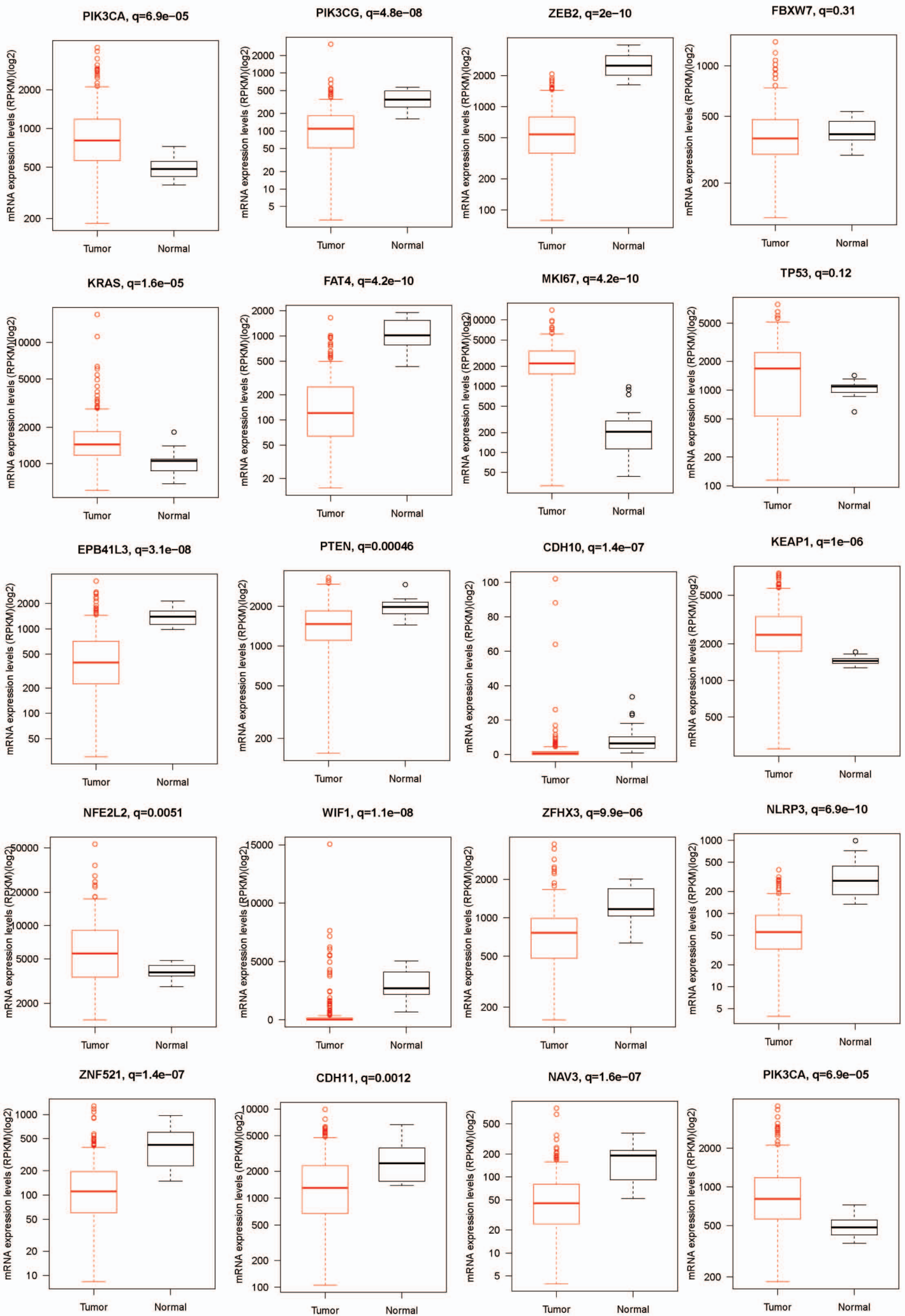


Whole Exome Sequencing Identifies Frequent Somatic Mutations in Cell-Cell Adhesion Genes in Chinese Patients with Lung Squamous Cell Carcinoma

Chenguang Li, Zhibo Gao, Fei Li, Xiangchun Li, Yihua Sun, Mengyun Wang, Dan Li, Rui Wang, Fuming Li, Rong Fang, Yunjian Pan, Xiaoyang Luo, Jing He, Liangtao Zheng, Jufeng Xia, Lixin Qiu, Jun He, Ting Ye, Ruoxin Zhang, Minghui He, Meiling Zhu, Haichuan Hu, Tingyan Shi, Xiaoyan Zhou, Menghong Sun, Shilin Tian, Yong Zhou, Qiaoxiu Wang, Longyun Chen, Guangliang Yin, Jingya Lu, Renhua Wu, Guangwu Guo, Yingrui Li, Xueda Hu, Lin Li, Asan, Qin Wang, Ye Yin, Qiang Feng, Bin Wang, Hang Wang, Mingbang Wang, Xiaonan Yang, Xiuqing Zhang, Huanming Yang, Li Jin, Cun-Yu Wang, Hongbin Ji, Haiquan Chen, Jun Wang, Qingyi Wei

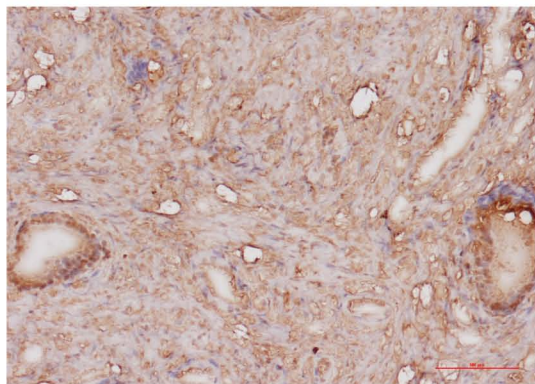


Supplementary Figure 2

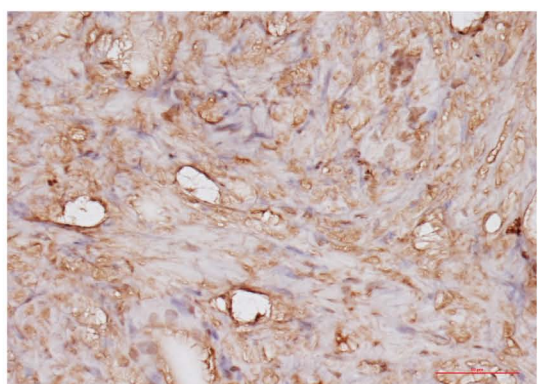
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.....A.....	FCD1C84ACXX:1:1313:13398:11903
.....A.....G.....G.....	FCD1C84ACXX:1:1211:6882:28472
.....G.....	FCC125BACXX:6:2302:7176:39104
.....	FCD1C84ACXX:1:1205:15619:50510
.....	FCD1C84ACXX:1:2103:15052:74944
.....G.....	FCC125BACXX:6:1208:4884:163989
.....	FCC125BACXX:6:2204:20401:46218
.....	FCD1C84ACXX:1:1212:17383:89626
.....C.....G.....	FCD1C84ACXX:1:2112:20414:32478
.....	FCC125BACXX:6:1106:9680:164443
.....A.....	FCC125BACXX:6:2304:14375:166096
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.....A.....	FCC125BACXX:6:1306:16346:92240
Mutation1: G>A	Mutation2: A>G	

