

Fig. S1: ANI-based species relationship within *B. cereus* sensu lato.

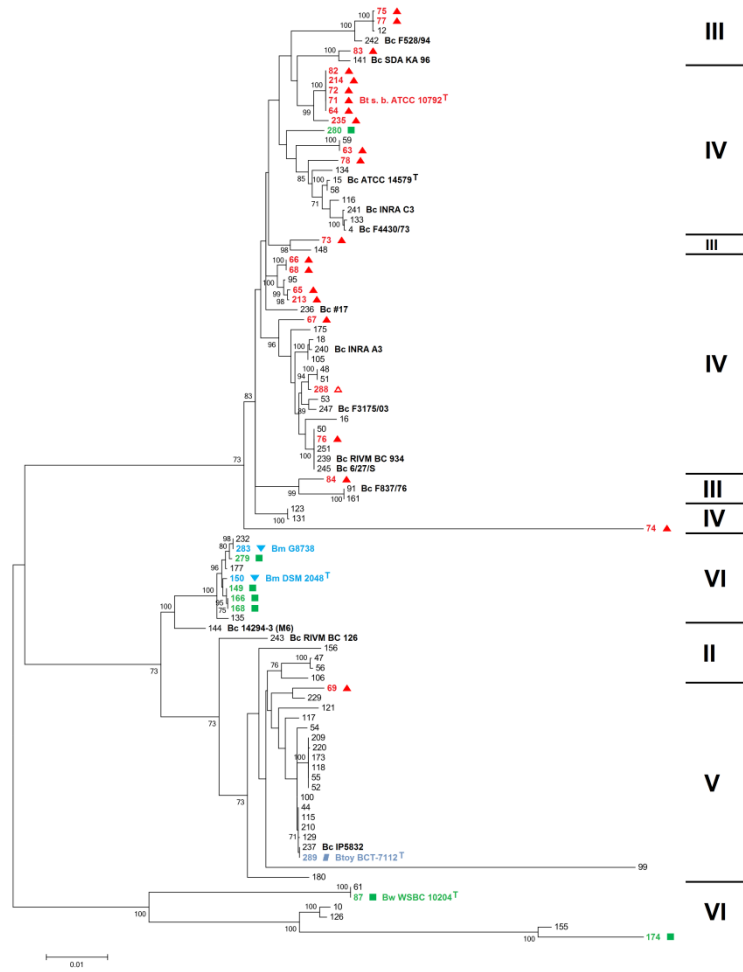
Neighbor network was calculated using ANI distances of 142 *B. cereus* sensu lato genomes. Entire genomes (completed or draft) including all available plasmids were used. Phylogenetic groups are designated according to [1]. *B. cereus* F528/94 (#242) is intermediate between clusters II and III.

Ancient duplication of *hbl*

The second *hbl* operon *hbl_a* is common in clusters II, V and VI and probably was acquired by HGT in cluster IV (Fig. 1, grey crosses; Fig. S2B, grey circles). Several strains seem to have lost their second *hbl* operon and it is not found at all in cluster III. Intra-operon recombination analysis (Table S4) revealed three significant recombination events that all include *B. cereus* MHI 226 (#140, II) as a parental sequence and took place within *hblD_a*. The duplication of *hblCDA* seems to be an ancient and unique event, because it occurs in all phylogenetic groups but III (loss of *hbl_a* at furcation of II and III) and VII (Fig. 1). The topologies of *hbl* and *hbl_a* phylogenies are similar but not identical, which could be explained by HGT (Fig. S2 and Fig. 4). Possibly due to directional selection *hblCDA_a* is as conserved as *hblCDA* (Fig. 4B). *Hbl_a* shows overall nucleotide sequence identity of 75 – 82 % towards the *hbl* genes, which are 89 – 100 % identical among themselves. *Hbl_a* are 93 – 100 % identical among each other. Six strains (#85, #97, #137, #140, #152, and #211) possess only *hblCDA*. Their version of *hbl* is homologous to *hblCDA_a* and they may have lost *hblCDAB*. *B. mycooides* Rock3-17 (#152, I) and *B. mycooides* Rock1-4 possess an *hblCDA* that differs from both *hbl* variants described above. *B. mycooides* Rock3-17 *hblCDA* shows 80 – 82 % identity to *hblCDA_a*, but 86 – 89 % identity to *hblCDA*. We suggest that these two strains have no *hbl_a*, but their *hblCDAB* developed differently and lost *hblB*.

