

FIGURES

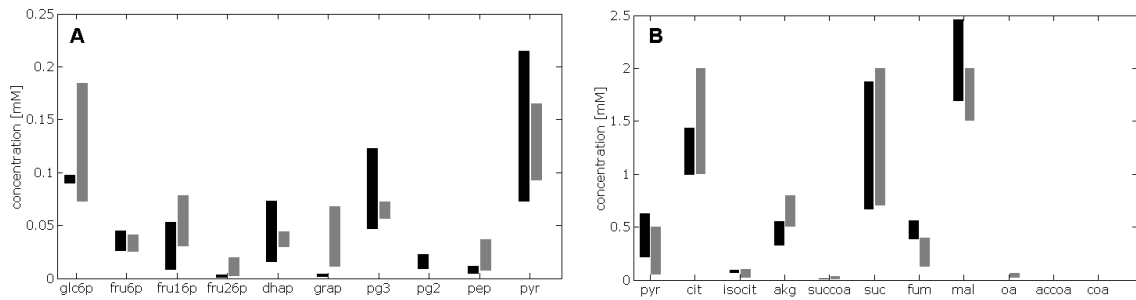


Figure S1 Calculated and measured concentrations of glycolytic metabolites (A) and intermediates of the TCA cycle (B).

Grey bars indicate experimentally determined concentration ranges taken from the literature (for glycolytic intermediates¹⁻¹³ and TCA-cycle intermediates^{5, 14-20}). Black bars indicate calculated metabolite concentration ranges for a normal energetic load (non-excited but spontaneously firing neuron) and in a highly activated state of the neuron (e.g. interneurons involved in gamma oscillations in hippocampal slices) where the activation is about 4-fold of the normal.²¹

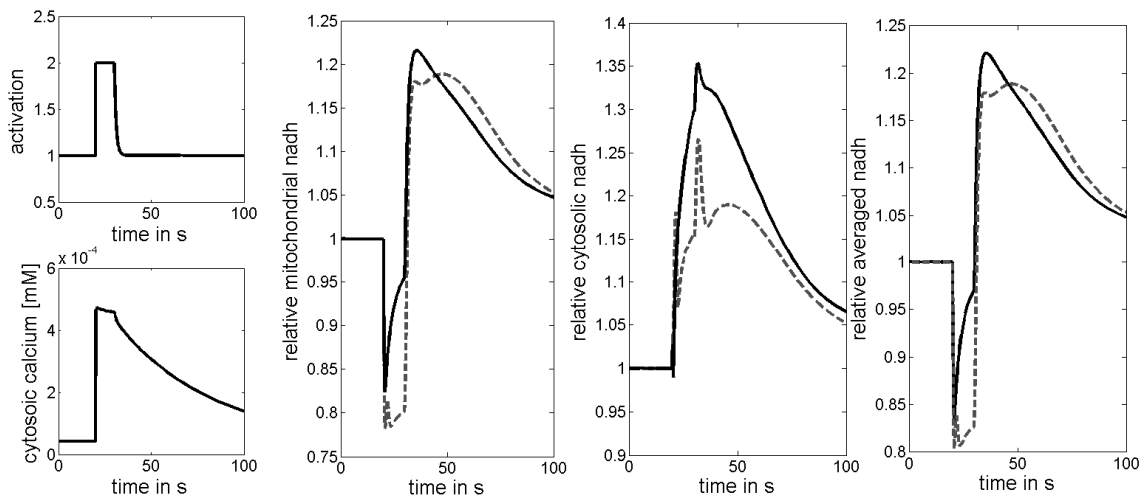


Figure S2 Comparison of the response of a more glycolytic cell (astrocyte) to a neuron with identical activation and cytosolic calcium transients

Dashed lines show astrocytic NADH transients, while solid lines show the NADH transients for the neuron. The relative glycolytic response is lower in the astrocyte due to the higher baseline level (not shown). However the abrupt changes in the energy demand are very pronounced in the astrocytic cytosolic NADH transients as the absolute contribution of the glycolytic pathway is higher and the buffering through the shuttle systems and the mitochondrial NADH pool is lower. Mitochondrial traces are similar, but the activation is lower in the astrocyte since the respiratory and citric acid cycle are closer to their maximal capacity in the basal state. The averaged transients (as measured by fluorescence) are very similar.

MODEL EQUATIONS

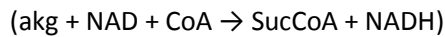
Kinetic rate equations for the individual enzymes were constructed on the basis of kinetic data gathered from the literature. Guiding principle was the construction of a comprehensive rate law that captures the regulatory features of the enzyme relevant to the chosen network and the intended purpose of the model. The example of the α -ketoglutarate dehydrogenase reaction below illustrates the strategy applied to select the relevant experimental information from the literature to establish a rate law that can be used as a module of the network model. First, the enzyme name and the literature used for the retrieval of kinetic parameters are given. Second, the stoichiometry of the reaction catalyzed by the enzyme is depicted. Third, the implemented rate equation and the numerical values of the kinetic parameters are given. Binding constants (K values) are given in mM, enzyme activity is given in mM/s.

The α -ketoglutarate dehydrogenase complex (KGDHC) catalyzes the irreversible reaction: $\text{akg} + \text{NAD} + \text{CoA} \rightarrow \text{SucCoA} + \text{NADH}$. The rate equation is of the Michaelis-Menten type with respect to reactants akg, NAD, and CoA. The K_m values are dynamic functions of metabolite concentrations. Activity of the enzyme is affected by the substrates akg, NAD and CoA while other metabolites as SucCoA, NADH, Ca, Mg, pH, ATP, ADP, P, NH_4 , various keto-acyl CoA esters act as allosteric regulators. Included in the rate equation

were only effectors that occur as variables of the network model. Effectors that are not part of the network model like Mg, NH_4^+ or keto-acyl CoA esters were omitted. Further reduction of the set of included effectors was achieved by taking into account the known metabolic alterations in the physiological regime considered. Under physiological conditions, the metabolites ATP, ADP, P as well as the proton concentration (pH value) do not exert regulatory control and were omitted as well. K_m values were either taken directly from the literature or obtained by fitting the rate law to published data. It was checked that used K_m values were consistent with values found in various publications. V_{\max} values for all enzymes are free parameters. They are adjusted to reproduce the correct systems behavior under the various conditions considered. They do not directly represent catalytic activity of the enzymes but absorb all activity influencing regulations considered to be constant within the model. For the KGDH, the V_{\max} value takes into account the effect of Mg, acyl esters, pH and otherwise unknown or neglected effectors.

Example:

α -ketoglutarate dehydrogenase complex^{16, 22-26}



$$v_{akdhc} = V_{\max}^{akdhc} \left(\frac{akg_{mito}}{akg_{mito} + K_m^{akdhc}} \right) \left(\frac{NAD_{mito}}{NAD_{mito} + K_m^{NAD} \cdot \left(1 + \frac{NADH_{mito}}{K_i^{NADH}} \right)} \right) \left(\frac{CoA}{CoA + K_m^{CoA} \cdot \left(1 + \frac{SucCoA}{K_i^{SucCoA}} \right)} \right)$$

$$V_{\max}^{akdhc} = 268.8$$

$$K_m^{akdhc} = \left(\frac{K_{m1}^{akdhc}}{\left(1 + \frac{Ca_{mito}}{K_{i-akg}^{Ca}} \right)} + K_{m2}^{akgh} \right) \left(1 + \frac{NADH_{mito}}{K_{i-akg}^{NADH}} \right)$$

$$K_{m1}^{akdhc} = 2.5$$

$$K_{m2}^{akdhc} = 0.13$$

$$K_m^{NAD} = 0.021$$

$$K_i^{NADH} = 0.0045$$

$$K_m^{CoA} = 0.0013$$

$$K_i^{SucCoA} = 0.0045$$

Glycolysis

Glucose transporter²⁷

(glc_ex ↔ glc_cyt)

$$v_{glt} = V_{\max-glt} \left(\frac{glc_{ext} - glc}{1 + \frac{glc}{K_m^{glc}} + \frac{glc_{ext}}{K_m^{glc_{ext}}}} \right)$$

$$V_{\max-glc} = 0.72$$

$$K_m^{glc} = 2.87$$

$$K_m^{glc_{ext}} = 2.87$$

Hexokinase²⁸⁻³¹

(Glc + ATP → Glc6P + ADP)

$$v_{hexk} = V_{\max-hexk} \left(\frac{glc}{glc + K_m^{glc}} \right) \left(\frac{atp}{atp + K_m^{atp} \left(1 + \frac{glc6p}{K_{i-atp}^{glc6p}} \right)} \right) \left(1 - \frac{glc6p}{K_i^{glc6p}} \right)$$

$$V_{\max-hexk} = 9.36$$

$$K_m^{glc} = 0.043$$

$$K_m^{atp} = 0.37$$

$$K_{i-atp}^{glc6p} = 0.074$$

$$K_i^{glc6p} = 0.1$$

Glucose-6-phosphate isomerase^{32,33}

(Glc6P ↔ Fru6P)

$$v_{g6piso} = V_{\max-g6piso} \left(\frac{glc6p - \frac{fru6p}{K_{eq-g6piso}}}{1 + \frac{glc6p}{K_m^{glc6p}} + \frac{fru6p}{K_m^{fru6p}}} \right)$$

$$V_{\max-g6piso} = 24.4$$

$$K_{eq-g6piso} = 0.5157$$

$$K_m^{glc6p} = 0.593$$

$$K_m^{fru6p} = 0.095$$

Phosphofructokinase I^{7,11,34}

(Fru6P + ATP → Fru16P + ADP)