Supplementary Figure 4

a

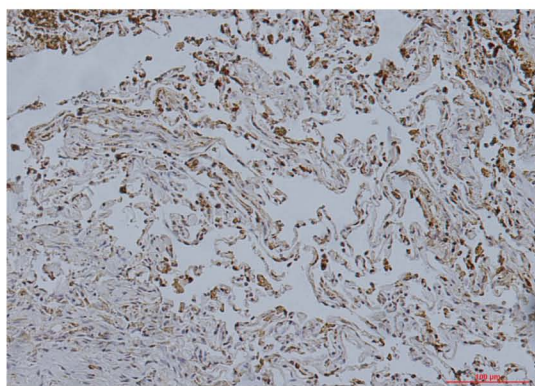


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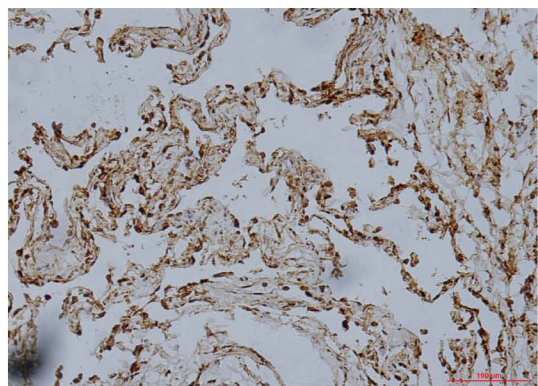


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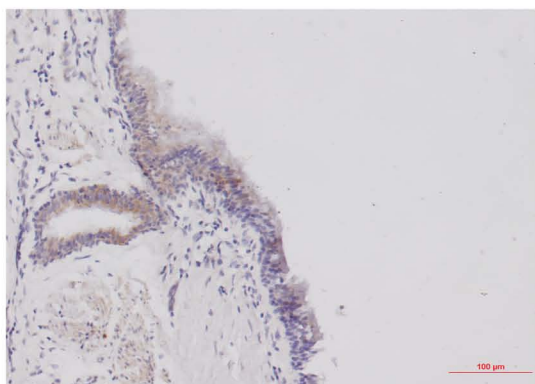


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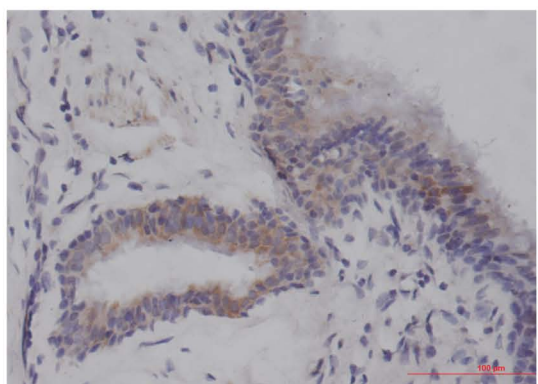


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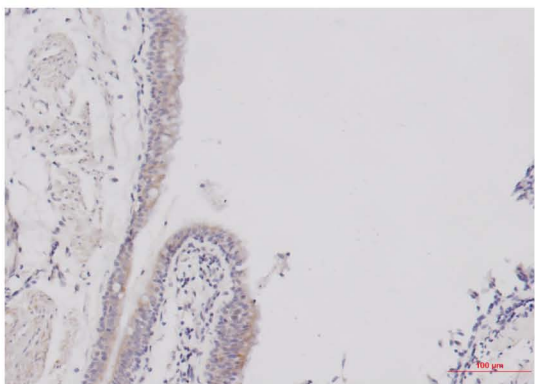
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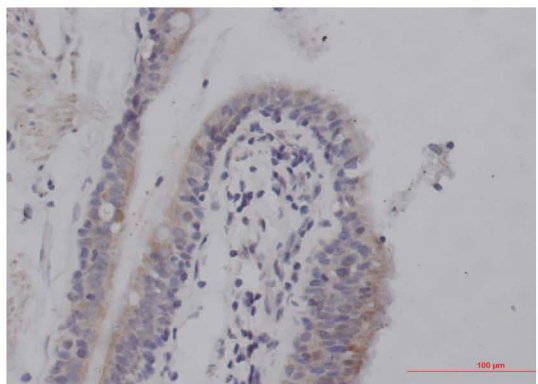
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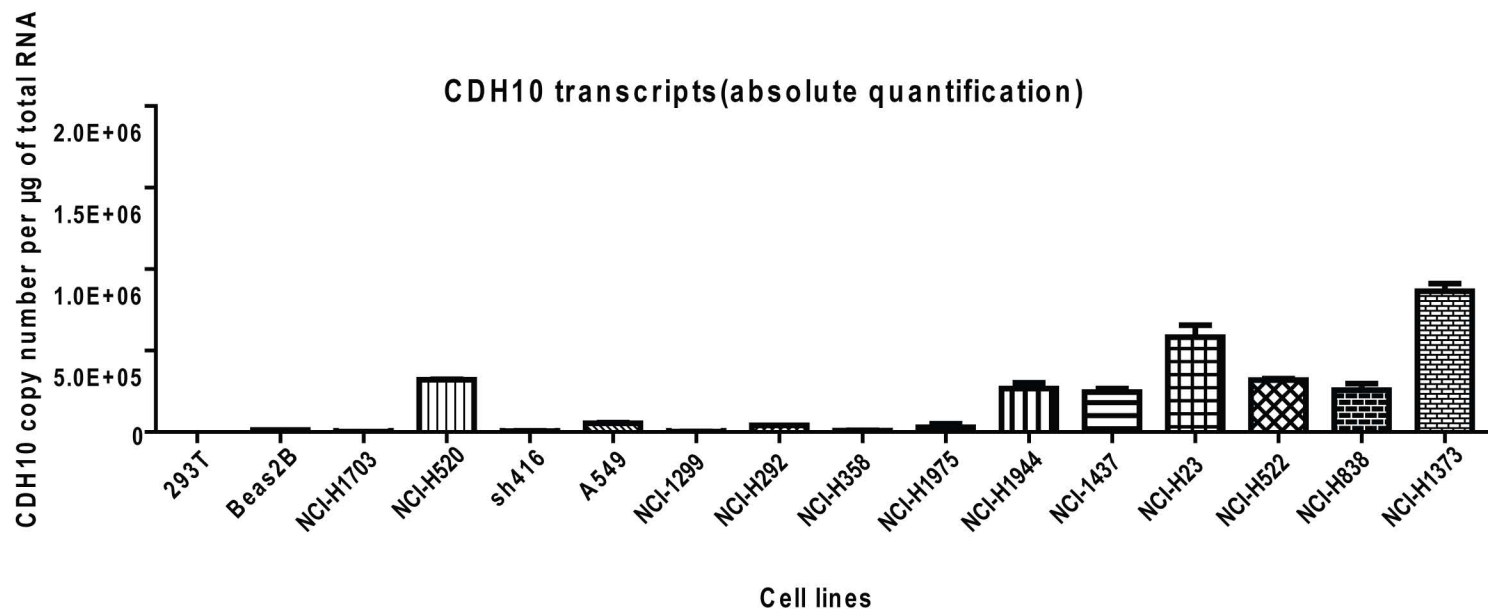