A

- Bacillus cereus (Bc)**
- ▲ **Bacillus thuringiensis (Bt)**
- △ **Bacillus bombysepticus**
- **Bacillus weihenstephanensis (Bw)**
- ▤ **Bacillus toyonensis (Btoy)**
- ▼ **Bacillus mycoides (Bm)**



B

- additional *hblCDA_a*
- only *hblCDA_a*
- ☆ only *hblCDA*

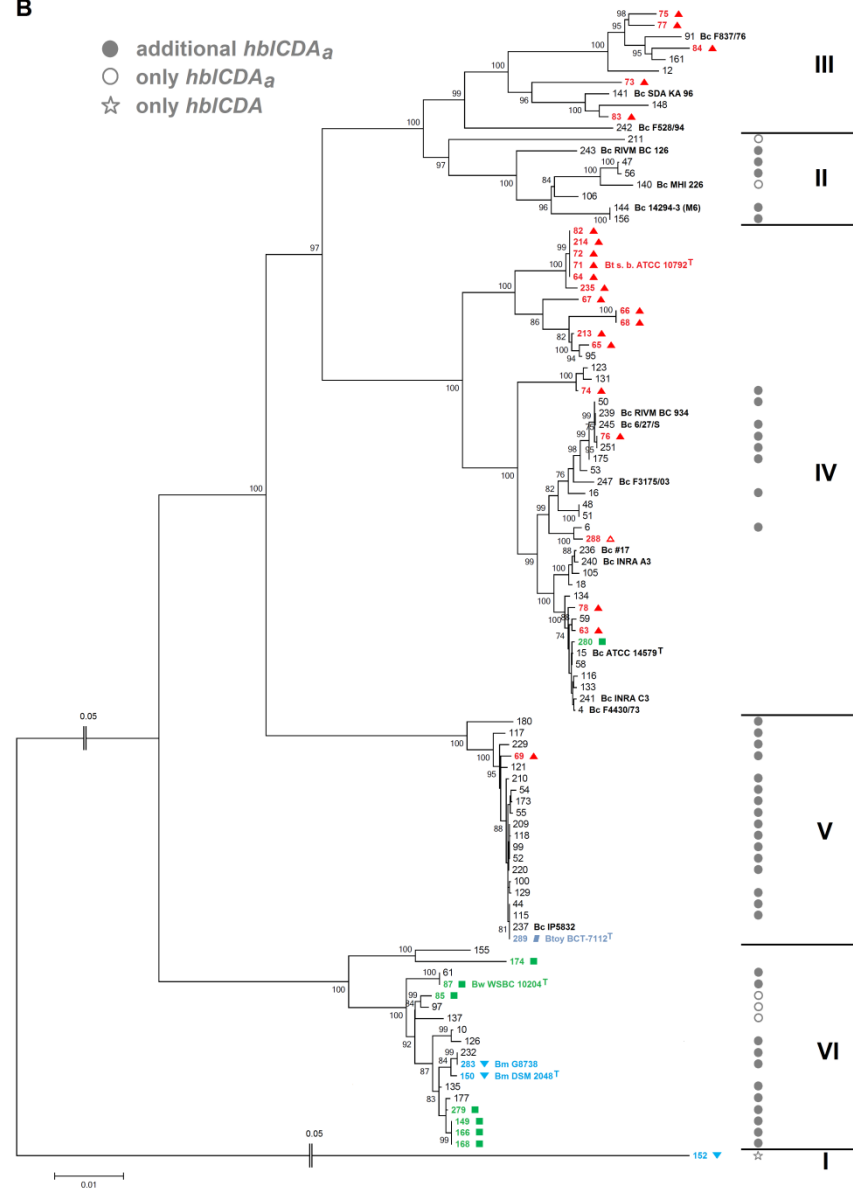


Fig. S2: *Hbl* in *B. cereus* sensu lato.

A: Phylogenetic tree (Maximum Likelihood Method) of the concatenated sequences of 94 *hblCDAB* genes. Seven strains within the set 142 contain only *hblCDA* (empty circle and star) or an incomplete *hbl* operon (#6, IV) and were excluded. **B:** Phylogenetic tree based on the seven housekeeping genes of 101 *hbl*-containing *B. cereus* sensu lato strains.

