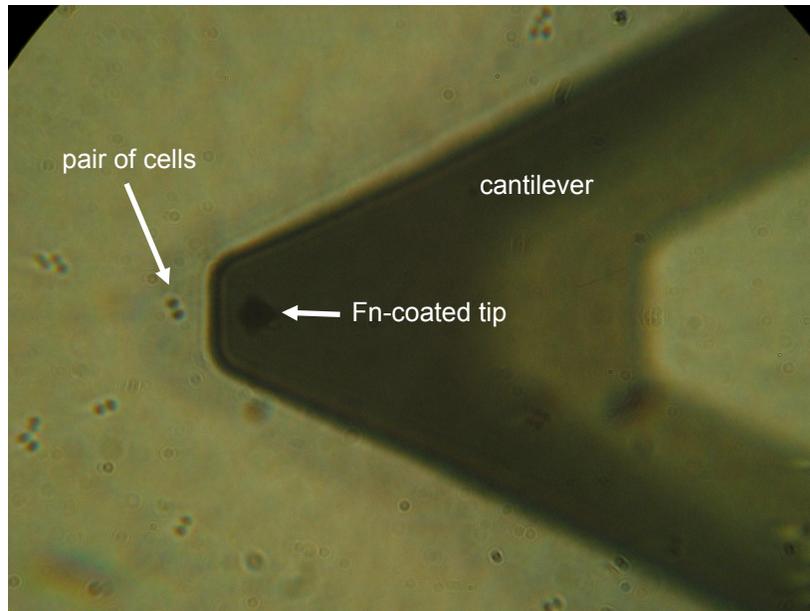


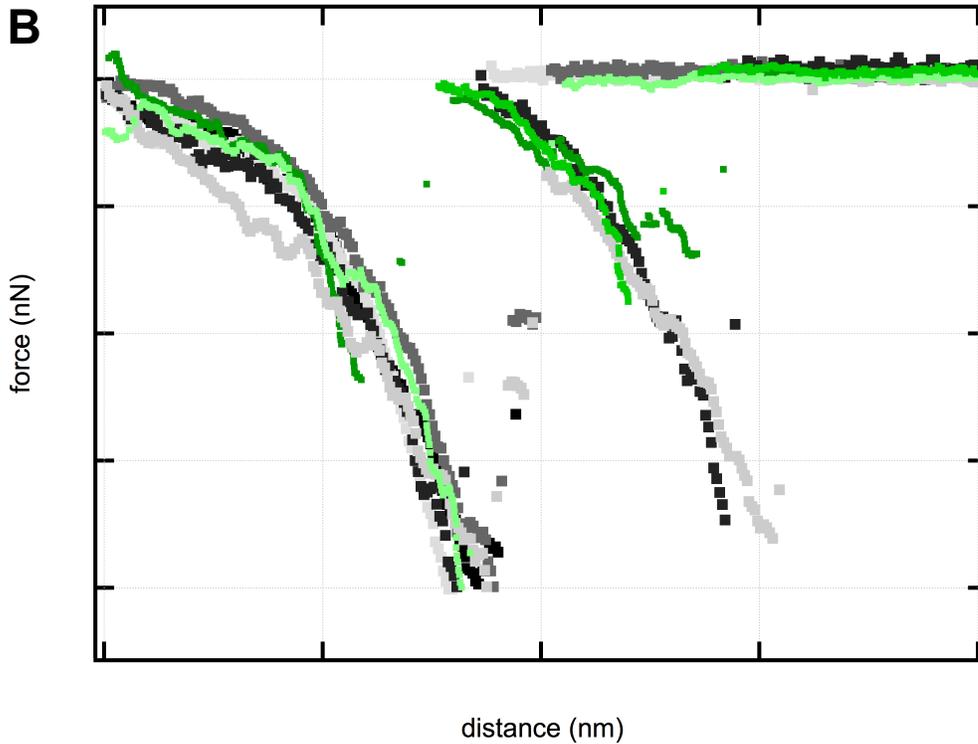
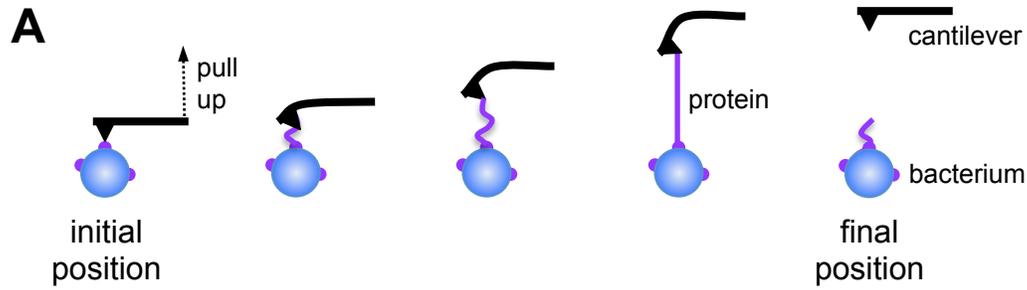
**Supplemental Table 1. Sequences of template and sequencing primers for *fnbA*.**