$$v_{pfl} = V_{\max-pfl} \left(\frac{fru6p}{fru6p + K_m^{fru6p} \cdot \left(1 - K_0 \frac{fru26p^{n_{fru26p}}}{fru26p^{n_{fru26p}} + (K_a^{fru26})^{n_{fru26p}}} \right)} \right) \left(\frac{atp}{atp + K_m^{atp}} \right) \left(1 - \frac{atp^n}{atp^n + (K_i^{atp})^n} \right) \left(\frac{fru26p}{fru26p + K_a^{fru26p}} \right)$$

$$V_{\max-pfkI} = 49.6$$

$$K_m^{fru6p} = 0.111$$

$$K_m^{atp} = 0.04$$

$$n = 1.8$$

$$K_i^{atp} = 1.2$$

$$K_0 = 0.55$$

$$K_a^{fru26p} = 0.0042$$

$$n_{fru26p} = 5.5$$

$$K_a^{fru26p} = 0.005$$

Fructose-1,6-bisphosphatase³⁵

(fru16bp → fru6p + Pi)

$$v_{fbpI} = V_{\max-fbpI} \left(\frac{fru6p}{fru6p + K_m^{fru6p}} \right)$$

$$V_{\max-fbpI} = 0.455$$

$$K_m^{fru6p} = 0.132$$

Phosphofruktokinase II³⁶⁻⁴¹

(Fru6P + ATP → Fru26P + ADP)

$$v_{pfkII} = V_{\max-pfkII} \left(\frac{fru6p}{fru6p + K_m^{fru6p}} \right) \left(\frac{atp}{atp + K_m^{atp}} \right) \left(\frac{amp}{amp + K_a^{amp}} \right) \left(\frac{adp}{adp + K_a^{adp}} \right)$$

$$V_{\max-pfkII} = 0.0026$$

$$K_m^{fru6p} = 0.027$$

$$K_m^{atp} = 0.055$$

$$K_a^{amp} = 0.073$$

$$K_a^{adp} = 0.056$$

Fructose-2,6-bisphosphatase^{36, 37}

(Fru26P → Fru6P + Pi)

$$v_{fru26pp} = V_{\max-fru26pp} \left(\frac{fru26p}{fru26p + K_m^{fru26p} \left(1 + \frac{fru6p}{K_i^{fru6p}} \right)} \right)$$

$$V_{\max-fru26pp} = 0.052$$

$$K_m^{fru26p} = 0.07$$

$$K_i^{fru6p} = 0.02$$

Aldolase⁴²

(Fru16P ↔ Grap + Dhap)

$$v_{aldo} = V_{\max-aldo} \left(\frac{fru16p - \frac{grap \cdot dhap}{K_{eq}^{aldo}}}{\left(1 + \frac{fru16p}{K_m^{fru16p}} \right) + \left(1 + \frac{grap}{K_m^{grap}} \right) \left(1 + \frac{dhap}{K_m^{dhap}} \right) - 1} \right)$$

$$V_{\max-aldo} = 46.8$$

$$K_{eq}^{aldo} = 0.0976$$

$$K_m^{fru16p} = 0.003$$

$$K_m^{grap} = 0.08$$

$$K_m^{dhap} = 0.03$$

Triosephosphate isomerase⁴³

(Dhap \leftrightarrow Grap)

$$v_{tpi} = V_{\max-tpi} \left(\frac{dhap - \frac{grap}{K_{eq}^{tpi}}}{1 + \frac{dhap}{K_m^{dhap}} + \frac{grap}{K_m^{grap}}} \right)$$

$$V_{\max-tpi} = 10^6$$

$$K_{eq}^{tpi} = 0.0545$$

$$K_m^{dhap} = 0.84$$

$$K_m^{grap} = 1.65$$

Glyceraldehyde 3-phosphate dehydrogenase⁴⁴⁻⁴⁶

(Grap + Pi + NAD \rightarrow Bpg13 + NADH)

$$v_{gapdh} = V_{\max-gapdh} \left(\frac{nad \cdot grap \cdot pi - \frac{bpg13 \cdot nadh}{K_{eq}^{gapdg}}}{\left(1 + \frac{nad}{K_m^{nad}}\right) \left(1 + \frac{grap}{K_m^{grap}}\right) \left(1 + \frac{pi}{K_m^{pi}}\right) + \left(1 + \frac{nadh}{K_m^{nadh}}\right) \left(1 + \frac{bpg13}{K_m^{bpg13}}\right) - 1} \right)$$

$$V_{\max-gapdh} = 72000$$

$$K_{eq}^{gapdh} = 0.0868$$

$$K_m^{nad} = 0.01$$

$$K_m^{nad} = 0.027$$

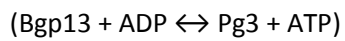
$$K_m^{grap} = 0.101$$

$$K_m^{pi} = 3.9$$

$$K_m^{nadh} = 0.008$$

$$K_m^{bpg13} = 0.0035$$

Phosphoglycerate kinase⁴⁷



$$v_{pgk} = V_{\max-pgk} \left(\frac{bpg13 \cdot adp - \frac{pg3 \cdot atp}{K_{eq}^{pgk}}}{\left(1 + \frac{bpg13}{K_m^{bpg13}}\right) \left(1 + \frac{adp}{K_m^{adp}}\right) + \left(1 + \frac{pg3}{K_m^{pg3}}\right) \left(1 + \frac{atp}{K_m^{atp}}\right) - 1} \right)$$

$$V_{\max-pg} = 396$$

$$K_{eq}^{pgk} = 1310$$

$$K_m^{bpg13} = 0.063$$

$$K_m^{adp} = 0.42$$

$$K_m^{pg3} = 0.67$$

$$K_m^{atp} = 0.25$$

Phosphoglycerate mutase^{48, 49}

(Pg3 \leftrightarrow Pg2)

$$v_{pgm} = V_{\max-pgm} \left(\frac{pg3 - \frac{pg2}{K_{eq}^{pgm}}}{\left(1 + \frac{pg3}{K_m^{pg3}}\right) + \left(1 + \frac{pg2}{K_m^{pg2}}\right) - 1} \right)$$

$$V_{\max-pgm} = 14400$$

$$K_{eq}^{pgm} = 0.1814$$

$$K_m^{pg3} = 0.22$$

$$K_m^{pg2} = 0.28$$

Enolase⁵⁰

(Pg2 \leftrightarrow Pep)

$$v_{eno} = V_{\max-eno} \left(\frac{pg2 - \frac{pep}{K_{eq}^{eno}}}{\left(1 + \frac{pg2}{K_m^{pg2}}\right) + \left(1 + \frac{pep}{K_m^{pep}}\right) - 1} \right)$$

$$V_{\max-eno} = 216000$$

$$K_{eq}^{eno} = 0.5$$

$$K_m^{pg2} = 0.05$$

$$K_m^{pep} = 0.15$$

Pyruvate kinase^{51, 52}

(Pep + ADP → Pyr + ATP)

$$v_{pk} = V_{\max-pk} \left(\frac{pep}{pep + K_m^{pep}} \right) \left(\frac{adp}{adp + K_m^{adp} \left(1 + \frac{atp}{K_i^{atp}} \right)} \right)$$

$$V_{\max-pk} = 23.76$$

$$K_m^{pep} = 0.074$$

$$K_m^{adp} = 0.42$$

$$K_i^{atp} = 4.4$$

Lactate dehydrogenase⁵³⁻⁵⁵

(Pyr + NADH ↔ Lac + NAD)

$$v_{ldh} = V_{\max-ldh} \left(\frac{pyr \cdot nadh - \frac{lac \cdot nad}{K_{eq}^{ldh}}}{\left(1 + \frac{pyr}{K_m^{pyr}} \right) \left(1 + \frac{nadh}{K_m^{nadh}} \right) + \left(1 + \frac{lac}{K_m^{lac}} \right) \left(1 + \frac{nad}{K_m^{nad}} \right) - 1} \right)$$

$$V_{\max-ldh} = 100000$$

$$K_{eq}^{ldh} = 8400$$

$$K_m^{pyr} = 0.36$$

$$K_m^{nadh} = 0.043$$

$$K_m^{lac} = 4.2$$

$$K_m^{nad} = 0.088$$

Monocarboxylate transporter (MCT)^{56, 57}

(Lac_cyt \leftrightarrow Lac_ex)

$$v_{mct} = V_{\max-mct} \left(\frac{lac - \frac{lac_{ex}}{K_{eq}^{mct}}}{\left(1 + \frac{lac}{K_m^{lac}}\right) + \left(1 + \frac{lac_{ex}}{K_m^{lac_{ex}}}\right) - 1} \right)$$

$$V_{\max-mct} = 5$$

$$K_{eq}^{mct} = \frac{h_{cyt}}{h_{ext}} = 1.737$$

$$K_m^{lac} = 1.1$$

$$K_m^{lac_{ex}} = 1.1$$

Creatine kinase⁴

(ATP + Cr \leftrightarrow ADP + CrP)

$$v_{ck} = V_{\max-ck} \left(adp \cdot crp - \frac{atp \cdot cr}{K_{eq-app}^{ck}} \right)$$

$$V_{\max-ck} = 0.0135$$

$$K_{eq-app}^{ck} = 7$$

The apparent equilibrium constant is lower than the thermodynamic equilibrium constant⁵⁸ as is known for muscle cells.⁵⁹

