

Table S1. *R. sphaeroides* and *E. coli* strains used in this study

Strains	Relevant features	References
<i>R. sphaeroides</i>		
2.4.1	<i>Rhodobacter sphaeroides</i> wild type	(van Niel, 1944)
2.4.1 Δ <i>iscR</i>	Sp ^r , <i>iscR</i> deletion strain	This study
2.4.1 Δ <i>iscR</i> _pBBR <i>iscR</i>	2.4.1 Δ <i>iscR</i> harbouring pBBR <i>iscR</i>	This study
2.4.1-pBE::P _{<i>iscR</i>} ::eCFP	Km ^r , 2.4.1 containing pBE::P _{<i>iscR</i>} ::eCFP, used for <i>in vivo</i> reporter system	This study
2.4.1 Δ <i>iscR</i> -pBE::P _{<i>iscR</i>} ::eCFP	2.4.1 Δ <i>iscR</i> containing pBBR <i>iscR</i> , Km ^r , used for <i>in vivo</i> reporter system	This study
<i>E. coli</i>		
JM109	Host strain for cloning procedures	(Yanisch-Perron et al., 1985)
S17-1	Strain for diparental conjugation, tra ⁺	(Simon et al., 1986)
M15	Strain for recombinant protein expression	Qiagen
M15-pQE30:: <i>iscR</i>	M15 containing pQE30:: <i>iscR</i> , Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _H93A	M15 containing pQE30:: <i>iscR</i> _H93A, Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _H121A	M15 containing pQE30:: <i>iscR</i> _H121A, Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _H127A	M15 containing pQE30:: <i>iscR</i> _H127A, Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _C142A	M15 containing pQE30:: <i>iscR</i> _C142A, Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _P143A	M15 containing pQE30:: <i>iscR</i> _P143A, Ap ^r , used for IscR overexpression	This study
M15-pQE30:: <i>iscR</i> _H93A/H121A/H127A	M15 containing pQE30:: <i>iscR</i> _H93A/H121A/H127A, Ap ^r , used for IscR overexpression	This study

Sp^r, spectinomycin-resistant; Ap^r, ampicillin-resistant; Km^r, kanamycin resistant; when required, antibiotics were used in the following concentrations: spectinomycin (10 μ g·ml⁻¹) and kanamycin (25 μ g·ml⁻¹) for *R. sphaeroides*; ampicillin (200 μ g·ml⁻¹) for *E. coli*

Table S2. Plasmids used in this study

Plasmid names	Relevant features	Source
pBBR1-MCS-2	Km ^r , broad-host-range cloning vector	(Kovach et al., 1995)
pBBR <i>iscR</i>	Km ^r , pBBR1-MCS-2 containing <i>iscR</i> gene	This study
pET28(a)	Km ^r , high expression vector	Novagen
pBE	Km ^r , Derivate of pBBR1-MCS-2	This study
pBE4352	Km ^r , pBE containing RSP_4352 promoter	This study
pBE4352::eCFP:eCFP	Km ^r , pBE4352 containing two eCFP	This study
pBE::P _{<i>iscR</i>} ::eCFP	Km ^r , pBBR containing <i>iscR</i> fragment for <i>cfp</i> fusion	This study
pHP45 Ω	Sp ^r , source of Ω -Sp ^r cassette	(Prentki et al., 1991)
pPHU281	Tc ^r , <i>lacZ'</i> <i>mob</i> (RP4)	(Hubner et al., 1991)
pPHU Δ 2.4. <i>iscR</i>	Tc ^r , pPHU281 containing <i>iscR</i> gene with flanking sites	This study
pPHU Δ 2.4.1 <i>iscR</i> :: Ω Sp	Tc ^r , Sp ^r , pPHU281 Δ 2.4.1 <i>iscR</i> containing spectinomycin cassette	This study
pJET1.2	Ap ^r , 2.97 kb, PCR cloning vector	Fermentas
pQE30	Ap ^r , 3.4 kb, 6xHis-tag overexpression vector	Qiagen
pQE30:: <i>iscR</i>	Ap ^r , pQE30 containing <i>iscR</i> fragment for overexpression	This study
pQE30:: <i>iscR</i> _H93A	Ap ^r , pQE30 containing a mutation of amino acid 93	This study
pQE30:: <i>iscR</i> _H121A	Ap ^r , pQE30 containing a mutation of amino acid 121	This study
pQE30:: <i>iscR</i> _H127A	Ap ^r , pQE30 containing a mutation of amino acid 127	This study
pQE30:: <i>iscR</i> _H93A/H121A/H127A	Ap ^r , pQE30 containing mutations of amino acids 93, 121 and 127	This study
pQE30:: <i>iscR</i> _C142A	Ap ^r , pQE30 containing a mutation of amino acid 142	This study
pQE30:: <i>iscR</i> _P143A	Ap ^r , pQE30 containing a mutation of amino acid 143	This study

