

## Supplementary tables

Genotype	Treatment	Leaf area (mm <sup>2</sup> )	Leaf number	Plant height (mm)	Fresh weight - FW (mg)	Dry weight - DW (mg)	RWC (%)	Water content (g g(DW) <sup>-1</sup> )	A (mM CO <sub>2</sub> plant <sup>-1</sup> h <sup>-1</sup> )	Fv/Fm	fPSII	qP	NPQ	SPAD
WT	Control	403 a	7.0 a	15.38 a	103.9 a	10.7 a	82.0 a	8.69 a	16.25 a	0.807 ab	0.446 a	0.951 ab	1.561 a	29.0 a
<i>h1.3</i>		365 a	6.5 a	5.38 b	89.5 a	9.3 a	77.4 b	8.45 a	14.06 a	0.817 ac	0.456 a	0.968 a	1.590 a	27.7 a
WT	Low light/ drought	62 b	5.4 b	6.50 b	8.5 b	0.8 b	84.3 a	9.69 ab	1.98 b	0.789 b	0.484 a	0.882 c	1.027 b	19.6 b
<i>h1.3</i>														
		142 c	6.3 ab	7.38 b	18.8 c	1.5 c	78.0 b	11.69 b	4.49 c	0.836 c	0.588 b	0.915 bc	0.931 c	20.6 b

**Table S1. Growth analysis and physiological parameters of wild-type (WT) and *h1.3* plants in the early-growth phase experiment.** Homogeneity groups are marked with letters (p<0.05, Tukey's HSD test).

Genotype	Treatment	Leaf area (cm <sup>2</sup> )	Leaf number	Plant height (cm)	Fresh weight - FW (mg)	Dry weight DW (mg)	RWC (%)	Water content (g g(DW) <sup>-1</sup> )	Pn (mM CO <sub>2</sub> plant <sup>-1</sup> h <sup>-1</sup> )	Fv/Fm	fPSII	qP	NPQ	SPAD
WT	Control	25 a	25 .9 a	19 .9 a	1797 a	190 a	84 .9 a	8 .52 abc	98 .35 a	0 .845 a	0 .670 abc	0 .949 ab	0 .731 a	39 .9 a
<i>h1.3</i>		24 a	30 .1 a	20 .3 a	1648 a	209 a	83 .2 a	8 .02 bc	63 .48 b	0 .841 abc	0 .684 a	0 .960 ac	0 .664 a	37 .5 ab
<i>h1.3 /H1.3</i>		26 a	29 .3 a	16 .6 b	1697 a	188 a	86 .0 a	9 .74 ab	99 .48 a	0 .834 c	0 .687 a	0 .976 c	0 .673 a	38 .1 ab
WT	Drought/low light	5 b	14 .4 b	10 .7 c	219 b	22 b	72 .3 b	7 .72 c	6 .75 c	0 .846 abc	0 .648 bcd	0 .923 d	0 .842 b	33 .5 b
<i>h1.3</i>		8 c	18 .9 c	7 .0 d	470 c	34 c	70 .2 c	7 .73 c	8 .52 d	0 .846 abc	0 .642 cd	0 .911 d	0 .708 a	40 .6 a
<i>h1.3 /H1.3</i>		5 b	14 .0 b	8 .8 cd	184 b	22 b	72 .9 b	8 .40 bc	5 .79 c	0 .839 abc	0 .627 d	0 .928 bd	0 .850 b	33 .2 b

**Table S2. Growth analysis and physiological parameters of wild-type (WT) and *h1.3* plants, and the *h1.3* mutant complemented with H1.3-GFP (*h1.3/H1.3*) in the late-growth phase experiment.** Homogeneity groups are indicated by letters (p<0.05, Tukey's HSD test).