Recombinational mechanism and boundaries of *hbl*

Since *hbl* occurs chromosomally as well as plasmid-bound, we analyzed the immediate vicinity of *hbl* and *hbl_a* with regard to potential indications of transposon activity in all 16 *hbl*-containing strains that were sequenced in this study. It has been speculated that *hbl* is part of a large 18 kb transposon [2-4]. A comparison of putative transposon regions including *hbl* or *hbl_a* is shown in Figure S3. Sequence analysis and annotation with RAST [5] revealed that half of the *hblCDAB* operons are inserted within the *uvrC* gene as described earlier [3], but in the rest neither insertion sites nor length of the inserting region or adjacent genes are conserved. The lowest common denominator of inserting regions from 16 *B. cereus* sensu lato strains consists only of a transcriptional regulator gene of the *araC* family and *hblCDAB* itself. Inverted repeats (IR and *bcr1*) [3, 6], which mark the insertion site interrupting *uvrC* as telltale signs of transposons, could not be found in half of the investigated strains. A transposase gene could only be detected in the vicinity of *hbl_a* of *B. cereus* 6/27/S (#245, IV), but not adjacent to *hbl* (Fig. S3). Studied *hbl_a* are located close to antibiotic resistance genes and do not contain known inverted repeats. Thus, one may speculate that the *hbl* operon is part of a highly degraded transposon which is in most cases not functional anymore.

Furthermore, the gene *pagA* encoding a protective antigen similar to a gene located on the *B. anthracis* pXO1 virulence plasmid, has been inserted into the chromosome of *B. cereus*, *B. mycooides* and *B. weihenstephanensis* (Fig. S3), proving that recombination between virulence plasmids and the bacterial chromosome occurs frequently. *Nhe* and *hbl* duplications occur chromosomally as well as plasmid-bound and, hence, are mobile within *B. cereus* sensu lato.

These results show that recombination within *B. cereus* sensu lato is limited only by preservation of gene/protein functionality. Consequently, the pathogenic potential of (psychrotolerant) environmental strains or probiotic strains can change rapidly with a single and simple exchange of genetic material. This observation may render the current risk assessment strategies questionable.

