

Supplementary Information

Figure S1. Inactivation of Imd pathway cannot suppress *GMR>Egr*-triggered small eye phenotype.

Light micrographs showing *Drosophila* adult eyes. The *GMR>Egr*-induced small eye phenotype (**a**, **d** and **g**) is not suppressed by knocking-down *imd* (**b**) or *relish* (**c**), or deleting both copies of endogenous *imd* (**f**) or *relish* (**i**), or expressing LacZ (**e** and **h**).

Figure S2. Ectopic expression of Grim induces loss-of-ACV phenotype.

(**a** and **b**) Light micrographs showing *Drosophila* adult wings. Compared with the control (**a**), limited expression of Grim driven by *ptc-GAL4* with *Tub-GAL80^{ts}* produces the loss-of-ACV phenotype (**b**). The lower panels show high magnification views of the boxed areas in the top panels. (**c**) Quantification of the ACV phenotype as shown in figures **a** and **b** (n=20, for each genotype). Error bars indicates standard deviation. Unpaired t test was used to calculate statistical significance, indicated with asterisks (***) ($P < 0.001$).

Figure S3. Expression of Hep triggers cell death in wing discs.

(**a** and **b**) Fluorescence micrographs of third instar larval wing discs are shown. Compared with the control (**a**), expression of Hep driven by *Sd-GAL4* induces evident cell death in larval wing discs indicated by AO staining (**b**). (**c**) Statistical analysis of cell death in wing discs (n=10) as shown in figures **a** and **b**. Error bars indicate standard deviation. Unpaired t test was used to calculate statistical significance, indicated with asterisks (***) ($P < 0.001$).

Figure S4. Loss of Toll signaling suppresses JNK-mediated cell death.

Light micrographs of *Drosophila* adult wings (**a-e**) or third instar larval wing discs (**g-k**) are shown. Compared with *Sd-GAL4* control (**a** and **g**), ectopic expression of Hep driven by *Sd-GAL4* results in a dramatic size reduction in adult wings (**b**) and larval wing discs (**h**), both of which are suppressed significantly by expressing Bsk^{DN} (**d** and

j) or a *dorsal* RNAi (**e** and **k**), but not that of GFP (**c** and **i**). (**f**) Statistical analysis of the adult wing size as shown in figures **a-e**. One-way ANOVA with Bonferroni multiple comparison test was used to compute *P*-values, significance was indicated with asterisks (***) $P < 0.001$. ns stands for not significant.

Figure S5. Inactivation of Toll pathway cannot suppress *GMR>Hid*-induced small eye phenotype.

Light micrographs showing *Drosophila* adult eyes. The *GMR>Hid*-triggered small eye phenotype (**a**) is not suppressed by knocking-down *dorsal* (**b**) or *Dif* (**c**).

Figure S6. Egr fails to induce *Dipt* expression in fat body cells.

Light micrographs of *Drosophila* third instar larval fat body are shown. *Dipt* expression in third instar larval fat body cells (**a**) cannot be activated by ectopic expression of Egr under the control of *Cg-GAL4* (**b**), but triggered by expression of Imd (**c**), detected by X-gal staining of *Dipt-LacZ*.

Figure S7. Gain of JNK signaling induces dorsal expression in eye discs.

Fluorescent microscope images showing eye discs dissected from third-instar larvae stained with anti-Dorsal (red). Compared with the control (**a**), the level of endogenous Dorsal was up-regulated by expressing dTAK1 (**b**). **a'-a''** and **b'-b''** show high magnification views of the boxed areas in **a** and **b**, respectively. White arrows indicate the regions where Dorsal staining overlaps with the nucleus. Nuclei were labeled with DAPI (blue). Imaging of prepared sample was conducted by a Leica confocal microscope (Leica SP5).

Figure S8. Gain of JNK signaling promotes dorsal nuclear-localization in fat body cells.

Fluorescent microscope images showing fat body cells dissected from third-instar larval stained with anti-dorsal (red). Nuclei were marked with DAPI (blue), cell membranes

were tagged by GFP (green). Compared with the control (**a**), expression of Egr (**b**) or RNAi inactivation of *puc* (**c**) in fat body promotes the nuclear accumulation of Dorsal. (**d**) Quantification of the nuclear/cytoplasmic ratio of Dorsal in fat body cells shown in **a-c**. Anti-Dorsal intensities were measured in pixels using Image J. Error bars showed standard deviation from measurement of at least 15 cells for each genotype. One-way ANOVA with Bonferroni multiple comparison test was used to calculate statistical significance, indicated with asterisks (***) $P < 0.001$)

Figure S9. Blocking caspases does not affect JNK-triggered Toll activation.