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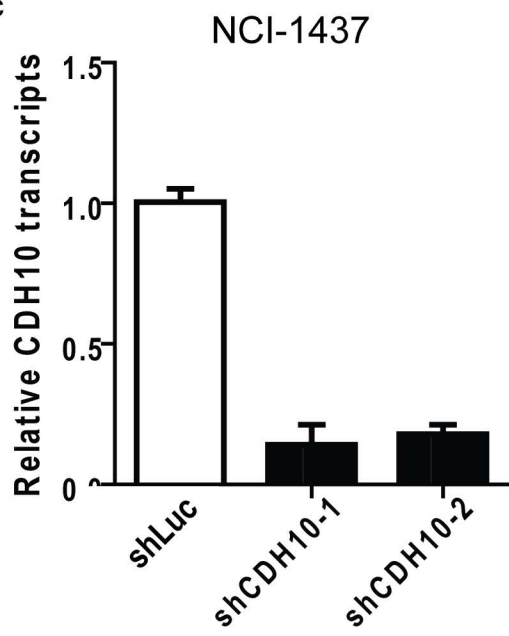
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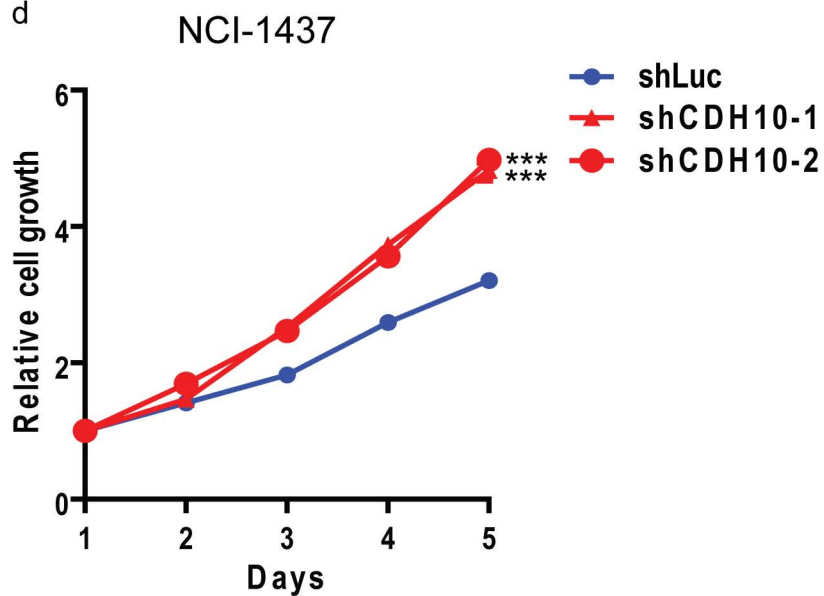
b

NCI-H522						NCI-H1373					
Scramble		shCDH10-1		shCDH10-2		Scramble		shCDH10-1		shCDH10-2	
GAPDH	CDH10	GAPDH	CDH10	GAPDH	CDH10	GAPDH	CDH10	GAPDH	CDH10	GAPDH	CDH10
16.596	24.061	15.899	25.801	16.067	25.506	16.188	23.371	15.76	25.189	15.937	24.888
16.458	23.977	15.854	25.784	16.091	25.542	16.264	23.412	16.044	25.289	15.583	24.714
	24.206		25.697		25.69		23.229		25.372		24.907

