

Figure S1: 81% of the total protein present in Bb Lyme antigen Grade 2 (American Research Products) is constituted by OspA. A dilution of a recombinant purified OspA protein (Genecopoeia) was analyzed by SDS PAGE and silver staining (100-to 1,000 ng, *Lanes 3-7*). The band intensities were quantified using ImageJ software and a calibration curve was built (insert graph). A linear regression curve was fitted (y=5.39x+12281, R² = 0.9873). 500 ng of Bb Lyme antigen Grade 2 were loaded on the gel (*Lane 2*) and compared to the recombinant OspA calibration curve (*Lanes 3-7*). From this procedure it derived that 81% of Bb Lyme antigen Grade 2 total protein content is OspA. In gel protein digestion and mass spectrometry (MS) analysis verified the predominant presence of tryptic peptides belonging to OspA in the bands at ~30 kDa and ~60kDa present both in the Bb Lyme antigen Grade 2 and in the recombinant OspA. In particular, tryptic peptides containing the epitope for mAb clone 0551 OspA236-239 were present in both bands (MS-MS spectra are reported in the insert).

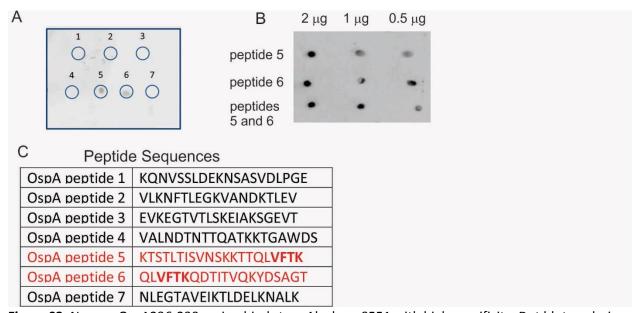


Figure S2. Narrow OspA236-239 region binds to mAb clone 0551 with high specificity. Dot blot analysis revealed that only 2 out of 7 synthetic peptides with sequence reported in Table S1 show reactivity with the anti OspA monoclonal antibody clone 0551 used in this study.

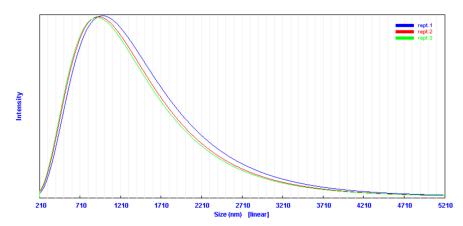


Figure S3. Light scattering analysis was used to determine the hydrodynamic diameter of the Nanotrap particles. As reported in Table S2, the hydrodynamic diameter of the Nanotrap particles was 1054.7 +/- 31.11 nm.

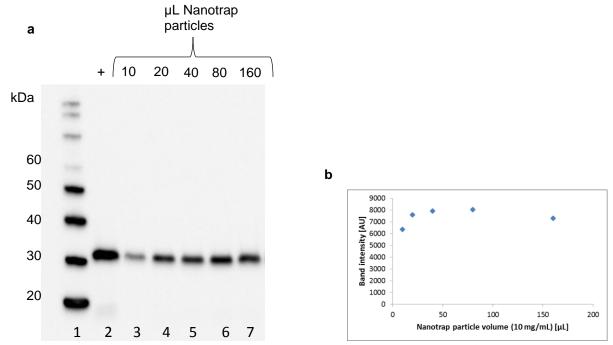


Figure S4. A V/v ratio (V=Nanotrap suspension volume, v=sample volume) of 1/10 was optimized in order to maximize Lyme antigen capturing. **a** 2 ng of Lyme antigen was spiked in 500 μL urine aliquots. Urine samples were incubated with increasing amount of Nanotrap particle suspension (10-160 μL of Nanotrap at 5 mg/mL concentration). *Lanes* 1) ladder; 2) Positive control (OspA 1ng); 3) 10μL of Nanotrap particles; 4) 20 μL Nanotrap particles; 5) 40μL Nanotrap particles; 6) 80 μL Nanotrap particles; 7) 160μL Nanotrap particles; **b** Band intensity was measured with ImageJ, plateau is reached with > 40μL of Nanotrap particles.