Fluorescence micrographs of third instar larvae (**a** and **b**), dissected fat body cells stained with anti-dorsal (red) (**c**) and eye discs (**d-i**) are shown. Nuclei were labeled with DAPI (blue). The up-regulated *Drs-GFP* expression and the nuclear accumulation of endogenous Dorsal induced by activated JNK signaling were caspase-independent (**a-c**). dTAK1+P35-expressing clones were tagged by GFP (green) and induced for 1h by heat shock at 37°C and recovered for 24h at 25°C. (**d-f**) The up-regulated expression of *Spz6-GFP* triggered by ectopic dTAK1 (**d**) can be suppressed by expressing Bsk^{DN} (**f**), but not by P35 (**e**). Cell membranes were stained by anti-Dlg antibody (red). (**g-i**) Compared with the control (**g**), the number of hemocytes attached to eye discs is not significantly changed by ectopic expression of Egr (**h**) or dTAK1 (**i**) driven by *GMR-GAL4*. Hemocytes were stained by anti-NimC1 antibody. Images **g-i** were taken in a Zeiss LSM780.

Figure S10. JNK signaling is not involved in Toll/NF-κB triggered cell death.

(**a-c**) Fluorescence micrographs of third instar larval eye discs are shown. Compared with the control (**a**), both *GMR>Dorsal* and *GMR>Dif* induce cell death in larval eye discs (**b** and **c**). (**d**) Statistic analysis of cell death in eye discs shown in (**a-c**). Average number of dying cells labelled by AO staining are counted. Error bars indicates standard deviation. One-way ANOVA with Bonferroni multiple comparison test was used to compute *P*-values, significance was indicated with asterisks (***) $P < 0.001$, $n=10$). (**e-l**) Light micrographs showing *Drosophila* adult eyes. Expression of Dorsal (**e**) or Dif (**i**)

under the control of the *GMR* promoter triggers cell death and generates rough eyes with reduced size, which is not suppressed by expressing of LacZ (**f** and **j**), Bsk^{DN} (**g** and **k**) or Puc (**h** and **l**).

Figure S11. The expression of *Drs-GFP* is induced by gain of Toll signaling in the fat body.

Fluorescence micrographs of third instar larvae are shown. The *Drs-GFP* is significantly up-regulated by gain of Toll signaling in the fat body.

Figure S12. Toll/NF- κ B triggered cell death is independent of caspase or necroptosis.

(**a-c**) Fluorescence micrographs of third instar larval eye discs are shown. Compared with the control (**a**), no increased caspase activity is observed in *GMR>Dorsal* or *GMR>Dif* eye discs (**b** and **c**). (**d**) Statistic analysis of caspase activation in eye discs shown in (**a-c**). Average number of cells labelled by anti-CC-3 staining are counted. Error bars indicates standard deviation. One-way ANOVA with Bonferroni multiple comparison test was used to compute *P*-values, significance was indicated with asterisks (***) $P < 0.001$, $n = 10$). (**e-p**) Light micrographs showing *Drosophila* adult eyes. The rough and small eye phenotypes of *GMR>Dorsal* (**e**) and *GMR>Dif* (**k**) are not suppressed by blocking caspases signaling (**f-h** and **l-n**) or knocking-down dTRAF2 (**i**, **j**, **o** and **p**).

Detailed Genotypes

Figure 1

(b) *GMR-GAL4/+*

(c) *UAS-Egr/+; GMR-GAL4/+*

(d) *UAS-Egr/+; GMR-GAL4/Df(3R)ED6232*

- (e) *UAS-Egr/+; GMR-GAL4/Df(3R)BSC496*
- (f) *UAS-Egr/+; GMR-GAL4/Df(3R)BSC524*
- (g) *UAS-Egr/+; GMR-GAL4/Df(3R)ED6255*
- (h) *UAS-Egr/Df(2L)BSC294; GMR-GAL4/+*
- (i) *UAS-Egr/+; GMR-GAL4/Toll^{r3}*

Figure 2

- (c) *GMR-GAL4/+*
- (d) *UAS-Egr/+; GMR-GAL4/+*
- (e) *UAS-Egr/+; GMR-GAL4/UAS-LacZ*
- (f) *UAS-Egr/imd^l; GMR-GAL4/+*
- (g) *UAS-Egr/Dif^l; GMR-GAL4/+*
- (h) *UAS-Egr/+; GMR-GAL4/UAS-Toll-IR (BL31044)*
- (i) *UAS-Egr/+; GMR-GAL4/UAS-tube-IR (NIG105520R3)*
- (j) *UAS-Egr/+; GMR-GAL4/UAS-pelle-IR (BL34733)*
- (k) *UAS-Egr/+; GMR-GAL4/UAS-dorsal-IR (V45998)*
- (l) *UAS-Egr/+; GMR-GAL4/UAS-Dif-IR (V30579)*

