	TCTCCATTCGGTTTCCCTTC
		reverse:	TAATCACTGGGTTGCTTCTGG
<i>Period</i>	ABF21088.1	forward:	CAGGGACTCGGGTGAGATGA
		reverse:	AGCACTGGTTGGATGGTAGG
<i>Phc</i>	XP_002432072.1	forward:	TACGCGAAGATGTGGTACAAGG
		reverse:	ACAAGTCCATCGGCGGTCTG
<i>SCAP</i>	XP_974766.1	forward:	AAAGCCAAAGCGAGCATAGCA
		reverse:	TACTGAACAAGCAGCCAGACC
<i>Tc</i>	XP_972068.2	forward:	TGTATTCCCAAGTCCGCTGTTT
		reverse:	TTGTTGATAGCTTCGCAAGAGT
<i>Tpi</i>	AY736358	forward:	ATTCGTTGTTGGTGGTAACTGGA
		reverse:	TTTGCCTGCCTCCCTCTCTTCT
<i>up3</i>		forward:	GTTGCGAGTAAGATAGCAGCAC
		reverse:	GTTGCGTGGCAGGAAGGTAA

**Table S2** Accessions of Z-linked loci and primer sequences used in this study. In cases where sequence quality was unsatisfactory, an internal (nested) primer was used for re-sequencing. The loci *down3* and *up3* refer to the regions 3 kb downstream and upstream of *Cyp303a1*, respectively, in which no known genes were identified.

<i>F<sub>ST</sub></i> values for pairwise comparisons of populations			
Locus	Dal X Orb	Dal X MV	Orb X MV
<i>Apt</i>	-0.077	0.020	0.013
<i>Cyp303a1</i>	-0.036	-0.012	0.018
<i>Period</i>	-0.017	-0.005	0.025
<i>Phc</i>	0.059	-0.028	-0.009
<i>Tpi</i>	-0.044	0.003	-0.176

**Table S3** *F<sub>ST</sub>* values for five loci sampled from Dalmore, Orbost and MacIntyre Valley populations using an unbiased estimator (Hudson *et al.*, 1992)

Locus	n	Number of sites (bp) <sup>a</sup>	Number of indel events		$\pi(i)$	
<i>Apt</i> (55)	Dalmore (8)	779–840	8	23	0.004	0.005
	Orbost (5)		4		0.003	
	M.Valley (41)		20		0.005	
<i>Cycle</i> (21)	M.Valley (20)	919		11		0.002
<i>Cyp303a1</i> (83) <sup>b</sup>	Dalmore (14)	774–780	13	20	0.005	0.006
	Orbost (10)		7		0.005	
	M.Valley (56)		17		0.006	
<i>Cyp305b1</i> (22)	M.Valley (21)	696		8		
<i>Period</i> (36)	Dalmore (13)	486–502	27	46	0.015	0.014
	Orbost (5)		12		0.011	
	M.Valley (17)		32		0.013	
<i>Phc</i> (36)	Dalmore (12)	723–754	28	38	0.011	0.010
	Orbost (7)		20		0.010	
	M.Valley (16)		25		0.009	
<i>SCAP</i> (12)	M.Valley (11)	974		20		
<i>Tc</i> (16)	M.Valley (15)	914		16		0.004
<i>Tpi</i> (33)	Dalmore (11)	551–577	18	21	0.009	0.007
	Orbost (2)		3		0.005	
	M.Valley (19)		14		0.005	

**Table S4** Indel diversity for nine loci surveyed in this study. Where estimates are presented in two columns under a single heading, the left column represents estimates for an individual population while the right column represents estimates after pooling sequences of all three populations. Figures in brackets after the locus name represent the total number of sequences surveyed including the reference strain.

\*  $p < 0.05$

<sup>a</sup> the number of sites is presented as a range due to the differing subsets of indel polymorphisms present in different populations. Since gapped sites are included in this analysis, the upper boundary represents the number of sites considered when alleles from all three populations are pooled.