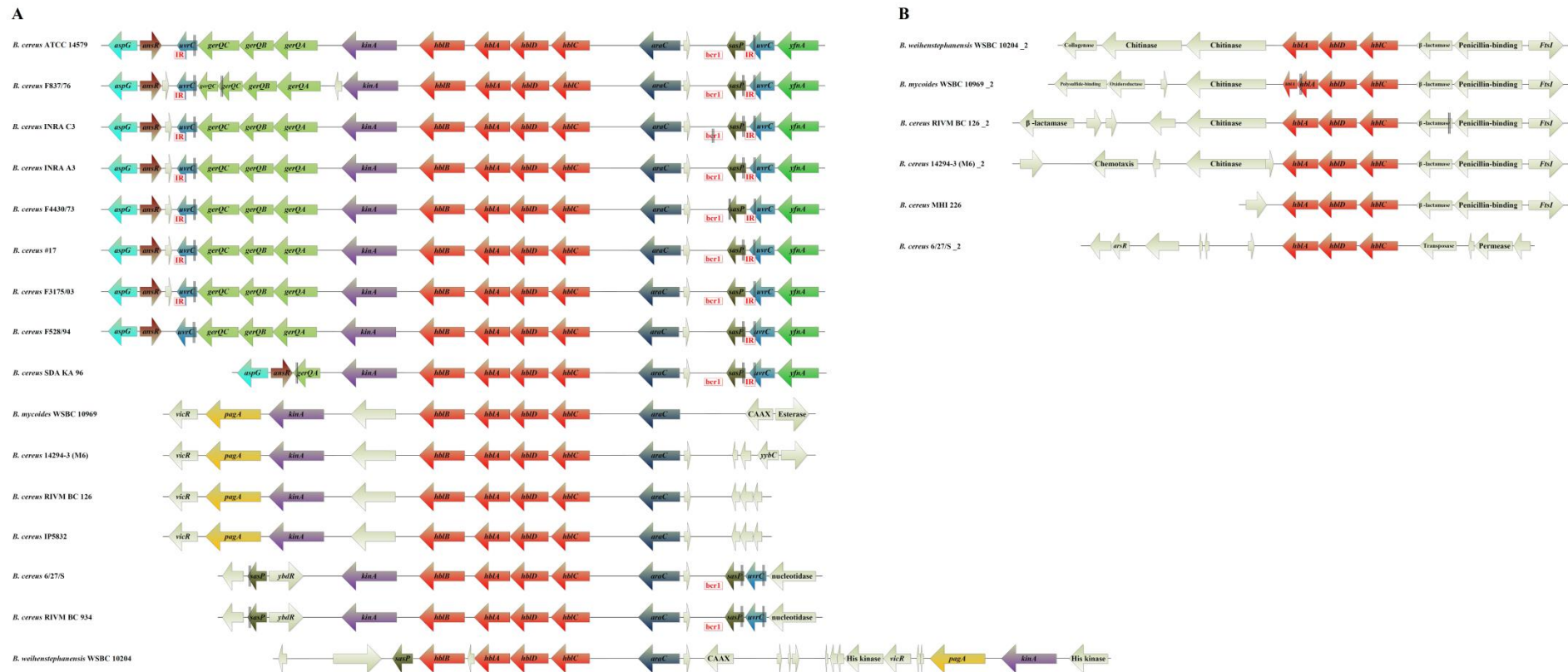


Fig. S3: Genomic organization of *hbl* operons and adjacent regions of 17 *B. cereus* sensu lato strains.

A: 15 of the strains sequenced in this study contain *hblCDAB*, they are shown in comparison to type strain *B. cereus* ATCC 14579.

B: Six of the strains sequenced in this study contain *hblCDA_n*.

Gene denomination according to annotation with RAST [5] and previous studies [3, 4, 6]: *ansR*: *ans* (L-aspartase, L-asparaginase, malate utilization/transporter) operon repressor protein, *araC*: AraC family transcriptional regulator TrrA, *arsR*: ArsR family transcriptional regulator, *aspG*: L-asparaginase, β -lactamase: β -

lactamase class A, CAAX: CAAX amino terminal protease family protein, Chemotaxis: Methyl-accepting chemotaxis protein, Esterase: Erythromycin-esterase type I, *ftsI*: cell division protein, *hblCDAB*: hemolysin BL, His kinase: Two component histidine kinase, *gerQABC*: spore germination proteins, *kinA*: sporulation kinase, nucleotidase: 5' Nucleotidase, Oxidoreductase: FAD-dependent pyridine nucleotide disulfide oxidoreductase, *pagA*: protective antigen (anthrax moiety, pXO1), Penicillin-binding: Penicillin-binding protein, Permease: Permease of the drug/metabolite transporter (DMT) family, Polysulfide-binding: Zn-dependent hydroxyacylglutathione hydrolase (polysulfide-binding protein), *sasP*: small acid soluble protein, Transposase: mobile element protein of the IS605 OrfB family transposase, *uvrC*: UvrC-like excinuclease subunit C, *vicR*: DNA-binding response regulator, *ybdR*: zinc-type alcohol dehydrogenase-like protein, *yfnA*: amino acid permease, *yycC*: uncharacterized protein, *bcr1*: *Bacillus cereus* repeat 1, IR: inverted repeat, ||: incomplete gene, not denominated arrow: hypothetical gene.

Fig. S4: *CytK* in *B. cereus* sensu lato.

A: Phylogenetic tree (Maximum Likelihood Method) based on 68 *cytK* gene sequences. **B:** Phylogenetic tree based on the concatenated sequence of seven housekeeping genes from 68 *cytK*-containing *B. cereus* sensu lato strains.

References

1. Guinebretiere MH, Thompson FL, Sorokin A, Normand P, Dawyndt P, Ehling-Schulz M, Svensson B, Sanchis V, Nguyen-The C, Heyndrickx M *et al*: **Ecological diversification in the *Bacillus cereus* Group**. *Environmental microbiology* 2008, **10**(4):851-865.
2. Okstad OA, Gominet M, Purnelle B, Rose M, Lereclus D, Kolsto AB: **Sequence analysis of three *Bacillus cereus* loci carrying PlcR-regulated genes encoding degradative enzymes and enterotoxin**. *Microbiology* 1999, **145** (Pt 11):3129-3138.
3. Han CS, Xie G, Challacombe JF, Altherr MR, Bhotika SS, Brown N, Bruce D, Campbell CS, Campbell ML, Chen J *et al*: **Pathogenomic sequence analysis of *Bacillus cereus* and *Bacillus thuringiensis* isolates closely related to *Bacillus anthracis***. *Journal of bacteriology* 2006, **188**(9):3382-3390.
4. Stenfors Arnesen LP, Fagerlund A, Granum PE: **From soil to gut: *Bacillus cereus* and its food poisoning toxins**. *FEMS microbiology reviews* 2008, **32**(4):579-606.
5. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M *et al*: **The RAST Server: rapid annotations using subsystems technology**. *BMC genomics* 2008, **9**:75.
6. Okstad OA, Hegna I, Lindback T, Rishovd AL, Kolsto AB: **Genome organization is not conserved between *Bacillus cereus* and *Bacillus subtilis***. *Microbiology* 1999, **145** (Pt 3):621-631.