Figure 3

- (a) *sev-GAL4/UAS-dTAK1*
- (b) *sev-GAL4/UAS-dTAK1; UAS-LacZ/+*
- (c) *sev-GAL4/UAS-dTAK1; UAS-bsk-IR/+*
- (d) *sev-GAL4/UAS-dTAK1; UAS-pelle-IR (BL34733)/+*
- (e) *sev-GAL4/UAS-dTAK1; UAS-dorsal-IR (V45998)/+*
- (f) *GMR-GAL4 UAS-Hep^{CA}/+*
- (g) *GMR-GAL4 UAS-Hep^{CA}/UAS-LacZ*
- (h) *GMR-GAL4 UAS-Hep^{CA}/UAS-bsk-IR*
- (i) *GMR-GAL4 UAS-Hep^{CA}/UAS-pelle-IR (BL34733)*
- (j) *GMR-GAL4 UAS-Hep^{CA}/UAS-dorsal-IR (V45998)*
- (k) *GMR-GAL4/UAS-Bsk*

- (l) *GMR-GAL4/UAS-Bsk; UAS-GFP/+*
- (m) *GMR-GAL4/UAS-Bsk; UAS-pelle-IR (BL34733)/+*
- (n) *GMR-GAL4/UAS-Bsk; UAS-dorsal-IR (V45998)/+*
- (o) *GMR-GAL4/UAS-Bsk; UAS-Dif-IR (V30579)/+*

Figure 4

- (a) *ptc-GAL4/+*
- (b) *ptc-GAL4/UAS-Egr^W*
- (c) *ptc-GAL4/UAS-Egr^W; UAS-LacZ/+*
- (d) *ptc-GAL4/UAS-Egr^W; UAS-Bsk^{DN}/+*
- (e) *ptc-GAL4/UAS-Egr^W; UAS-Toll-IR (BL31044)/+*
- (f) *ptc-GAL4/UAS-Egr^W; UAS-dorsal-IR (V45998)/+*
- (g) *ptc-GAL4/UAS-Egr^W; UAS-Dif-IR (V30579)/+*
- (h) *ptc-GAL4/UAS-Hep*
- (i) *ptc-GAL4/UAS-Hep; UAS-LacZ/+*
- (j) *ptc-GAL4/UAS-Hep; UAS-Bsk^{DN}/+*
- (k) *ptc-GAL4/UAS-Hep; UAS-Toll-IR (BL31044)/+*
- (l) *ptc-GAL4/UAS-Hep; UAS-dorsal-IR (V45998)/+*
- (m) *ptc-GAL4/+; Toll^{EP(3)1051}/+*

Figure 5

- (a) *Sd-GAL4/+*
- (b) *Sd-GAL4/+; UAS-dJun/+*
- (c) *Sd-GAL4/+; UAS-dFos/+*
- (d) *Sd-GAL4/+; UAS-dFoxO/+*
- (e) *Sd-GAL4/+; UAS-dFoxO/+; UAS-GFP/+*
- (f) *Sd-GAL4/+; UAS-dFoxO/+; UAS-Toll-IR (BL31044)/+*
- (g) *Sd-GAL4/+; UAS-dFoxO/+; UAS-dorsal-IR (V45998)/+*
- (h) *Sd-GAL4/+; UAS-dFoxO/+; UAS-Dif-IR (V30579)/+*
- (j) *Sd-GAL4/+; UAS-Hep/+*

- (k) *Sd-GAL4/+; UAS-Hep/+; H99/+*
- (l) *Sd-GAL4/+; UAS-Hep/+; UAS-dIAP1/+*
- (m) *Sd-GAL4/+; UAS-Hep/+; UAS-Dronc^{DN}/+*
- (o) *Sd-GAL4/+; hid-LacZ/+*
- (p) *Sd-GAL4/+; UAS-Hep/+; hid-LacZ/+*
- (q) *Sd-GAL4/+; UAS-Hep/+; hid-LacZ/UAS-dorsal-IR (V45998)*
- (r) *Sd-GAL4/+; UAS-Hep/+; hid-LacZ/UAS-Dif-IR (V30579)*

Figure 6

- (a and h) *ptc-GAL4/UAS-puc-IR (V3108)*
- (b and i) *ptc-GAL4/UAS-puc-IR; UAS-LacZ/+*
- (c and j) *ptc-GAL4/UAS-puc-IR; UAS-Bsk^{DN}/+*
- (d and k) *ptc-GAL4/UAS-puc-IR; UAS-Toll-IR (BL31044)/+*
- (e and l) *ptc-GAL4/UAS-puc-IR; UAS-tube-IR (NIG10520R3)/+*
- (f and m) *ptc-GAL4/UAS-puc-IR; UAS-dorsal-IR (V45998)/+*
- (g and n) *ptc-GAL4/UAS-puc-IR; UAS-Dif-IR (V30579)/+*