<sup>b</sup> includes two sequences from a laboratory-maintained colony

Site	<i>Del200</i>	<i>Ins200</i>	<i>H. assulta</i>	<i>H. punctigera</i>	<i>H. zea</i>
1	ATAA	-	x	x	GTAA
2	G(1A)	C	C	x	C
3	C	T	T	x	T
4	T	C	C	x	T
5	G	A/T	T	x	T
6	T(1G)	A	A/C	x	A
7	C	G	G	x	G
8	A	G	G	x	G
9	G	A	G	x	G
10	C(1G)	A	C	x	C
11	G	T	T	x	T
12	G	C	G/A	x	C
13	T	A	T	x	T
14	G	T	G	x	T
15	A	G	G	x	G
16	T	C	T/-	x	C
17	G	T	T/-	x	T
18	C	G	C	x	C
19	C	A	C	x	C
20	A	G	G	x	-
21	A	G	G	x	-
22	A	G	A	x	G
23	A	C	C	x	C
24	T	A/C	T	T	T
25	A	C	C	C	C
26	A	T	T	T	T
27	T	C	C	C	C
28	C	A	A	A	C
29	T	A	A	A	G
30	AACG	-	AATG	AATN	-
31	-	A	GA	-	A
32	A	C	A	A	G

33	A	C	A	A	G
34	A	G	A	A	T
35	G	A	A	A	A
36	G	A	AG	-	G

Number of sites with state identical or closer to the

<i>Del200</i> haplogroup	10	14	5
<i>Ins200</i> haplogroup	20	19	6
Ambiguous unknown:	6	3	25

**Table S5** State of the outgroup consensus sequences at sites where fixed differences occur between the *H. armigera* insertion and deletion haplogroups. '-' represents a gap in the alignment flanked by matching nucleotides whereas 'x' represents a gap that forms part of a deletion extending beyond the boundaries of the sites where the fixed differences occur. All sites marked with an 'x' are deemed ambiguous. Sites with a '-' are considered unambiguous if they match one of the two states in the *H. armigera* alleles e.g. site 20 in *H. zea* is considered ambiguous but site 30 is not. Sites which require more than one change from either allele are also deemed ambiguous e.g. site 30 in *H. assulta* is considered to be closer to the insertion state but the same site in *H. punctigera* is deemed ambiguous because a single event is insufficient to produce a match to either state. Similarly, site 31 in *H. assulta* is ambiguous because only one event (a mono- or di-nucleotide insertion) is sufficient to produce a match to either state. Where polymorphisms are present in the outgroup sequences, they are deemed unambiguous if at least one of the states is identical to that of *H. armigera* and all possible states are observed to be exclusive of the alternate *H. armigera* allele. Sites 2, 5, 6, 10 and 24 are, strictly speaking, not fixed differences, but have been included because no overlapping states have been observed between the two *H. armigera* alleles.

Organism	$\pi$ ( $10^2$ )	Approximate distance (bp) at which $E(r^2) \approx 0.2$	References
<b>Lepidoptera</b>			
<i>Bombyx mori</i>	1	>1600 <sup>a</sup>	Xia <i>et al.</i> (2009); Guo <i>et al.</i> (2011)
<i>Bombyx mandarina</i>	1–2	20–200 <sup>a</sup>	Xia <i>et al.</i> (2009); Guo <i>et al.</i> (2011)
<i>Heliconius erato</i>	2	<500	Counterman <i>et al.</i> (2010)
<i>Heliconius melpomene</i>	1	>500	Baxter <i>et al.</i> (2010)
<i>Helicoverpa armigera</i>	3	10–200 <sup>b</sup>	this study
<i>Melitaea cinxia</i>	n.a	3000	Ahola <i>et al.</i> (2014)
<b>Other insects</b>			
<i>Acyrtosiphon pisum</i>	0.6	1000	Brisson <i>et al.</i> (2009)
<i>Anopheles arabiensis</i>	0.2–0.3	<200	Marsden <i>et al.</i> (2014)
<i>Anopheles gambiae</i>	0.8–25	<200	Wilding <i>et al.</i> (2009); Harris <i>et al.</i> (2010)
<i>Drosophila melanogaster</i>	0.6–2	30 <sup>c</sup> –640 <sup>d</sup>	Langley <i>et al.</i> (2012); Pool <i>et al.</i> (2012)

**Table S6** Estimates of nucleotide diversity,  $\pi$  and linkage disequilibrium,  $E(r^2)$  in different species. Unless otherwise indicated, LD was imputed from diploid sequences.

