Discovery of Small Molecules that Inhibit the Disordered Protein, p27^{Kip1}

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Supplementary Figure 1. Representative 1D NMR results showing that small molecules bind to p27-KID. **(a, b)** Representative 1D NMR results for two Group 1 molecules binding to p27-KID. 1D ¹H spectra are shown for the "hit" molecules and for the pool of five compounds (pool) in which they were originally screened. Also illustrated are WaterLOGSY (WL) and saturation transfer difference (STD) spectra for the compounds in the presence (w/) and absence (w/o) of p27-KID. The chemical structures of the two compounds are illustrated at the left.



Supplementary Figure 2. Individual ¹⁵N ($\Delta\delta_N$) chemical shift perturbations obtained through analysis of 2D ¹H-¹⁵N "in-phase" HSQC NMR titrations of SJ710 (**b**, Group 1) and SJ403 (**c**, Group 2), respectively, into ¹⁵N-p27-KID (100 μ M).



Supplementary Figure 3. Results of 2D ¹H-¹⁵N HSQC analysis of all small molecules that exhibited binding to ¹⁵N-p27-KID. Individual ¹H chemical shift perturbation histograms are shown for the molecules identified as "hits" in the initial fragment library screen (**a**), in the follow-up screen of chemically similar compounds in the same libraries (**b**), and from the inhouse high-throughput screening library identified on the basis of the field alignment models (**c**). The threshold for identifying specific interactions with p27-KID residues was defined as two standard deviations above the average the perturbation values (represented by a dotted black line in each graph). The experimental spectral resolution in the ¹H dimension (3.5 Hz) is represented by the dotted magenta line. The molar ratios of ¹⁵N-p27-KID (100 μ M) to inhibitor used for titrations were 1:2.5 (red), 1:5 (green), and 1:10 (blue).



Supplementary Figure 3 (continued)



Supplementary Figure 3 (continued)



Supplementary Figure 4. Amino acids with aromatic side chains do not bind specifically to p27. (a, c) Overlaid 2D ¹H-¹⁵N HSQC NMR spectra of ¹⁵N-p27-KID (100 µM) in the absence (red) and presence of (a) tryptophan (3 mM, green) and (c) tyrosine (3 mM, blue). (b, d) Expanded regions (represented by the blue boxes) including a subset of the p27-KID residues involved in interaction with fragment hits (highlighted in Fig. 2b, e) show no perturbations after addition of tryptophan (b) and tyrosine (d), respectively.



Supplementary Figure 5. Results of 2D NMR analysis of mutant forms of p27-KID to determine the contributions of tryptophan and tyrosine residues to interactions with small molecules. Histograms of ¹H chemical shift perturbations observed in 2D ¹H-¹⁵N HSQC spectra with increasing concentrations of SJ403 for, (**a**) ¹⁵N-p27-KID-W₆₀A, (**b**) ¹⁵N-p27-KID-W₇₆A, (**c**) ¹⁵N-p27-KID-W₆₀A-W76A, (**d**) ¹⁵N-p27-KID-W₆₀F, (**e**) ¹⁵N-p27-KID-W₇₆F, (**f**) ¹⁵N-p27-KID-W₆₀F-W₇₆F, (**g**) ¹⁵N-p27-KID-F₈₇A, (**h**) ¹⁵N-p27-KID-Y₈₈A, (**i**) ¹⁵N-p27-KID-Y₈₉A; and of SJ319843 for (**j**) ¹⁵N-p27-KID-F₈₇A, (**k**) ¹⁵N-p27-KID-Y₈₈A, (**l**) ¹⁵N-p27-KID-Y₈₉A, and (**m**) ¹⁵N-p27-KID-R₉₀A. Ratios of ¹⁵N-p27-KID to small molecules of 1:5 (green) and 1:10 (blue) were used.



Supplementary Figure 6. (a) Comparison of amino acid sequences of the D2 subdomains of p27-KID and p21-KID; (b, c) Overlaid 2D ¹H-¹⁵N HSQC NMR spectra of ²H/¹⁵N-p21-KID (20 μ M) in the absence (red) and presence (green) of 1 mM SJ319843 (b, Group 1) and 1 mM SJ403 (c, Group 2), respectively; glycine region was omitted for improved clarity.