Figure 7

- (a) *Drs-GFP/+; Cg-GAL4/+*
- (b) *Drs-GFP/+; Cg-GAL4/+; UAS-dTAK1/+*
- (c) *Drs-GFP/+; Cg-GAL4/UAS-puc-IR (V3108)*
- (d) *w¹¹¹⁸ hs-Flp/+; act>y+>GAL4 UAS-GFP/UAS-dTAK1*

Figure 8

- (b) *Spz6-GFP/+; GMR-GAL4/UAS-mRFP*
- (c) *Spz6-GFP/+; GMR-GAL4/+*
- (d) *Spz6-GFP/+; GMR-GAL4/UAS-Egr^{KB}*
- (e) *UAS-dTAK1/Spz6-GFP; GMR-GAL4/+*
- (f) *UAS-Hep^{CA}/Spz6-GFP; GMR-GAL4/+*

Figure 9

- (a) *GMR-GAL4/+*
- (b) *GMR-GAL4/UAS-LacZ*
- (c) *UAS-Toll^{10B}/+; GMR-GAL4/+*
- (d) *GMR-GAL4/UAS-Pelle*
- (e) *GMR-GAL4/UAS-cactus-IR* (NIG5848R3)
- (f) *UAS-Egr^W/+; GMR-GAL4/+*
- (g) *UAS-Egr^W/+; GMR-GAL4/UAS-LacZ*
- (h) *UAS-Egr^W/UAS-Toll^{10B}; GMR-GAL4/+*
- (i) *UAS-Egr^W/+; GMR-GAL4/UAS-Pelle*
- (j) *UAS-Egr^W/+; GMR-GAL4/UAS-cactus-IR* (NIG5848R3)

Figure S1

- (a) *UAS-Egr/+; GMR-GAL4/+*
- (b) *UAS-Egr/UAS-imd-IR* (NIG5576R2); *GMR-GAL4/+*
- (c) *UAS-Egr/UAS-rel-IR* (V49413); *GMR-GAL4/+*
- (d) *GMR-GAL4 UAS-Egr^{KB}/+*
- (e) *GMR-GAL4 UAS-Egr^{KB}/UAS-LacZ*
- (f) *imd¹/imd¹; GMR-GAL4 UAS-Egr^{KB}/+*
- (g) *GMR-GAL4 UAS-Egr/+*
- (h) *GMR-GAL4 UAS-Egr/+; UAS-LacZ/+*
- (i) *GMR-GAL4 UAS-Egr/+; rel^{E38}/rel^{E38}*

Figure S2

- (a) *ptc-GAL4/+; Tub-GAL80^{ts}/+*
- (b) *ptc-GAL4/UAS-Grim; Tub-GAL80^{ts}/+*

Figure S3

- (a) *Sd-GAL4/+*
- (b) *Sd-GAL4/+; UAS-Hep/+*

Figure S4

- (a and g) *Sd-GAL4/+*
- (b and h) *Sd-GAL4/+; UAS-Hep/+*
- (c and i) *Sd-GAL4/+; UAS-Hep/+; UAS-GFP/+*
- (d and j) *Sd-GAL4/+; UAS-Hep/+; UAS-Bsk^{DN}/+*
- (e and k) *Sd-GAL4/+; UAS-Hep/+; UAS-dorsal-IR (V45998)/+*

Figure S5

- (a) *UAS-Hid/+; GMR-GAL4/+*
- (b) *UAS-Hid/+; GMR-GAL4/+; UAS-dorsal-IR (V45998)/+*
- (c) *UAS-Hid/+; GMR-GAL4/+; UAS-Dif-IR (V30579)/+*

Figure S6

- (a) *Dipt-LacZ/+; Cg-GAL4/+*
- (b) *Dipt-LacZ/+; Cg-GAL4/UAS-Egr*
- (c) *Dipt-LacZ/+; Cg-GAL4/+; UAS-Imd/+*

Figure S7

- (a) *GMR-GAL4/+*
- (b) *UAS-dTAK1/+; GMR-GAL4/+*

Figure S8

- (a) *Cg-GAL4/+*
- (b) *Cg-GAL4/UAS-Egr^W*
- (c) *Cg-GAL4/UAS-puc-IR (V3108)*

Figure S9

- (a) *Drs-GFP/+; Cg-GAL4/UAS-puc-IR (V3108)*
- (b) *Drs-GFP/+; Cg-GAL4/UAS-puc-IR; UAS-P35/+*