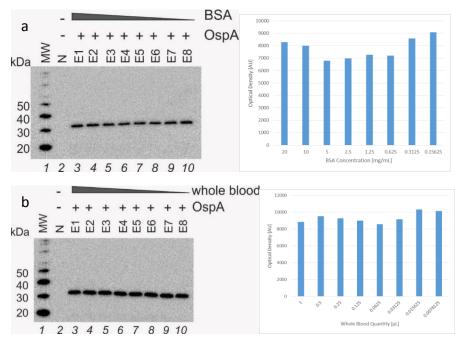


Figure S5. Interfering substances: the presence of a high amount of protein and blood in the urine does not interfere with Lyme antigen capture and detection. **a** 320 pg of Lyme antigen was spiked in samples of 40 mL of human urine. We tested the interference of albumin present in excess up to 10^8 fold. Increasing amounts of bovine serum albumin ranging from 0.31 mg/mL to 20 mg/mL were added to 40 mL of OspA containing human urine; urine samples were processed with Nanotrap particles. Ability of the Nanotrap particles to sequester OspA is not affected by increasing concentration of competing proteins in urine. *Lanes*: 1) ladder; 2) volunteer human urine in absence of OspA antigen (negative control); 3) OspA + BSA 20 mg/mL; 4) OspA + BSA 10 mg/mL; 5) OspA + BSA 5 mg/mL; 6) OspA + BSA 2.5 mg/mL; 7) OspA + BSA 1.25 mg/mL; 8) OspA + BSA 0.625 mg/mL; 9) OspA + BSA 0.31 mg/mL; 10) OspA + BSA 0.15 mg/mL. **b** Lyme antigen 320 pg was spiked in urine samples (40 mL). Increasing amounts of whole blood from 0.015 μL to 1 μL was added to the urine samples; urine samples were processed with Nanotrap particles and analyzed using western blot. Lanes: 1) ladder; 2) volunteer human urine in absence of OspA antigen (negative control); 3) OspA + 1 μL whole blood; 4) OspA + 0.5 μL whole blood; 5) OspA + 0.25 μL whole blood; 6) OspA + 0.125 μL whole blood; 7) OspA + 0.062 μL whole blood; 8) OspA + 0.031 μL whole blood; 9) OspA + 0.015 μL whole blood; 10) OspA + 0.007 μL whole blood.

Note: Bilirubin, which we screen against in the urine against in the urine dipstick prior to analysis, does not cause any interference with the Nanotrap based test for the following two reasons. Firstly, the Nanotrap particle urine OspA test uses chemiluminescence solid phase based detection whereas bilirubin is known to interfere with absorbance readings at $\lambda \sim 456$ nm [Clin Biochem Rev Vol 29 Suppl (i) August 2008 I S43] in solution-phase homogeneous assays. Secondly, the molecular weight of bilirubin is ~500 Da. This means that bilirubin is too small to be retained in the SDS PAGE system (Tris Gly 4-20%) we use to detect the OspA antigen. Bilirubin would migrate out of the gel during the electrophoretic run necessary to separate the proteins, which have a much higher molecular weight (10,000 – 100,000 Da). The presence of abnormally high proteins in the urine of patients, as happens in the in the rare disorder of monoclonal gammopathy (3% incidence in general population >50 yo, decreasing with decreasing age), does not interfere with the Nanotrap OspA test as demonstrated in

Figure S5. The exquisite specificity of the test is ensured by the specificity of the anti OspA monoclonal antibody and by the two-step test that includes a competition assay for each positive result. This competition assures that immunoreactivity with the antigen can be clearly differentiated from the rare possibility of background caused by immunoglobulins in the urine.

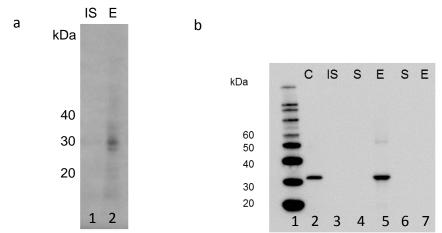


Figure S6. a Nanotrap particle preprocessing step is necessary to detect an OspA specific band in the urine of patient PD113, clinically positive for Lyme disease. Initial solution (IS) before Nanotrap particle processing. Eluate (E) after Nanotrap particle processing. **b** Positive and negative controls run with all Lyme patient samples. *Lane 2* contains borrelia protein lysate (2 ng) in human urine. *Lane 5* is a positive control 4 ng spiked in 40 mL of urine sample. *Lane 6* and 7 are negative controls of 40 mL of volunteer urine processed through the Nanotrap particles. C=borrelia lysate control, IS=initial solution, S=supernatant, E=eluate.

