Supplemental data

Material and methods

Flow cytometry

Mouse ES cells $(1x10^6 \text{ cells in 6 well plates})$ were transfected with shRNA or overexpression plasmids. On the 4^{th} day after selection with puromycin or zeocin, cells suspension in medium and adherent on wells were harvested and fixed with 70% ethanol for 30 minutes. Then these fixed cells were washed with PBS and treated with propidium iodide (20 μ g/ml) and RNase A (0.2 mg/ml) for 30 minutes. The ratio of sub-G1 was determined and analyzed by CellQuest software (Becton Dickinson).

Quantification of Alkaline phosphatase (ALP) activity

Mouse ES cells incubated with 10% Alamar blue for Alamar blue assay were washed with PBS three times and fixed with 4% formaldehyde for 10 minutes. After fixation, cells were re-washed with PBS three times and incubated with the ALP substrate, p-Nitrophenyl phosphate (#N-7653; Sigma-Aldrich), within 10 minutes. Then the ALP activities were measured at OD405 in the substrate solution. To obtain the ALP/AB (alkaline phosphatase activity/Alamar blue activity) value, the ALP activities acquired were divided by the Alamar blue values.

Figure Legends

Figure S1. Down-regulation of DLK enhances mouse ES cell renewal

(A) Apoptosis of cells was slightly decreased in sh-DLK mouse ES cells compared to sh-Control mouse ES cells. Sub-G1 level of D3 mouse ES cells transfected with sh-Control, sh-DLK-1 or sh-DLK-2 was analyzed on 4th day after transfection. (B) Relative cell numbers increased in sh-DLK transfectants in R1 ES cells. Alamar blue assays were performed. (C) The transfection of two independent sh-DLKs increased expression of Nanog protein in R1 mouse ES cells. Expression of self-renewal markers, Nanog and Oct4 proteins in mouse ES cells transfected with sh-Control, sh-DLK-1 or sh-DLK-2 were measured by Western blotting. The error bars in the figures represent standard error of the mean (mean±SEM). *P* values were obtained from two-tailed Student's *t*-tests (***, *P* < 0.0001; **, *P* < 0.001; *, *P* < 0.05).

Figure S2. DLK Protein expression levels in mouse ES cells, EBs, MEFs and in

775 the withdrawal of LIF.

(A) DLK protein expression was upregulated in mouse EBs compared to mouse ES cells and MEFs. DLK protein was detected in wild type D3 mouse ES cells, MEFs and mouse EBs (5th, 10th, 15th and 20th days). No obvious difference in DLK protein was observed in mouse ES cells and MEFs. (B) A decrease of phosphorylated Akt was

observed upon the removal of LIF. The protein expression levels of DLK, 780 phosphorylated Akt, and total Akt were detected by Western blot analysis.

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Figure S3. DLK activity increases upon the treatment of PI3K/Akt signaling

inhibitor in R1 mouse ES cells

(A) LY-294002 treatment down-regulated Akt activity and expression amounts of Nanog. Western blotting was performed to detect DLK, phospho-Akt at S473, phospho-Akt at T308, total Akt, Nanog and actin after R1 mouse ES cells were treated with 0, 0.25 and 1.0 μM of LY-294002. Under the same condition, (B) DLK activity increased does-dependently upon the treatment of LY-294002. Western blotting and kinase assay using MBP as substrates were carried out as DLK protein was immunoprecipitated.

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Figure S4. Up-regulation of DLK in mouse ES cells induces cell death and the

down-regulation of Nanog protein

Mouse ES cells were transfected with GFP, DLK wild type and DLK mutants (DLK S584A, DLK T659A, or DLK S584A/T659A) plasmids. (A) Apoptotic cells increased significantly in D3 mouse ES cells transfected with DLK mutant plasmids. Sub-G1 levels of cells were analyzed on 4th day after transfection. Cells were fixed and stained with propidium iodine. (B) Expression of wild type and mutants of DLK did not affect expression levels of phospho-Akt and Akt. Western blot analyses were performed. (C) Expression of wild type and mutants of DLK did not alter the ALP/AB value. After Alamar blue assay and alkaline phosphatase (ALP) activity were performed, values represented to alkaline phosphatase activity were divided by values obtained from Alamar blue assay in each cells. (D) Relative cell numbers decreased in R1 cells transfected with DLK mutant plasmids. (E) Nanog and Oct4 proteins were downregulated upon transfection of DLK mutant plasmids in R1 cells. Western blot analyses were performed. The error bars in the figures represent standard error of the mean (mean \pm SEM). P values were obtained from two-tailed Student's t-tests (***, P < 0.0001; **, P < 0.001; *, P < 0.05).

