HIV Flow Cytometry Supplement Data

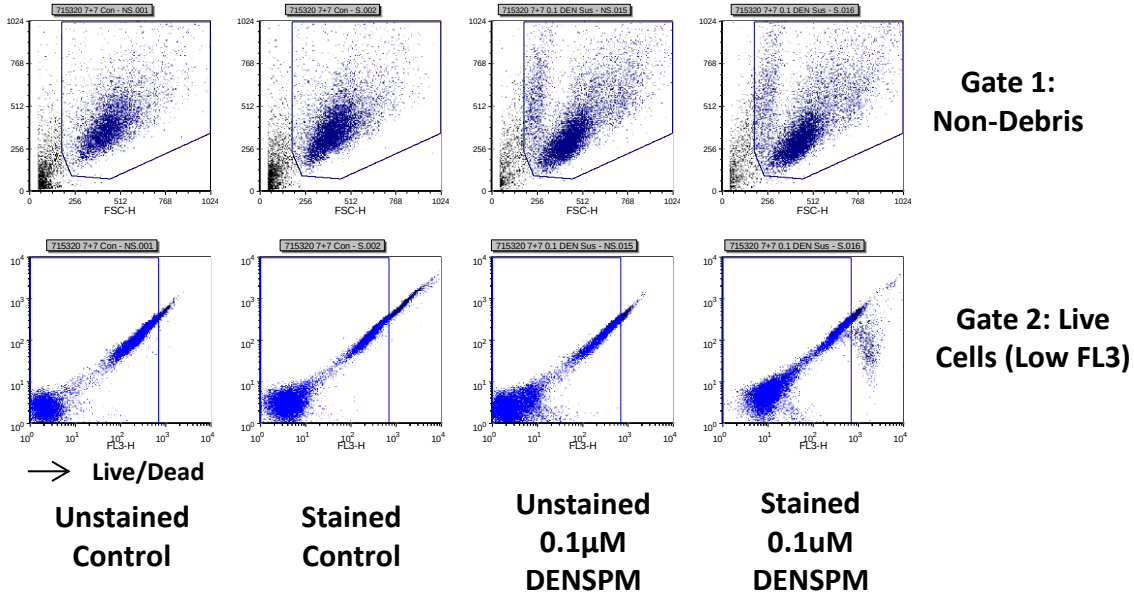

Fig. S1 Gating strategy for FACS analysis. Freshly isolated human monocytes were cultured for 6 days to allow full differentiation. Some of the cells were then treated with DENSPM for another week to induce cell death. They were then harvested and stained with the LIVE/DEAD® Fixable Red Dead Cell Stain (Invitrogen). Untreated cells and unstained cells were included as the negative controls. For gating, debris was first excluded by forward and side scatters. On the FL2/3 plots, the dead cell population was clearly visible in DENSPM treated versus untreated cells and a second gate was draw accordingly to exclude these cells. The diagonal strip of cells shown on the FL2/3 plots comes from the strong autofluorescence of the differentiated macrophages which feature rough and irregular surfaces.