Table S1. Sequences of peptides tested for antibody binding.

Peptide ID	Sequence	Reactivity with mAb 0551
OspA peptide 1	KQNVSSLDEKNSASVDLPGE	Negative
OspA peptide 2	VLKNFTLEGKVANDKTLEV	Negative
OspA peptide 3	EVKEGTVTLSKEIAKSGEVT	Negative
OspA peptide 4	VALNDTNTTQATKKTGAWDS	Negative
OspA peptide 5	KTSTLTISVNSKKTTQL VFTK	Positive
OspA peptide 6	QL VFTK QDTITVQKYDSAGT	Positive
OspA peptide 7	NLEGTAVEIKTLDELKNALK	Negative
OspA219-253	KTSTLTISVNSKKTTQL VFTK QDTITVQKYDSAGT	Positive
OspA219-235	KTSTLTISVNSKKTTQL	Negative
OspA240-253	QDTITVQKYDSAGT	Negative

Table S2. The mAb epitope used herein (red rectangle) is conserved in common pathogenic species of *Borrelia*. BAA22342.1 in *Borrelia garinii* Taxonomy ID 29519, ADD14639.1 in *Borrelia burgdorferi* taxonomy ID 139, WP_012665647.1 in *Borrelia valaisiana* taxonomy ID 62088, WP_012665647.1 in *Borrelia* sp. SV1 taxonomy ID 498741 YP_003110622.1 in *Borrelia burgdorferi* 297 taxonomy ID 521009, NP_045688.1 in *Borrelia burgdorferi* B31 taxonomy ID 224326, WP_014023199.1 in *Borrelia bissettii* taxonomy ID 64897, ADG02035.1 in *Borrelia afzelii* taxonomy ID 29518, AAN65460.1 in *Borrelia spielmanii* taxonomy ID 88916. BLAST analysis was performed on the sequence KTSTLTISVNSKTTQLVFTKQDTITVQKYDSAGT (combination of peptide 5 and 6) with the following organisms: *Treponema pertenue* (taxonomy ID 168), *Leptospiraceae* (taxonomy ID 170), *Treponema* (taxonomy ID 157), *Spirochaetes* (taxonomy ID 203691) excluding *Borrelia* (taxonomy ID 138), *Homo sapiens* (taxonomy ID 9606), Epstein-Barr virus EBV (taxid:10376), Human cytomegalovirus (taxid:10359), herpes simplex virus 1 HSV-1 (taxid:10298), hepatitis C virus HCV (taxid:11103), Babesia txid5864, Anaplasma txid768, Ehrlichieae txid942, Bartonella txid773, Rickettsias txid766. No significant similarity found. No homology was identified.