<b>Primer direction and identifier</b>	<b>Sequence</b>
<b>Forward</b>	
<i>fnbA</i> -F	5'-CCGAACAATATAGACTTGCATTTATTAAG-3'
<i>fnbA</i> -F1	5'-GAACACAAATGGTAGGACATCCAGAGCAAC-3'
<i>fnbA</i> -F2	5'-CAATAGAAGAAACGGATTCATCAGCTATTG-3'
<i>fnbA</i> -F3	5'-CTGGTTTAGGAACTGAAAATGGTCACG-3'
<i>fnbA</i> -F4	5'-CAGTCATTTCGAGGAAGATACAGAAAAAGAC-3'
<b>Reverse</b>	
<i>fnbA</i> -R	5'-CATCAGACATAAACCAATGAAGCAATCAG-3'
<i>fnbA</i> -R1	5'-CTTCATATTCAACAACATCAGCGTGATG-3'
<i>fnbA</i> -R2	5'-CCCGTGACCATTTTCAGTTCCTAAACC-3'
<i>fnbA</i> -R3	5'-GTTGTATCTTCAATCGTTTGTTGACCTTC-3'
<i>fnbA</i> -R4	5'-CTTTACCTTGTTCCACTGGTTTAGAAGG-3'

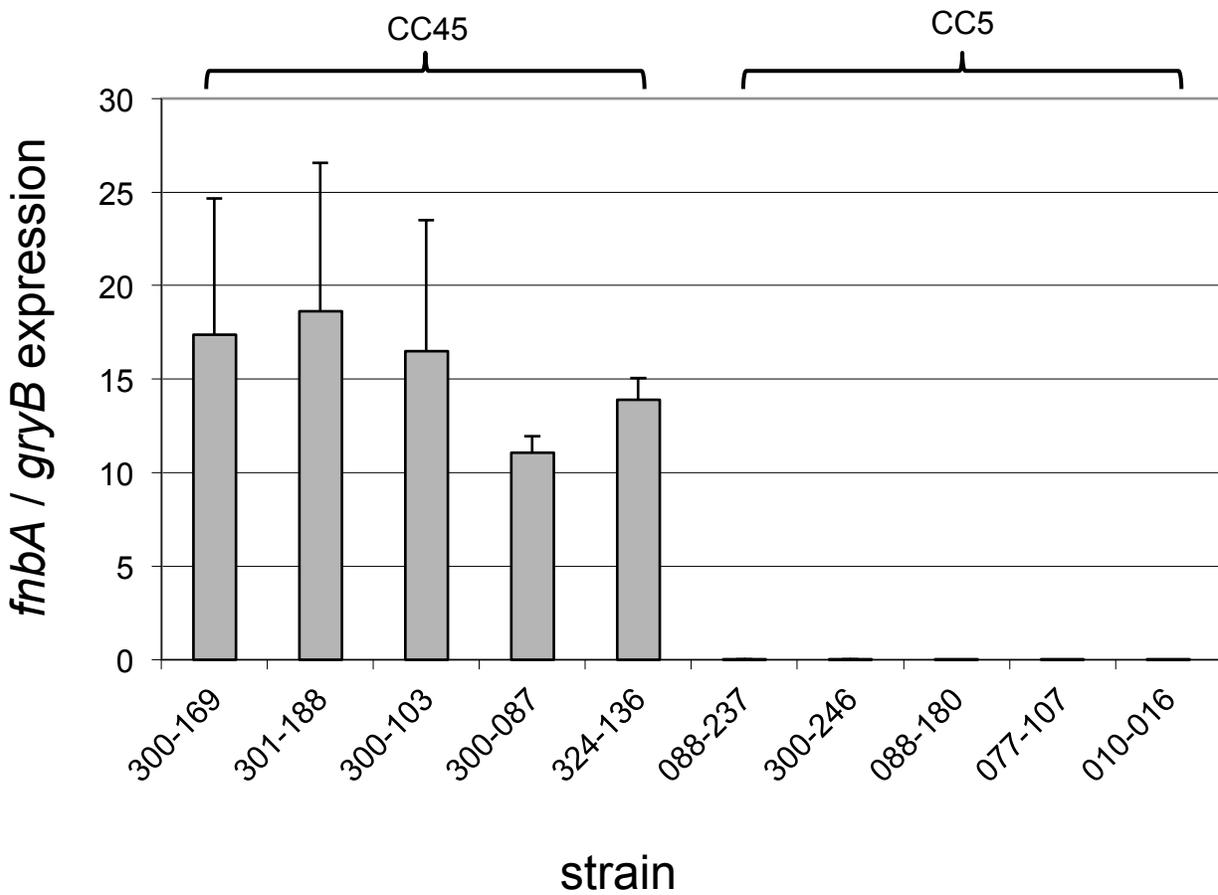


**Supplemental Figure 1. Optical microscope image of AFM experiment.**

Binary fission pair of *S. aureus* cells about to be probed with a fibronectin-coated tip, which is integrated into the end of the force-sensing cantilever. The cells are supported on a glass slide. The entire system is immersed in buffer.



