Supplemental Information

The external PASTA domain of the essential serine/threonine protein kinase PknB regulates mycobacterial growth

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Content

Table S1. Plasmids and strains generated for over-expression studies.

 Table S2.
 Primers used in the study.

Table S3.Growth of *M. smegmatis* strains over-expressing *pkn*B variants.

Table S4. Muropeptides and sugars tested in TMP growth assays.

Table S5. List of proteins differently present in the membrane fractions of TMP and MIND (submitted as a separate excel file). Peptides from the TMP and MIND samples were labelled with different tandem mass tags. During MS/MS fragmentation each tag generates a reporter-ion with a different m/z value. The peaks areas of the reporter-ion can thus be compared to give a relative quantitative value. The data were normalized separately within each acquisition run. Intensities for each peptide identification were normalized within the assigned protein. The reference channel (MIND) was normalized to produce a 1:1 fold change. All normalization calculations were performed using medians to multiplicatively normalize data. The values shown thus correspond to the log₂ fold change for the TMP sample normalized against the MIND (reference) sample.

Figure S1. Figure S2. Figure S3.



Figure S1. Growth of *M. smegmatis* MIND and TMP strains in Sauton's medium supplemented with hygromycin and tetracycline after *in vitro* passage. Mycobacteria were grown in Sauton's medium to stationary phase and used for inoculation of microtitre plates as described in Materials and Methods.



Figure S2. Superimposition of [¹H, ¹⁵N] HSQC spectra from PknB_PASTA in 25 mM sodium acetate pH 4.6 in the presence (black) or absence (blue) of 25 mM MgCl₂, recorded at 37°C.



Figure S3. Analytical gel filtration chromatograms of PknB_PASTA (50 μ l at 50 μ M) with (blue) or without (red) 25 mM MgSO₄. Both profile are almost identical indicating that the protein is not subject to any major conformational changes or oligomerization in presence of a large excess of MgSO₄.