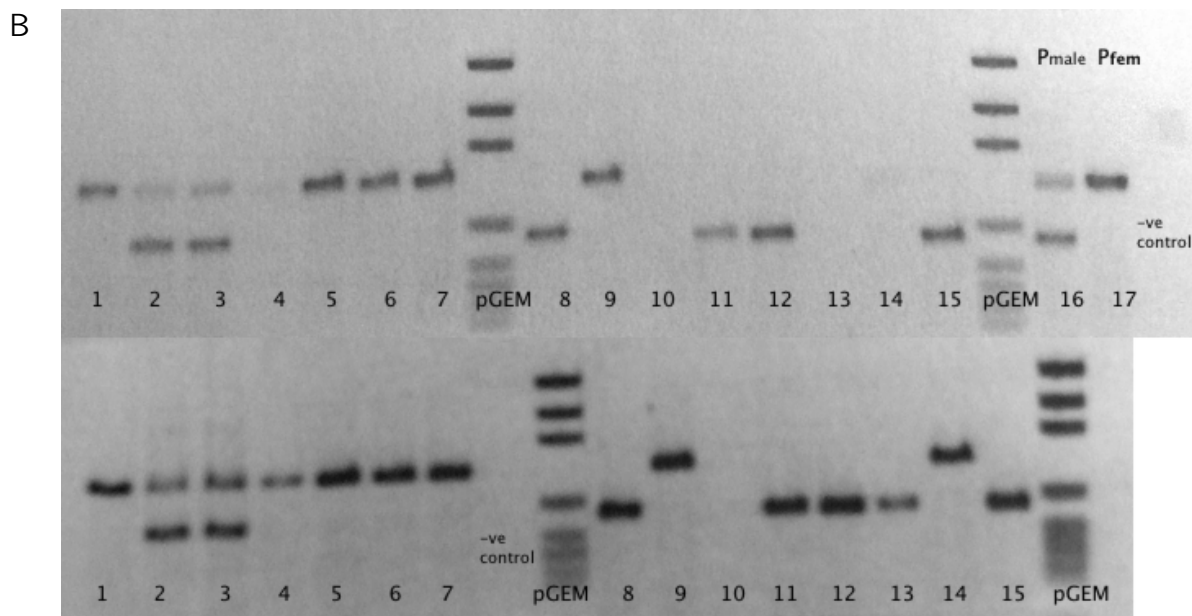
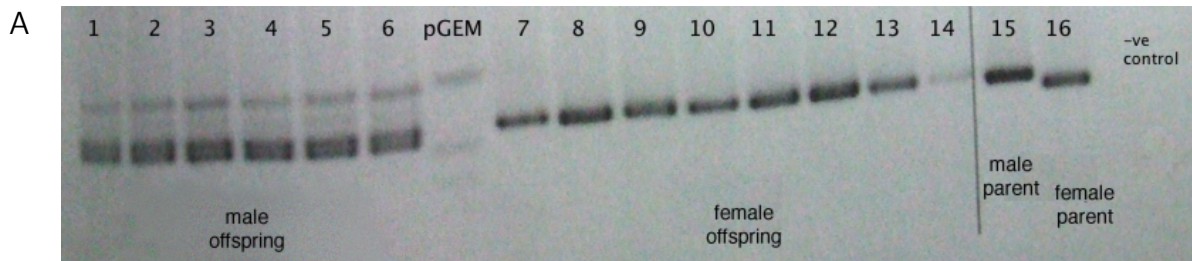
<sup>a</sup> haploid by cloning amplicons prior to sequencing

