

Supplemental Information

Supplemental Tables

Table S1. Conventional PCR primers used in SF1 and Scr 3C capture

NAME OF ELEMENT	OUTSIDE PRIMER FOR NESTED PCR (5'>3')	INSIDE PRIMER FOR NESTED PCR (5'>3')
SCR-P	CATCGGGGCTTAACGATTTTG	TTTATCGGTCTGTTTCGGGG
SF1	ACCCGAGCTGTCGGTTATTA	CTGAAATTTTCGAAAGTATGAA
R1	TCGAGGGTTCCTTACCTCT	CGACGCAAGCACACAGA
R2	CAGGAGCTAAAAATGCGTTCA	TTCGCAACATTTTGGCCTG
R3	GAAAGGTTGCTCAGAACAGGG	CCGCACAAATTACATAATCACG
R4	CGACAAAAGACGCCAAGACA	ATGCGAATGGGAGAAATGCG
R5	TGAAAGGAGTTACTCGCAACT	TAATTTAGCAGGTCGGGCG
R6	CACGATCGATCGCCAATCAT	GCCAAACAGGTGTGTCTGG
R7	TGG CGC TCA AAA GCG TT	GGCCTGGCTTCGACTTTT
R8	AATGTCGTATCACACAGGCG	GGCAATCCGTCACTCACATT
R9	GCGACAAGTGTGTGCGTAG	GCGCTTTGCCTTTGGACAT
R10	GCCTTCTATTCGGGTATCTCT	ATAACCACTGCAGTCGGTTAG
R11	TGTCAGAAATCCACTCGAGC	TCCATCTCTCGATCCATCAA

Table S2. Primers for SF1-based 3C capture qPCR

SF1	CTGAAATTTTCGAAAGTATGAA
FTZ-P/R-2-Q	TGATAATGGCCAAGAGACTT
FTZ-E/R-1-Q	ATGCACACTTGGAAAAAGTT
R1-Q	CAGTCCGAGAGTATCCTTTT
R2-Q	ATTTTGGCCTGACTAGGG
R3-Q	GTGCCGCACAAATTACATA
R4-Q	CTTTACTGGGGCGGATCA
R5-Q	TGAAACAATGATGAGGTTGA
R6-Q	TCCAGTCGATTAAAGTGAGC
R7-Qd	TAACGAACGATCAGCAATTA
R7-Qp	CTGGACGTGGATGGATAAT
R8-Q	ATGGTCAGGGGTGTAATATG
R9-Q	GAAAAGGAGGAACTGCACA
R10-Q	ATCGCTTTTTTGGGAGTTCTA
R11-Qd	TATTTAAATCGTTCGGTTGC
R11-Qp	TACTTTACACGGGTTTTGTT
R12-Qp	TGGGTAAC TCAAACACAT
R12-Qd	GACGAGGGATCAGGGAAT

Table S3. qPCR primers used in ftz promoter-based 3C capture

NAME OF ELEMENT	OUTSIDE PRIMER FOR NESTED PCR (5'>3')	INSIDE PRIMER FOR NESTED PCR (5'>3')
R3	GAAAGGTTGCTCAGAACAGGG	CCGCACAAATTACATAATCACG
R2	CAGGAGCTAAAAATGCGTTCA	TTCGCAACATTTTGGCCTG
R1	TCGAGGGTTCCTTACCTCT	CGACGCAAGCACACAGA
F1	CAAAGAGGCAGCGCAAACA	AAGCCCAAACACTCAGCTG
F3	TAATTGCCCGCGTATCCTTCG	CCCCTTCCTCCTTCCTTCGTT
F4	CATCATGCCCTTCCGTAGTCG	CGCCACACACTTTTCGTTACAAG
F5	GTCCTTGACTGCCATAAAGTCAG	AGTTTCAAGAAATTCTCTGTTTCGCT
SF1	ACCCGAGCTGTCGGTTATTA	CTGAAATTTGAAAGTATGAA
SCR-1	ACGAATGACCTATGCCATTTATCT	CAAATCGATTTTGGGTGTGTCT
SCR-P	CATCGGGGCTTAACGATTTTG	TTTATCGGTCTGTTTCGGGG