Malate-Aspartate Shuttle

Cytosolic malate dehydrogenase⁶⁰⁻⁶²

(Mal_in + NAD_in ↔ Oa_in + NADH_in)

$$v_{mdh}^{cyt} = V_{\max-mdh}^{cyt} \left(\frac{mal \cdot nad - \frac{oa \cdot nadh}{K_{eq}^{mdh}}}{\left(1 + \frac{mal}{K_m^{mal}}\right) \left(1 + \frac{nad}{K_m^{nad}}\right) + \left(1 + \frac{oa}{K_m^{oa}}\right) \left(1 + \frac{nadh}{K_m^{nadh}}\right) - 1} \right)$$

$$V_{\max-mdh}^{cyt} = 10^4$$

$$K_{eq}^{mdh} = 10^{-4}$$

$$K_m^{nad} = 0.05$$

$$K_m^{mal} = 0.77$$

$$K_m^{oa} = 0.04$$

$$K_m^{nadh} = 0.05$$

Mitochondrial malate dehydrogenase

(Mal_mito + NAD_mito ↔ Oa_mito + NADH_mito)

→ see section CAC

Cytosolic aspartate aminotransferase⁶³

(Asp_cyt + akg_cyt ↔ oa_cyt + glu_cyt)

$$v_{aat}^{cyt} = V_{\max-aat}^{cyt} \left(\frac{asp \cdot akg - \frac{oa \cdot glu}{K_{eq}^{aat}}}{\left(1 + \frac{asp}{K_m^{asp}}\right) \left(1 + \frac{akg}{K_m^{akg}}\right) + \left(1 + \frac{oa}{K_m^{oa}}\right) \left(1 + \frac{glu}{K_m^{glu}}\right) - 1} \right)$$

$$V_{\max-aat}^{cyt} = 32$$

$$K_{eq}^{aat} = 0.147$$

Mitochondrial aspartate aminotransferase⁶³

(Asp_mito + ak_g_mito \leftrightarrow oa_mito + glu_mito)

$$v_{aat}^{mito} = V_{\max-aaat}^{mito} \left(asp \cdot ak_g - \frac{oa \cdot glu}{K_{eq}^{aat}} \right)$$

$$V_{\max-aaat}^{mito} = 32$$

$$K_{eq}^{aat} = 0.147$$

Aspartate/glutamate carrier⁶⁴

(Asp_mito + glu_cyt + h_cyt \leftrightarrow Asp_cyt + glu_mito + h_mito)

$$v_{asp-glu-c.} = V_{\max-asp-glu-c.} \left(\frac{asp_{mito} \cdot glu_{cyt} - \frac{asp_{cyt} \cdot glu_{mito}}{K_{eq}^{asp-glu-c.}}}{(asp_{mito} + K_m^{asp_{mito}})(glu_{cyt} + K_m^{glu_{cyt}}) + (asp_{cyt} + K_m^{asp_{cyt}})(glu_{mito} + K_m^{glu_{mito}})} \right)$$

$$V_{\max-asp-glu-c.} = 3.2 \cdot 10^3$$

$$dG_{asp-glu-c} = -V_{mm} + \frac{1000 \cdot R \cdot T}{F} \log \left(\frac{h_{cyt}}{h_{mito}} \right)$$

$$K_{eq}^{asp-glu-c.} = \exp \left(\frac{F \cdot dGp}{1000 \cdot R \cdot T} \right)$$

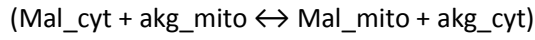
$$K_m^{asp_{mito}} = 0.05$$

$$K_m^{glu_{cyt}} = 2.8$$

$$K_m^{asp_{cyt}} = 0.05$$

$$K_m^{glu_{mito}} = 2.8$$

Malate/ α -ketoglutarate carrier^{65, 66}



$$v_{\text{mal-akg-c.}} = V_{\text{max-mal-akg-c.}} \left(\frac{\text{mal}_{\text{cyt}} \cdot \text{akg}_{\text{mito}} - \text{mal}_{\text{mito}} \cdot \text{akg}_{\text{cyt}}}{\left(\text{mal}_{\text{cyt}} + K_m^{\text{mal}_{\text{cyt}}} \right) \left(\text{akg}_{\text{mito}} + K_m^{\text{akg}_{\text{mito}}} \right) + \left(\text{mal}_{\text{mito}} + K_m^{\text{mal}_{\text{mito}}} \right) \left(\text{akg}_{\text{cyt}} + K_m^{\text{akg}_{\text{cyt}}} \right)} \right)$$

$$V_{\text{max-mal-akg-c.}} = 32$$

$$K_m^{\text{mal}_{\text{cyt}}} = 1.36$$

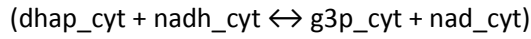
$$K_m^{\text{akg}_{\text{cyt}}} = 0.1$$

$$K_m^{\text{mal}_{\text{mito}}} = 0.71$$

$$K_m^{\text{akg}_{\text{mito}}} = 0.2$$

Glycerol-3-Phosphate Shuttle

Cytosolic glycerol-3-phosphate dehydrogenase^{67, 68}



$$v_{\text{g3pdh}}^{\text{cyt}} = V_{\text{max-g3pdh}}^{\text{cyt}} \left(\frac{\text{dhap}_{\text{cyt}} \cdot \text{nadh}_{\text{cyt}} - \frac{\text{g3p}_{\text{cyt}} \cdot \text{nad}_{\text{cyt}}}{K_{\text{eq-g3pdh}}^{\text{cyt}}}}{\left(1 + \frac{\text{dhap}_{\text{cyt}}}{K_m^{\text{dhap}_{\text{cyt}}}} \right) \cdot \left(1 + \frac{\text{nadh}_{\text{cyt}}}{K_m^{\text{nadh}_{\text{cyt}}}} \right) + \left(1 + \frac{\text{g3p}_{\text{cyt}}}{K_m^{\text{g3p}_{\text{cyt}}}} \right) \cdot \left(1 + \frac{\text{nad}_{\text{cyt}}}{K_m^{\text{nad}_{\text{cyt}}}} \right) - 1} \right)$$

$$V_{\text{max-g3pdh}}^{\text{cyt}} = 3.2 \cdot 10^4$$

$$K_{\text{eq-g3pdh}}^{\text{cyt}} = 3257.3$$

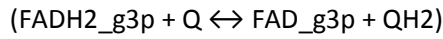
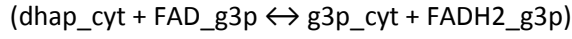
$$K_m^{\text{dhap}} = 0.17$$

$$K_m^{\text{nadh}} = 0.01$$

$$K_m^{\text{g3p}} = 0.3$$

$$K_m^{nad} = 0.03$$

Mitochondrial glycerol-3-phosphate dehydrogenase^{19, 67, 68}



$$v_{g3pdh}^{mito} = V_{max-g3pdh}^{mito} \left(\frac{dhap_{cyt} \cdot FAD_{g3pdh} - \frac{g3p_{cyt} \cdot FADH_{2g3pdh}}{K_{eq-g3pdh}^{mito}}}{\left(1 + \frac{dhap_{cyt}}{K_m^{dhap}}\right) + \left(1 + \frac{g3p_{cyt}}{K_m^{g3p}}\right)} \right)$$

$$v_{g3pdh-Q} = V_{max}^{g3pdh-Q} \cdot \left(FADH_{2pdhc} \cdot Q_{mito} - \frac{1}{K_{eq-g3p-fad-Q}} FAD_{pdhc} \cdot QH_{2mito} \right)$$

$$V_{max-g3pdh}^{cyt} = 6.4 \cdot 10^4$$

$$V_{max}^{g3pdh-Q} = 3.2 \cdot 10^6$$

$$K_{eq-g3pdh}^{mito} = \exp\left(\frac{(2 \cdot Em_{dhap/g3p} - 2 \cdot Em_{FAD_{g3p}}) \cdot F}{1000 \cdot R \cdot T}\right)$$

$$K_{eq-g3-fad-Q}^{mito} = \exp\left(\frac{(2 \cdot Em_{FAD_{g3p}} + 2 \cdot Em_Q) \cdot F}{1000 \cdot R \cdot T}\right)$$

$$Em_{dhap/g3p} = 190mV$$

$$Em_{FAD_{g3p}} = 210mV$$

Oxidative Phosphorylation

ATP synthetase³³

(ATP \leftrightarrow ADP + Pi)

$$v_{syn} = V_{max}^{syn} \cdot \left(ADP_{mito} \cdot P_{mito} - \frac{ATP_{mito}}{K_{eq-syn}} \right)$$

$$V_{max}^{syn} = 1.8 \cdot 10^{-16} (dG^n)$$

$$dG = -V_{mm} + \frac{R \cdot T}{1000 \cdot F} \ln \left(\frac{h_{cyt}}{h_{mito}} \right)$$

$$n = 3$$

$$K_{eq-syn} = \exp \left(\frac{dG_0^{syn}}{R \cdot T} - k \cdot U \right) \cdot \left(\frac{h_{cyt}}{h_{mito}} \right)^k$$

$$dG_0^{syn} = 30500$$

$$k = 3$$

The dG containing term describes activation by the proton motive force.

ADP/ATP exchanger⁶⁹

(ATP_mito + ADP_cyt \leftrightarrow ADP_mito + ATP_cyt)

$$v_{ATP-exchanger} = V_{max}^{ATP-exchanger} \cdot \left(\frac{1 - \frac{ATP_{in} \cdot ADP_{mito}}{ADP_{in} \cdot ATP_{mito}} \exp(U)}{\left(1 + \frac{ATP_{in}}{ADP_{in}} \exp(S_{V_{mm}} \cdot U) \left(1 + \frac{ADP_{mito}}{ATP_{mito}} \right) \right)} \right)$$

$$V_{max}^{ATP-exchanger} = 5.4 \cdot 10^{-5}$$

$$S_{V_{mm}} = 0.3$$

ATP Consumption

ATP consumption is modeled as a Michaelis-Menten equation with low K_m value.

$$v_{ATP-use} = V_{\max-ATP-use} \frac{ATP_{in}}{ATP_{in} + K_m^{ATP}} (1 + activation)$$

$$K_m^{ATP} = 1$$

The activation describes the additional ATP consumption upon excitation. The value for the activation is 1 in undisturbed state.