<sup>b</sup> haploid by sequencing sex chromosomes in hemizygous individuals

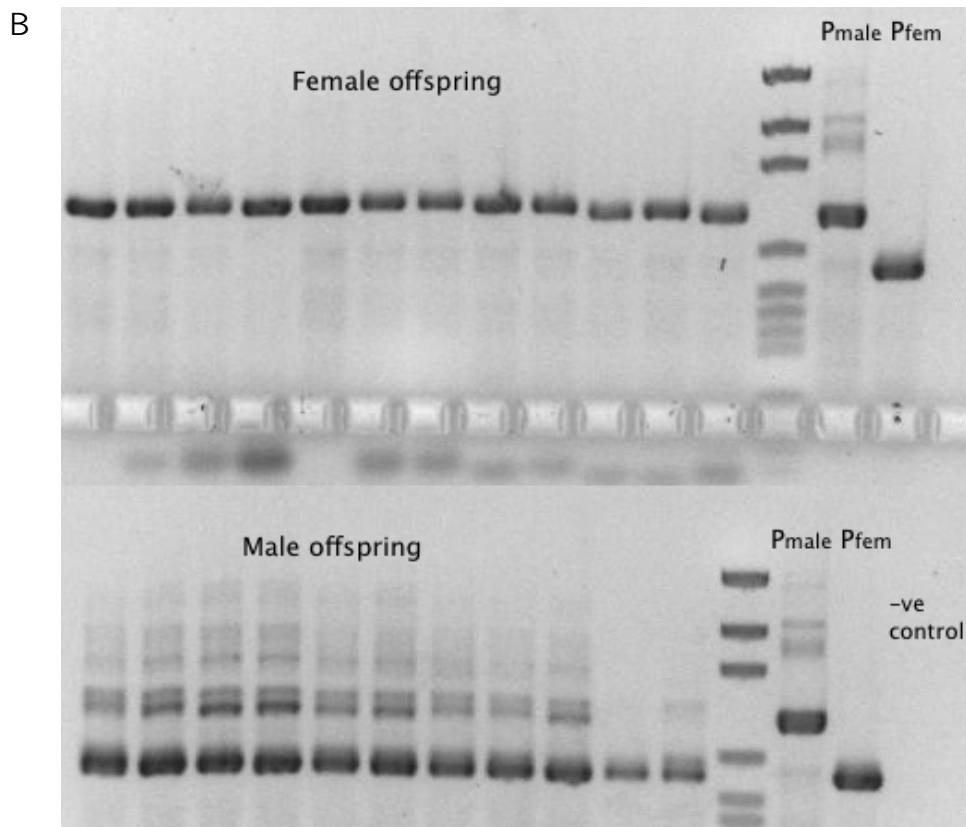
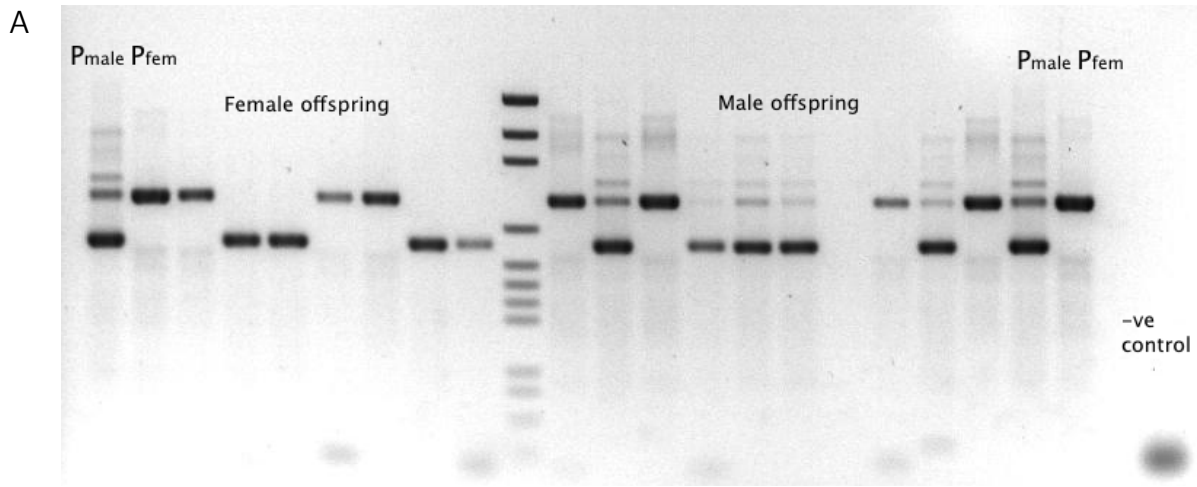
<sup>c</sup> haploid embryos