Table S4. Other primers used in for the *Drosophila* distance-frequency curve

CATCGCTTTTTTCGTTTCGCAT	ATGCCCGTCTGATGACGACAA
GCTTTACTCATGCCATTCTGTGT	GTAAAAGCAAAGCCAGAATGCG
GTTTGTTTACGGTTCGTAGCTGA	GGAAGATCAGTGTTATTGCGCT
CGGGTGGCTCATAGTTCATAG	TCCTGTGGCACTTTACTAGAACTT
CCATTAATTTTCGCCGCCATTT	GTGCATACAGGCGATTGAGAA
CCTGGCAAAAAGTTGAATGGGA	CAGCCGGGCAAACCGTCTATG
CCGTGTCCTTCTCAGCATTTT	GATGTTGGCATCGGTGCTCCA
GAAGTCGCGTTCGGCCAAATG	GGGAAAGCTTGATGCAACCAT
CTCTCGCTCTCGTGGTTTCCT	CAATTGCACAGAAAAAGTTTCGCT
TCGAGGATGTGAGAGTACGGAT	CCCCGAACTGTGATAAACGGA
ATTACTCAAGCATGGCACCCA	TCTTAGAGCATTCTGAATGAAGGA
GGCAATCCCTTCTATACTTTCCTT	AAATCGGGTTTTTCCTTGCCG
AATTTGGAATGTGTCCCGCTG	ACCTACACACTTGGTAGCATAATCA
GGCTTAATGAGTGTAAGCCACA	CACGCGAAATGTCCGCTTACA
GGTCCTAAGCAACTCCTGAAAA	TGACTTCTTGGCAGCAATGCA
CTGGGGGATACGAGTGTAAGA	CGCCCTGGATCTGAAGACATA
CCCAGCTTAAGTGCTTAACCA	ATCTGTAGGGTTGGCCGTATT
CCCGCTTATGAAATCGGCATA	CCGTATATCTTGTTGCAGGCG
TCATGGCCACGAAAATGTCCA	GATTCGGAACCTTAGTACGTTTTCT
GTGGCATCGTTTTTGGTTGTTG	
CTCACGTGTTTCTGACGTACG	
ACGTACATATCTGGTTAAGC	
CACCTAAATTGGGGTAACTG	
TTAGAACCAAAAAGGAGCTGGG	
AGTTTGCCGGTAATGTTCT	
TAAAGGCGCTGCTTTTGCAAG	
TAAGCTGGAAAACCTTACCTCACC	
GGATTAGTTTGAGTCCGATCAGG	
TTTCTCCATTTTCGATCCCGAG	
GCCAATGATCTGAACGATGCG	

Table S5. PCR primers used for cloning STEs in enhancer-blocking transgenes

SF2B	TGCGGCCGCCCATCCTCTTGTGAGGCTGG
	TGCGGCCGCCTGATTGACGAATTGCGTGCG
R2	GAATTCGTCAATGTCAAAATCAGAC
	TTCTGTTTTTGCTTCCAGTGTG
R6	GCGGCCGCGAAATAAATGAATTCTGGT
	AGAATTCACCCGCCAATGTGT
R9	ATAAGAATATGCATCCCGCTCCTCGATTGTGTGAT
	ATAAGAATATGCATTGATTTGAGAGTCCGCTG
R10	GAATTCGAAGTCGAACTTTCGC
	TCAATTGTGCAAAAAAAGCCA
1.5 kb spacer	TTGCATTAAGAAGAGGTGACTG
	GCGGCCGCTCTAGATACCGAATTT
attB	CCAATGCATGTCGACGATGTAGGTCACG
	AACTGCAGTTCGGCTTGTGACATGC

Supplemental Figure Legend

Figure S1.

Photo of PCR products after electrophoresis on 2% agarose gel and stained with ethidium bromide. Right: product from 3C capture between SF1 and R1-R11, followed by nested PCR. Middle and left: PCR product from positive control 3C using ligated genomic DNA after EcoRI digestion (GCL), and negative control 3C without crosslinking (NCL).

Figure S2

Relative frequency of 3C capture between SF1 and R-1, R1 and R2 in 4h-8h (orange bars) and 10h-14h (purple bars) embryos were quantitated by qPCR (for details see methods and Fig. 4 legend).