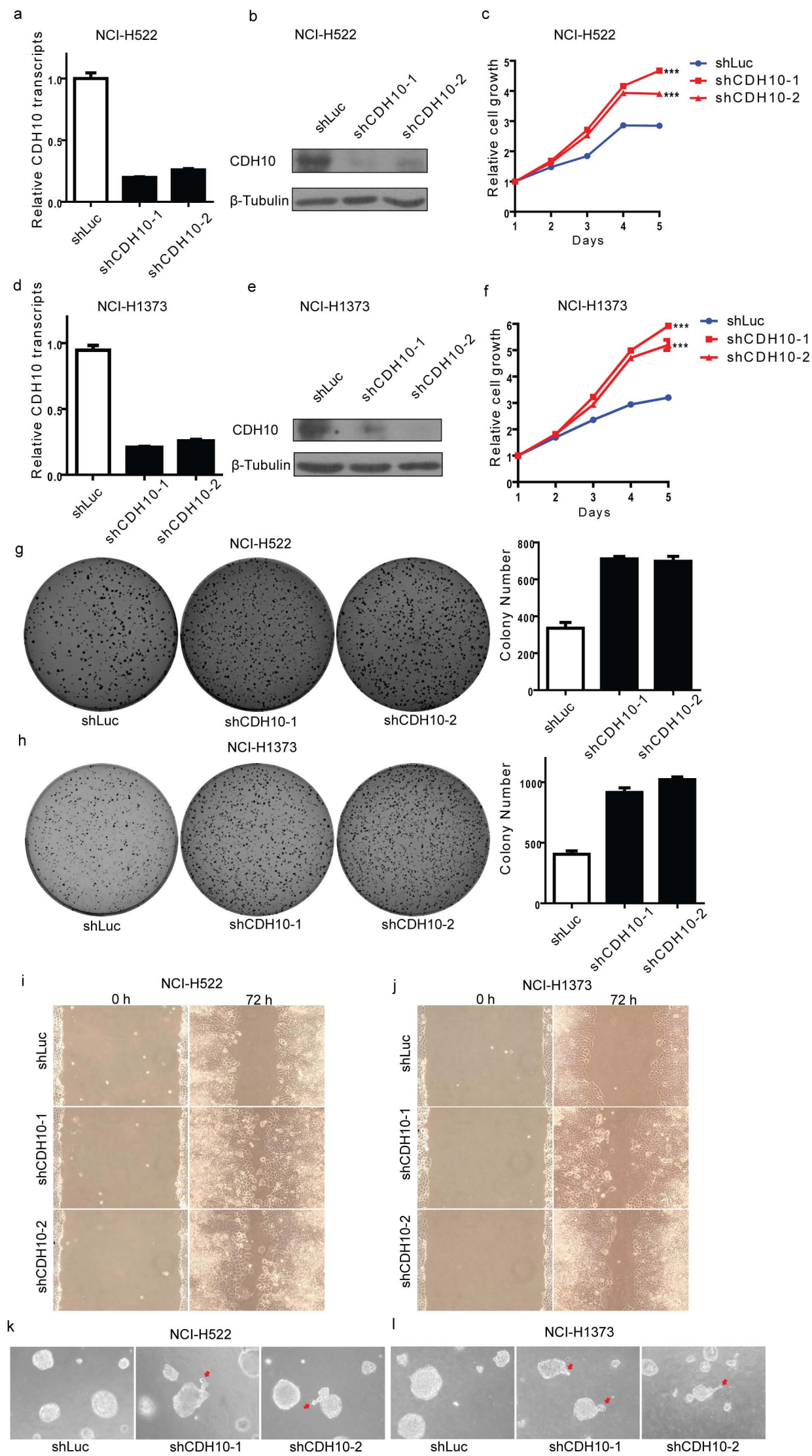
c



d

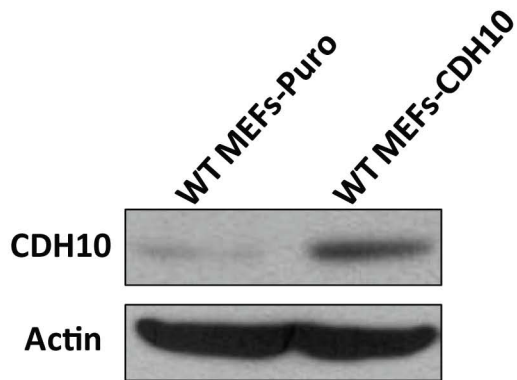


Supplementary Figure 6

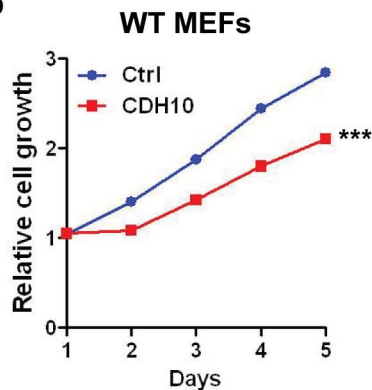


Supplementary Figure 7

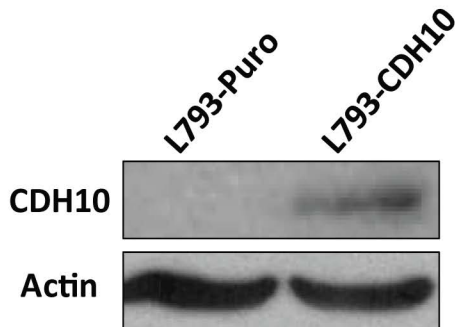
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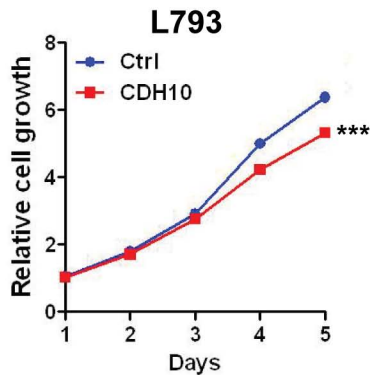
b



c

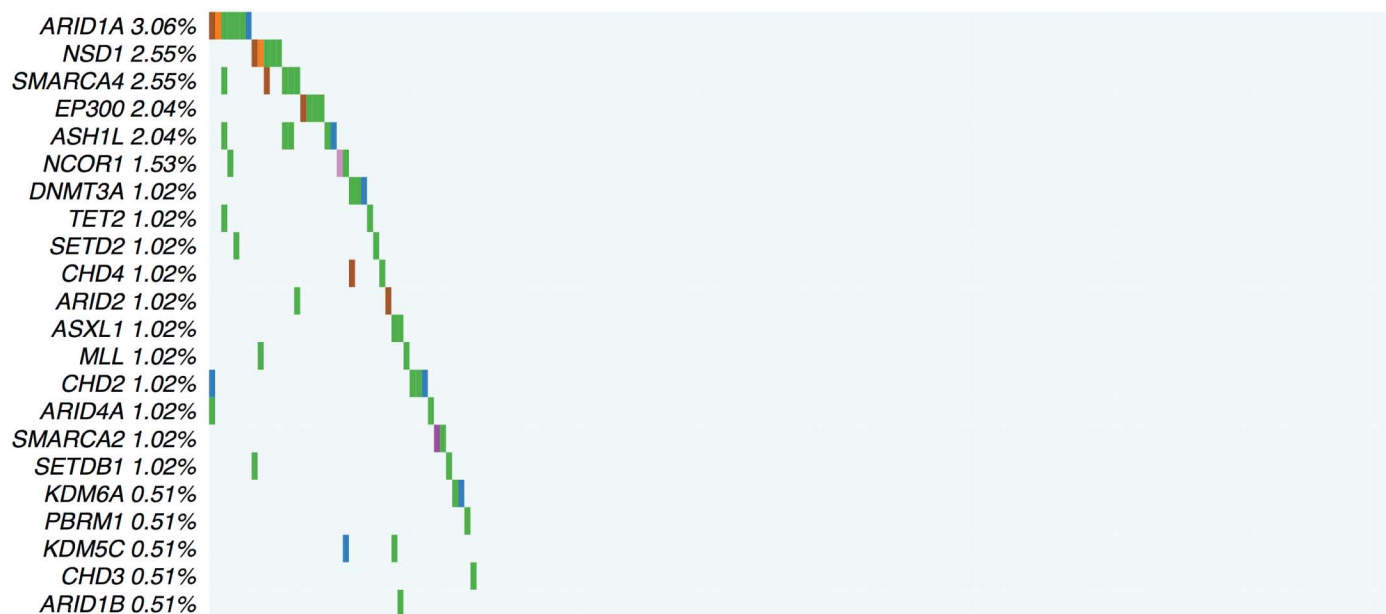


d



Supplementary Figure 8

a Likely driver chromatin regulatory factor genes, overall mutation frequency 27.3%



b SWI/SNF, overall mutation frequency 13.6%

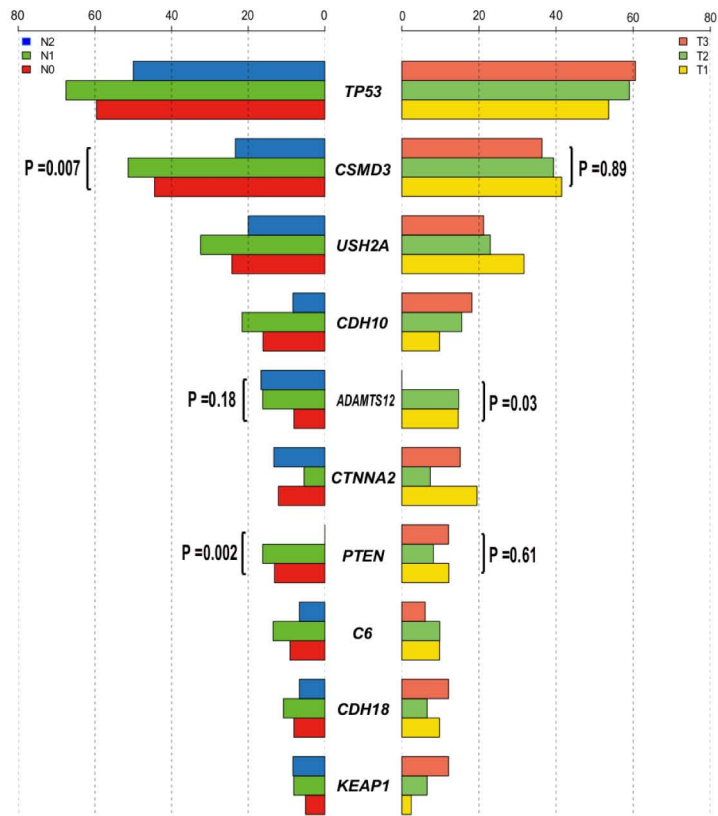
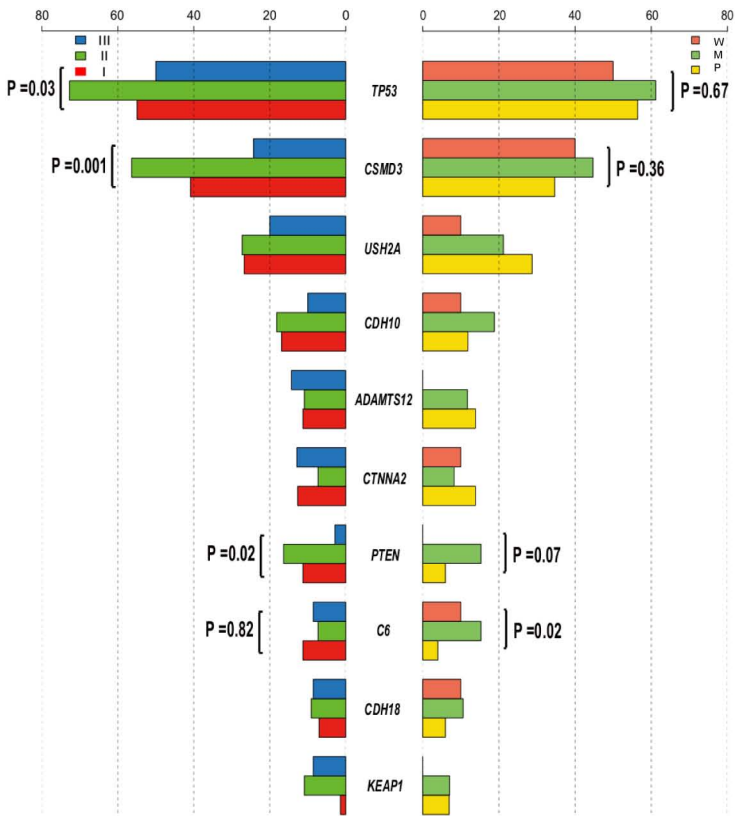


