

# Untangling the biological effects of cerium oxide nanoparticles: the role of surface valence states

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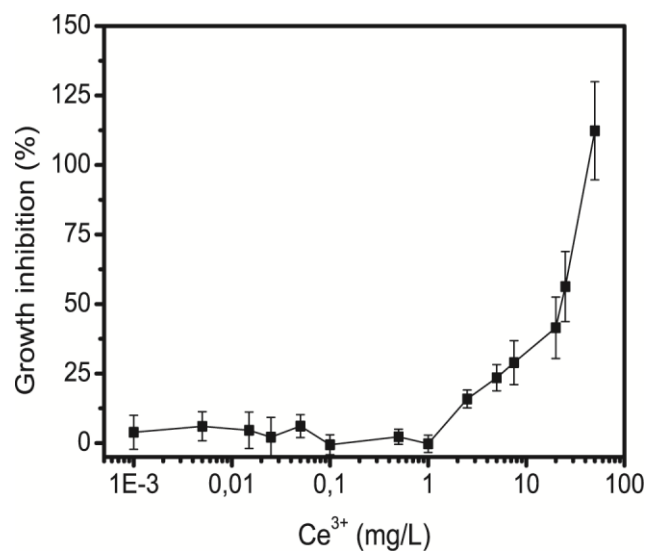
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**Supplementary Table S1.-** Spontaneous cerium dissolution in the exposure media for all CNPs was tested by performing ICP-MS analyses of ultrafiltrated samples (10 mg/l).

<b>Sample</b>	<b>Concentration (mg/l)</b>	<b>Stand. Dev</b>
CNP1	0,00045	4.13%
CNP2	0,00001	29.22%
CNP3	0,00081	8.95%
CNP4	0,00003	4.05%
CNP5	n.d.*	-
CNP6	n.d.*	-
CNP7	n.d.*	-

\*No detected

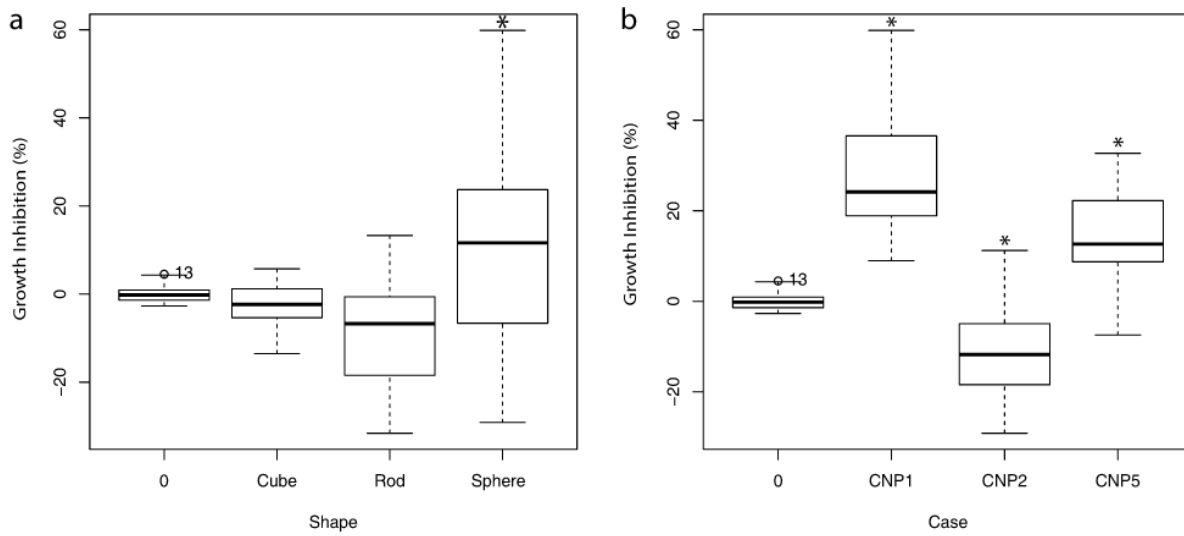
**Supplementary Figure S1.-** Effect of free ion  $Ce^{3+}$  on growth of *P. subcapitata*.



**Supplementary Table S2.-** Summary table of correlation analyses between algal growth inhibition and different tested factors, showing the adjusted  $R^2$  values and their associated  $p$ -values (ANOVA,  $F < P$ ).  $\alpha = 0.05$  is established as significance criterion (marked by asterisks).

<b>Tested factors</b>	<b><math>R^2</math></b>	<b><math>p</math>-value</b>
Surface $\text{Ce}^{3+}$	0.706	0.047*
$\zeta$ -Potential	0.693	0.049*
Nominal Size	0.007	0.384
Effective Size	0.152	0.281

**Supplementary Figure S2.** Box plots which show agal growth inhibition as function of nanoparticle shape (cube: CNP4, rod: CNP3 and sphere: CNP1, CNP2 and CNP5; Figure S2a) and just as function of spheric-shaped nanoceria (CNP1, CNP2 and CNP5; Figure S2b). A one way ANOVA coupled with Tukey's HSD (honestly significant difference) post-hoc test was performed for comparison of means. Statistically significant differences ( $\alpha = 0.05$ ) are marked by asterisks.