Figure S1, Wu

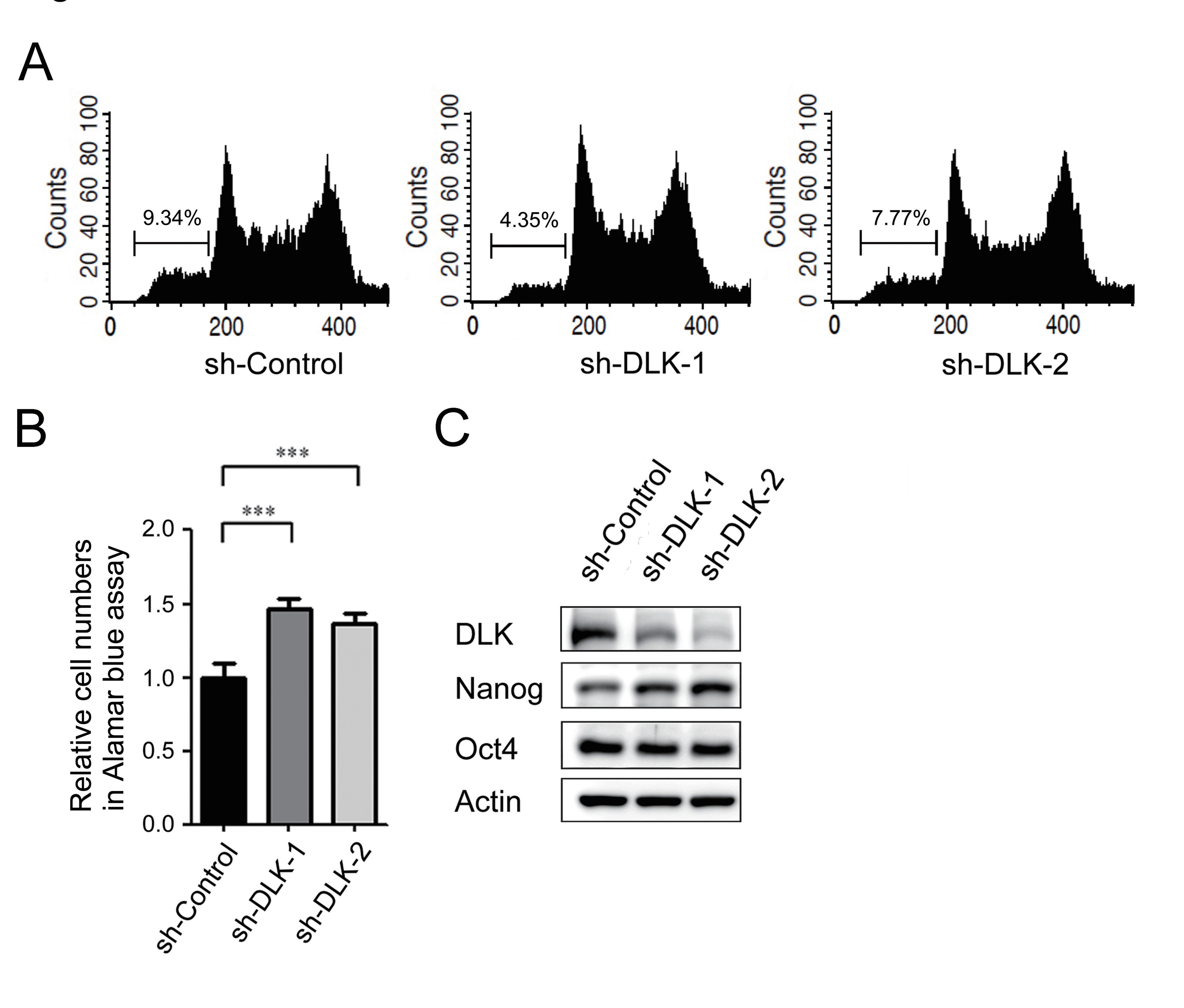


Figure S2, Wu

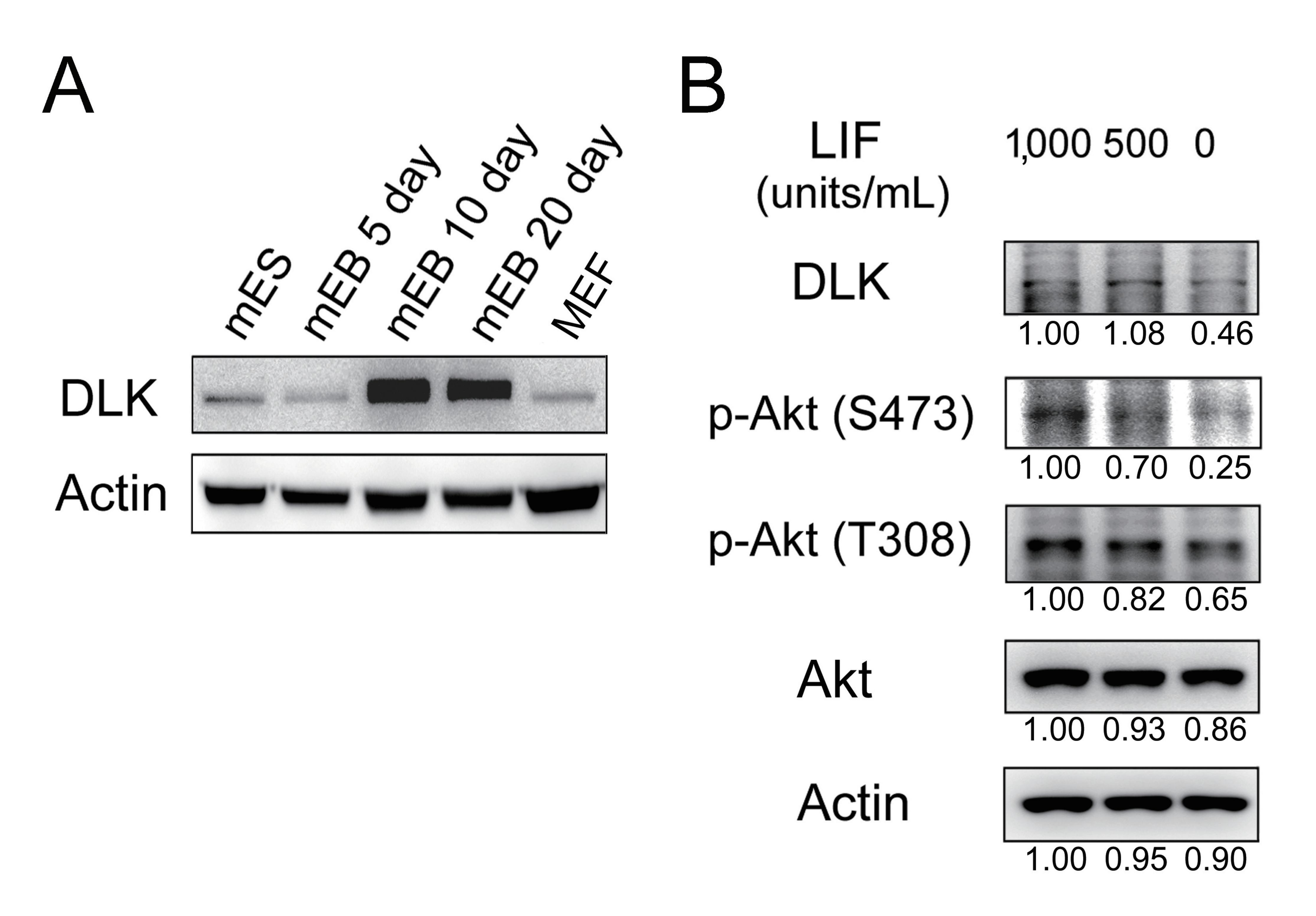


Figure S3, Wu

LY-294002 LY-294002 MBP DLK 0.90 1.00 kinase assay p-Akt (S473) 0.33 p-Akt (T308) immunoprecipitation 0.68 0.86 1.00 Akt 0.86 1.00 Nanog

1.00

Figure S4, Wu




Table 1. Primers used in quantitative real-time PCR

Primer name	Primer sequence
DLK forward	5'-AGCACCAAGATGTCCTTTGC
DLK reverse	5'-ATGATGGCGATTTCGTG
Nanog forward	5'-ATGCGGACTGTTCTCA
Nanog reverse	5'-GCAATGGATGCTGGGATACT
Oct4 forward	5'-ATGAGAACCTTCAGGAGATATG
Oct4 reverse	5'-CTCGTTGGGAATACTCAATACT
GAPDH forward	5'-ACCACAGTCCATGCCATCAC
GAPDH reverse	5'-TCCACCACCTGTTGCTGTA