Electrophysiology

$$U = \frac{V_{mm} \cdot F}{1000 \cdot R \cdot T}$$

$$F = 96490.0 \frac{C}{mol}$$

$$R = 8.314 \frac{J}{K \cdot mol}$$

$$T = 310 \text{ K}$$

Electro diffusion

The passive efflux of sodium ions, potassium ions, chloride ions and protons is modeled by the Goldman-Hodgkin equation.

Potassium⁷⁰

$$I_{K_{mito}}^{ed} = A_m \cdot P_{K_{mito}} \cdot U \cdot F \cdot \left(\frac{K_{in} - K_{mito} \cdot \exp(U)}{1 - \exp(U)} \right)$$

$$P_{Pot_{mito}} = 2 \cdot 10^{-10} \frac{m}{s}$$

Sodium

$$I_{Na_{mito}}^{ed} = A_m \cdot P_{Na_{mito}} \cdot U \cdot F \cdot \left(\frac{Na_{in} - Na_{mito} \cdot \exp(U)}{1 - \exp(U)} \right)$$

$$P_{Na_{mito}} = 1 \cdot 10^{-10} \frac{m}{s}$$

Protons^{71, 72}

$$I_{H_{mito}}^{ed} = -A_m \cdot P_{H_{mito}} \cdot U \cdot F \cdot \left(\frac{H_{in} - H_{mito} \cdot \exp(U)}{1 - \exp(U)} \right)$$

$$P_{H_{mito}} = 2 \cdot 10^{-4} \frac{m}{s}$$

Pumps⁷³

The pumping of sodium, potassium and phosphate is modeled as electro-neutral proton driven antiport.

$$V_{Phos-exchanger} = V_{max}^{Phos-exchanger} (P_{in} \cdot H_{in} - P_{mito} \cdot H_{mito})$$

$$V_{max}^{Phos-exchanger} = 43.3485$$

$$I_{Na_{mito}}^{pump} = V_{max}^{Na-pump} (Na_{in} \cdot H_{mito} - Na_{mito} \cdot H_{in})$$

$$V_{max}^{Na-pump} = 5 \cdot 10^{-3}$$

$$I_{Pot_{mito}}^{pump} = V_{max}^{Pot-pump} (K_{in} \cdot H_{mito} - K_{mito} \cdot H_{in})$$

$$V_{max}^{K-pump} = 7.5 \cdot 10^{-4}$$

Protons are pumped by the corresponding processes in complex I, complex III and complex IV.

$$I_{H_{mito}}^{pump} = -(4v_{cx1} + 2v_{cx2} + 2v_{cx4})$$

Calcium⁷³⁻⁷⁹

$$I_{Ca_{mito}}^{ed} = A_m \cdot 2 \cdot U \cdot F \cdot \left(\frac{Ca_{in} - Ca_{mito} \cdot \exp(2 \cdot U)}{1 - \exp(2 \cdot U)} \right) \left(P_{ca}^{RMC} \left(1 - \frac{ca_{cyt}}{ca_{cyt} + K_i^{ca_{cyt}}} \right) + P_{Ca}^{Mcu} \left(\frac{Ca_{in}^n}{Ca_{in}^n + (K_{m-Ca}^{Mcu})^n} \right) \cdot \left(\frac{Ca_{in}^{n_a}}{Ca_{in}^{n_a} + (K_a^{Ca})^{n_a}} \right) \right)$$

$$P_{ca}^{RMC} = 2 \cdot 10^{-6} \frac{m}{s}$$

$$K_i^{ca_{cyt}} = 0.0001$$

$$P_{Ca}^{Mcu} = 2 \cdot 10^{-2} \frac{m}{s}$$

$$K_{m-Ca}^{Mcu} = 19.2$$

$$n = 0.6$$

$$K_a^{Ca} = 0.0003$$

$$n_a = 5$$

$$I_{Ca-Na}^{pump} = \left(\frac{Ca_{mito}}{Ca_{mito} + K_m^{Ca_{mito}}} \right) \left(\frac{(Na_{in})^n}{(Na_{in})^n + (K_m^{Na_{in}})^n} \right) \left(Ca_{mito} \cdot Na_{in}^{n_{Na}} - \frac{1}{K_{eq}^{Ca-Na}} Ca_{in} \cdot Na_{mito}^{n_{Na}} \right)$$

$$K_{eq}^{Ca-Na} = \exp\left(-\frac{V_{mm} \cdot F}{1000 \cdot R \cdot T}\right)$$

$$K_m^{Na_{in}} = 8$$

$$K_m^{Ca_{mito}} = 0.0096$$

$$n^{Na} = 3$$

$$n = 2.8$$

$$I_{Ca-H}^{pump} = \left(\frac{Ca_{mito}}{Ca_{mito} + K_m^{Ca_{mito}}} \right) \left(Ca_{mito} \cdot (H_{in})^n - \frac{1}{K_{eq}^{Ca-H}} Ca_{in} \cdot (H_{mito})^n \right)$$

$$K_{eq}^{Ca-H} = \exp\left(-\frac{V_{mm} \cdot F}{1000 \cdot R \cdot T}\right)$$

$$K_m^{Ca_{mito}} = 0.01$$

$$n^H = 3$$

Currents

$$I_{Na_{mito}} = I_{Na_{mito}}^{pump} + I_{Na_{mito}}^{ed} - n^{Na} \cdot I_{Ca-Na}^{pump}$$

$$I_{Pot_{mito}} = I_{Pot_{mito}}^{pump} + I_{Pot_{mito}}^{ed}$$

$$I_{H_{mito}} = I_{H_{mito}}^{ed} + I_{H_{mito}}^{pump} + I_{Na_{mito}}^{pump} + I_{Pot_{mito}}^{pump} - n^H \cdot I_{Ca-H}^{pump} + v_{Phos-exchanger} + 3 \cdot v_{syn}$$

$$I_{Ca_{mito}} = I_{Ca-Na}^{pump} + I_{Ca-H}^{pump} + I_{Ca_{mito}}^{ed}$$

Membrane potential

The mitochondrial membrane is modeled by the capacitor equation.

$$v_{V_{mm}} = \frac{1}{c_m \cdot A_m} \left(-I_C + I_K + I_H + I_{Na} + 2 \cdot I_{Ca} - v_{Phos-exchanger} + v_{ATP-exchanger} \right)$$

$$c_m = 0.9 \cdot 10^{-6} \text{ farad/cm}^2$$

$$A_m = 3.7 \cdot 10^{-5} \text{ cm}^2$$

Complex I¹⁹

(NADH + Q + 4 H_mito <-> NAD + QH₂ + 4 H_in)

$$v_{cxl} = V_{max}^{cxl} \cdot \left(\text{nadh}_{mito} \cdot Q - \frac{1}{K_{eq}^{cxl}} \text{nad}_{mito} \cdot QH_2 \right)$$

$$V_{max}^{cxl} = 2.25$$

$$K_{eq}^{cxi} = \exp\left(2 \cdot Em_N + 2 \cdot Em_Q + \frac{4 \cdot V_{mm} \cdot F}{1000 \cdot R \cdot T} \left(\frac{h_{mito}}{h_{in}}\right)\right)^4$$

Complex III¹⁹

$$v_{cxIII} = V_{max}^{cxIII} \cdot \left(QH_2 \cdot cyt_{ox}^n - \frac{1}{K_{eq}^{cxIII}} Q \cdot cyt_{red}^n \right)$$

$$V_{max}^{cxIII} = 2.25 \cdot 10^4$$

$$K_{eq}^{cxIII} = \exp\left(-2 \cdot Em_Q + 2 \cdot Em_{cytc} + \frac{2 \cdot V_{mm} \cdot F}{1000 \cdot R \cdot T} \left(\frac{h_{mito}}{h_{in}}\right)\right)^4$$

Complex IV⁸⁰⁻⁸²

(2 cyt_{red} + O₂ + 4 H_{mito} → H₂O + 2 cyt_{ox} + 2 H_{in})

$$v_{C_4} = V_{max}^{Cx4} \cdot \frac{cyt_{red}^n}{cyt_{red}^n + (K_m^{cyt_{red}})^n} \frac{O_{2mito}}{O_{2mito} + K_m^{O_2}} \exp\left(-\frac{dG_H \cdot F}{1000 \cdot R \cdot T}\right)^2$$

$$V_{max}^{Cx4} = 32.5$$

$$K_m^{O_2} = 0.001$$

$$K_m^{cyt_{red}} = 0.001$$

$$n = 2$$

The factor $\exp\left(-\frac{dG_H \cdot F}{1000 \cdot R \cdot T}\right)^2$ was included to ensure proper activation of complex IV with increased

demand, but complex IV activity might actually be regulated by the intra- and/or extramitochondrial

ATP/ADP ratio and/or calcium, but exact kinetics are unknown.