- (c) $w^{1118} hs-Flp/+; act>y+>GAL4 UAS-GFP/UAS-dTAK1; UAS-P35/+$
- (d) $UAS-dTAK1/Spz6-GFP; GMR-GAL4/+$
- (e) $UAS-dTAK1/Spz6-GFP; GMR-GAL4/UAS-P35$
- (f) $UAS-dTAK1/Spz6-GFP; GMR-GAL4/UAS-Bsk^{DN}$
- (g) $GMR-GAL4/+$
- (h) $GMR-GAL4/UAS-Egr^{KB}$
- (i) $UAS-dTAK1/+; GMR-GAL4/+$

Figure S10

- (a) $GMR-GAL4/+$
- (b and e) $GMR-GAL4/UAS-Dorsal$
- (c and i) $GMR-GAL4/UAS-Dif$
- (f) $GMR-GAL4/UAS-Dorsal; UAS-LacZ/+$
- (g) $GMR-GAL4/UAS-Dorsal; UAS-Bsk^{DN}/+$
- (h) $GMR-GAL4/UAS-Dorsal; UAS-Puc/+$
- (j) $GMR-GAL4/UAS-Dif; UAS-LacZ/+$
- (k) $GMR-GAL4/UAS-Dif; UAS-Bsk^{DN}/+$
- (l) $GMR-GAL4/UAS-Dif; UAS-Puc/+$

Figure S11

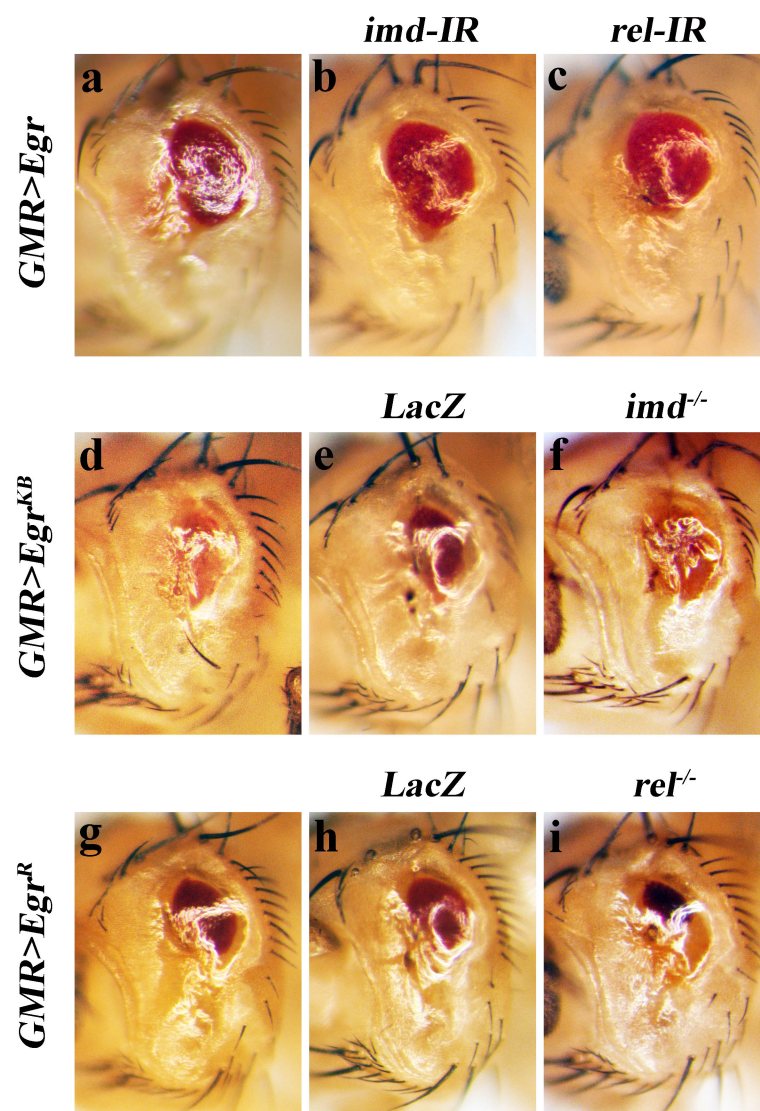
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Figure S12