Figure S3

STEs correspond to developmentally regulated chromatin domain boundary in the *Scr-Antp* genomic region. Screen crops of H3K9Me3 and H3K4Me1 ChIP-seq profile in the *Scr-Antp* region from different embryonic stages (<http://www.genome.gov/modENCODE>). EcoRI sites are shown on top. Vertical shaded bars indicate CBEs in the regions (red, SF1; orange, R1, grey R2, R6 and R10).

Figure S4.

Binding profile of known insulator factors on STEs. Screen crop of ChIP-seq profile of several known insulator proteins at and near SF1 and STE sequences (<http://www.genome.gov/modENCODE>).

Figure S5.

NEE-blocking in labial thoracic tissues in R2 and R6 transgenic embryos.

Representative images of embryos from independent transgenic lines containing R2 or R6 element in the pFWNHZ transgene, stained with the anti-lacZ (**A-L**), or anti-lacZ and anti-Scr (**P-R**) RNA probes. Anterior region of embryos are shown in ventral lateral views, anterior to the left and dorsal up. **A-D**). Variation of NEE blocking levels in pFWNHZ-R2 or R6 embryos (see **M**). **E-H**). Example of strong NEE-block in pFWNHZ-R2 embryos. **I-L**). Example of strong NEE-block in pFWNHZ-R6 embryos. **M**). Quantitation of NEE-blocking in the labial thoracic tissues (gap) in embryos from multiple pFWNHZ-R2 and pFWNHZ-R6 transgenic lines, stained with the lacZ probe (see methods for details). **N-Q**). Embryos from pFWNHZ-R6 lines double stained with anti-lacZ and anti-Scr probes.