CAC

Pyruvate exchanger⁸³

(Pyr_in + H_mito <-> Pyr_mito + H_in)

$$v_{pyr-exchanger} = V_{max}^{Pyr-exchanger} \cdot \frac{(Pyr_{in} \cdot H_{in} - Pyr_{mito} \cdot H_{mito})}{\left(1 + \frac{Pyr_{in}}{K_m^{Pyr_{in}}}\right) \left(1 + \frac{Pyr_{mito}}{K_m^{Pyr_{mito}}}\right)}$$

$$V_{max}^{Pyr-exchanger} = 128$$

$$K_m^{Pyr_{in}} = 0.15$$

$$K_m^{Pyr_{mito}} = 0.15$$

Pyruvate dehydrogenase complex^{26, 84, 85}

(Pyr_mito + CoA + FAD_pdhc -> AcCoA + CO₂ + FADH₂_pdhc)

(FADH₂_pdhc + NAD_mito <-> FAD_pdhc + NADH_mito)

$$v_{pdhc_fad} = V_{max}^{pdhc_fad} \left(1 + A_{max}^{Ca_{mito}} \frac{Ca_{mito}}{Ca_{mito} + K_a^{Ca_{mito}}}\right) \left(\frac{Pyr_{mito}}{Pyr_{mito} + K_m^{Pyr}}\right) \left(\frac{FAD_{pdhc}}{FAD_{pdhc} + K_m^{FAD_{pdhc}}}\right) \left(\frac{CoA_{mito}}{CoA_{mito} + K_m^{CoA_{mito}} \cdot \left(1 + \frac{ACoA_{mito}}{K_i^{ACoA}}\right)}\right)$$
$$v_{pdhc_nad} = V_{max}^{pdhc_nad} \cdot \frac{\left(FADH_{2_pdhc} \cdot NAD_{mito} - \frac{1}{K_{eq_pdhc_fad_nad}} FAD_{pdhc} \cdot NADH_{mito}\right)}{\left(NAD_{mito} + K_m^{NAD_{mito}}\right)}$$

$$V_{max}^{pdhc_fad} = 13.1$$

$$V_{max}^{pdhc_nad} = 1e4$$

$$A_{max}^{Ca_{mito}} = 1.7$$

$$K_a^{Ca_{mito}} = 10^{-3}$$

$$K_m^{Pyr} = 0.068$$

$$K_m^{NAD_{mito}} = 0.041$$

$$K_m^{CoA_{mito}} = 0.0047$$

$$K_i^{ACoA_{mito}} = 0.0004$$

$$K_m^{FAD_{pdhc}} = 0.00001$$

$$K_{eq-pdhc_fad_nad} = \exp\left(\frac{(2 \cdot Em_{FAD} + 2 \cdot Em_{NAD}) \cdot F}{1000 \cdot R \cdot T}\right)$$

$$Em_m^{FAD} = 297mV$$

Citrate synthetase⁸⁶

(Oxa + AcCoA → Cit)

$$v_{cs} = V_{max}^{cs} \left(\frac{Oxa_{mito}}{Oxa_{mito} + K_m^{Oxa} \left(1 + \frac{Cit_{mito}}{K_i^{Cit}} \right)} \right) \left(\frac{AcCoA_{mito}}{AcCoA_{mito} + K_m^{AcCoA} \cdot \left(1 + \frac{CoA_{mito}}{K_i^{CoA}} \right)} \right)$$

$$V_{max}^{cs} = 1.28 \cdot 10^3$$

$$K_m^{Oxa} = 0.0045$$

$$K_i^{Cit} = 3.7$$

$$K_m^{accoa} = 0.005$$

$$K_i^{CoA} = 0.025$$

$$K_{eq}^{Aco} =$$

Aconitase⁸⁷

(Cit ↔ IsoCit)

$$v_{Aco} = V_{\max}^{Aco} \frac{Cit_{mito} - \frac{IsoCit_{mito}}{K_{eq}^{Aco}}}{1 + \frac{Cit_{mito}}{K_m^{Cit}} + \frac{IsoCit_{mito}}{K_m^{IsoCit}}}$$

$$V_{\max}^{Aco} = 1.6 \cdot 10^6$$

$$K_{eq}^{Aco} = 0.067$$

$$K_m^{Cit} = 0.48$$

$$K_m^{IsoCit} = 0.12$$

NAD-dependent isocitrate dehydrogenase⁸⁸⁻⁹²

(IsoCit + NAD → akg + NADH)

$$v_{icdh} = V_{\max}^{icdh} \left(\frac{IsoCit_{mito}^{n^{IsoCit}}}{IsoCit_{mito}^{n^{IsoCit}} + (K_m^{IsoCit})^{n^{IsoCit}}} \right) \left(\frac{NAD_{mito}}{NAD_{mito} + K_m^{NAD} \cdot \left(1 + \frac{NADH_{mito}}{K_i^{NADH}} \right)} \right)$$

$$V_{\max}^{icdh} = 64$$

$$n^{IsoCit} = 1.9$$

$$K_m^{IsoCit} = \frac{K_{m1}^{IsoCit}}{\left(1 + \left(\frac{Ca_{mito}}{K_a^{Ca}} \right)^{n^{Ca}} \right)} + K_{m2}^{IsoCit}$$

$$K_{m1}^{IsoCit} = 0.11$$

$$K_{m2}^{IsoCit} = 0.06$$

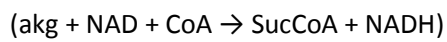
$$K_a^{Ca} = 0.0074$$

$$n^{Ca} = 2$$

$$K_m^{NAD} = 0.091$$

$$K_i^{NADH} = 0.041$$

α -ketoglutarate dehydrogenase complex^{16, 22-26}



$$v_{akdhc_fad} = V_{\max}^{akdhc_fad} \left(\frac{akg_{mito}}{akg_{mito} + K_m^{akdhc}} \right) \left(\frac{FAD_{kgdhc}}{FAD_{mito} + K_m^{FAD_{kgdhc}}} \right) \left(\frac{CoA}{CoA + K_m^{CoA} \cdot \left(1 + \frac{SucCoA}{K_i^{SucCoA}} \right)} \right)$$

$$v_{pdhc_nad} = V_{\max}^{pdhc_nad} \cdot \frac{\left(FADH_{2_kgdhc} \cdot NAD_{mito} - \frac{1}{K_{eq_kgdhc_fad_nad}} FAD_{kgdhc} \cdot NADH_{mito} \right)}{\left(NAD_{mito} + K_m^{NAD} \cdot \left(1 + \frac{NADH_{mito}}{K_i^{NADH}} \right) \right)}$$

$$V_{\max}^{akdhc_nad} = 134.4$$

$$V_{\max}^{akdhc_fad} = 1e4$$

$$K_m^{akdhc} = \left(\frac{K_{m1}^{akdhc}}{\left(1 + \frac{Ca_{mito}}{K_{i-akg}^{Ca}} \right)} + K_{m2}^{akgh} \right) \left(1 + \frac{NADH_{mito}}{K_{i-akg}^{NADH}} \right)$$

$$K_{m1}^{akdhc} = 2.5$$

$$K_m^{NAD} = 0.021$$

$$K_i^{NADH} = 0.0045$$

$$K_m^{CoA} = 0.0013$$

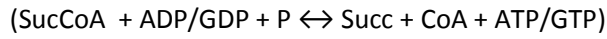
$$K_i^{SucCoA} = 0.0045$$

$$K_m^{FAD_{kgdhc}} = 0.00001$$

$$K_{eq-kgdhc_fad_nad} = \exp\left(\frac{(2 \cdot Em_{FAD} + 2 \cdot Em_{NAD}) \cdot F}{1000 \cdot R \cdot T}\right)$$

$$Em_m^{FAD} = 297mV$$

Succinyl-CoA synthetase⁹³⁻⁹⁶



$$v_{succoa-atp} = V_{max}^{succoa-atp} \left(1 + A_{max}^P \cdot \left(\frac{P_{mito}^{n^P}}{P_{mito}^{n^P} + (K_m^P)^{n^P}} \right) \right) \left(\frac{SucCoA_{mito} \cdot ADP_{mito} \cdot P_{mito} - \frac{Succ_{mito} \cdot CoA_{mito} \cdot ATP_{mito}}{K_{eq-succoas}}}{\left(1 + \frac{SucCoA_{mito}}{K_m^{SucCoA}} \right) \left(1 + \frac{ADP_{mito}}{K_m^{ADP}} \right) \left(1 + \frac{P_{mito}}{K_m^P} \right) + \left(1 + \frac{Succ_{mito}}{K_m^{Succ}} \right) \left(1 + \frac{CoA_{mito}}{K_m^{CoA}} \right) \left(1 + \frac{ATP_{mito}}{K_m^{ATP}} \right) - 1} \right)$$

$$V_{max-succoas-atp} = 1.92 \cdot 10^4$$

$$K_{eq-succoas} = 3.8$$

$$A_{max}^P = 1.2$$

$$K_m^P = 2.5$$

$$n^P = 3$$

$$K_m^{SucCoA} = 0.041$$

$$K_m^{ADP} = 0.25$$

$$K_m^P = 0.72$$

$$K_m^{Succ} = 1.6$$

$$K_m^{CoA} = 0.056$$

$$K_m^{ATP} = 0.017$$

$$v_{succoa-glp} = V_{\max}^{succoa-glp} \left(1 + A_{\max}^P \cdot \left(\frac{P_{\text{mito}}^P}{P_{\text{mito}}^{n^P} + (K_m^P)^{n^P}} \right) \right) \left(\frac{SucCoA_{\text{mito}} \cdot GDP_{\text{mito}} \cdot P_{\text{mito}} - \frac{Succ_{\text{mito}} \cdot CoA_{\text{mito}} \cdot GTP_{\text{mito}}}{K_{eq-succoas}}}{\left(1 + \frac{SucCoA_{\text{mito}}}{K_m^{SucCoA}} \right) \left(1 + \frac{GDP_{\text{mito}}}{K_m^{GDP}} \right) \left(1 + \frac{P_{\text{mito}}}{K_m^P} \right) + \left(1 + \frac{Succ_{\text{mito}}}{K_m^{Succ}} \right) \left(1 + \frac{CoA_{\text{mito}}}{K_m^{CoA}} \right) \left(1 + \frac{GTP_{\text{mito}}}{K_m^{GTP}} \right) - 1} \right)$$