<i>Observation</i>	<i>H1 subtype</i>	<i>p-value</i>
H1 enrichment on transposons compared to the whole genome	Main variants	$p < 10^{-167}$
	H1.3 LL	$10^{-27}$
H1 depletion on introns compared to the whole genome	Main variants	$p < 10^{-166}$
	H1.3 LL	$10^{-87}$
H1 depletion on 5'ends compared to whole genes	Main variants	$p < 10^{-230}$
	H1.3 LL	
H1 enrichment on exons compared to whole genes	Main variants	$p < 10^{-4}$
	H1.3 LL	0.009

**Table S3. Statistics for H1 enrichment and depletion at genic features.** Supplementary to Figure 5A and Figure S15.

	DNA methylation context		
	CG	CHG	CHH
<b>Mutant - WT</b>			
<b><i>h1.1h1.2h1.3</i></b>	[0.131, 0.151] <sup>a</sup> 2.009e-26 [-0.005, 0.05] <sup>c</sup> 2.154e-84 <sup>d</sup>	[0.111, 0.181] <sup>a</sup> 5.407e-256 <sup>b</sup> [-0.021, 0.12] <sup>c</sup> 0.0 <sup>d</sup>	[0.068, 0.097] <sup>a</sup> 1.570e-101 [-0.016, 0.012] <sup>c</sup> 4.966e-70 <sup>d</sup>
<b><i>h1.3</i> (control)</b>	[0.073, 0.091] <sup>a</sup> 2.483e-66 [-0.001, -0.005] <sup>c</sup> 0.005 <sup>d</sup>	[0.061, 0.102] <sup>a</sup> 9.906e-164 [-0.001, -0.013] <sup>c</sup> 1.041e-10 <sup>d</sup>	[0.039, 0.067] <sup>a</sup> 1.14e-121 [0.001, -0.002] <sup>c</sup> 0.003 <sup>d</sup>
<b><i>h1.3 (stress)</i></b>	[0.063, 0.081] <sup>a</sup> 2.75e-75 [-0.004, -0.001] <sup>c</sup> 0.072 <sup>d</sup>	[0.056, 0.092] <sup>a</sup> 1.318e-172 [-0.007, -0.005] <sup>c</sup> 0.161 <sup>d</sup>	[0.042, 0.067] <sup>a</sup> 1.252e-127 [-0.008, -0.008] <sup>c</sup> 0.808 <sup>d</sup>
<b>Response to stress (Stress - control)</b>			
<b>WT</b>	[0.052, 0.077] <sup>a</sup> 1.555e-134 [0.007, 0.012] <sup>c</sup> 4.853e-05 <sup>d</sup>	[0.054, 0.09] <sup>a</sup> 1.027e-170 [0.012, 0.020] <sup>c</sup> 3.929e-07 <sup>d</sup>	[0.040, 0.067] <sup>a</sup> 4.516e-121 [0.013, 0.028] <sup>c</sup> 1.281e-43 <sup>d</sup>
<b><i>h1.3</i></b>	[0.059, 0.083] <sup>a</sup> 1.216e-102 [0.004, 0.017] <sup>c</sup> 1.731e-17 <sup>d</sup>	[0.061, 0.101] <sup>a</sup> 2.563e-164 [0.007, 0.028] <sup>c</sup> 4.918e-33 <sup>d</sup>	[0.041, 0.069] <sup>a</sup> 4.188e-146 [0.004, 0.022] <sup>c</sup> 4.895e-58 <sup>d</sup>

**Table S4. Statistics for changes in methylation of euchromatic and heterochromatic transposable elements (TEs).** Euchromatic TEs – 6499 TEs with the lowest H3K9me2 level; heterochromatic TEs – 5188 TEs with the highest H3K9me2 level. Letters indicate the following: a – standard deviations, b – Levene (Brown-Forsythe) test p-value, c – mean values, d – T-test. The first value in brackets is for euchromatic TEs and the second is for heterochromatic TEs.