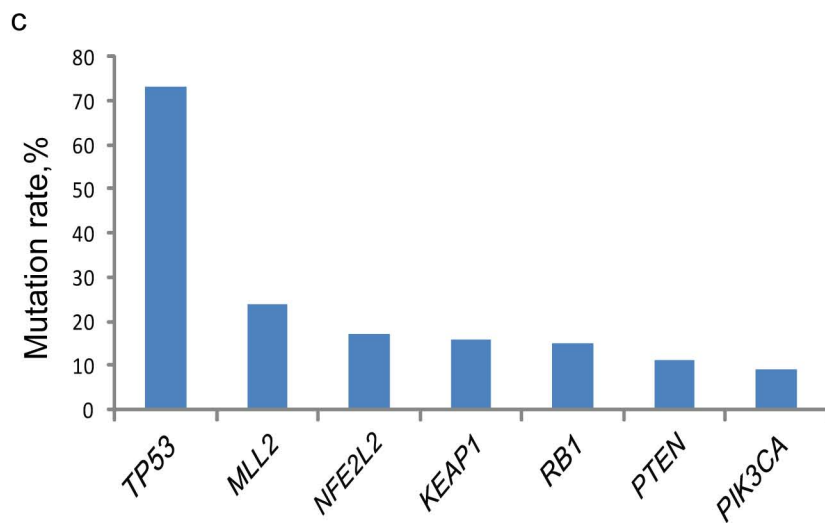
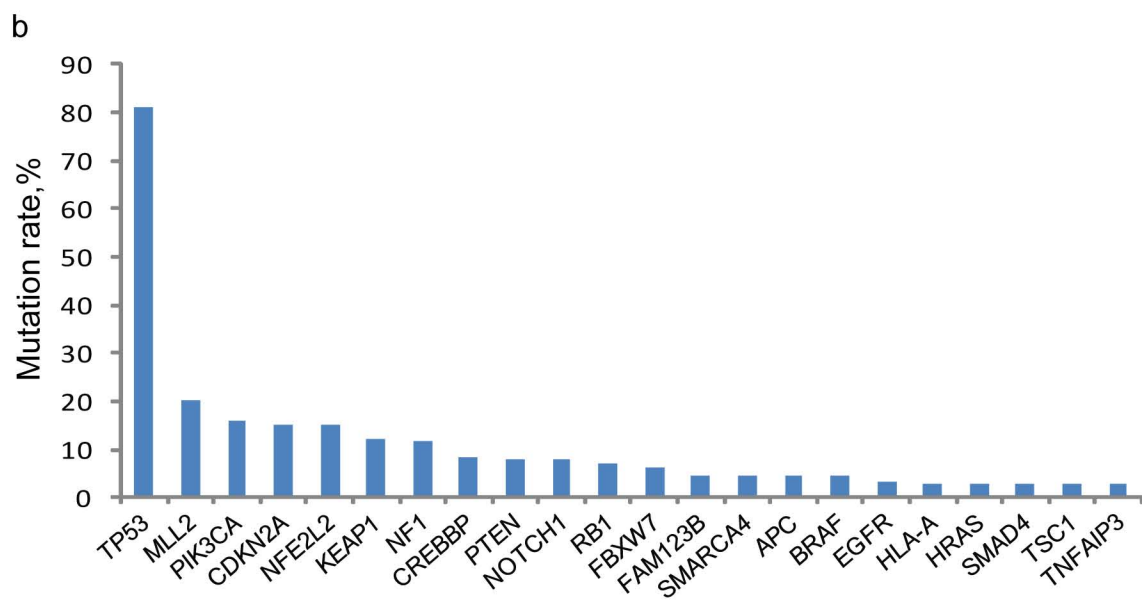
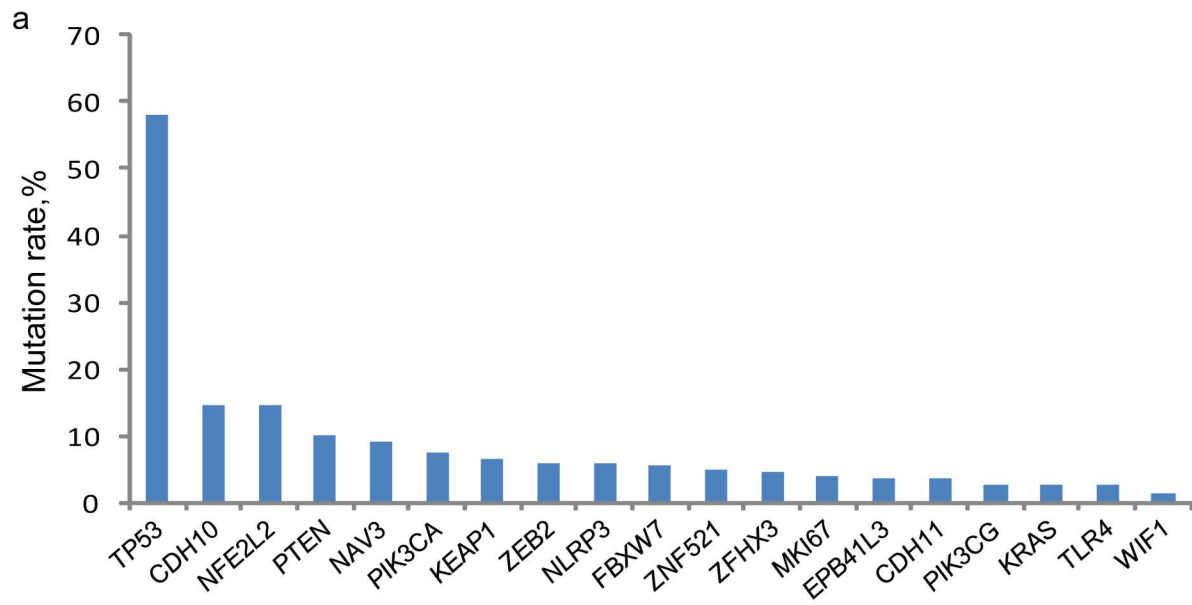
c PRC, overall mutation frequency 7.1%