Supplementary Figure 7. 2D NMR analysis of the displacement of ²H/¹³C/¹⁵N-p27-D2 from Cdk2/cyclin A by SJ403. 2D ¹H-¹⁵N TROSY-HSQC spectra of ²H/¹³C/¹⁵N-p27-D2/Cdk2/cyclin A in the absence (**a**) and presence of 3 mM SJ403 (**b**). In (**c**), the spectra in (**a**) and (**b**) are overlaid to emphasize resonance perturbations associated with the binding of SJ403 to p27-D2. Unperturbed resonances appear in yellow color. Examples of resonances of p27-D2 bound to Cdk2/cyclin A that decreased in intensity upon interaction with SJ403 are labeled with a superscripted "b". Examples of resonances corresponding to free p27-D2 that increased in intensity in the presence of SJ403 are marked by a superscripted "f". (**d**, **e**) Chemical shift perturbations observed for resonances of isolated ¹⁵N-p27-D2 in the presence of 3 mM SJ403 (**d**) and for the population of free p27-D2 observed for the sample containing ²H/¹³C/¹⁵N-p27-D2, Cdk2/cyclin A, and 3 mM SJ403. The locations of sub-domains within p27-D2 are indicated by the shaded boxes.



Supplementary Figure 8. Graphic representation of the displacement of p27-D2 by SJ403. The initial equilibrium is perturbed by the addition of SJ403, resulting in increased population of free p27-D2, which is bound to the small molecule, SJ403.



Supplementary Figure 9. Results of Cdk2 activity assays for Cdk2/cyclin A (200 pM) in the presence of (**a**) increasing concentrations of p27-D2, (**b**) p27-D2 (200 nM) and increasing concentrations of SJ403, and (**c**) increasing concentrations of SJ403. The panels show phosphoimager results after SDS-PAGE analysis of ³²P incorporation from ATP into the substrate, Histone H1. A single set of representative results are shown; all experiments were performed in triplicate.



Supplementary Figure 10. Compact (C; panels a) and Extended (E; panel b) conformations from the p27-D2 MD simulation partitioned using both the timeseries information and distance distributions from Fig. 9. The times at which these representative structures were selected are indicated in Fig. 9a. The atoms of W_{60} , W_{76} and Y_{88} are illustrated as sticks in each structure. The blurring represents the conformations of molecules within 0.1 ns before and after the sampled time in the trajectory. Panel (c) summarizes the distribution of conformers along the distances between W₆₀-W₇₆ Cβ, W₆₀-Y₈₈ Cβ and W₇₆- Y_{88} C β atoms. Each conformer is colored according to its timestamp from the simulation, with the color drawn uniformly from fifty bins between 0 (blue) microseconds and 0.4 (red) microseconds. The conformers from the respective compact (C) states are shown as three ellipses. The first ellipse (light blue color) represents conformers from C1 and C2, where the distance between W_{60} - Y_{88} C β and W_{76} - Y_{88} C β atoms were small. The conformers from C3-7 (marked by green ellipses) are shown in regions where either the W_{60} - Y_{88} C β distance was small (and the W_{76} - Y_{88} C β distance was large) or vice-versa. The extended (E) conformers are highlighted in E1-E12, which were taken from other regions of the simulation (Fig. 9a).



Supplementary Figure 11. Illustration of the structure of aromatic residues within sub-domain D2 and key binding residues with sub-domain D1 of p27 from the p27-KID/Cdk2/cyclin A structure. (a) View of entire structure. (b–d) Expanded views of sub-domain D1 binding to cyclin A (b) and of the binding of sub-region D2.1-D2.2 (c) and D2.3 (d) to Cdk2.



p27-KID

b

$$\label{eq:gshm_e_22} \begin{split} \texttt{GSHM-e}_{22} \texttt{HPKPSACRNLFGPVDHEELTRDLEKHCRDME} \\ \texttt{EASQRKWNFDFQNHKPLEGKYEWQEVEKGSLPE} \\ \texttt{FYYRPPRPPKGACKVPAQE}_{105} \end{split}$$