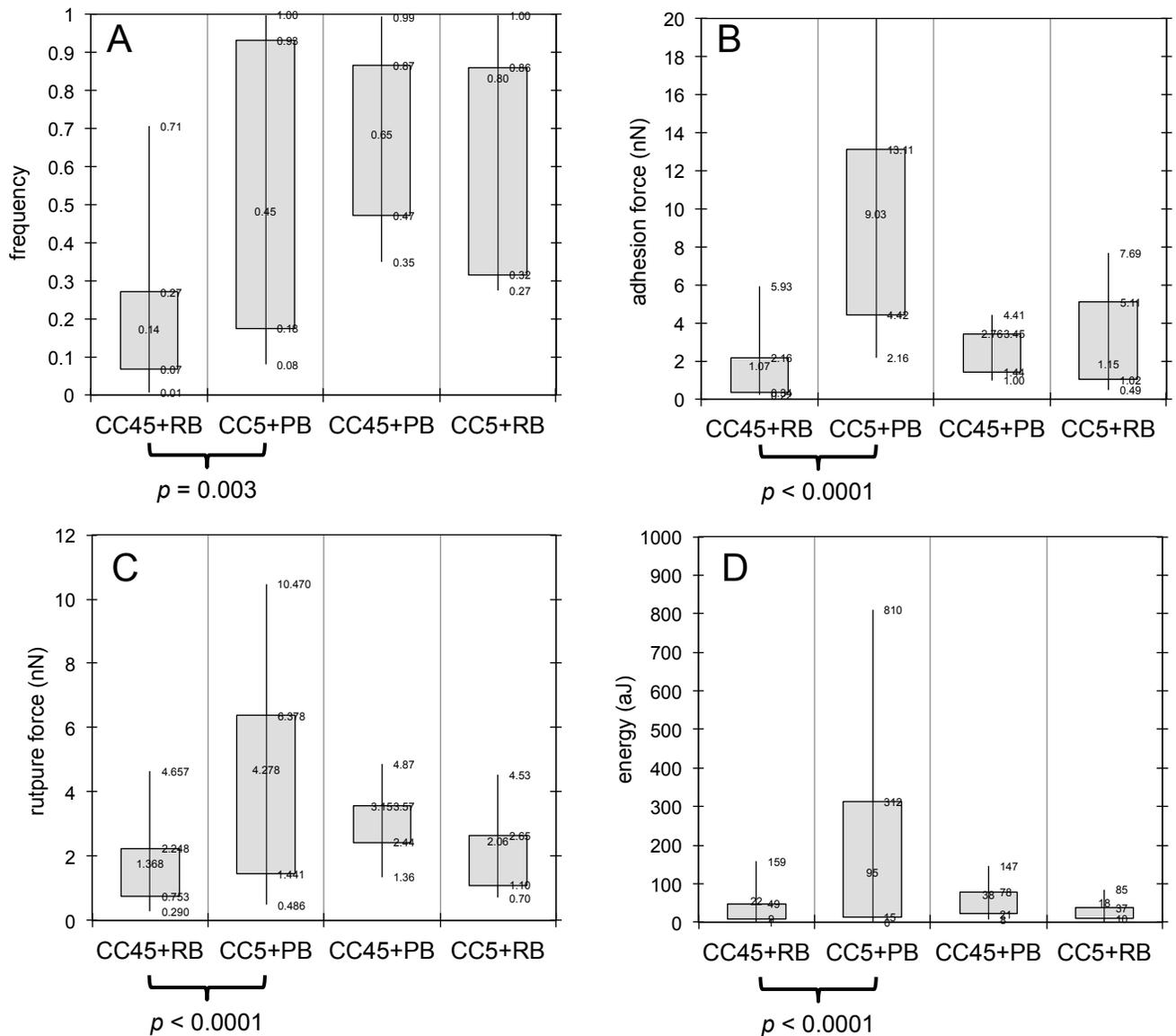
**Supplemental Figure 2. Control experiment using AFM on *Lactococcus lactis*.** (A) Schematic diagram (not to scale) showing a fibronectin-coated tip that was used to probe *L. lactis* strains engineered to express either *fnbA* or *fnbB* genes. Fibronectin on the tip was mechanically separated from FnBPs on a bacterium by pulling the cantilever away from the cell. Unraveling of a protein-protein complex resulted in a “sawtooth shaped” force spectrum. (B) Black- and gray-colored curves were obtained on *L. lactis* expressing FnBPA or FnBPB on their outer cell wall. Green-colored curves are some of the spectra obtained on the CC5 and CC45 clinical isolates of *S. aureus* as described in the manuscript. These spectra are not offset vertically (as is the case for Figure 1 in the manuscript) to highlight the overlapping similarities in the curves for *L. lactis* and *S. aureus*. Native *L. lactis* is an ideal control specimen because it is Gram-positive like *S. aureus*, but it does not naturally bind to fibronectin and lacks *fnbA* and *fnbB* genes.



**Supplemental Figure 3. qRT-PCR of *fnbA* at 3 hour post-incultation.** The effect of clonal variation on *fnbA* transcription by qRT-PCR, normalized by *gryB*, at 3 h post-inoculation.



**Supplemental Figure 4. Western blots of fibronectin binding proteins.** Western blot showing protein bands for FnBPs in the ten *S. aureus* isolates arranged from left to right in the same order as the RT-PCR data. A negative control was also performed with *S. aureus* DU5883, a double mutant that cannot express *fnbA* and *fnbB*.



**Supplemental Figure 5. AFM binding metrics for a fibronectin-coated probe on *S. aureus* grouped by both clonotype and persistence of infection.** Box and whisker plots of (A) frequency of binding, (B) adhesion force, (C) rupture force, and (D) energy (or work) of binding for a fibronectin-coated probe on *S. aureus* grouped as CC45+RB versus CC5+PB. CC corresponds to the clonal complex, RB indicates resolving bacteremia and PB is persistent bacteremia. Force is in nanoNewtons ( $10^{-9}$  N) and energy/work is in attoJoules ( $10^{-18}$  J). Box ends represent the first and third quartiles. Whisker ends represent the 9<sup>th</sup> and 91<sup>st</sup> percentile.