

# ADVANCED FUNCTIONAL MATERIALS

## Supporting Information

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**A Highly Elastic and Rapidly Crosslinkable Elastin-Like  
Polypeptide-Based Hydrogel for Biomedical Applications**

*Yi-Nan Zhang, Reginald K. Avery, Queralt Vallmajo-Martin,  
Alexander Assmann, Andrea Vegh, Adnan Memic, Bradley D.  
Olsen, Nasim Annabi,\* and Ali Khademhosseini\**

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The complete protein sequence is as follows:

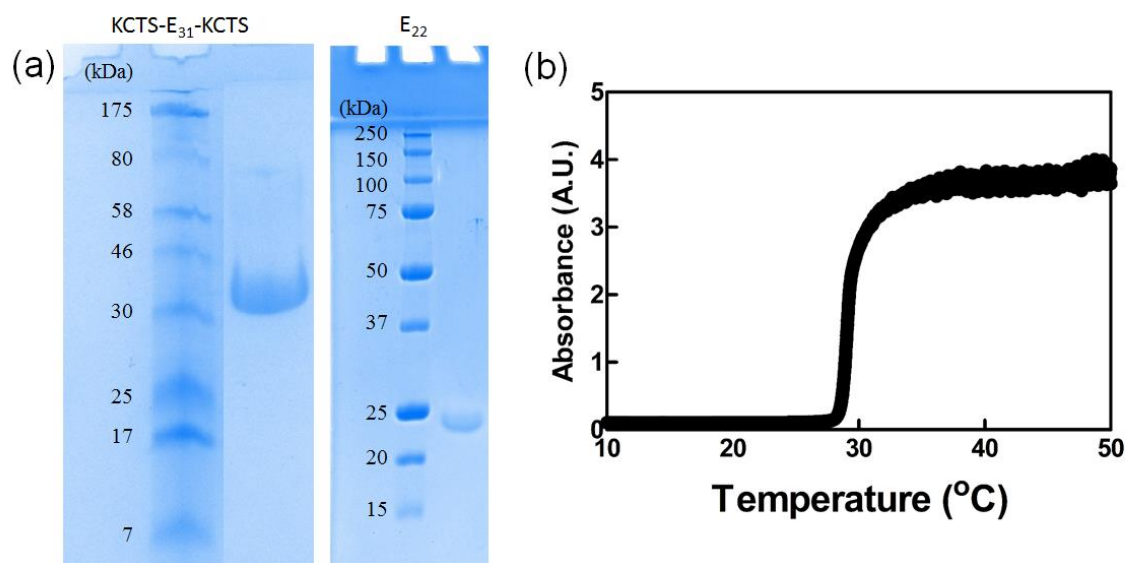
MGWGSKCTSASGLVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGV  
VPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGV  
VPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGV  
VPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGV  
VPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVPGVGV  
PGVGVPGVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVIPGVGV  
GVGVPGVGVPGVGVPGVGVIPGVGVPGVGVPGVGETTSKCTS\*

The ELP was designed as a pentapeptide repeat of hydrophobic amino acids, with abbreviated sequence:

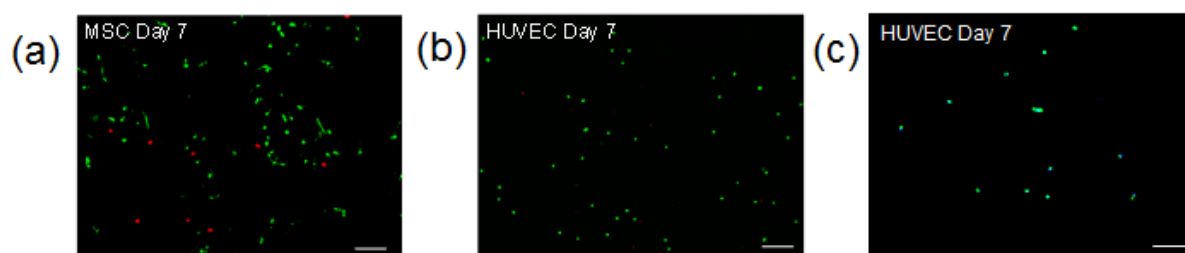
[[VPGVG]<sub>4</sub>IPGVG]<sub>14</sub>.