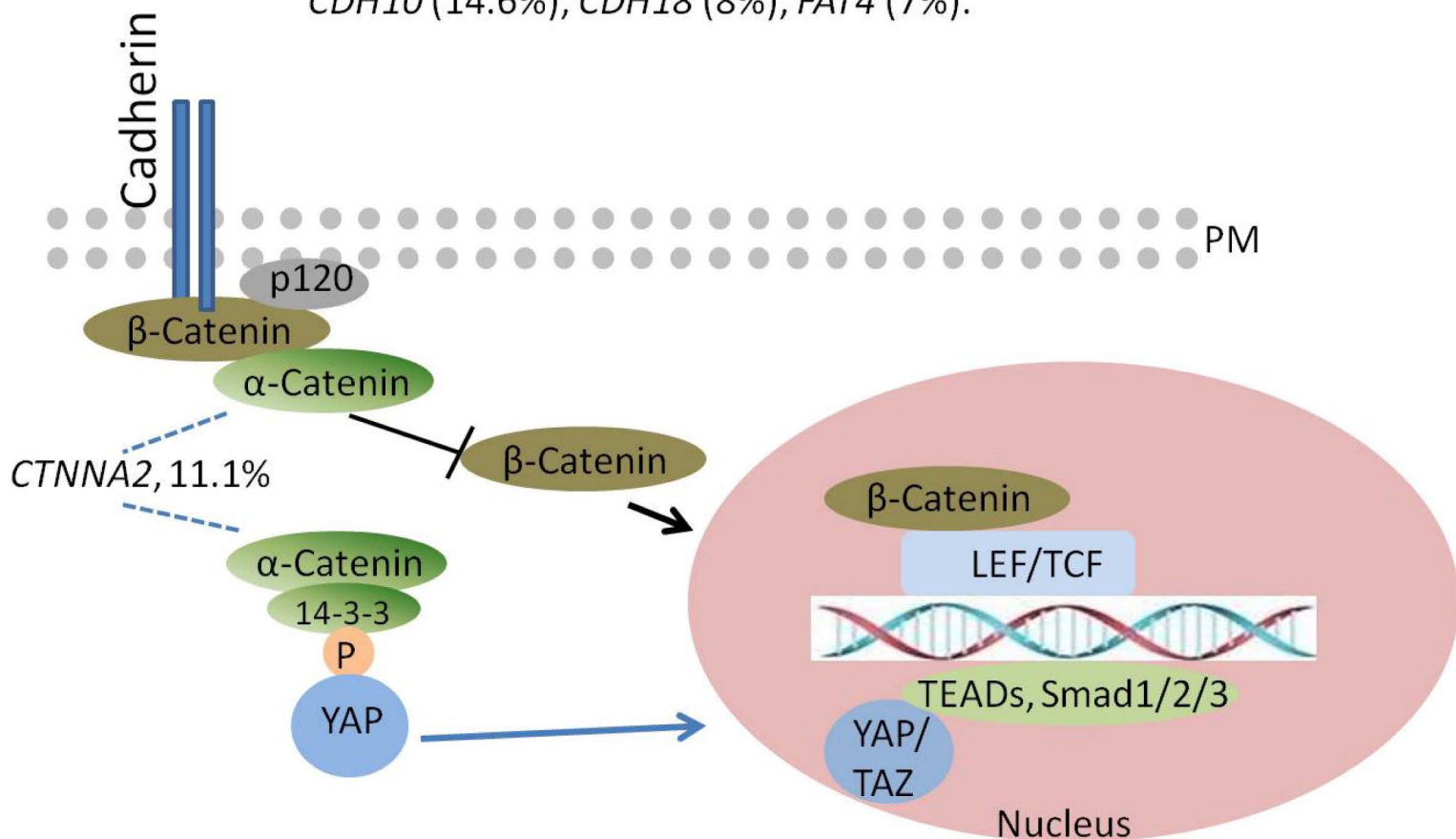
■ Nonsense ■ Inframe shift
■ Splice site ■ Missense
■ Frame shift ■ Syn.

Supplementary Figure 9





69.2% of lung SQCC samples harbor non-silent mutations of Cadherin superfamily genes, including *CDH10* (14.6%), *CDH18* (8%), *FAT4* (7%).



Supplementary Figure Legends

Supplementary Figure 1. mRNA expression level of 20 SMGs in TCGA LUSC cohort.

The tumor-normal lung mRNA expression level of 20 SMGs identified in our study were queried from the TCGA LUSC cohort.

Supplementary Figure 2. Bi-allelic inactivation analysis of CDH10 in sample 1019LC.

Bi-allelic inactivating mutations in CDH10 (extracted by samtools tview), 2 SNVs were encompassed green and violet rectangles, black dot and comma represent nucleotide from forward and reverse strands, respectively.

Supplementary Figure 3. CDH10 expression level in human tissues. (a) The absolute transcript values of CDH10 in 1 µg RNA of normal human lung, brain and prostate samples were assessed by quantitative PCR. The CDH10 plasmid (PCDH-puro-CDH10) was used to draw the standard curve for CDH10 absolute transcript measurement. (b) The CDH10 expression was assessed by Western-blot in protein lysate from normal human lung and brain tissues. To make comparison between lung and brain tissues, 80 and 10 µg of normal lung and brain tissue lysates were loaded, respectively. Average level of brain samples were normalized as 1.

Supplementary Figure 4. CDH10 immunohistochemistry staining of prostate (a), pulmonary alveoli (b) and airway epithelial (c), the scale bar was 100µm.

Supplementary Figure 5. CDH10 expression level in cell lines, knockdown efficiency of shCDH10 in cell line NCI-H522 and NCI-H1373 and MTT assay result of NCI-1437. (a)

The absolute transcript values of CDH10 in 1 μ g RNA of each cell line were assessed by quantitative PCR. The CDH10 plasmid (PCDH-puro-CDH10) was used to draw the standard curve for CDH10 absolute transcript measurement. (b) Raw data of quantitative PCR analysis of CDH10 expression in cell lines with CDH10 knockdown. Two cell lines with highest CDH10 expression were chosen for shCDH10 expression. Δ Ct were shown. Each experiment was repeated at least three times. (c) CDH10 knockdown efficiency was assessed in NCI-1437 cells through the real time PCR. (d) The MTT assay was performed in NCI-1437 cells with or without CDH10 knockdown, and CDH10 knockdown promoted cell proliferation. *** P<0.001.

Supplementary Figure 6. CDH10 knockdown significantly promotes lung cancer cell proliferation, anchorage-independent cell growth, cell migration and invasion. (a)

CDH10 knockdown efficiency was assessed in NCI-H522 cells through the real time PCR. (b) Knockdown of CDH10 in NCI-H522 detected by Western-blot. (c) The MTT assay was performed in NCI-H522 cells with or without CDH10 knockdown, and CDH10 knockdown promoted cell proliferation. *** P<0.001. (d) CDH10 knockdown efficiency was assessed in NCI-H1373 cells through the real time PCR. (e) Knockdown of CDH10 in NCI-H1373 cells detected by Western-blot. (f) The MTT assay was performed in NCI-H1373 cells with or without CDH10 knockdown, and CDH10 knockdown promoted cell proliferation. *** P<0.001. (g-h) The soft agar assay was performed in NCI-H522 (g) or NCI-H1373 (h) cells with or without CDH10 knockdown, and CDH10 knockdown promoted anchorage-independent cell

growth in both cell lines. (i-j) The wound healing assay was performed in NCI-H522 (i) and NCI-H1373 (j) cell lines, and CDH10 knockdown promoted cell migration in both cell lines. The magnification is 200×. (k-l) Cell invasion assay in matrigel was performed in NCI-H522 (k) or NCI-H1373 (l) cells, and CDH10 knockdown promoted cell invasion in both cell lines. Red arrows indicate the cell protrusion. Each experiment was repeated at least three times. The original magnification is 200X.

Supplementary Figure 7. CDH10 overexpression significantly inhibits cell proliferation.

(a). Overexpression of CDH10 in WT MEFs detected by western-blot. (b). Overexpression of CDH10 in WT MEFs significantly inhibited cell proliferation. (c). Overexpression of CDH10 in L793 cell line detected by western-blot. (d). Overexpression of CDH10 in L793 cell line significantly inhibited cell proliferation. Each experiment was repeated at least three times. (WT, wild-type; MEFs, mouse embryonic fibroblasts; ***, $P < 0.001$)

Supplementary Figure 8. Mutations from Chromatin regulatory factor genes identified

in lung SQCCs. Mutation and mutually exclusive analysis of likely driver Chromatin regulatory factor genes (a), SWI/SNF (b) and PRC protein complex genes(c).