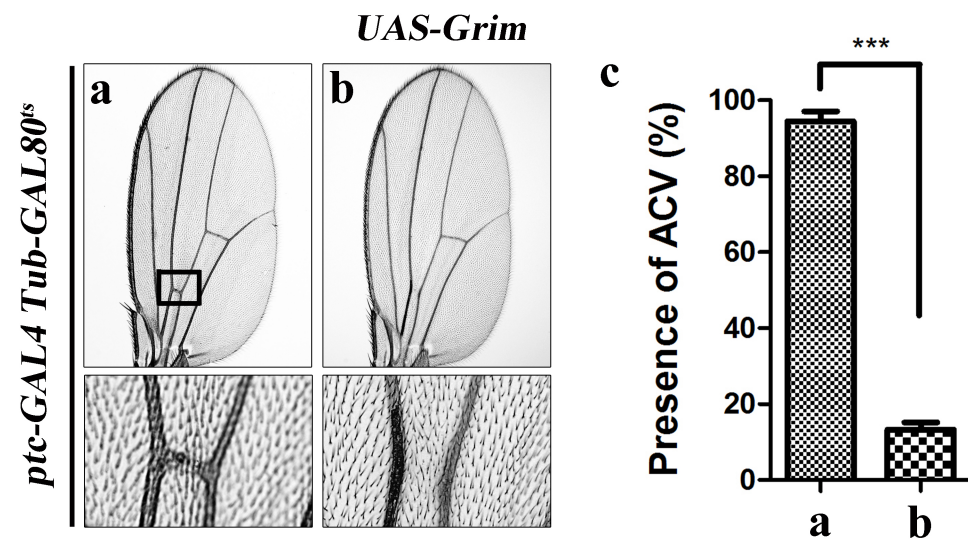
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- (c and k) $GMR-GAL4/UAS-Dif$
- (f) $GMR-GAL4/UAS-Dorsal; H99/+$
- (g) $GMR-GAL4/UAS-Dorsal; UAS-Dronc^{DN}/+$

- (h) *GMR-GAL4/UAS-Dorsal; UAS-P35/+*
- (i) *dTRAF2^{EX1.1}/+; GMR-GAL4/UAS-Dorsal*
- (j) *GMR-GAL4 UAS-Dorsal/UAS-dTRAF2-IR*
- (l) *GMR-GAL4/UAS-Dif; H99/+*
- (m) *GMR-GAL4/UAS-Dif; UAS-Dronc^{DN}/+*
- (n) *GMR-GAL4/UAS-Dif; UAS-P35/+*
- (o) *dTRAF2^{EX1.1}/+; GMR-GAL4/UAS-Dif*
- (p) *GMR-GAL4 UAS-Dif/UAS-dTRAF2-IR*