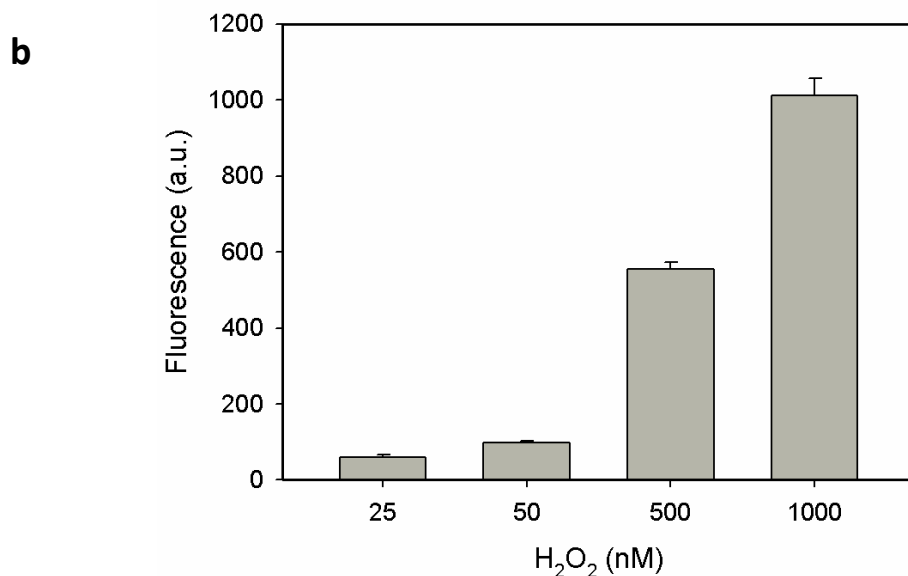
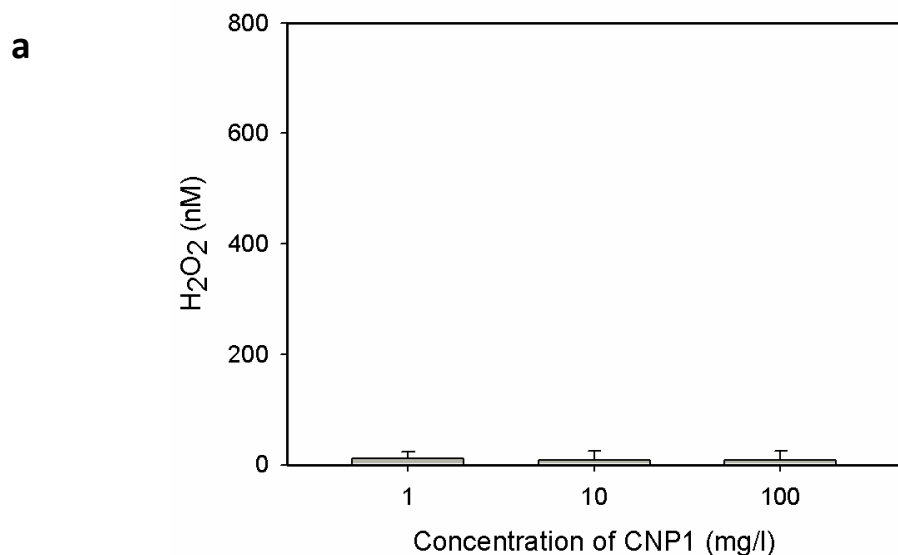


**Supplementary Table S3.-**  $\zeta$ -potential and effective diameter of CNP1 and CNP2 with the addition of Fe or phosphate.

10 mgL <sup>-1</sup> OECD medium						
Sample Name	0.1 mM Fe			0.2 mM Phosphate		
	$\zeta$ -potential (mV)	Effective Diameter (nm)	PDI*	$\zeta$ -potential (mV)	Effective Diameter (nm)	PDI*
CNP1	10.15 ±	787.3	0.979	-19.90 ±	817.2	0.52
	0.889			0.557		6
CNP2	-15.30 ±	1073	0.362	-10.14 ±	450.7	0.57
	1.9			1.06		3

\*PDI=Polydispersity index

**Supplementary Figure S3.-** Detection of H<sub>2</sub>O<sub>2</sub> using the Amplex® Red Hydrogen Peroxide/Peroxidase Assay Kit. Reactions containing 50 μM Amplex® Red reagent, 0.1 U/mL HRP and the indicated amount of CNP1 (a) or H<sub>2</sub>O<sub>2</sub> as standard curve (b) were incubated for 30 minutes at room temperature. The fluorescence (excitation 535 nm, emission 595 nm) was measured on a Fluorostar Omega plate reader (BMG LABTECH GmbH, Germany). Background fluorescence, determined for a control reaction (H<sub>2</sub>O), has been subtracted from each value.



**Supplementary Table S4.-** Composition of the OECD TG 201 medium.

Component	OECD	
	mg/L	mM
NaHCO <sub>3</sub>	50.0	0.595
NaNO <sub>3</sub>		
NH <sub>4</sub> Cl	15.0	0.280
MgCl <sub>2</sub> ·6(H <sub>2</sub> O)	12.0	0.0590
CaCl <sub>2</sub> ·2(H <sub>2</sub> O)	18.0	0.122
MgSO <sub>4</sub> ·7(H <sub>2</sub> O)	15.0	0.0609
K <sub>2</sub> HPO <sub>4</sub>		
KH <sub>2</sub> PO <sub>4</sub>	1.60	0.00919
FeCl <sub>3</sub> ·6(H <sub>2</sub> O)	0.0640	0.000237
Na <sub>2</sub> EDTA·2(H <sub>2</sub> O)	0.100	0.000269
H <sub>3</sub> BO <sub>3</sub>	0.185	0.00299
MnCl <sub>2</sub> ·4(H <sub>2</sub> O)	0.415	0.00210
ZnCl <sub>2</sub>	0.00300	0.0000220
CoCl <sub>2</sub> ·6(H <sub>2</sub> O)	0.00150	0.00000630
Na <sub>2</sub> MoO <sub>4</sub> ·2(H <sub>2</sub> O)	0.00700	0.0000289
CuCl <sub>2</sub> ·2(H <sub>2</sub> O)	0.00001	0.00000006