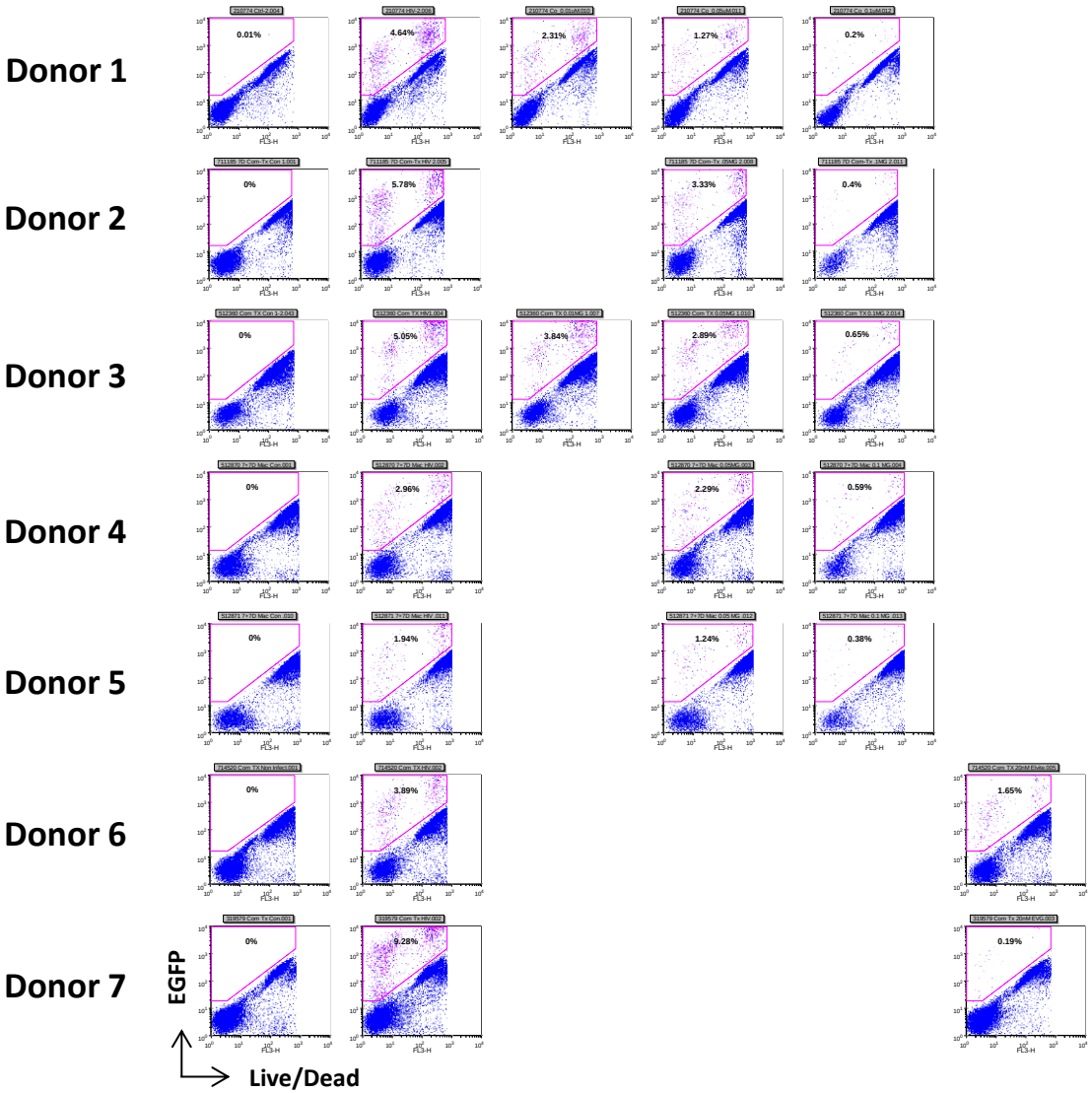

Fig. S2 FACS data for EGFP expression shown in Figure 4A.