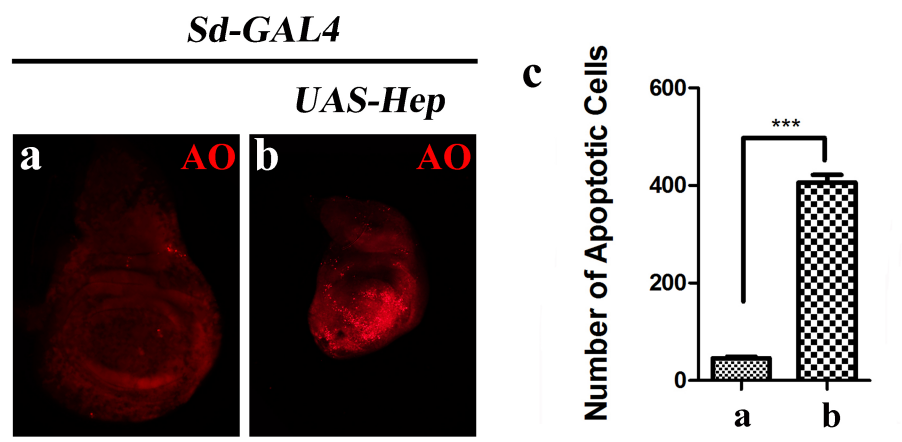
Wu et al., Figure S1

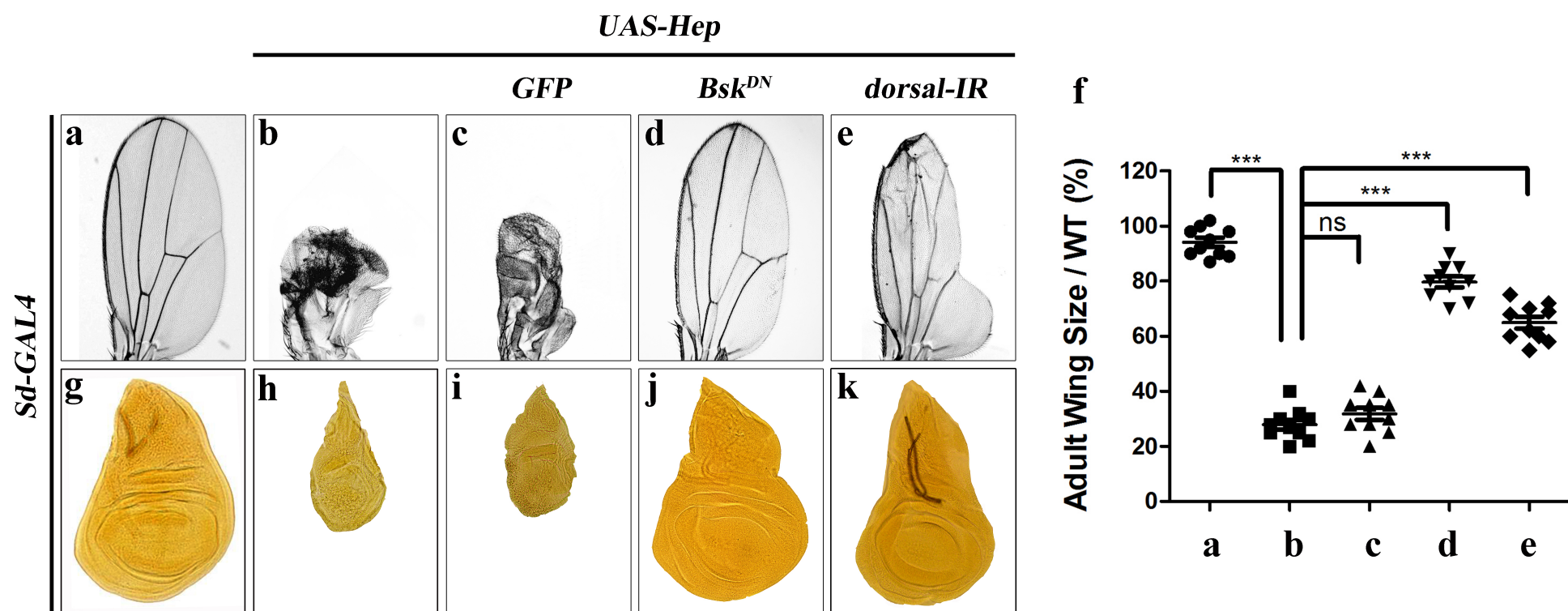


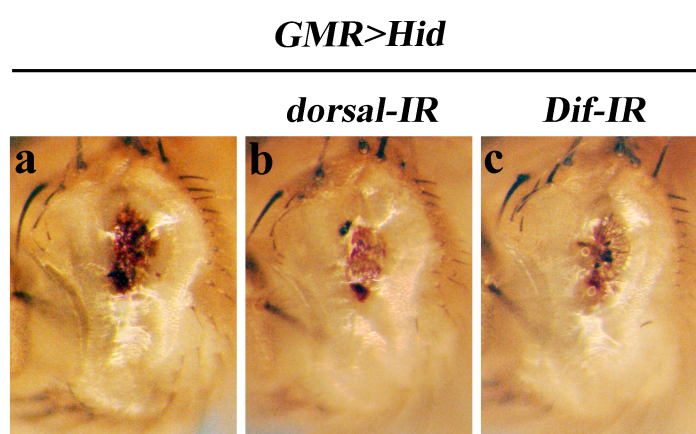
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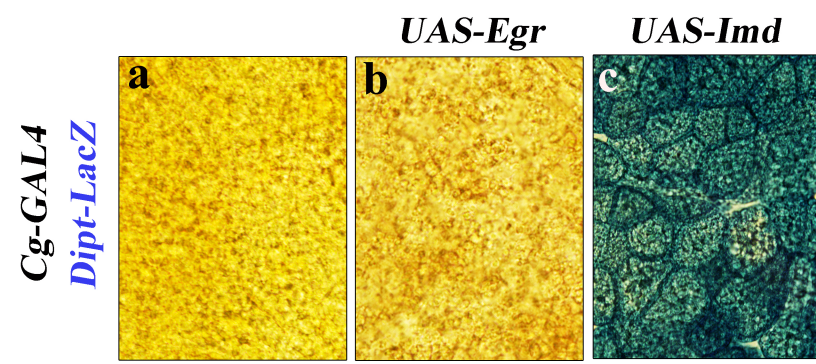
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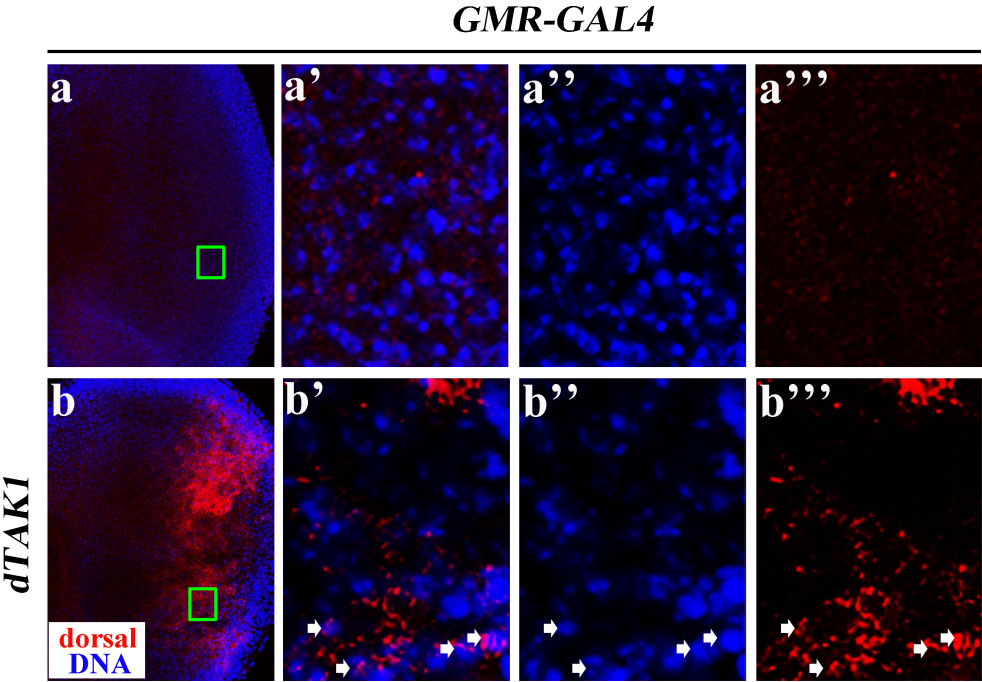