BAA22342.1	1	LGIGLILALIACKQNVSSLDEKNSVSVDLPGEMKVLVSKEKDKDGKYSLMA
ADD14639.1	1	ILALIACKONVSSLDEKNSVSVDLPGEMKVLVSKEKNKDGKYDLIA
WP 012672372.1	1	MKKYLLGIGLILALIACKONVSSLDEKNSVSVDLPGEIKVLVSKEKNKDGKYSLMA
WP 012665647.1	1	MKKYLLGIGLILALIACKQNVSSLDEKNSASVDLPGEMKVLVSKEKDKDGKYSLVA
YP 003110622.1	1	MKKYLLGIGLILALIACKONVSSLDEKNSVSVDLPGEMKVLVSKEKNKDGKYDLIA
NP_045688.1	1	MKKYLLGIGLILALIACKQNVSSLDEKNSVSVDLPGEMKVLVSKEKNKDGKYDLIA
WP_014023199.1	1	MKKYLLGIGLILALIACKQNVSGLDEKNSVSVDLPGEMKVLVSKEKDKDGKYSLMA
ADG02035.1	1	ALIACKQNVSSLDEKNSASVDLPGEMKVLVSKEKDKDGKYSLKA
AAN65460.1	1	ILALIACKQNVSGLDEKNSTSVDVPGELKVLVSKEKDKDGKYSLMA
BAA22342.1	52	TVDKLELKGTSDKSNGSGILEGVKTDKSKAKLTISDDLSKTTFEVFKEDGKTLVSR
ADD14639.1	47	TVDKLELKGTSDKNNGSGVLEGVKADKSKVKLTISDDLGQTTLEVFKEDGKTLVSK
WP 012672372.1	57	TVDKLELKGTSDKNNGSGVLEGVKADKSKVKLTVSDDLGOTTLEVLKEDGKTLVSR
	57	
WP_012665647.1		TVDKVELKGTSDKNNGSGTLEGVKDDKSKVKLTISDDLGETKLETFKEDG-TLVSR
YP_003110622.1	57	TVDKLELKGTSDKNNGSGVLEGVKADKSKVKLTISDDLGQTTLEVFKEDGKTLVSK
NP_045688.1	57	TVDKLELKGTSDKNNGSGVLEGVKADKSKVKLTISDDLGQTTLEVFKEDGKTLVSK
WP 014023199.1	57	TVDKLELKGTSDKNNGSGILEGVKADKSKVKLTVSEDLSTTTLEVLKEDGKTLVSK
ADG02035.1	45	TVDKIELKGTSDKDNGSGVLEGTKDDKSKAKLTIADDLSKTTFELFKEDGKTLVSR
AAN65460.1	47	TVDKLELKGTSDKNDGSGVLEGVKADKSKVKLTISDHLSKTTFEVFKEDGKTLVSR
		1,5/1010101010100001000101011101110111101
BAA22342.1	108	KVNSKDKSSTEEKFNAKGELSEKVVTRANGNRLEYTEI-KSDGSGKAKEVLKDFTL
ADD14639.1	103	KVTSKDKSSTEEKFNEKGEVSEKIITRADGTRLEYTEI-KSDGSGKAKEVLKG-YV
WP 012672372.1	113	KVTSKDKSSTEEKFNEKGELAEKIMTRANGTRLEYTEI-KSDGSGKAKEVLKD-YV
WP_012665647.1	112	KVNFKDKSFTEEKFNEKGEVSEKILTRSNGTTLEYSQMtDAENATKAVETLKNGIK
YP_003110622.1	113	KVTSKDKSSTEEKFNEKGEVSEKIITRADGTRLEYTEI-KSDGSGKAKEVLKG-YV
NP 045688.1	113	KVTSKDKSSTEEKFNEKGEVSEKIITRADGTRLEYTGI-KSDGSGKAKEVLKG-YV
WP 014023199.1	113	KTTSKDKSSTEEKFNDKGELAEKTIVRANGTRLEYTEV-KSDGSGKAKETLKD-YA
ADG02035.1	101	KVSSKDKTSTDEMFNEKGELSAKTMTRENGTKLEYTEM-KSDGTGKAKEVLKN-FT
AAN65460.1	103	NVNSKDKSSTKEKFNEKGELSEKTLVRANGTKLEYTEI-KSDGTGKAKEVLKD-FT
1111100100.1	100	NVNONDNOOTNENTIVENOODEEENTEVIVINOTNEETTET NOOTORENEVEND TT
BAA22342.1	163	LEGTLTADKTTLTIQEGSVTLSKNIAKSGEITVELNDTDSS-GDKKSGOWNSSTST
ADD14639.1	157	LEGTLTAEKTTLVVKEGTVTLSKNISKSGEVSVELNDTDSSAATKKTAAWNSGTST
WP 012672372.1	167	LEGTLTAEKTTLVVKEGTVTLSKHISKSGEVTAELNDTESSSATKKTAAWNSGTST
WP_012665647.1	168	LPGNLVGGKTTLKITEGTVTLSKHIAKSGEVTVEINDTSSTPNTKKTGKWDARNST
YP_003110622.1	167	LEGTLTAEKTTLVVKEGTVTLSKNISKSGEVSVELNDTDSSAATKKTAAWNSGTST
NP_045688.1	167	LEGTLTAEKTTLVVKEGTVTLSKNISKSGEVSVELNDTDSSAATKKTAAWNSGTST
WP 014023199.1	167	LEGTLTAEKATLVVKEGTVTLSKHISKSGEVTAELNDTDSAQATKKTGKWDAGTST
ADG02035.1	155	LEGKVANDKVTLEVKEGTVTLSKEIAKSGEVTVALNDTNTTQATKKTGAWDSKTST
AAN65460.1	157	LEGTLANEKATLTVKEGTVTLSKNIDKSGEVTVALNDTDSTAATKKTGAWDSKTST
BAA22342.1	218	LTISAKSKKTKDIVFTKODTITVQKYDSAGTNLEGSAVEIKTLDELKNALK
ADD14639.1	213	LTITVNSKKTKDIVFTKENTITVQQYDSNGTKLEGSAVEITKLDEIKNALR
WP 012672372.1	223	LTITVNSKKTKDIVFTKENTITVQKYDTAGTNLEGSAVEIKKLDELKNALK
WP_012665647.1	224	LTIIVDSKNKTKIVFTKODTITVOSYNPAGNKLEGTAVEIKTLOELKNALK
YP_003110622.1	223	LTITVNSKKTKDIVFTKENTITVQQYDSNGTKLEGSAVEITKLDEIKNALK
NP_045688.1	223	LTITVNSKKTKDIVFTKENTITVQQYDSNGTKLEGSAVEITKLDEIKNALK
WP_014023199.1	223	LTISVNSKKTKNIVFTKODTITVQKYDSAGTNLEGTAVEIKTLDELKNALK
ADG02035.1	211	LTISVNSKKTTQIVFTK DTITVQKYDSAGTNLEGTAVEIKTLDELKNA
AAN65460.1	213	LTITVNSKKTKDIVFTKODTITVQKYDSAGTNLEGSAVEIK