	DNA methylation context		
	CG	CHG	CHH
<b>Mutant - WT</b>			
<b><i>h1.1h1.2h1.3</i></b>	[0.059, 0.138] <sup>a</sup> 1.799e-159 <sup>b</sup> [0.0014, 0.0053] <sup>c</sup> 0.092 <sup>d</sup>	[0.047, 0.135] <sup>a</sup> 0.0 [0.0016, 0.0400] <sup>c</sup> 2.683e-81 <sup>d</sup>	[0.032, 0.075] <sup>a</sup> 5.226e-165 <sup>b</sup> [0.0010, -0.0040] <sup>c</sup> 6.924e-05 <sup>d</sup>
<b><i>h1.3</i> (control)</b>	[0.032, 0.076] <sup>a</sup> 9.687e-155 <sup>b</sup> [-0.0004, -0.0049] <sup>c</sup> 0.0008 <sup>d</sup>	[0.027, 0.072] <sup>a</sup> 1.118e-217 <sup>b</sup> [0.0000, -0.0028] <sup>c</sup> 0.016 <sup>d</sup>	[0.018, 0.049] <sup>a</sup> 4.204e-187 <sup>b</sup> [-0.0002, 0.0018] <sup>c</sup> 0.008 <sup>d</sup>
<b><i>h1.3</i> (stress)</b>	[0.028, 0.069] <sup>a</sup> 7.868e-152 <sup>b</sup> [-0.0028, -0.0021] <sup>c</sup> 0.566 <sup>d</sup>	[0.025, 0.067] <sup>a</sup> 6.684e-247 <sup>b</sup> [-0.0032, -0.0038] <sup>c</sup> 0.581 <sup>d</sup>	[0.019, 0.051] <sup>a</sup> 2.576e-174 <sup>b</sup> [-0.0031, -0.0067] <sup>c</sup> 3.154e-06 <sup>d</sup>
<b>Response to stress (Stress - control)</b>			
<b>WT</b>	[0.023, 0.053] <sup>a</sup> 1.387e-177 <sup>b</sup> [0.0041, 0.0063] <sup>c</sup> 0.018 <sup>d</sup>	[0.023, 0.063] <sup>a</sup> 2.373e-214 <sup>b</sup> [0.005, 0.01] <sup>c</sup> 1.716e-07 <sup>d</sup>	[0.019, 0.036] <sup>a</sup> 2.036e-58 <sup>b</sup> [0.0047, 0.0092] <sup>c</sup> 6.661e-16 <sup>d</sup>
<b><i>h1.3</i></b>	[0.026, 0.065] <sup>a</sup> 1.292e-228 <sup>b</sup> [0.0019, 0.0089] <sup>c</sup> 3.938e-11 <sup>d</sup>	[0.025, 0.071] <sup>a</sup> 1.088e-237 <sup>b</sup> [0.002, 0.009] <sup>c</sup> 5.322e-12 <sup>d</sup>	[0.019, 0.047] <sup>a</sup> 7.899e-182 <sup>b</sup> [0.0016, 0.0083] <sup>c</sup> 1.988e-18 <sup>d</sup>

**Table S5. Statistics for changes in methylation of euchromatic and heterochromatic genes.**

Euchromatic genes – 17,026 genes with the lowest H3K9me2 level; heterochromatic genes – 1308 genes with the highest H3K9me2 level. Letters indicate the following: a – standard deviations, b – Levene (Brown-Forsythe) test p-value, c – mean values, d – T-test. The first value in brackets is for euchromatic genes and the second is for heterochromatic genes.

<b>CG</b>	<b><i>h1.3</i></b>	<b>WT stress</b>	<b><i>h1.3</i> stress</b>
<b>WT</b>	0.0008	3.2593e-33	
<b><i>h1.3</i></b>			1.6916e-17
<b>WT stress</b>			8.9125e-12

<b>CHG</b>	<b><i>h1.3</i></b>	<b>WT stress</b>	<b><i>h1.3</i> stress</b>
<b>WT</b>	9.4357e-18	1.5502e-197	
<b><i>h1.3</i></b>			1.9566e-122
<b>WT stress</b>			1.0462e-49

<b>CHH</b>	<b><i>h1.3</i></b>	<b>WT stress</b>	<b><i>h1.3</i> stress</b>
<b>WT</b>	0.3106	<10e-256	
<b><i>h1.3</i></b>			<10e-256
<b>WT stress</b>			<10e-256

**Table S6. Statistics (T-test) for differences in the average global methylation level between genotypes and conditions in three different DNA methylation contexts.** Supplementary to Figure 8A.