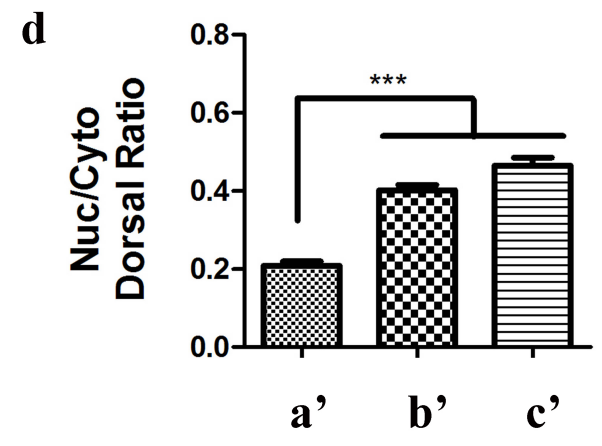
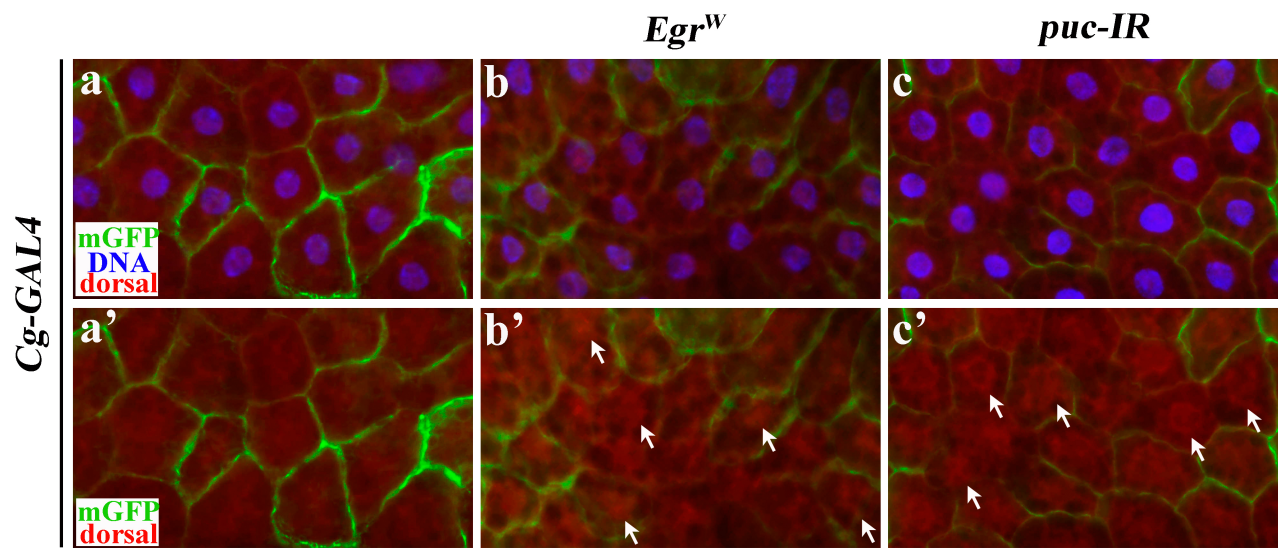




Wu et al., Figure S6







Wu et al., Figure S9