Table S3. Light scattering analysis of Nanotrap particles

Rept#	Mean [nm]	P.I.	Diff.Coef	Counts/s	Baseline
			[m²/s]		error
1	1097.4	-1.065	4.47e-13	1.02e+5	4.50%
2	1042.8	-1.230	4.70e-13	1.01e+5	1.21%
3	1024.0	-1.422	4.79e-13	9.95+4	0.33%
Average	1054.7 +/-	-1.239 +/-			
	31.11	0.146			

Table S4. Quantification of the amount of Remazol brilliant blue (RBB) dye covalently bound to the Nanotrap particles and percentage of reacted acrylic acid (AAc) moles. This information was obtained and recorded for every batch of produced Nanotrap particles (example batch # RM37B4 is reported here).

Parameter	Numerical value	Unit of Measure
Weight of 20 mL of freezed	110	mg
dried poly(NIPAm-co-AAc)		
Nanotrap*		
Weight of 20 mL of freeze	153	mg
dried poly(NIPAm/RBB)		
Nanotrap		
Δ weight	43	mg
Molar quantity of RBB	0.0686	mmol
covalently bound to the		
Nanotrap		
Molar quantity of AAc in the	0.2430	mmol
poly(NIPAm-co-AAc)		
Nanotrap		
Percentage of mol RBB	28	%
bound to AAc / tot mol AAc		

^{*}Note: The weight is relative to the poly(NIPAm-co-AAc) Nanotrap before RBB covalently binding.

Table S5. Clinical and diagnostic information of patients suspected of having early stage Lyme disease N=51 (N= 117 healthy volunteers were recruited under informed consent and included in the study).

Treatment: Dx = doxycycline, Pd = prednisone, RC = rocepherin, sv = synovectomy, st = steroids, Am = amoxil, Zm = Zithromax. Urine collection timing and presence of symptoms: B = before treatment, PT = during or after treatment, symptoms present at the time urine was collected.

Patient	t, symptoms present Clinical	Treatment	Serology results	Urine	OspA urine
ID#	manifestation	readment	(ELISA, IgG, IgM) N=negative, P=positive, ND=not done	collection timing	test (N=negative; P=positive confirmed by competition)
1	joint pain, malaise, neck pain, headache, nausea, dizziness	Dx	N, ND, ND	PT	N
3	EM, bell's palsy, myalgia	Dx, Pd	P, N, N	PT	Р
8	myalgia, fever, joint pain	Dx, Rc, Sv	P, P, N	PT	Р
11	EM, Bell's palsy, joint pain	Dx, Rc	N, ND, ND	PT	Р
13	EM, fever, joint pain, neck pain, fatigue	Dx	P, N, P	PT	P
14	joint pain	St, Dx	P, P, ND	PT	Р
15	EM, myalgia, fever, joint pain, malaise, fatigue	Rc, Pc	P, N, P	PT	P
17	joint pain	Dx, Rc	P, P, N	PT	Р
18	EM, fever, joint pain	Dx	P, P, P	PT	Р
21	headaches	Dx	N, N, N	PT	Р
23	EM, fever, anthralgiatis	Am	P, P, P	В	Р
24	EM, fever, anthralgias	Zm	P, N, P	PT	Р
25	EM, fever, neck pain, fatigue	Dx	P, N, P	В	Р
26	EM, anthralgiatis	Dx, Rc, Am	N, N, N	PT	Р
27	EM, fatigues, anthralgiatis	Dx, Zm	ND, ND, ND	PT	Р
102	EM		N, ND, ND	PT	Р
103	fatigue, joint pain, pos for Lyme on lumber puncture	Dx	P, P, N	PT	Р
103.1	No symptoms	Rc	P, P, N	-	N
105	joint pain, arthritis	Dx	P, N, N	PT	Р