$$K_{eq-succoas} = 3.8$$

$$A_{\max}^P = 1.2$$

$$K_m^P = 2.5$$

$$n^P = 3$$

$$K_m^{SucCoA} = 0.086$$

$$K_m^{GDP} = 0.007$$

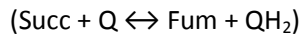
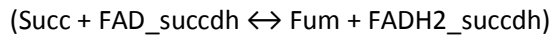
$$K_m^P = 2.26$$

$$K_m^{Succ} = 0.49$$

$$K_m^{CoA} = 0.036$$

$$K_m^{GTP} = 0.036$$

Succinate dehydrogenase⁹⁷⁻⁹⁹



$$v_{\text{succdh_fad}} = V_{\max-\text{succdh_fad}} \left(\frac{Succ_{\text{mito}} \cdot Q_n - \frac{Fum_{\text{mito}} \cdot (QH_2)_n}{K_{eq-succdh}}}{Succ_{\text{mito}} + K_m^{Succ} \left(1 + \frac{Mal_{\text{mito}}}{K_i^{Mal}} \right)} \right)$$

$$v_{succdh_nad} = V_{\max}^{succdh_nad} \cdot \frac{\left(FADH_{2_pdhc} \cdot NAD_{mito} - \frac{1}{K_{eq_pdhc_fad_nad}} FAD_{pdhc} \cdot NADH_{mito} \right)}{\left(NAD_{mito} + K_m^{NAD_{mito}} \right)}$$

$$V_{\max-succdh} = 1.6 \cdot 10^5$$

$$V_{\max}^{succdh_nad} = 10^{12}$$

$$K_{eq-succdh_fad} = \exp\left(\frac{25 \cdot F}{R \cdot T}\right) \cdot \exp\left(\frac{-Em_{FAD-succdh} \cdot F}{R \cdot T}\right)$$

$$K_{eq-succdh_nad} = \exp\left(\frac{Em_{FAD-succdh} \cdot F}{R \cdot T}\right)$$

$$K_m^{Succ} = 1.6$$

$$K_i^{Mal} = 2.2$$

$$Em_{FAD-succdh} = 100mV$$

Fumarase^{100, 101}

(Fum ↔ Mal)

$$v_{fum} = V_{\max-fum} \left(\frac{Fum_{mito} - \frac{Mal_{mito}}{K_{eq-fum}}}{1 + \frac{Fum_{mito}}{K_m^{Fum}} + \frac{Mal_{mito}}{K_m^{Mal}}} \right)$$

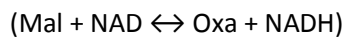
$$V_{\max-fum} = 6.4 \cdot 10^7$$

$$K_{eq-fum} = 4.4$$

$$K_m^{Fum} = 0.14$$

$$K_m^{Mal} = 0.3$$

Malate dehydrogenase^{61, 88, 102}



$$v_{mdh} = V_{\max-mdh} \left(\frac{\text{Mal}_{\text{mito}} \cdot \text{NAD}_{\text{mito}} - \frac{\text{Oxa}_{\text{mito}} \cdot \text{NADH}_{\text{mito}}}{K_{eq-mdh}}}{\left(1 + \frac{\text{Mal}_{\text{mito}}}{K_m^{\text{Mal}}}\right) \left(1 + \frac{\text{NAD}_{\text{mito}}}{K_m^{\text{NAD}}}\right) + \left(1 + \frac{\text{Oxa}_{\text{mito}}}{K_m^{\text{Oxa}}}\right) \left(1 + \frac{\text{NADH}_{\text{mito}}}{K_m^{\text{NADH}}}\right) - 1} \right)$$

$$V_{\max-mdh} = 3.2 \cdot 10^4$$

$$K_{eq-mdh} = 1 \cdot 10^{-4}$$

$$K_m^{\text{Mal}} = 0.145$$

$$K_m^{\text{NAD}} = 0.06$$

$$K_m^{\text{Oxa}} = 0.017$$

$$K_m^{\text{NADH}} = 0.044$$

REFERENCES

1. Sacktor B, Wilson JE, Tiekert CG. Regulation of Glycolysis in Brain in Situ during Convulsions. *Journal of Biological Chemistry* 1966; 241(21): 5071-&.
2. Gimenez C, Valdivie.F, Mayor F. Glycolysis in Brain and Liver of Rats with Experimentally Induced Phenylketonuria. *Biochemical Medicine* 1974; 11(1): 81-86.
3. Szutowicz A, Stepien M, Lysiak W, Angielski S. Purification and Kinetic-Properties of Atp - Citrate Oxaloacetate Lyase from Rat-Brain. *Journal of Neurochemistry* 1975; 25(1): 11-20.
4. Merrill DK, Guynn RW. Electroconvulsive Seizure - Investigation into Validity of Calculating Cytoplasmic Free [Nad+]/[Nadh][H+] Ratio from Substrate Concentrations of Brain. *Journal of Neurochemistry* 1976; 27(2): 459-464.
5. Gorell JM, Law MM, Lowry OH, Ferrendelli JA. Levels of Cerebral Cortical Glycolytic and Citric-Acid Cycle Metabolites during Hypoglycemic Stupor and Its Reversal. *Journal of Neurochemistry* 1977; 29(2): 187-191.
6. Miller AL, Corrdry DH. Brain Carbohydrate-Metabolism in Developing Rats during Hypercapnia. *Journal of Neurochemistry* 1981; 36(3): 1202-1210.
7. Vora S, Oskam R, Staal GEJ. Isoenzymes of Phosphofructokinase in the Rat - Demonstration of the 3 Non-Identical Subunits by Biochemical, Immunochemical and Kinetic-Studies. *Biochemical Journal* 1985; 229(2): 333-341.

8. Hoyer S, Krier C. Ischemia and the Aging Brain - Studies on Glucose and Energy-Metabolism in Rat Cerebral-Cortex. *Neurobiology of Aging* 1986; 7(1): 23-29.
9. Yamamoto M, Hamasaki N, Maruta Y, Tomonaga M. Fructose 2,6-Bisphosphate Changes in Rat-Brain during Ischemia. *Journal of Neurochemistry* 1990; 54(2): 592-597.
10. Ambrosio S, Ventura F, Rosa JL, Bartrons R. Fructose 2,6-Bisphosphate in Hypoglycemic Rat-Brain. *Journal of Neurochemistry* 1991; 57(1): 200-203.
11. Kasten TP, Mhaskar Y, Dunaway GA. Regulation of Brain 6-Phosphofructo-1-Kinase - Effects of Aging, Fructose-2,6-Bisphosphate, and Regional Subunit Distribution. *Molecular and Cellular Biochemistry* 1993; 120(1): 61-68.
12. Vannucci RC, Brucklacher RM. Cerebral Mitochondrial Redox States during Metabolic Stress in the Immature Rat. *Brain Research* 1994; 653(1-2): 141-147.
13. Zwingmann C, Leibfritz D, Hazell AS. Brain energy metabolism in a sub-acute rat model of manganese neurotoxicity: An ex vivo nuclear magnetic resonance study using [1-C-13]glucose. *Neurotoxicology* 2004; 25(4): 573-587.
14. Simpson DP, Angielsk.S. Regulation by Bicarbonate Ion of Intramitochondrial Citrate Concentration in Kidney Mitochondria. *Biochimica Et Biophysica Acta* 1973; 298(1): 115-123.
15. Folbergr.J, Ljunggre.B, Norberg K, Siesjo BK. Influence of Complete Ischemia on Glycolytic Metabolites, Citric-Acid Cycle Intermediates, and Associated Amino-Acids in Rat Cerebral-Cortex. *Brain Research* 1974; 80(2): 265-279.
16. Smith CM, Bryla J, Williams.Jr. Regulation of Mitochondrial Alpha-Ketoglutarate Metabolism by Product Inhibition at Alpha-Ketoglutarate Dehydrogenase. *Journal of Biological Chemistry* 1974; 249(5): 1497-1505.
17. Hansford RG, Johnson RN. Steady-State Concentrations of Coenzyme a-Sh and Coenzyme-a Thioester, Citrate, and Isocitrate during Tricarboxylate Cycle Oxidations in Rabbit Heart-Mitochondria. *Journal of Biological Chemistry* 1975; 250(21): 8361-8375.
18. Benzi G, Arrigoni E, Marzatico F, Villa RF. Influence of Some Biological Pyrimidines on the Succinate Cycle during and after Cerebral-Ischemia. *Biochemical Pharmacology* 1979; 28(17): 2545-2550.
19. Brown GC, Brand MD. Proton/electron stoichiometry of mitochondrial complex I estimated from the equilibrium thermodynamic force ratio. *Biochem J* 1988; 252(2): 473-9.
20. Gibala MJ, MacLean DA, Graham TE, Saltin B. Tricarboxylic acid cycle intermediate pool size and estimated cycle flux in human muscle during exercise. *American Journal of Physiology-Endocrinology and Metabolism* 1998; 275(2): E235-E242.
21. Huchzermeyer C, Berndt N, Holzhutter HG, Kann O. Oxygen consumption rates during three different neuronal activity states in the hippocampal CA3 network. *J Cereb Blood Flow Metab* 2013; 33(2): 263-71.
22. McCormack JG, Denton RM. The effects of calcium ions and adenine nucleotides on the activity of pig heart 2-oxoglutarate dehydrogenase complex. *Biochem J* 1979; 180(3): 533-44.
23. Lai JC, Cooper AJ. Brain alpha-ketoglutarate dehydrogenase complex: kinetic properties, regional distribution, and effects of inhibitors. *J Neurochem* 1986; 47(5): 1376-86.
24. Luder AS, Parks JK, Frerman F, Parker WD, Jr. Inactivation of beef brain alpha-ketoglutarate dehydrogenase complex by valproic acid and valproic acid metabolites. Possible mechanism of anticonvulsant and toxic actions. *J Clin Invest* 1990; 86(5): 1574-81.
25. Faff-Michalak L, Albrecht J. The two catalytic components of the 2-oxoglutarate dehydrogenase complex in rat cerebral synaptic and nonsynaptic mitochondria: comparison of the response to in vitro treatment with ammonia, hyperammonemia, and hepatic encephalopathy. *Neurochem Res* 1993; 18(2): 119-23.