Sp^r, spectinomycin-resistant; Ap^r, ampicillin-resistant; Tc^r, tetracycline-resistant; Km^r, kanamycin resistant

Table S3. Oligonucleotides used in this study

Name	Sequence	Purpose
RT_RSP_0040_A	TCGAACGACACCAACACC	Forward primer for RSP_0040 (<i>fliS</i>) real-time RT-PCR
RT_RSP_0040_B	CACCTTCATGCCGTTGAA	Reverse primer for RSP_0040 (<i>fliS</i>) real-time RT-PCR
RT_RSP_0257_A	ACACCTACGGCAACTTCC	Forward primer for RSP_0257 (<i>pufL</i>) real-time RT-pCR
RT_RSP_0257_B	ATCGAGTAGCCGACCAGA	Reverse primer for RSP_0257 (<i>pufL</i>)) real-time RT-PCR
RT_RSP_0288_A	CGATCAGGCGGTGGTGGT	Forward primer for RSP_0288 (<i>bchL</i>) real-time RT-PCR
RT_RSP_0288_B	CGTCGAGGTCCGGCATGT	Reverse primer for RSP_0288 (<i>bchL</i>) real-time RT-PCR
RT_RSP_0443_A	GGTGGAAGAGACGCTCAA	Forward primer for RSP_0443 (<i>iscR</i>) real-time RT-PCR
RT_RSP_0443_B	ATAGACATGCGCCGAGAC	Reverse primer for RSP_0443 (<i>iscR</i>) real-time RT-PCR
RT_RSP_0920_A	GGCAGCTTCGAGGAAGTG	Forward primer for RSP_0920 (<i>exbB</i>) real-time RT-PCR
RT_RSP_0920_B	GTCTGGCTCTGCGAGATG	Reverse primer for RSP_0920 (<i>exbB</i>) real-time RT-PCR
RT_RSP_1545_A	CTTCGAGCGGAGAT	Forward primer for RSP_1545 real-time RT-PCR
RT_RSP_1545_B	GCCGCAGGAACATCG	Reverse primer for RSP_1545 real-time RT-PCR
RT_RSP_1547_A	GTCTGCCACTGCATGGGCAT	Forward primer for RSP_1547 (<i>bfd</i>) real-time RT-PCR
RT_RSP_1547_B	GTTTCATTCCTGCGGTTGTGG	Reverse primer for RSP_1547 (<i>bfd</i>) real-time RT-PCR
RT_RSP_1669_A	ATCGCGGAAGAGACCCAGAG	Forward primer for RSP_1669 (<i>rpoZ</i>) real-time RT-PCR
RT_RSP_1669_B	GAGCAGCCCATCTGATCCT	Reverse primer for RSP_1669 (<i>rpoZ</i>) real-time RT-PCR
RT_RSP_2913_A	CATCAGCCTCGGCAACAC	Forward primer for RSP_2913 (<i>afuA</i>) real-time RT-PCR
RT_RSP_2913_B	TGGTTCGTCTCGGCGTAG	Reverse primer for RSP_2913 (<i>afuA</i>) real-time RT-PCR
RT_RSP_3416_A	CGAGATGATCCGGCT	Forward primer for RSP_3416 real-time RT-PCR
RT_RSP_3416_B	GTCGGTCGCGGAATA	Reverse primer for RSP_3416 real-time RT-PCR
RT_RSP_3417_A	AGACACGCAAGAGCG	Forward primer for RSP_3417 real-time RT-PCR
RT_RSP_3417_B	GCCACCGATTCCACA	Reverse primer for RSP_3417 real-time RT-PCR
RT_RSP_4275_A	ACACCTCCTCAGCT	Forward primer for RSP_4275 (<i>fecl</i>) real-time RT-PCR
RT_RSP_4275_B	AGACGCAGGAAGATG	Reverse primer for RSP_4275 (<i>fecl</i>) real-time RT-PCR
RT_RSP_6006_A	ATGACCGACCCGATGGAG	Forward primer for RSP_6006 (<i>hemP</i>) real-time RT-PCR
RT_RSP_6006_B	AGTAGATCTGCCGCTCGAG	Reverse primer for RSP_6006 (<i>hemP</i>) real-time RT-PCR
RT_RSP_6020_A	CGGCGAGGTCAAGGTGAT	Forward primer for RSP_6020 (<i>feoA2</i>) real-time RT-PCR
RT_RSP_6020_B	TGCAGCGGAGTGACGAAG	Reverse primer for RSP_6020 (<i>feoA2</i>) real-time RT-PCR
<i>suf1</i> RT_A	CGAAGCTCACGGCTTCACCT	Forward primer for RT-PCR
<i>suf1</i> RT_B	AGGCGCCATTGTCTGAGATAG	Reverse primer for RT-PCR