108	EM, fever, joint pain	Dx	P, N, P	В	Р
108.1	no symptoms	Dx	P, N, P	-	N
113	EM, fever, neck pain, fatigue		P, N, P	PT	Р
116	joint paint, fatigue		N, ND, ND	-	N
117	Tick bite		N, NA	В	Р
118	EM fatigue		P, P, P	В	Р
119	joint pain, arthritis, fatigue, neurologic	Dx	P, N, P	PT	P
120	EM, fever, malaise	Dx	P, N, P	В	Р
120.1	No symptoms	Dx	P, N, P	-	N
121	EM	Dx	N, NA, NA	В	Р
133	joint pain, fever, fatigue		N, N, N	-	N
139	joint pain, fatigue	Dx	P, N, P	В	Р
139.1	No symptoms		P, N, P	-	Р
142	rash joint pain		P, N, P	В	Р
148	joint pain, fatigue		P, P, P	В	Р
151	fatigue	Dx	P, N, N	PT	Р
169	EM, neck stiffness, migraine		Equivocal	В	Р
180	EM, tick bite	Dx	N, N, N	В	Р
180.1	no symptoms	Dx	P, N, P	-	N
601	EM	Az	Equiv	В	Р
601.1	no symptoms	Az	Equiv	-	N
602	EM	Dx	N, ND, ND	В	Р
603	EM	Dx	P, P, P	В	Р
604	EM	Dx	ND	В	Р
605	EM	Dx	ND	В	Р
606	EM	Dx	ND	В	Р
607	EM	Dx	Equiv	В	Р
608	EM	Dx	ND	В	Р
623	EM	Dx	N, ND, ND	В	Р
623.1	EM	Dx	N, ND, ND	-	Р
623.2	EM	Dx	N, ND, ND	-	N
623.3	EM	Dx	N, ND, ND	-	N
623.4	EM	Dx	N, ND, ND	-	N
623.5	EM	Dx	N, ND, ND	-	N

Table S6: Post treatment patients being evaluated for recurrent or persistent disseminated Lyme disease derived from a Lyme endemic geographic region.