26. Moxley MA, Beard DA, Bazil JN. A pH-dependent kinetic model of dihydrolipoamide dehydrogenase from multiple organisms. *Biophysical journal* 2014; 107(12): 2993-3007.
27. Maher F, Davies Hill TM, Simpson IA. Substrate specificity and kinetic parameters of GLUT3 in rat cerebellar granule neurons. *Biochemical Journal* 1996; 315: 827-831.
28. Thompson MF, Bachelar HS. Cerebral-Cortex Hexokinase - Comparison of Properties of Solubilized Mitochondrial and Cytoplasmic Activities. *Biochemical Journal* 1970; 118(1): 25-&.
29. Purich DL, Fromm HJ. Kinetics and Regulation of Rat Brain Hexokinase. *Journal of Biological Chemistry* 1971; 246(11): 3456-&.
30. Chakraba U, Kenkare UW. Dimerization of Brain Hexokinase Induced by Its Regulator Glucose-6-Phosphate. *Journal of Biological Chemistry* 1974; 249(18): 5984-5988.
31. Garfinkel L, Garfinkel D, Matsiras P, Matschinsky B. Kinetic-Properties of Hexokinase as Assembled with a Microcomputer Database. *Biochemical Journal* 1987; 244(2): 351-357.
32. Gaitonde MK, Murray E, Cunningham VJ. Effect of 6-Phosphogluconate on Phosphoglucose Isomerase in Rat-Brain *In vitro* and *In vivo*. *Journal of Neurochemistry* 1989; 52(5): 1348-1352.
33. Berg JM, J. L. Tymoczko and L. Stryer *Biochemistry*, New York, NY, W. H. Freeman, 2012.
34. Gekakis N, Johnson RC, Jerkins A, Mains RE, Sul HS. Structure, Distribution, and Functional Expression of the Phosphofructokinase-C Isozyme. *Journal of Biological Chemistry* 1994; 269(5): 3348-3355.
35. Majumder AL, Eisenberg F, Jr. Unequivocal demonstration of fructose-1,6-bisphosphatase in mammalian brain. *Proceedings of the National Academy of Sciences of the United States of America* 1977; 74(8): 3222-5.
36. Elmaghrabi MR, Correia JJ, Heil PJ, Pate TM, Cobb CE, Pilkis SJ. Tissue Distribution, Immunoreactivity, and Physical-Properties of 6-Phosphofructo-2-Kinase Fructose-2,6-Bisphosphatase. *Proceedings of the National Academy of Sciences of the United States of America* 1986; 83(14): 5005-5009.
37. Ventura F, Rosa JL, Ambrosio S, Pilkis SJ, Bartrons R. Bovine Brain 6-Phosphofructo-2-Kinase Fructose-2,6-Bisphosphatase - Evidence for a Neural-Specific Isozyme. *Journal of Biological Chemistry* 1992; 267(25): 17939-17943.
38. Marsin AS, Bertrand L, Rider MH, Deprez J, Beauloye C, Vincent MF *et al*. Phosphorylation and activation of heart PFK-2 by AMPK has a role in the stimulation of glycolysis during ischaemia. *Current Biology* 2000; 10(20): 1247-1255.
39. Kessler R, Eschrich K. Splice isoforms of ubiquitous 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase in human brain. *Molecular Brain Research* 2001; 87(2): 190-195.
40. Oakhill JS, Chen ZP, Scott JW, Steel R, Castelli LA, Ling NM *et al*. beta-Subunit myristoylation is the gatekeeper for initiating metabolic stress sensing by AMP-activated protein kinase (AMPK). *Proceedings of the National Academy of Sciences of the United States of America* 2010; 107(45): 19237-19241.
41. Oakhill JS, Steel R, Chen ZP, Scott JW, Ling N, Tam S *et al*. AMPK Is a Direct Adenylate Charge-Regulated Protein Kinase. *Science* 2011; 332(6036): 1433-1435.
42. Penhoet EE, Kochman M, Rutter WJ. Molecular and Catalytic Properties of Aldolase C. *Biochemistry* 1969; 8(11): 4396-&.
43. Guix FX, Ill-Raga G, Bravo R, Nakaya T, de Fabritiis G, Coma M *et al*. Amyloid-dependent triosephosphate isomerase nitrotyrosination induces glycation and tau fibrillation. *Brain* 2009; 132: 1335-1345.
44. Ryzlak MT, Pietruszko R. Purification and Characterization of Aldehyde Dehydrogenase from Human-Brain. *Archives of Biochemistry and Biophysics* 1987; 255(2): 409-418.

45. Schuster R, Holzutter HG. Use of Mathematical-Models for Predicting the Metabolic Effect of Large-Scale Enzyme-Activity Alterations - Application to Enzyme Deficiencies of Red-Blood-Cells. *European Journal of Biochemistry* 1995; 229(2): 403-418.
46. Kish SJ, Lopes-Cendes I, Guttman M, Furukawa Y, Pandolfo M, Rouleau GA *et al.* Brain glyceraldehyde-3-phosphate dehydrogenase activity in human trinucleotide repeat disorders. *Archives of Neurology* 1998; 55(10): 1299-1304.
47. Sharma HK, Rothstein M. Altered Brain Phosphoglycerate Kinase from Aging Rats. *Mechanisms of Ageing and Development* 1984; 25(3): 285-296.
48. Ikura K, Narita H, Sasaki R, Chiba H. Immunochemical and Enzymatic Properties of "Bisphosphoglyceromutase-Phosphatase and Phosphoglyceromutase from Human Erythrocytes. *European Journal of Biochemistry* 1978; 89(1): 23-31.
49. Batke J, Nazaryan KB, Karapetian NH. Complex of Brain D-Phosphoglycerate Mutase and Gamma-Enolase and Its Reactivation by D-Glycerate 2,3-Bisphosphate. *Archives of Biochemistry and Biophysics* 1988; 264(2): 510-518.
50. Suzuki F, Umeda Y, Kato K. Rat-Brain Enolase Isozymes - Purification of 3 Forms of Enolase. *Journal of Biochemistry* 1980; 87(6): 1587-1594.
51. Schwark WS, Singhal RL, Ling GM. Metabolic Control Mechanisms in Mammalian Systems - Regulation of Pyruvate Kinase in Rat Cerebral Cortex. *Journal of Neurochemistry* 1971; 18(1): 123-&.
52. Srivastava LK, Baquer NZ. Purification and Properties of Rat-Brain Pyruvate-Kinase. *Archives of Biochemistry and Biophysics* 1985; 236(2): 703-713.
53. Nisselbaum JS, Bodansky O, Packer DE. Comparison of Actions of Human Brain Liver + Heart Lactic Dehydrogenase Variants on Nucleotide Analogues + on Substrate Analogues in Absence + in Presence of Oxalate + Oxamate. *Journal of Biological Chemistry* 1964; 239(9): 2830-&.
54. Bonavita V, Amore G, Avellone S. Molecular and Kinetic Properties of Lactate Dehydrogenase in Degenerating Peripheral Nerve. *Journal of the Neurological Sciences* 1966; 3(4): 340-&.
55. Kocha T, Fukuda T, Isobe T, Okuyama T. Large-Scale Purification of Bovine Brain Lactate-Dehydrogenase by Affinity-Chromatography on Immobilized Colchicine. *Journal of Biochemistry* 1990; 107(1): 138-143.
56. Nedergaard M, Kraig RP, Tanabe J, Pulsinelli WA. Dynamics of Interstitial and Intracellular Ph in Evolving Brain Infarct. *American Journal of Physiology* 1991; 260(3): R581-R588.
57. McKenna MC, Tildon JT, Stevenson JH, Hopkins IB, Huang XL, Couto R. Lactate transport by cortical synaptosomes from adult rat brain: Characterization of kinetics and inhibitor specificity. *Developmental Neuroscience* 1998; 20(4-5): 300-309.
58. Teague WE, Dobson GP. Effect of Temperature on the Creatine-Kinase Equilibrium. *Journal of Biological Chemistry* 1992; 267(20): 14084-14093.
59. Sahlin K, Harris RC, Hultman E. Creatine-Kinase Equilibrium and Lactate Content Compared with Muscle Ph in Tissue Samples Obtained after Isometric-Exercise. *Biochemical Journal* 1975; 152(2): 173-180.
60. Mueggler PA, Wolfe RG. Malate-Dehydrogenase - Kinetic Studies of Substrate Activation of Supernatant Enzyme by L-Malate. *Biochemistry* 1978; 17(22): 4615-4620.
61. Gelpi JL, Dordal A, Montserrat J, Mazo A, Cortes A. Kinetic studies of the regulation of mitochondrial malate dehydrogenase by citrate. *Biochem J* 1992; 283 (Pt 1): 289-97.
62. Malik P, McKenna MC, Tildon JT. Regulation of Malate-Dehydrogenases from Neonatal, Adolescent, and Mature Rat-Brain. *Neurochemical Research* 1993; 18(3): 247-257.