GSHM-R₅₈KWNFDFQNHKPLEGKYEWQEVEKGSLPE FYYRPPRPPKGACKVPAQE₁₀₅



Supplementary Figure 12. (a) Chemical features of molecules in the two fragment libraries used to screen for binding to p27-KID. Features of the Maybridge Ro3 library are shown in red and those of the **In-house** fragment library in blue. **(b-e)** Amino acid sequences and 2D ¹H-¹⁵N HSQC spectra (showing resonance assignments) for p27-KID (**b**, **c**) and p27-D2 (**d**, **e**), respectively. Puple asterisk (*) indicate resonances arising from minor conformers. In (**c**), the regions marked by colored boxes are expanded and shown at a lower contour level in the insets at the right.

ID	Chemical structure	Binding Group	AlogP	tPSA	H-bond acceptor	H-bond donor	MW (Da)
CC34414	O NH2	Group 1	2.19	35.25	2	2	159.18
SJ000053521	F ₃ C NH ₂	Group 1	2.97	67.14	3	2	244.24
SJ000483069	NH ₂	Group 1	2.76	56.21	4	2	264.32
SJ000023358	ОН	Group 1	3.09	46.53	3	1	216.23
SJ000572774		Group 1	2.24	20.31	2	0	197.23
SJ000248846		Group 1	3.17	79.45	4	1	264.34
SJ000572486		Group 1	2.75	49.41	4	1	294.34
SJ000319768		Group 1	2.02	58.64	5	1	296.32
SJ000208978		Group 2	0.07	72.08	8	0	291.30

Supplementary Table 1. Names, chemical structure, Group designation, and chemical parameters for molecules identified as "hits" in the initial fragment library screen (**a**), in the follow-up screen of chemically similar compounds in the same libraries (**b**), and from the in-house high-throughput screening library identified on the basis of the field alignment models (**c**).

а

ID	Chemical structure	Binding mode	AlogP	tPSA	H-bond acceptor	H-bond donor	MW (Da)
SJ000572513		Group 1	2.75	49.41	4	1	294.34
SJ000572710	O C C C C C C C C C C C C C C C C C C C	Group 1	2.77	55.76	4	1	260.28
SJ000319843		Group 1	2.15	58.64	5	1	296.32
SJ000319683		Group 1	1.67	58.64	5	1	282.29
SJ000572487		Group 1	2.33	58.64	5	1	296.32
SJ000572443		Group 1	2.15	55.56	4	2	254.28
SJ000572401		Group 2	1.1	62.85	7	0	287.31
SJ000572403		Group 2	0.83	62.85	7	0	275.30
SJ000572405		Group 2	1.35	62.85	7	0	289.33
SJ000572407		Group 2	0.83	62.85	7	0	275.30
SJ000572409		Group 2	1.1	62.85	7	0	287.31
SJ000572542		Group 2	0.92	62.85	7	0	275.30
SJ000852806		Group 2	2.08	62.85	7	0	317.39
SJ000271822		Group 2	0.285	71.06	7	0	248.24

С								
	ID	Chemical structure	Binding Group	AlogP	tPSA	H-bond acceptor	H-bond donor	MW (Da)
	SJ000017408	S NH2	Group 1	1.79	84.22	3	2	218.27
	SJ000023360		Group 1	3.05	83.72	3	0	221.3
	SJ000023362	S S S S S S S S S S S S S S S S S S S	Group 1	3.664	83.72	3	0	247.34
	SJ000024853	HOTOGO	Group 1	2.76	72.83	5	1	276.28
	SJ000034522		Group 1	2.623	90.69	8	2	298.24
	SJ000039342		Group 1	2.63	71.84	7	0	355.39
	SJ000039843		Group 1	3.01	127.1	6	0	355.4
	SJ000045744	HO NO	Group 1	2.94	56.33	4	2	306.36
	SJ000013100	N N N	Group 2	2.02	44.12	4	0	214.22
	SJ000082958		Group 2	2.69	88.32	7	1	362.38
	SJ000572844		Group 2	0.97	62.74	6	0	247.25
	SJ000852808		Group 2	1.317	66.09	8	0	346.43

Supplementary Table 1 (continued)