<sup>d</sup> haploid by using inbred lines

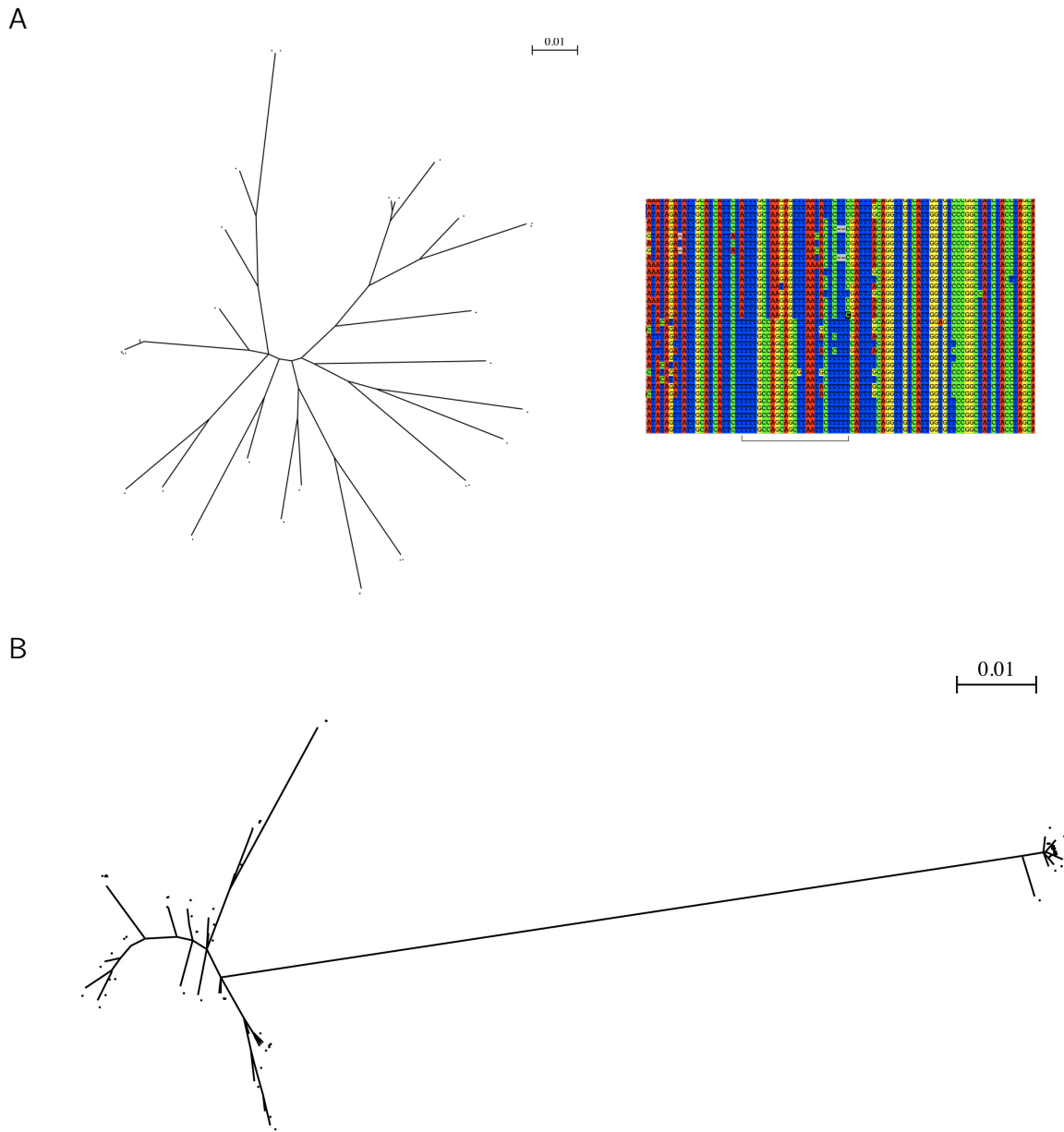




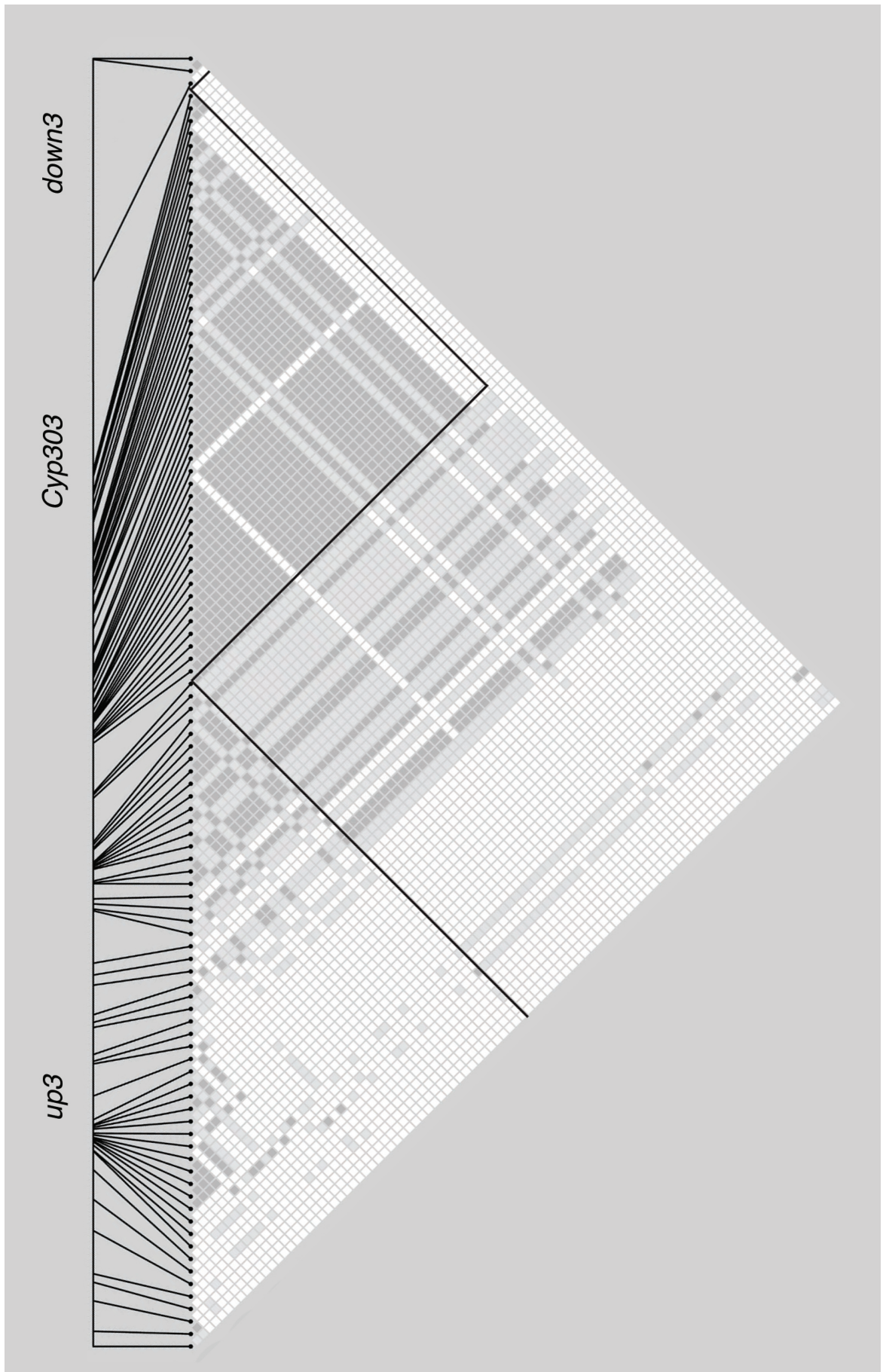
**Figure S1** Pedigree analysis of (A) *Period*. All male offspring (lanes 1–6) present double bands inherited from both parents while all female offspring (7–14) inherit a single band from the male parent (15). The female parent has a slightly shorter product (16) and her allele is only seen in male offspring. Pedigree analysis of (B) *Phc*. The upper gel shows male offspring (lanes 1–7), female offspring (8–15), male parent (16) and female parent (17). The male parent is heterozygous. The lower gel stems from a technical replicate (different PCR) to account for lack of product in samples 4, 10, 13 and 14. Male offspring exhibit heterozygosity (2,3) while female offspring (excluding sample 10) exhibit hemizyosity.

