

Welsh_FigS2

GRO-seq Analysis of Pitx2 Locus Nascent Transcription



Welsh_FigS3





Welsh FigS5



Welsh_FigS6



Welsh_FigS7



Supplemental Figure Legends

Figure S1. Reproducible domains of asymmetric enhancer activity of hs926 by in vivo transgenic analysis. Related to Figure 1 and 3. (A) Human hs926 sequence cloned into the Hsp68-LacZ reporter construct (top) shows reproducible enhancer activity in asymmetric domains of transgenic embryos (bottom). Scale bar 100µm (B) Prediction of *Pitx2* binding sites within e926 conserved between human and mouse (top) or human and chicken (bottom) (http://rvista.dcode.org/).

Figure S2. Annotation of asymmetric *Pitx2* locus transcription in vivo using GRO-

seq. Related to Figure 2. (A) GRO-seq of HH21 whole embryo characterizes nascent transcription across the *Pitx2* locus. (B) Whole embryo, HH12 head, and HH12 left & right hemisected samples were collected in addition to HH21 left and right DM samples to monitor enrichment and specificity of asymmetric transcription. (C) Transcription of bilaterally expressed *Pitx2ab* isoforms is observed in HH21 whole embryo and HH12 head. Elevated *Pitx2* expression is observed in the HH12 left hemisected sample compared to the right, while only the left DM sample detects exclusive *Pitx2c* expression. Dashed vertical lines mark *Pitx2ab* and *Pitx2c* TSS.

Figure S3. Annotation of active regulatory elements at the *Pitx2* locus in vivo using **dREG.** Related to Figure 2. Top: dREG annotation of the *Pitx2* locus from GRO-seq of HH21 whole embryo sample identifies a considerable number of distal regulatory elements distributed across the gene desert in addition to peaks more closely associated with coding genes. Bottom: dREG peaks in the left DM sample demonstrates enrichment

of asymmetric *Pitx2c* specific peaks and in the right DM shows absence of ASE activity. Dashed vertical lines mark *Pitx2ab* and *Pitx2c* TSS; grey box marks the ASE.

Figure S4. Reproducible L-R differences in chicken and mouse FISH data. Related to Figure 4 and 5. (A) Increased proximity in the left DM between cC4orf32-Pitx2 and Playrr-Pitx2 in chicken, or Playrr-Pitx2 in mouse, is reproducible across biological replicates. Box plots show the lower quartile, median, upper quartile, and whiskers show ± 1.5 times the interquartile range. Contour plotting the density of the underlying distribution of the data are color coded for left (tan) and right (green) DM. (B) Box plots showing Playrr-Pitx2 interprobe distances in the left DM of Pitx2 -/- embryos is indistinguishable from the right DM of WT embryos. (C) TaqMan QRT-PCR of Playrr and Pitx2 isoform expression levels in Pixt2 -/- (n=3) compared to WT (n=3), error bars \pm S.E.M.

Figure S5. Cumulative density curves of chicken and mouse FISH data. Related to Figure 4 and 5. Empirical cumulative distribution frequency (ECDF) plots generated in R show cumulative density (y-axis) of FISH interprobe distance (x-axis) for each probe pair in the left or right DM measured in chicken and mouse.

Figure S6. Conserved and divergent features of the *Pitx2* **locus in chicken and mouse.** Related to Figure 4, 5, and 6 (A) Increased separation of *mC4orf32* from *Playrr* and *Pitx2* in mice (compare Fig. 4 and 5), and TAD structure of the *Pitx2* locus in mESCs (Fig. 6) showing that *Pitx2* and *Enpep* are positioned within separate regulatory domains is accompanied by divergent chicken vs. mouse *C4orf32* and *Enpep* DM expression. Scale bars, 100µm (B) Summary models of conserved and divergent locus topology. (C)

Comparison of conserved sequences from the mouse *Pitx2* locus with zebrafish, chicken, and human, supports that evolutionary divergence is greatest at the proximal end of the gene desert. Note that Zebrafish lack orthologs of *C4ORF32*, e926/*Playrr*, and *ENPEP*.

Figure S7. ChIP-seq analysis of mouse embryonic stem cells (mESCs). Related to Figure 6. RNA Pol II ChIP-seq data demonstrate that exclusive expression of the asymmetric *Pitx2c* isoform in mESCs is accompanied by binding at the ASE enhancer by positive regulators of transcriptional activation such as Med12, p300 and chromatin marks including H3K79 methylation deposited by the DOT1L transcriptional elongation complex via association with RNA Pol II.