Fig S1

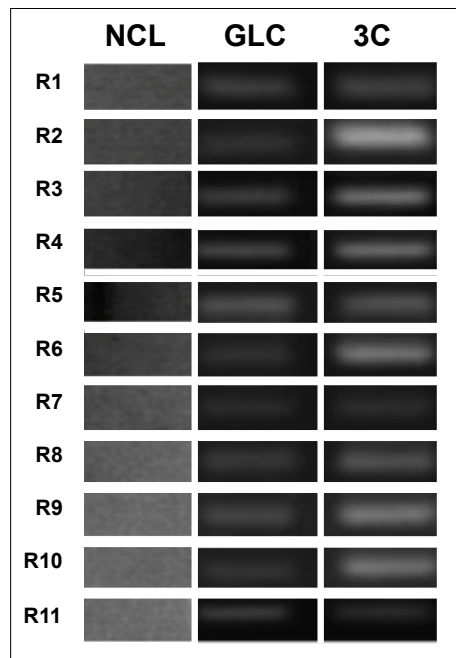


Fig S2

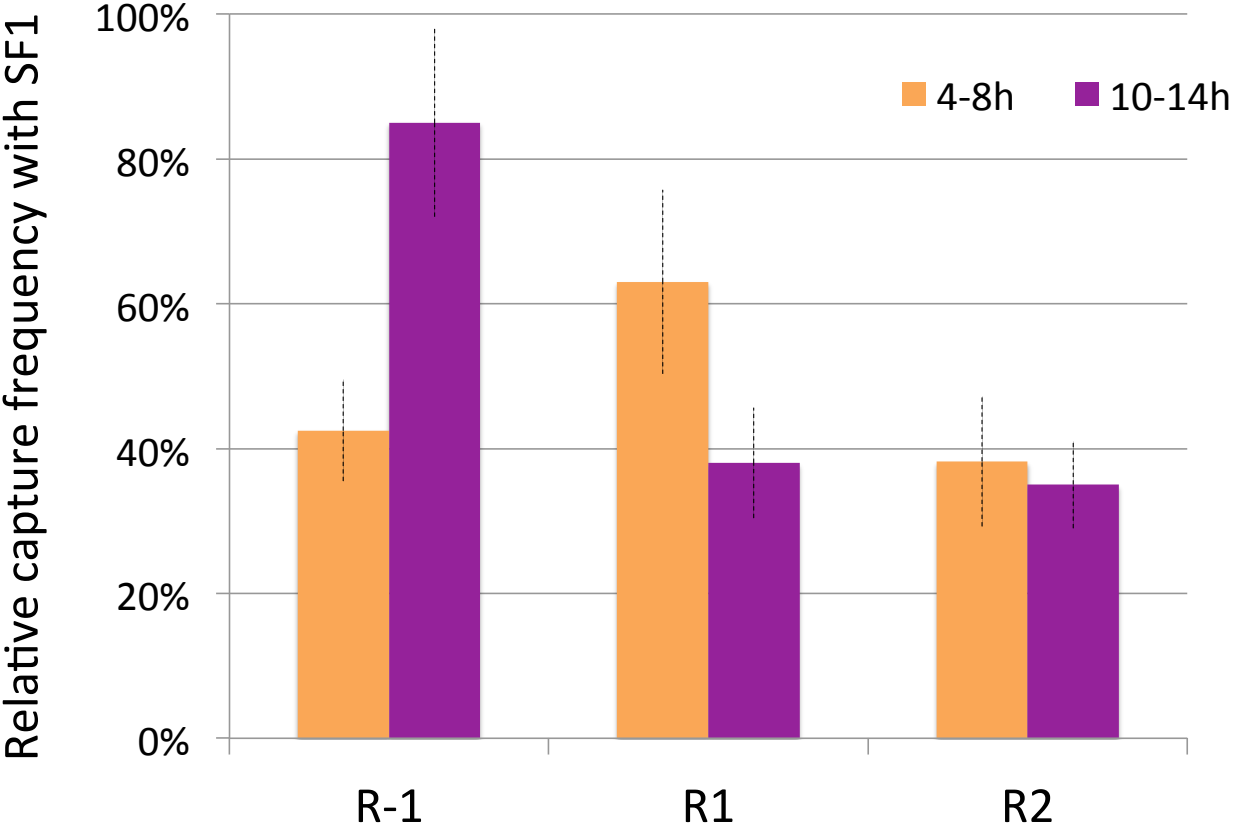


Fig S3