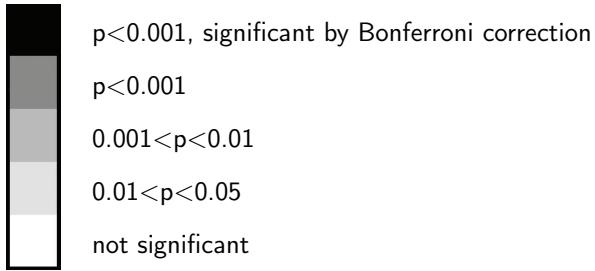


**Figure S2** Pedigree analysis on the *Cyp303a1* insertion and deletion variants. The male parent of family 1 (A) is heterozygous while that of family 2 (B) is homozygous for the insertion. In both cases, female offspring exhibit hemizyosity whereas male offspring may be homozygous or heterozygous.



**Figure S3** Unrooted maximum-likelihood trees for (A) *Tpi* and (B) *Cyp303a1* after exclusion of the 25-bp stretch and 200-bp indel, respectively. The long internal branch is still apparent in *Cyp303a1*. The fixed differences between the insertion and deletion alleles cannot be accounted for by a consecutive stretch of nucleotides that would constitute a single event. In contrast, the alignment in (A) illustrates that excluding a single 25-bp stretch in *Tpi* (underlined with black bar) largely eliminates the long internal branch.





**Figure S4 (preceding page)** LD plotted as significance of the  $r^2$  value for *Cyp303a1* and its flanking regions. The sequence under consideration was generated by concatenating the *up3*, *Cyp303a1* and *down3* loci ( $n=11$ ). The range of each locus is bounded by black lines. Approximately 3 kb separate *Cyp303a1* from its flanking sequences. The downstream locus contained very few segregating sites, hence the small area. LD is observable between the 3' end of the upstream locus and *Cyp303a1*.