<i>suf2</i> RT_A	CGACCTCGAGGCGCGGATCAAAGG	Forward primer for RT-PCR
<i>suf2</i> RT_B	GCGAAGTCCATCTCGATCTCGGTTTC	Reverse primer for RT-PCR
<i>suf3</i> RT_A	CTCGGAGAGGAAGAGGCCGGCTC	Forward primer for RT-PCR
<i>suf3</i> RT_B	GTGCCGACCGAGACCGAGTCGAAAG	Reverse primer for RT-PCR
<i>suf4</i> RT_A	AGACGCAGGTGATGCAGATG	Forward primer for RT-PCR
<i>suf4</i> RT_B	GAACCGCAGTCTCTTCCAGT	Reverse primer for RT-PCR
<i>suf5</i> RT_A	TCTCGGCGCATGTCTATGTC	Forward primer for RT-PCR
<i>suf5</i> RT_B	CGTCGCATTCCAGTCGAGAT	Reverse primer for RT-PCR
0443_upA	GAGCCGCCGAATTCGGGGTGC	Forward primer for RSP_0443 cloning
0443_upB	GAGCGCGGGATCCACCAGCG	Reverse primer for RSP_0443 cloning
0443_downA	GCGTGCAGGATCCTTGACCAACCG	Forward primer for RSP_0443 cloning
0443_downB	GAGACAGCCTCAAGCTTCACGTCG	Reverse primer for RSP_0443 cloning
<i>IscR</i> _complA	TAATCTAGAACCATTCCACCTGGGCG	Forward primer for RSP_0443 complementation
<i>IscR</i> _complB	CGCTGGATCCTTCACTCTGGC	Reverse primer for RSP_0443 complementation
<i>IscR</i> -His_fwd	GGCGGGAACCGGGATCCAAACTCTC	Forward primer for <i>IscR</i> overexpression
<i>IscR</i> -His_rev	CGACAGCCAAGCTTTCAGTCTCTCGTC	Forward primer for <i>IscR</i> overexpression
<i>IscR</i> _repA	TCTAGAAAATCACTTCGGGCATCGC	Forward primer RSP_0443 for <i>in vivo</i> reporter system
<i>IscR</i> _repB	GGATCCTTTCGGTTCGCGCAAATC	Reverse primer RSP_0443 for <i>in vivo</i> reporter system
<i>IscR</i> _H93A_A	GCGATGGCTACCGGTGCTGGTGCAAGTG	Forward primer inserting a mutation to H93
<i>IscR</i> _H93A_B	GAGACGGTCAATGCGATGGCTACCGGTG	Reverse primer inserting a mutation to H93
<i>IscR</i> _H121A_A	CTCGGCGGCTGTCTATGTCTTCTGAC	Forward primer inserting a mutation to H121
<i>IscR</i> _H121A_B	GTGGGAGGGCCTCTCGGCGGCTGTCTAT	Reverse primer inserting a mutation to H121
<i>IscR</i> _H127A_A	CTTCTGCGCCAGACCCGTCTGTGCGAC	Forward primer inserting a mutation to H127
<i>IscR</i> _H127A_B	GCATGTCTATGTCTTCTGCGCCAGACC	Reverse primer inserting a mutation to H127
<i>IscR</i> _C142A_A	GCGTCCAGCCCCGGCGGTGCCGGCGCTG	Forward primer inserting a mutation to C142
<i>IscR</i> _C142A_B	CAAAAACGAGATGCGTCCAGCCCCGGCG	Reverse primer inserting a mutation to C142
<i>IscR</i> _P143A_A	GCGTCCAGCCCCGGCGGTGCCGGCGCTG	Forward primer inserting a mutation to P143
<i>IscR</i> _P143A_B	AAACGAGATGCGTCCATGCGCGGCGGTG	Reverse primer inserting a mutation to P143
<i>iscR</i> _up_fwd	CGCGGCGTAATCTTGACAAAAACG	Forward primer for generation of the <i>iscR</i> promoter region
<i>iscR</i> _up_rev	CGACACGTCGACAAGCGAGACAAG	Reverse primer for generation of the <i>iscR</i> promoter region
<i>hemP</i> _up_fwd	CGCATAAGTCGACCGAAAGAATCAG	Forward primer for generation of the <i>hemP</i> promoter region
<i>hemP</i> _up_rev	CCGTCGACAAGGATCCGGGCC	Reverse primer for generation of the <i>hemP</i> promoter region

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