The complete nucleotide sequence of the completed gene is as follows, with the cysteine containing sequence (KCTS) italicized and the ELP gene in normal type:

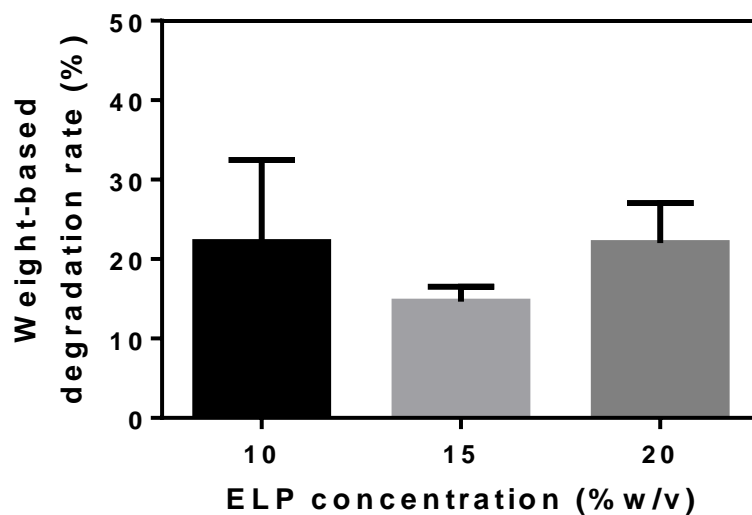
GGATCCAAATGTACCAGCGCTAGCGGTCTCGTTGGTGTACCTGGTGTGGCGTCCC  
GGGTGTAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGT  
GTACCTGGTGTGGCGTCCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTGGTGTACCGGGT  
TAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTGGTGTACCGGGT  
AGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGC  
GTCCCAGGCGTTAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCG  
TTGGTGTACCTGGTGTGGCGTCCCAGGCGTTAGGTATCCCAGGCGTTGGTGTACC  
GGGTGTAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTAGGT  
ATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGTGTACCTGGT  
TTGGCGTCCCAGGCGTTAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCC  
AGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTAGGTATCCCAGGCGTTGGT  
GTACCGGGTGTAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCG  
TAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGTGTACC  
TGGTGTGGCGTCCCAGGCGTTAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGG  
GTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTAGGTATCCCAGGCG  
TTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCC  
GGGTGTAGGTATCCCAGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGT  
GTACCTGGTGTGGCGTCCCAGGCGTTAGGTATCCCAGGCGTTGGTGTACCGGGT  
TAGGCGTTCCAGGCGTTGGTGTACCTGGTGTGGCGTCCCAGGCGTTAGGTATCCC  
AGGCGTTGGTGTACCGGGTGTAGGCGTTCCAGGCGTTGGTGAGACCACTAGTTAA  
ATGAATAAATGCACGTCCTAAAAGCTT



**Figure S1.** (a) Protein gel of KCTS-E<sub>31</sub>-KCTS and E<sub>22</sub> proteins after purification. (b) UV-Vis plot showing the transition temperature of a 1% (w/v) solution of ELP.



**Figure S2.** *In vitro* cell seeding on ELP hydrogels. (a-b) Calcein-AM (green)/ethidium homodimer (red) LIVE/DEAD assay on ELP hydrogels seeding with MSCs (a) or HUVECs (b) at day 7 of culture (scale bar = 200  $\mu$ m). (c) Phalloidin (green)/DAPI (blue) staining for F-actin/cell nuclei of ELP with HUVECs at day 7 of incubation (scale bar = 100  $\mu$ m).



**Figure S3.** Enzymatic degradation rate of ELP gels *in vitro* with digestion with proteinase K for 6 hours.

**Table S1.** Mechanical characterization of photocrosslinked ELP hydrogels.

ELP concentration (% (w/v))	Elastic modulus (kPa)	Stress at break (kPa)	Strain at break (%)	Compressive modulus (kPa)	Energy loss (%)
10	1.28 ± 0.17	6.46 ± 0.35	419 ± 25	3.01 ± 0.44	35.13 ± 2.55
15	1.72 ± 0.11	7.71 ± 0.53	395 ± 10	6.15 ± 0.28	42.10 ± 4.37
20	2.21 ± 0.36	10.09 ± 1.81	388 ± 12	13.05 ± 1.20	51.15 ± 2.90