## SUPPLEMENTARY REFERENCES

- Ahola V, Lehtonen R, Somervuo P, Salmela L, Koskinen P, Rastas P, Valimaki N, Paulin L, Kvist J, Wahlberg N, Tanskanen J, Hornett EA, Ferguson LC, Luo S, Cao Z, de Jong MA, Duploux A, Smolander OP, Vogel H, McCoy RC, Qian K, Chong WS, Zhang Q, Ahmad F, Haukka JK, Joshi A, Salojarvi J, Wheat CW, Grosse-Wilde E, Hughes D, Katainen R, Pitkanen E, Ylinen J, Waterhouse RM, Turunen M, Vaharautio A, Ojanen SP, Schulman AH, Taipale M, Lawson D, Ukkonen E, Makinen V, Goldsmith MR, Holm L, Auvinen P, Frilander MJ and Hanski I (2014). The Glanville fritillary genome retains an ancient karyotype and reveals selective chromosomal fusions in Lepidoptera. *Nat Commun* **5**
- Baxter SW, Nadeau NJ, Maroja LS, Wilkinson P, Counterman BA, Dawson A, Beltran M, Perez-Espona S, Chamberlain N, Ferguson L, Clark R, Davidson C, Glithero R, Mallet J, McMillan WO, Kronforst M, Joron M, French Constant RH and Jiggins CD (2010). Genomic hotspots for adaptation: The population genetics of Mullerian mimicry in the *Heliconius melpomene* clade. *PLoS Genet* **6**: e1000794
- Brisson J, Nuzhdin S and Stern D (2009). Similar patterns of linkage disequilibrium and nucleotide diversity in native and introduced populations of the pea aphid, *Acyrtosiphon pisum*. *BMC Genetics* **10**: 22
- Counterman BA, Araujo-Perez F, Hines HM, Baxter SW, Morrison CM, Lindstrom DP, Papa R, Ferguson L, Joron M, French Constant RH, Smith CP, Nielsen DM, Chen R, Jiggins CD, Reed RD, Halder G, Mallet J and McMillan WO (2010). Genomic hotspots for adaptation: The population genetics of Mullerian mimicry in *Heliconius erato* **6**
- Guo Y, Shen YH, Sun W, Kishino H, Xiang ZH and Zhang Z (2011). Nucleotide diversity and selection signature in the domesticated silkworm, *Bombyx mori*, and wild silkworm, *Bombyx mandarina*. *Journal of Insect Science* **11**: 155

- Harris C, Rousset F, Morlais I, Fontenille D and Cohuet A (2010). Low linkage disequilibrium in wild *Anopheles gambiae s.l.* populations. *BMC Genetics* **11**: 81
- Hudson RR, Slatkin M and Maddison WP (1992). Estimation of levels of gene flow from DNA sequence data. *Genetics* **132**: 583–589
- Langley CH, Stevens K, Cardeno C, Lee YCG, Schrider DR, Pool JE, Langley SA, Suarez C, Corbett-Detig RB, Kolaczkowski B, Fang S, Nista PM, Holloway AK, Kern AD, Dewey CN, Song YS, Hahn MW and Begun DJ (2012). Genomic variation in natural populations of *Drosophila melanogaster*. *Genetics* **192**: 533–598
- Marsden CD, Lee Y, Kreppel K, Weakley A, Cornel A, Ferguson HM, Eskin E and Lanzaro GC (2014). Diversity, differentiation, and linkage disequilibrium: prospects for association mapping in the malaria vector *Anopheles arabiensis*. *G3 (Bethesda)* **4**: 121–131
- Pool JE, Corbett-Detig RB, Sugino RP, Stevens KA, Cardeno CM, Crepeau MW, Duchon P, Emerson JJ, Saelao P, Begun DJ and Langley CH (2012). Population genomics of sub-Saharan *Drosophila melanogaster*: African diversity and non-African admixture. *PLoS Genet* **8**: e1003080
- Wilding C, Weetman D, Steen K and Donnelly M (2009). High, clustered, nucleotide diversity in the genome of *Anopheles gambiae* revealed through pooled-template sequencing: implications for high-throughput genotyping protocols. *BMC Genomics* **10**: 320
- Xia Q, Guo Y, Zhang Z, Li D, Xuan Z, Li Z, Dai F, Li Y, Cheng D, Li R, Cheng T, Jiang T, Becquet C, Xu X, Liu C, Zha X, Fan W, Lin Y, Shen Y, Jiang L, Jensen J, Hellmann I, Tang S, Zhao P, Xu H, Yu C, Zhang G, Li J, Cao J, Liu S, He N, Zhou Y, Liu H, Zhao J, Ye C, Du Z, Pan G, Zhao A, Shao H, Zeng W, Wu P, Li C, Pan M, Li J, Yin X, Li D, Wang J, Zheng H, Wang W, Zhang X, Li S, Yang H, Lu C, Nielsen R, Zhou Z, Wang J, Xiang Z and Wang J (2009). Complete resequencing of 40 genomes reveals domestication events and genes in silkworm (*Bombyx*). *Science* **326**: 433–436