Supplementary Figure 9. Clinical association of frequently mutated genes in lung

SQCCs. Clinical-pathological association of the highlighted genes with mutations observed in lung SQCCs, with frequencies of mutations in the highlighted genes by the TNM stage (tumor, lymph node metastasis and distant metastasis), differentiation (W, well differentiated; M,

moderately differentiated; P, poorly differentiated), N stage (lymph node metastasis alone) and T stage (tumor stage alone). For each stage or differentiation, the frequency of mutations in a given gene was calculated as the proportion of tumors harboring non-silent mutations in the gene among all tumors of the indicated stage or differentiation. The significance of the correlations between mutations and tumor stage or differentiation was determined by Fisher's exact test, two-sided.

Supplementary Figure 10. Comparison of significantly mutated genes identified in three Lung SQCC studies (Adjusted to same analysis tools). (a) Significantly mutated genes in the present study; (b) The TCGA Lung SQCC study; (c) The Korean Lung SQCC study.

Supplementary Figure 11. Overview of the cadherin-catenin and YAP signaling and its regulation. α -catenin binds to β -catenin and inhibits the signaling through the canonical Wnt pathway. YAP acts downstream of α -catenin to control transcription. In this study, 69.2% of lung SQCCs were found to harbor non-silent mutations in cadherin superfamily genes. CTNNA2 was found to be mutated in 11.1% of lung SQCCs.

Supplementary Table 1. Clinical information of 100 Lung SQCC patients

Sample ID	Age (years)	Gender	Smoking status	Differentiation	TNM*	Stage*
7	58	Male	Smoker	M	T3N2M0	IIIa
11	61	Male	Smoker	M	T2aN0M0	I b
21	65	Male	Smoker	M	T2aN0M0	I b
25	50	Male	Smoker	M	T2aN0M0	I b
27	49	Male	Smoker	P	T2aN2M0	IIIa
30	59	Male	Smoker	P	T2aN0M0	I b
36	59	Male	Smoker	P	T2aN2M0	IIIa
39	48	Male	Smoker	M	T2aN2M0	IIIa
67	68	Male	Smoker	P	T2aN0M0	I b
94	57	Male	Smoker	M	T2aN0M0	I b
96	69	Male	Smoker	M	T1aN0M0	I a
124	61	Male	Smoker	P	T3N2M0	IIIa
127	53	Male	Smoker	M	T2aN1M0	II a
130	72	Male	Smoker	M	T2aN0M0	I b
133	64	Male	Smoker	P	T2bN0M0	II a
134	71	Male	Smoker	P	T3N2M0	IIIa
137	63	Male	Smoker	M	T1bN0M0	I a
144	54	Male	Smoker	P	T2bN2M0	IIIa
145	57	Male	Non-smoker	M	T1bN0M0	I a
159	59	Male	Smoker	M	T3N2M0	IIIa
160	48	Male	Smoker	W	T3N1M0	IIIa
170	61	Male	Smoker	P	T2aN3M0	IIIb
171	79	Male	Smoker	P	T1aN0M0	I a
185	61	Male	Smoker	P	T1bN0M0	I a
188	50	Female	Non-smoker	P	T2bN2M0	IIIa
192	48	Male	Smoker	P	T2bN0M0	II a
196	57	Male	Smoker	M	T2aN2M0	IIIa
207	61	Male	Smoker	M	T2bN0M0	II a
209	62	Male	Smoker	P	T2aN0M0	I b
214	78	Male	Smoker	M	T1bN0M0	I a
225	63	Male	Smoker	P	T1aN0M0	I a
234	55	Male	Smoker	M	T1aN0M0	I a
257	60	Male	Smoker	M	T3N0M0	II b
281	65	Male	Smoker	M	T1bN2M0	IIIa
292	54	Male	Smoker	M	T2aN0M0	I b
298	55	Female	Non-smoker	M	T2aN2M0	IIIa
299	50	Male	Smoker	M	T2aN1M0	II a
301	60	Male	Smoker	P	T2bN2M0	IIIa
303	60	Male	Smoker	P	T2aN2M0	IIIa
310	51	Male	Smoker	P	T4N0M0	IIIa
313	57	Male	Smoker	P	T2aN1M0	II a
315	52	Male	Smoker	M	T2aN1M0	II a
321	62	Male	Smoker	M	T3N1M0	IIIa
341	55	Male	Smoker	P	T2bN2M0	IIIa
343	65	Female	Non-smoker	P	T2aN2M0	IIIa
348	64	Male	Smoker	M	T3N0M0	II b
361	55	Male	Smoker	M	T2aN0M0	I b
367	54	Male	Smoker	P	T1bN1M0	II a
371	58	Male	Smoker	P	T2aN1M0	II a

375	70	Male	Smoker	W	T2aN0M0	I b
378	60	Male	Smoker	P	T3N1M0	IIIa
387	48	Female	Non-smoker	P	T2bN2M0	IIIa
394	54	Male	Smoker	P	T1aN0M0	I a
400	62	Male	Smoker	P	T1aN0M0	I a
408	45	Male	Smoker	P	T2aN1M0	II a
416	56	Female	Non-smoker	P	T2aN2M0	IIIa
417	68	Male	Smoker	P	T1bN0M0	I a
428	57	Male	Smoker	M	T2aN0M0	I b
431	60	Male	Smoker	M	T2aN0M0	I b
433	51	Male	Smoker	M	T2aN0M0	I b
438	70	Male	Smoker	P	T2bN2M0	IIIa
451	63	Male	Smoker	M	T1bN0M0	I a
467	68	Male	Non-smoker	P	T3N0M0	II b
474	67	Male	Smoker	P	T2aN2M0	IIIa
482	61	Male	Smoker	P	T1bN0M0	I a
503	57	Male	Smoker	M	T2aN0M0	I b
504	69	Male	Smoker	M	T2aN0M0	I b
520	63	Male	Smoker	P	T1aN2M0	IIIa
523	51	Male	Smoker	P	T2bN2M0	IIIa
527	42	Female	Non-smoker	P	T2aN2M0	IIIa
534	75	Male	Non-smoker	M	T2bN2M0	IIIa
537	70	Male	Smoker	W	T3N1M0	IIIa
538	73	Male	Smoker	W	T2bN2M0	IIIa
545	59	Male	Smoker	P	T2aN2M0	IIIa
553	59	Male	Smoker	P	T2aN0M0	I b
554	39	Male	Smoker	M	T2aN0M0	I b
601	49	Male	Smoker	P	T2bN2M0	IIIa
611	63	Male	Non-smoker	M	T3N1M0	IIIa
623	50	Male	Smoker	P	T2aN0M0	I b
699	64	Male	Smoker	P	T2aN0M0	I b
700	62	Male	Smoker	M	T1aN1M0	II a
715	58	Male	Smoker	P	T2bN0M0	II a
755	62	Male	Smoker	P	T2aN0M0	I b
757	55	Male	Smoker	P	T2aN1M0	II a
804	59	Male	Smoker	P	T2aN0M0	I b
814	67	Male	Smoker	M	T2aN2M0	IIIa
821	47	Male	Smoker	M	T1bN2M0	IIIa
830	54	Male	Smoker	M	T2aN2M0	IIIa
856	72	Male	Non-smoker	P	T1aN0M0	I a
863	52	Male	Non-smoker	M	T1aN0M0	I a
869	61	Female	Smoker	P	T3N2M0	IIIa
873	63	Male	Smoker	M	T2bN2M0	IIIa
882	69	Male	Smoker	P	T1bN0M0	I a
886	51	Female	Non-smoker	P	T2aN2M0	IIIa
888	69	Male	Smoker	P	T2bN0M0	II a
896	69	Male	Smoker	P	T1aN0M0	I a
901	61	Male	Smoker	M	T3N0M0	II b
909	64	Female	Non-smoker	P	T2aN2M0	IIIa
916	55	Male	Smoker	M	T2aN2M0	IIIa
935	56	Male	Smoker	M	T2bN2M0	IIIa

*The TNM stage and stage grouping are according to UICC 2009 NSCLC staging system. W, well differentiated; M, moderately differentiated; P, poorly differentiated.