Name	Sequence 5'→ 3'	Aim
156AW_L	GAAAACCACCACTCATCCTCCATACTTCA	<i>h1.3</i> genotyping
157AW_R	CCTTCTTCACTTCCCTCTCTTTCTTGTGTTG TTC	
Ds5-4_R	TACGATAACGGTCGGTACGG	<i>h1.1</i> genotyping
164AW_ins-L	TTCTTTGATTATAAGGGATTTGCCGATT T	
165AW_gen_R	CAGTCTTAGGTTAACGGCGATTGTTCTT	
166AW_gen_L	AAACGAATTTCATCGAGTATATCACGAGTT ATGG	
167AW_gen_L	CGTTACAATCGCATCTTAATCATCTGCAT C	<i>h1.2</i> genotyping
168AW_ins_R	CATCATACTCATTGCTGATCCATGTAGATT TCC	
169AW_gen_R	GCACACCAATAACAATAACACAAAAAGCACCT ATC	
13MP_H1.3_SalI_L	<u>GTCGACAAGTCATGCAGAACTCCATTG</u>	
14MP_H1.3_BamHI_R	<u>GGATCCAGCAGCGGAAGCTTCATG</u>	Cloning the promoter and coding region (without stop codon) of H1.3
19MP_H1.3utr_EcoRI_L	<u>GAATTCTGATCACTGAGGGAGGAGGAG</u>	Cloning the 3'UTR of H1.3
10MP_H1.3utr_NcoI_R	<u>CCATGGACGAGACTTGGGAGCAAGA</u>	qPCR, expression of H1.1
H1.1qPCR3-R	CCCTAAGGCGGCAGCGGAGA	
H1.1qPCR3-L	GCAGTGGCACGAGGAACGGG	qPCR, expression of H1.2
H1.2_qPCR_L	GTCTGTTGCTGCTGTTCCA	
H1.2_qPCR_R	CCGGAGCCTTCTTAGTCACA	qPCR, expression of H1.3
H1.3_qPCR_L	CGGATCAAGCCCTATGCTA	
H1.3_qPCR_R	CCTGCTGCCTCGTTATCATC	qPCR, expression of SPCH
ml83_SPCH_F	TAAGCCCAAGAAAACCGCCT	
ml84_SPCH_R	ATGAGTGGTAGTTGCGGTGG	qPCR, expression of MKK7/MK K9
ml85_MKK7/MKK9_F	ATTTCCCTTGCTTCCGCCG	
ml86_MKK7/MKK9_R	CGGGTTCTCCAAAACACACC	qPCR, expression of TMM
ml91_TMM_F	TAAGCCCAAGAAAACCGCCT	
ml92_TMM_R	ATGAGTGGTAGTTGCGGTGG	qPCR, expression of JAZ1
ml99_JAZ1_F	TTCGTGGTAGCCGGAGATT	
ml100_JAZ1_R	CGTGAGTTGCCTAAAGTTCCA	expression

		of JAZ1
ml101_EXPA11_F	GCCGCTCTTTATTGCGGT	qPCR, expression of EXPA11
ml102_EXPA11_R	GCTCCACCCATTGTTCCAGA	
ml105_CSLA03_F	AGTTGTGGCAGAGAAGAAAGC	qPCR, expression of CSLA03
ml106_CSLA03_R	CCACAGGACTTGAAACCAACC	
ml107_AGP21_F	CCAAGCCAACTTCTGATGC	qPCR, expression of AGP21
ml108_AGP21_R	ACCAGATGCCAAAGCAACAA	
ml109_WAKL10_F	GACATCCCGTACCCGTTGG	qPCR, expression of WAKL10
ml110_WAKL10_R	CCTACGTTAAAGAACCGGGCGA	
ml113_EXPA6_F	GGCAATGTTGGGCTTGGTT	qPCR, expression of EXPA6
ml114_EXPA6_R	ACCGTAGAAAGTGGCGTGAG	
ml115_FAD3_F	CATCAAACCCTTCCTCACAC	qPCR, expression of FAD3
ml116_FAD3_R	GTTCACATTGGTGCCTGGT	
ml117_ERF38_F	AGACGACGGAGAGAAGAGTG	qPCR, expression of ERF38
ml118_ERF38_R	TGGAGAAAGTACCGAGCCAG	
ml119_ANNAT4_F	ATACTGCGGTGGTATGTGG	qPCR, expression of ANNAT4
ml120_ANNAT4_R	TGTGCATGAGACCTAACGA	
ml121_CA1_F	CCTGCTCCTATCATTGCC	qPCR, expression of CA1
ml122_CA1_R	GCTGTGATCTGCTCCACCTT	
ml125_FAD5_F	CAGCGACGGAGGGAGATTAC	qPCR, expression of FAD5
ml126_FAD5_R	GCCCCAAAAGCAACCGAAACA	
ml129_CWLP_F	TTGCGACTGTACCCCTCCTA	qPCR, expression of CWLP
ml130_CWLP_R	AGGCGGTTGACTGGATGTT	
ml131_CSLG2_F	AAAAACCAGTTGCCTCCGT	qPCR, expression of CSLG2
ml132_CSLG2_R	CTGTATGGAATGGTTCGCCG	
ml133_FAD6_F	ACTCTCGCCTCCTACCACT	qPCR, expression of FAD6
ml134_FAD6_R	TCAAACCTCTGGCGGAAC	
ml135_HCF136_F	TCAATGGCGTCTGCAACT	qPCR, expression of HCF136
ml136_HCF136_R	ATTGGTACAGAAGCTCCGC	
ml137_G6PD1_F	AGGGCGTGGAGGTTACTTG	qPCR, expression of G6PD1
ml138_G6PD1_R	CCACATCTCAAGCCGCAAC	
ml139_RBCS3B_F	CGAGTTAGAGCACGGAAACACT	qPCR, expression of RBCS3B
ml140_RBCS3B_R	ATTGGACTTGACGGGTGTGT	
ml141_RBCS1B_F	GACAACACCCGTCAAGTCCA	qPCR,