Patient ID# Clinical Manifestation (IgG, IgM) (N=negative; P=positive confirmed by competition) 300 Neurocognitive ND N 306 Neurologic ND N 309 Neurologic ND N 310 Joint pain ND N 315 Joint pain ND N 318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 321 ND N 322 ND N 323 ND N 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND N 329 Joint pain ND N 331 neurologic ND N
P=positive confirmed by competition) 300 Neurocognitive ND N 306 Neurologic ND N 309 Neurologic ND N 310 Joint pain ND N 315 Joint pain ND P 318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 321 ND N 322 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND N 329 Joint pain ND N
confirmed by competition) 300 Neurocognitive ND N 306 Neurologic ND N 309 Neurologic ND N 310 Joint pain ND N 315 Joint pain ND P 318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 321 ND P 322 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND N 329 Joint pain ND N
300NeurocognitiveNDN306NeurologicNDN309NeurologicNDN310Joint painNDN315Joint painNDP318Joint painNDN319Bladder painNDN320NeurologicNDN323NDP324asymptomaticNDN327Bladder painNDN328Bladder painNDP329Joint painNDN
300NeurocognitiveNDN306NeurologicNDN309NeurologicNDN310Joint painNDN315Joint painNDP318Joint painNDN319Bladder painNDN320NeurologicNDN323NDP324asymptomaticNDN327Bladder painNDN328Bladder painNDP329Joint painNDN
306NeurologicNDN309NeurologicNDN310Joint painNDN315Joint painNDP318Joint painNDN319Bladder painNDN320NeurologicNDN323NDP324asymptomaticNDN327Bladder painNDN328Bladder painNDP329Joint painNDN
309 Neurologic ND N 310 Joint pain ND N 315 Joint pain ND P 318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 323 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND P 329 Joint pain ND N
310 Joint pain ND N 315 Joint pain ND P 318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 323 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND P 329 Joint pain ND N
318 Joint pain ND N 319 Bladder pain ND N 320 Neurologic ND N 323 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND P 329 Joint pain ND N
319 Bladder pain ND N 320 Neurologic ND N 323 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND P 329 Joint pain ND N
320NeurologicNDN323NDP324asymptomaticNDN327Bladder painNDN328Bladder painNDP329Joint painNDN
323 ND P 324 asymptomatic ND N 327 Bladder pain ND N 328 Bladder pain ND P 329 Joint pain ND N
324asymptomaticNDN327Bladder painNDN328Bladder painNDP329Joint painNDN
327Bladder painNDN328Bladder painNDP329Joint painNDN
327Bladder painNDN328Bladder painNDP329Joint painNDN
329 Joint pain ND N
331 neurologic ND N
333 Bladder pain ND N
334 Joint pain ND P
336 Fatigue ND N
337 Joint pain ND P
339 neurologic ND P
340 Neurologic ND P
341 Joint pain ND P
343 Myalgia ND N
344 Neurolocognitive ND N
346 EM N,P P
347 Neurologic ND P
348 Neurologic ND N
349 Neurocognitive ND P
350 Neurologic ND P
353 Neurologic ND N
355 Neurologic ND N
356 Neurologic ND P
360 Joint pain ND P
361 fatigue ND N
363 Joint pain ND N
364 Joint pain ND N
377 Joint pain ND N
378 Neurologic ND N
379 Neurologic ND P
380 Fatigue ND P
381 Joint pain ND P

382	Joint pain	ND	Р
383	Neurologic	ND	Р
384	Joint pain	N,N	Р
386	Joint pain	ND	N
388	Joint pain, fatigue	ND	Р
389	Joint pain	ND	P
390	Joint pain	N,N	P
391	Joint pain	N, P	N
392	Joint pain	N, N	N
393	Joint pain	ND ND	P
394	Joint pain	N, N	N
397	Joint pain	ND ND	N
400	EM	N, N	N
401	LIVI	ND ND	N
402		ND	N
402		ND	N
404		ND	N
406	Neurocognitive	ND	N
400	Joint pain		P
	Joint Pain	N,N	P
410		N,N	
413	latal and	ND	N
414	Joint pain	ND	N
416		ND	N
417		ND	N
419		ND	P
421		ND	N
424	Neurologic	N,N	N
425		N,N	P
426		ND	N
427	EM, joint pain	N,N	Р
429	Neurologic	ND	N
430	Neurocognitive	ND	N
431	Neurologic	ND	N
433	Joint pain	ND	N
435	Neurocognitive	ND	N
437		ND	Р
438	Joint pain	ND	Р
439	Joint pain	ND	Р
443	Myocarditis	ND	Р
446	Neurocognitive	ND	N
452	Myalgia	ND	N
458	Joint pain	ND	N
459		ND	N
465		ND	N
466	Joint pain	ND	N
469	1		
			P

478		ND	N
487	Neurocognitive,	ND	N
	Fatigue		
488	Neurocognitive,	ND	Р
	Neurological		
489	Joint pain, Fatigue	ND	Р
490		ND	Р
491		ND	N
492	Fatigue	ND	Р
493	Neurocognitive	ND	N
495	Neurocognitive	ND	Р
497		ND	Р
498	Neurocognitive	ND	N
499		ND	N
500		ND	N
501	Joint pain	ND	Р

- 1. Casjens SR, Fraser-Liggett CM, Mongodin EF, Qiu WG, Dunn JJ, Luft BJ, Schutzer SE: **Whole genome sequence of an unusual Borrelia burgdorferi sensu lato isolate.** *J Bacteriol* 2011, **193:**1489-1490.
- 2. Stanek G, Reiter M: **The expanding Lyme Borrelia complex--clinical significance of genomic species?** *Clin Microbiol Infect* 2011, **17:**487-493.