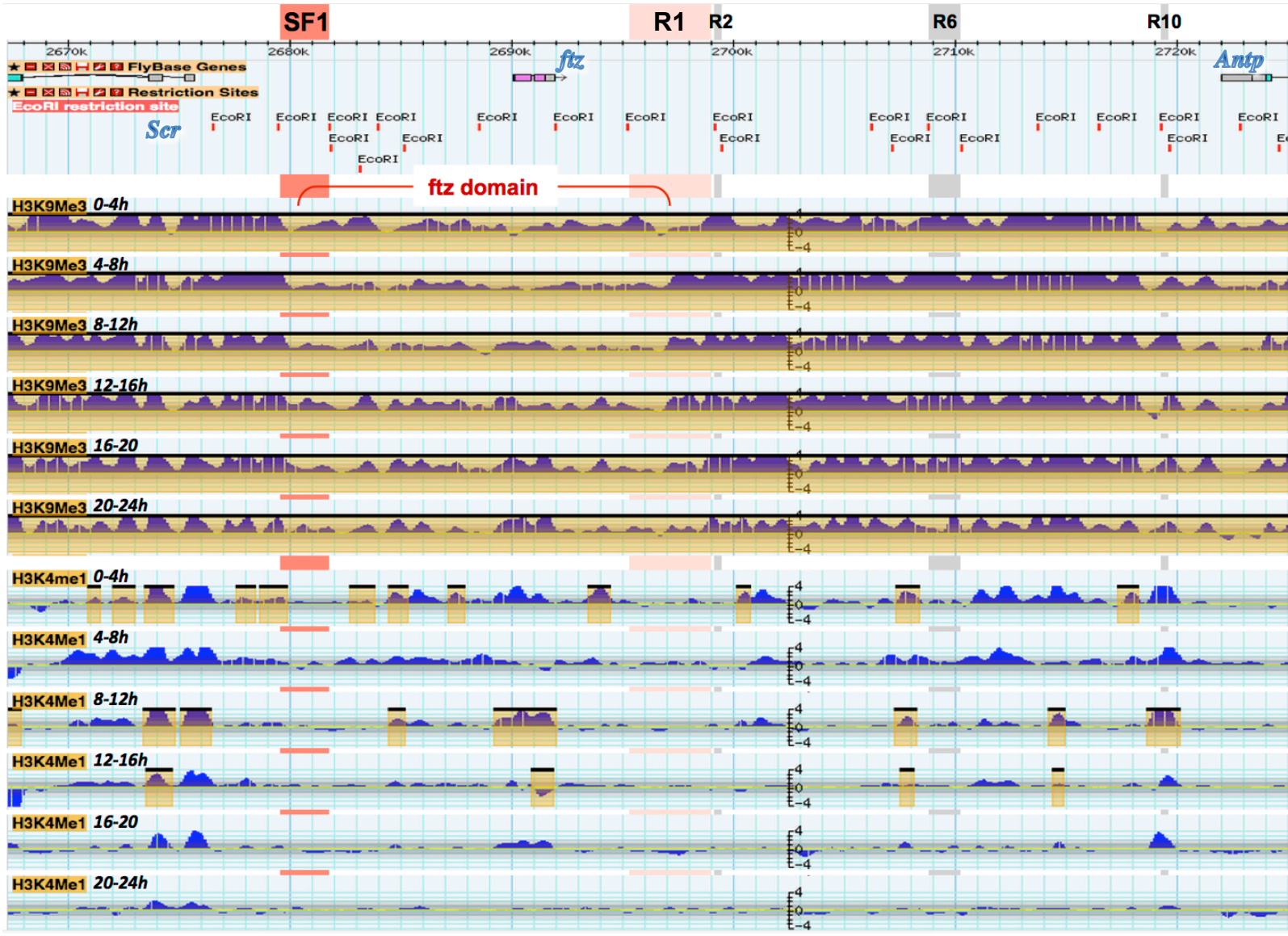


Fig. S4

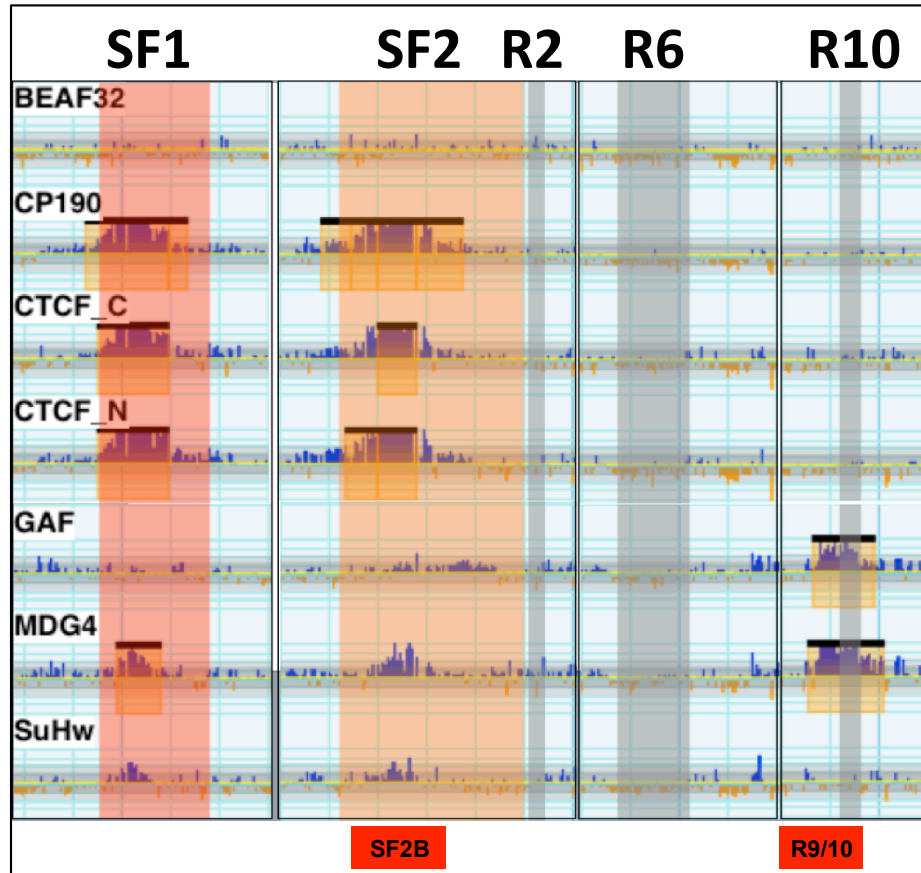


Fig. S5