ml142_RBCS1B_R	TGGGTTCCGGATAGTCAACATT	expression of RBCS1B
ml143_GAMT2_F	TACAACGGCGATACTGCC	qPCR, expression of GAMT2
ml144_GAMT2_R	CTCTGGGAAAAAGCCGGTCA	
ml145_ABI2_F	TCGCTGTTCCATTAGACCA	qPCR, expression of ABI2
ml146_ABI2_R	CGCCAGAACAAAGAACTTCCG	
ml147_AGP1_F	TCTTCCGCAGTGGCTCAATC	qPCR, expression of AGP1
ml148_AGP1_R	GAGCCGGTGAGATCCGTCTA	
ml149_ANNAT2_F	GTGCGTCTGCTTCGTCTCA	qPCR, expression of ANNAT2
ml150_ANNAT2_R	TGAGAGACGCCATTGTTGGG	
ml151_XTH6_F	GTTCATTGGGTTCCGCTC	qPCR, expression of XTH6
ml152_XTH6_R	ATGTGAGATTCCGACCAGGC	
ml155_COR27_F	GGTAACGGATGAGCCCCAAA	qPCR, expression of COR27
ml156_COR27_R	CAACGCTGATTGGTCGTGG	
ml157_COR15B_F	GCCAATGAAACTGCGACTGA	qPCR, expression of COR15B
ml158_COR15B_R	TGAATCAGGACTTGTGGCA	
ml161_AT4G30650_F	GGCGAGCAACATGGAAGTTTT	qPCR, expression of AT4G30650
ml162_AT4G30650_R	TGCAACAGCCACGTTGAGA	
ml163_MUTE_L	CGAAAGACCCTAAACCGACCA	qPCR, expression of MUTE
ml164_MUTE_R	GGTAGTGGCGGCTCCTAAAC	
ml165_CYCA2,3_L	CCCCAAAGAGCGGATTCAAGG	qPCR, expression of CYCA2,3
ml166_CYCA2,3_R	TGGCCGAGACACAGCATTTC	
183KR_FAMA_qPCR_L	AGACTGCGGAGAACAAAGTCG	qPCR, expression of FAMA
184KR_FAMA_qPCR_R	TCTTAATCAGCTGTCCGGC	
207KR_ERECTA_qPCR_L	CGTGGAACGTAACCGGAGAA	qPCR, expression of ERECTA
208KR_ERECTA_qPCR_R	ATGATCGATGCTTGGTCCCC	
ml1_UBC_F	CTGCGACTCAGGGAATCTTCTAA	qPCR, expression of UBC
ml2_UBC_R	TTGTGCCATTGAATTGAACCC	

**Table S7. Sequences of oligonucleotide primers used in this study.** Restriction enzyme sites are underlined.