Supplementary Table 2. Clinical information of additional Lung SQCCs for TCS validation

Sample ID	Age (years)	Gender	Smoking status	Differentiation	TNM*	Stage*
1019	83	Female	Non-smoker	P	T2aN0M0	I b
1021	55	Male	Smoker	M	T2aN0M0	I b
1038	74	Male	Smoker	P	T2aN1M0	II a
1043	48	Male	Smoker	P	T1bN0M0	I a
1080	59	Female	Non-smoker	P	T3N0M0	II b
1088	67	Male	Smoker	P	T2aN0M0	I b
1096	52	Male	Smoker	M	T3N0M0	II b
1108	76	Male	Smoker	M	T2bN0M0	II a
1115	67	Male	Smoker	M	T2aN1M0	II a
1125	60	Male	Smoker	M	T2aN2M0	IIIa
1149	56	Male	Smoker	W	T2aN0M0	I b
1167	48	Male	Smoker	M	T2aN1M0	II a
1175	64	Male	Smoker	P	T3N0M0	II b
1185	58	Male	Smoker	P	T2aN2M0	IIIa
1194	58	Male	Smoker	P	T2aN0M0	I b
1217	70	Male	Smoker	M	T1aN0M0	I a
1223	58	Male	Smoker	P	T1bN2M0	IIIa
1233	56	Male	Smoker	M	T1bN2M0	IIIa
1239	67	Male	Smoker	W	T3N2M0	IIIa
1241	59	Male	Smoker	W	T4N0M0	IIIa
1246	75	Female	Smoker	M	T2bN2M0	IIIa
1250	59	Male	Smoker	P	T2aN1M0	II a
1251	60	Male	Non-smoker	M	T2bN1M0	II b
1252	57	Male	Smoker	P	T2aN0M0	I b
1257	54	Male	Smoker	P	T2aN1M0	II a
1264	71	Male	Non-smoker	M	T2aN1M0	II a
1270	56	Male	Smoker	P	T3N2M0	IIIa
1276	72	Male	Smoker	M	T2bN0M0	II a
1285	79	Male	Smoker	W	T1aN0M0	I a
1292	71	Male	Smoker	P	T1bN0M0	I a
1294	58	Male	Smoker	M	T2bN0M0	II a
1296	70	Male	Smoker	M	T2bN0M0	II a
1297	51	Male	Smoker	M	T2aN1M0	II a
1300	62	Male	Smoker	P	T2aN0M0	I b
1314	63	Male	Smoker	P	T3N0M0	II b
1322	62	Male	Smoker	P	T2bN1M0	II b
1323	63	Male	Smoker	P	T2aN0M0	I b
1325	51	Male	Smoker	M	T2aN1M0	II a
1338	78	Male	Smoker	M	T3N0M0	II b
1344	69	Male	Non-smoker	P	T3N2M0	IIIa
1347	57	Male	Smoker	P	T2aN2M0	IIIa
1348	60	Male	Smoker	M	T1aN1M0	II a
1351	65	Male	Smoker	P	T2aN2M0	IIIa
1355	60	Male	Smoker	P	T3N2M0	IIIa
1361	64	Male	Smoker	P	T3N0M0	II b
1368	59	Male	Smoker	P	T2aN2M0	IIIa
1369	63	Male	Smoker	P	T2bN0M0	II a
1370	68	Male	Smoker	W	T2bN0M0	II a
1378	67	Male	Smoker	M	T2aN0M0	I b

1382	60	Male	Smoker	W	T1bN0M0	I a
1387	62	Male	Smoker	M	T3N1M0	IIIa
1389	70	Male	Smoker	M	T2aN1M0	II a
1395	57	Male	Smoker	M	T2aN1M0	II a
1396	79	Male	Smoker	P	T2aN0M0	I b
1405	63	Female	Non-smoker	P	T3N1M0	IIIa
1433	66	Male	Smoker	M	T2aN0M0	I b
1436	51	Male	Smoker	P	T1aN0M0	I a
1447	54	Male	Smoker	M	T2aN0M0	I b
1461	64	Female	Non-smoker	M	T2aN2M0	IIIa
1477	45	Female	Non-smoker	P	T3N2M0	IIIa
1485	56	Male	Smoker	P	T2aN2M0	IIIa
1488	49	Female	Non-smoker	P	T1bN2M0	IIIa
1489	49	Male	Smoker	P	T3N2M0	IIIa
1507	67	Male	Smoker	M	T3N0M0	II b
1530	76	Male	Smoker	M	T2aN0M0	I b
1535	70	Male	Smoker	P	T1bN0M0	I a
1540	87	Male	Smoker	P	T1bN0M0	I a
1556	45	Male	Smoker	P	T2aN2M0	IIIa
1557	58	Male	Smoker	P	T2aN1M0	II a
1567	71	Male	Smoker	M	T1bN0M0	I a
1570	54	Male	Smoker	M	T1bN1M0	II a
1587	56	Male	Smoker	M	T2aN2M0	IIIa
1589	54	Male	Smoker	P	T2bN1M0	II b
1598	53	Male	Smoker	M	T2bN0M0	II a
1630	53	Male	Smoker	M	T2aN2M0	IIIa
1660	44	Male	Smoker	M	T1bN0M0	I a
1673	56	Male	Smoker	P	T1bN1M0	II a
1678	62	Male	Smoker	P	T2aN0M0	I b
1679	60	Male	Smoker	P	T2aN1M0	II a
1681	59	Male	Smoker	P	T2aN1M0	II a
1689	60	Male	Smoker	P	T1bN0M0	I a
1691	72	Male	Smoker	M	T2bN0M0	II a
1723	57	Male	Smoker	M	T2bN2M0	IIIa
1730	56	Male	Smoker	P	T2aN0M0	I b
1740	56	Male	Smoker	M	T2bN1M0	II b
1745	60	Male	Smoker	P	T2aN0M0	I b
1754	43	Male	Smoker	M	T1aN0M0	I a
1762	46	Male	Smoker	M	T1bN0M0	I a
1763	62	Male	Smoker	M	T3N0M0	II b
1780	62	Male	Smoker	M	T2aN0M0	I b
1785	74	Male	Smoker	M	T2bN0M0	II a
1796	71	Male	Smoker	P	T2aN0M0	I b
1797	74	Male	Smoker	M	T2bN2M0	IIIa
1813	51	Male	Smoker	P	T2aN2M0	IIIa
1837	55	Female	Non-smoker	P	T2aN0M0	I b
1848	67	Male	Smoker	P	T3N0M0	II b
1855	62	Male	Smoker	M	T3N1M0	IIIa
1862	63	Male	Smoker	P	T1aN0M0	I a

TCS, Target-Capture Sequencing. *The TNM stage and stage grouping are according to UICC 2009 NSCLC staging system. W, well differentiated; M, moderately differentiated; P, poorly