63. Kishore N, Tewari YB, Goldberg RN. An investigation of the equilibrium of the reaction {L-aspartate(aq) plus 2-oxoglutarate(aq) = oxaloacetate(aq) plus L-glutamate(aq)}. *Journal of Chemical Thermodynamics* 1998; 30(11): 1373-1384.
64. Dierks T, Riemer E, Kramer R. Reaction-Mechanism of the Reconstituted Aspartate Glutamate Carrier from Bovine Heart-Mitochondria. *Biochimica Et Biophysica Acta* 1988; 943(2): 231-244.
65. Indiveri C, Dierks T, Kramer R, Palmieri F. Reaction-Mechanism of the Reconstituted Oxoglutarate Carrier from Bovine Heart-Mitochondria. *European Journal of Biochemistry* 1991; 198(2): 339-347.
66. De Palma A, Prezioso G, Spagnoletta A, Genchi G, Scalera V. The oxoglutarate/malate carrier of rat brain mitochondria operates by a uniport exchange mechanism. *Journal of Bioenergetics and Biomembranes* 2010; 42(5): 371-379.
67. McGinnis JF, de Vellis J. Purification and characterization of rat brain glycerol phosphate dehydrogenase. *Biochimica et biophysica acta* 1974; 364(1): 17-27.
68. Cai J, Pietzsch M, Theobald U, Rizzi M. Fast purification and kinetic studies of the glycerol-3-phosphate dehydrogenase from the yeast *Saccharomyces cerevisiae*. *Journal of biotechnology* 1996; 49(1-3): 19-27.
69. Bohnensack R. The role of the adenine nucleotide translocator in oxidative phosphorylation. A theoretical investigation on the basis of a comprehensive rate law of the translocator. *J Bioenerg Biomembr* 1982; 14(1): 45-61.
70. Nicolli A, Redetti A, Bernardi P. The K⁺ conductance of the inner mitochondrial membrane. A study of the inducible uniport for monovalent cations. *The Journal of biological chemistry* 1991; 266(15): 9465-70.
71. Krishnamoorthy G, Hinkle PC. Non-Ohmic Proton Conductance of Mitochondria and Liposomes. *Biochemistry* 1984; 23(8): 1640-1645.
72. Rolfe DFS, Hulbert AJ, Brand MD. Characteristics of Mitochondrial Proton Leak and Control of Oxidative-Phosphorylation in the Major Oxygen-Consuming Tissues of the Rat. *Biochimica Et Biophysica Acta-Bioenergetics* 1994; 1188(3): 405-416.
73. Bernardi P. Mitochondrial transport of cations: Channels, exchangers, and permeability transition. *Physiological Reviews* 1999; 79(4): 1127-1155.
74. Gunter TE, Pfeiffer DR. Mechanisms by which mitochondria transport calcium. *The American journal of physiology* 1990; 258(5 Pt 1): C755-86.
75. Hilgemann DW, Collins A, Matsuoka S. Steady-state and dynamic properties of cardiac sodium-calcium exchange. Secondary modulation by cytoplasmic calcium and ATP. *The Journal of general physiology* 1992; 100(6): 933-61.
76. Hilgemann DW, Matsuoka S, Nagel GA, Collins A. Steady-state and dynamic properties of cardiac sodium-calcium exchange. Sodium-dependent inactivation. *The Journal of general physiology* 1992; 100(6): 905-32.
77. Matsuoka S, Hilgemann DW. Steady-state and dynamic properties of cardiac sodium-calcium exchange. Ion and voltage dependencies of the transport cycle. *The Journal of general physiology* 1992; 100(6): 963-1001.
78. Gunter TE, Buntinas L, Sparagna G, Eliseev R, Gunter K. Mitochondrial calcium transport: mechanisms and functions. *Cell calcium* 2000; 28(5-6): 285-96.
79. Kirichok Y, Krapivinsky G, Clapham DE. The mitochondrial calcium uniporter is a highly selective ion channel. *Nature* 2004; 427(6972): 360-4.
80. Napiwotzki J, Kadenbach B. Extramitochondrial ATP/ADP-ratios regulate cytochrome c oxidase activity via binding to the cytosolic domain of subunit IV. *Biol Chem* 1998; 379(3): 335-9.

81. Arnold S, Kadenbach B. The intramitochondrial ATP/ADP-ratio controls cytochrome c oxidase activity allosterically. *FEBS Lett* 1999; 443(2): 105-8.
82. Long J, Gao F, Tong L, Cotman CW, Ames BN, Liu J. Mitochondrial decay in the brains of old rats: ameliorating effect of alpha-lipoic acid and acetyl-L-carnitine. *Neurochem Res* 2009; 34(4): 755-63.
83. Halestrap AP. The mitochondrial pyruvate carrier. Kinetics and specificity for substrates and inhibitors. *Biochem J* 1975; 148(1): 85-96.
84. Blass JP, Lewis CA. Kinetic properties of the partially purified pyruvate dehydrogenase complex of ox brain. *Biochem J* 1973; 131(1): 31-7.
85. Land JM, Clark JB. Effect of phenylpyruvate on pyruvate dehydrogenase activity in rat brain mitochondria. *Biochem J* 1973; 134(2): 539-44.
86. Matsuoka Y, Srere PA. Kinetic studies of citrate synthase from rat kidney and rat brain. *The Journal of biological chemistry* 1973; 248(23): 8022-30.
87. Guarriero-Bobyleva V, Volpi-Becchi MA, Masini A. Parallel partial purification of cytoplasmic and mitochondrial aconitate hydratases from rat liver. *Eur J Biochem* 1973; 34(3): 455-8.
88. Lai JC, Clark JB. Isocitrate dehydrogenase and malate dehydrogenase in synaptic and non-synaptic rat brain mitochondria: a comparison of their kinetic constants. *Biochem Soc Trans* 1978; 6(5): 993-5.
89. Willson VJ, Tipton KF. Purification and characterization of ox brain NAD⁺-dependent isocitrate dehydrogenase. *J Neurochem* 1979; 33(6): 1239-47.
90. Willson VJ, Tipton KF. Allosteric properties of ox brain nicotinamide--adenine dinucleotide dependent isocitrate dehydrogenase. *J Neurochem* 1980; 34(4): 793-9.
91. Rutter GA, Denton RM. Regulation of NAD⁺-linked isocitrate dehydrogenase and 2-oxoglutarate dehydrogenase by Ca²⁺ ions within toluene-permeabilized rat heart mitochondria. Interactions with regulation by adenine nucleotides and NADH/NAD⁺ ratios. *Biochem J* 1988; 252(1): 181-9.
92. Rutter GA, Denton RM. Rapid purification of pig heart NAD⁺-isocitrate dehydrogenase. Studies on the regulation of activity by Ca²⁺, adenine nucleotides, Mg²⁺ and other metal ions. *Biochem J* 1989; 263(2): 445-52.
93. Lynn R, Guynn RW. Equilibrium constants under physiological conditions for the reactions of succinyl coenzyme A synthetase and the hydrolysis of succinyl coenzyme A to coenzyme A and succinate. *The Journal of biological chemistry* 1978; 253(8): 2546-53.
94. Krivanek J, Novakova L. A novel effect of vanadium ions: inhibition of succinyl-CoA synthetase. *Gen Physiol Biophys* 1991; 10(1): 71-82.
95. Johnson JD, Muhonen WW, Lambeth DO. Characterization of the ATP- and GTP-specific succinyl-CoA synthetases in pigeon. The enzymes incorporate the same alpha-subunit. *The Journal of biological chemistry* 1998; 273(42): 27573-9.
96. Phillips D, Aponte AM, French SA, Chess DJ, Balaban RS. Succinyl-CoA synthetase is a phosphate target for the activation of mitochondrial metabolism. *Biochemistry* 2009; 48(30): 7140-9.
97. Gutman M, Silman N. The steady state activity of succinate dehydrogenase in the presence of opposing effectors.II. Reductive activation of succinate dehydrogenase in presence of oxaloacetate. *Mol Cell Biochem* 1975; 7(3): 177-85.
98. Vinogradov AD, Kotlyar AB, Burov VI, Belikova YO. Regulation of succinate dehydrogenase and tautomerization of oxaloacetate. *Adv Enzyme Regul* 1989; 28: 271-80.
99. Quinlan CL, Orr AL, Perevoshchikova IV, Treberg JR, Ackrell BA, Brand MD. Mitochondrial complex II can generate reactive oxygen species at high rates in both the forward and reverse reactions. *The Journal of biological chemistry* 2012; 287(32): 27255-64.

100. Bock RM, Alberty RA. Studies of the Enzyme Fumarase .1. Kinetics and Equilibrium. *Journal of the American Chemical Society* 1953; 75(8): 1921-1925.
101. Kobayashi K, Yamanishi T, Tuboi S. Physicochemical, catalytic, and immunochemical properties of fumarases crystallized separately from mitochondrial and cytosolic fractions of rat liver. *J Biochem* 1981; 89(6): 1923-31.
102. Jespersen N. Thermochemistry of Reaction Catalyzed by Malate Dehydrogenase. *Thermochimica Acta* 1976; 17(1): 23-27.