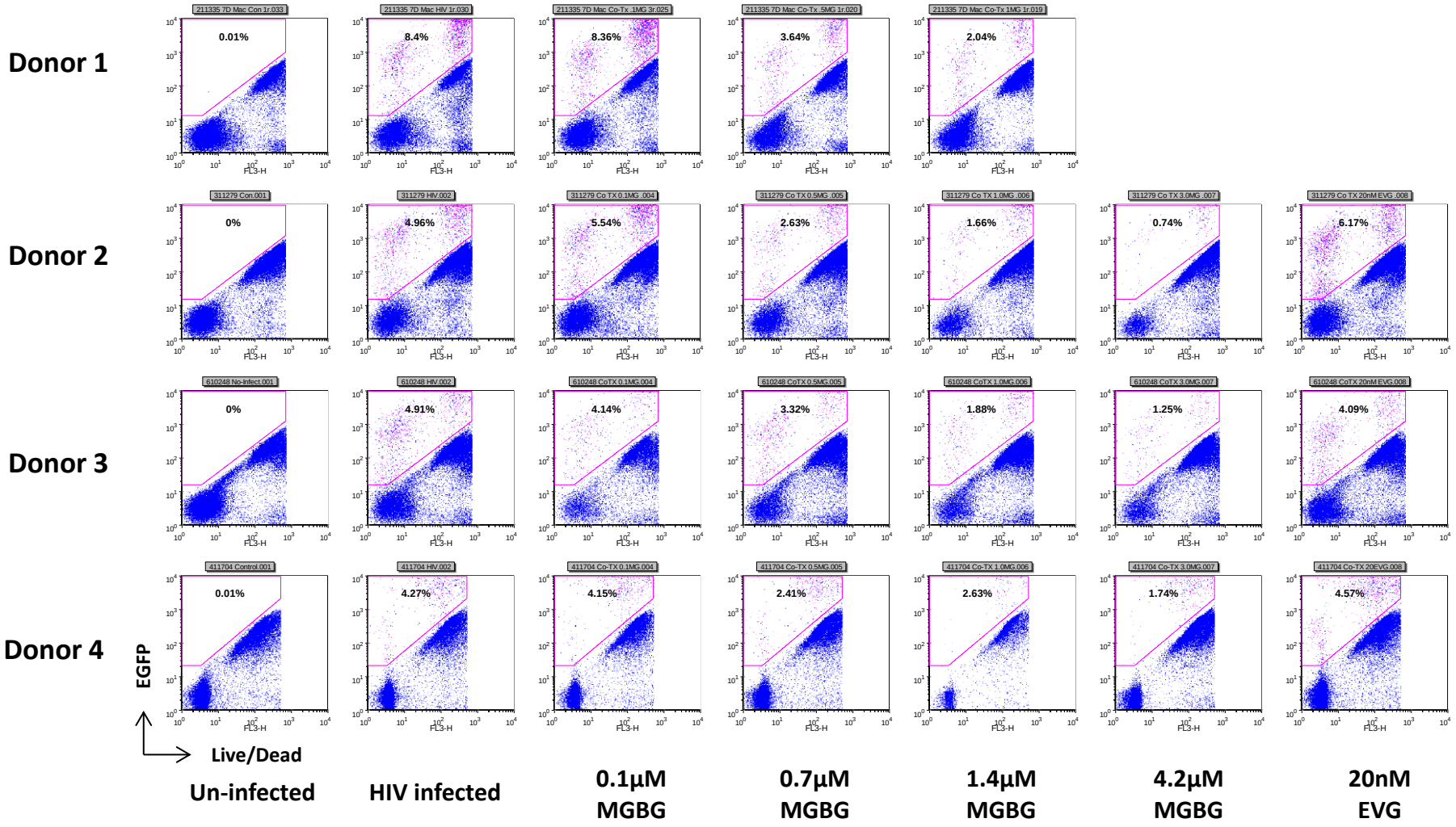

Fig. S3 FACS data for EGFP expression shown in Figure 4B.

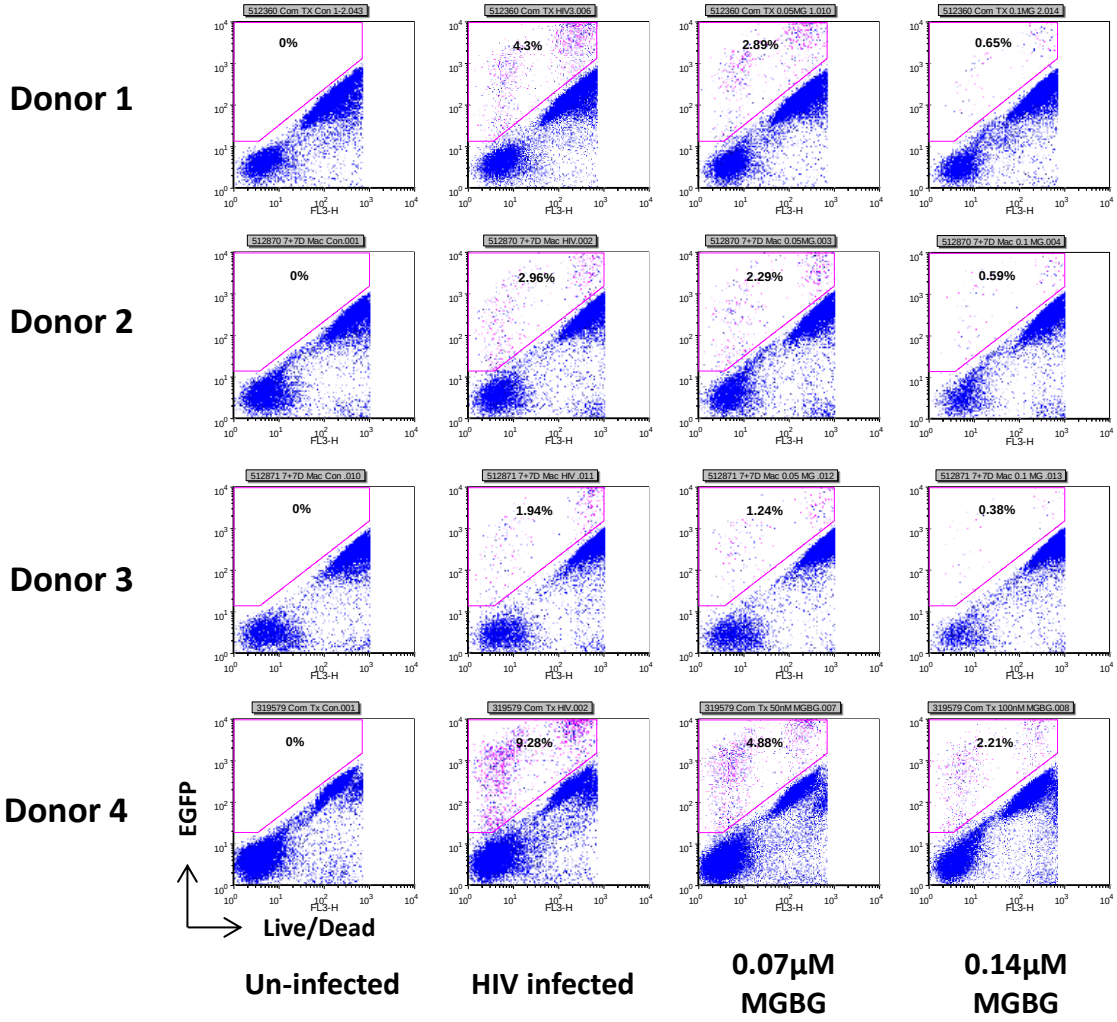

Fig. S4 FACS data for EGFP expression shown in Figure 6A.

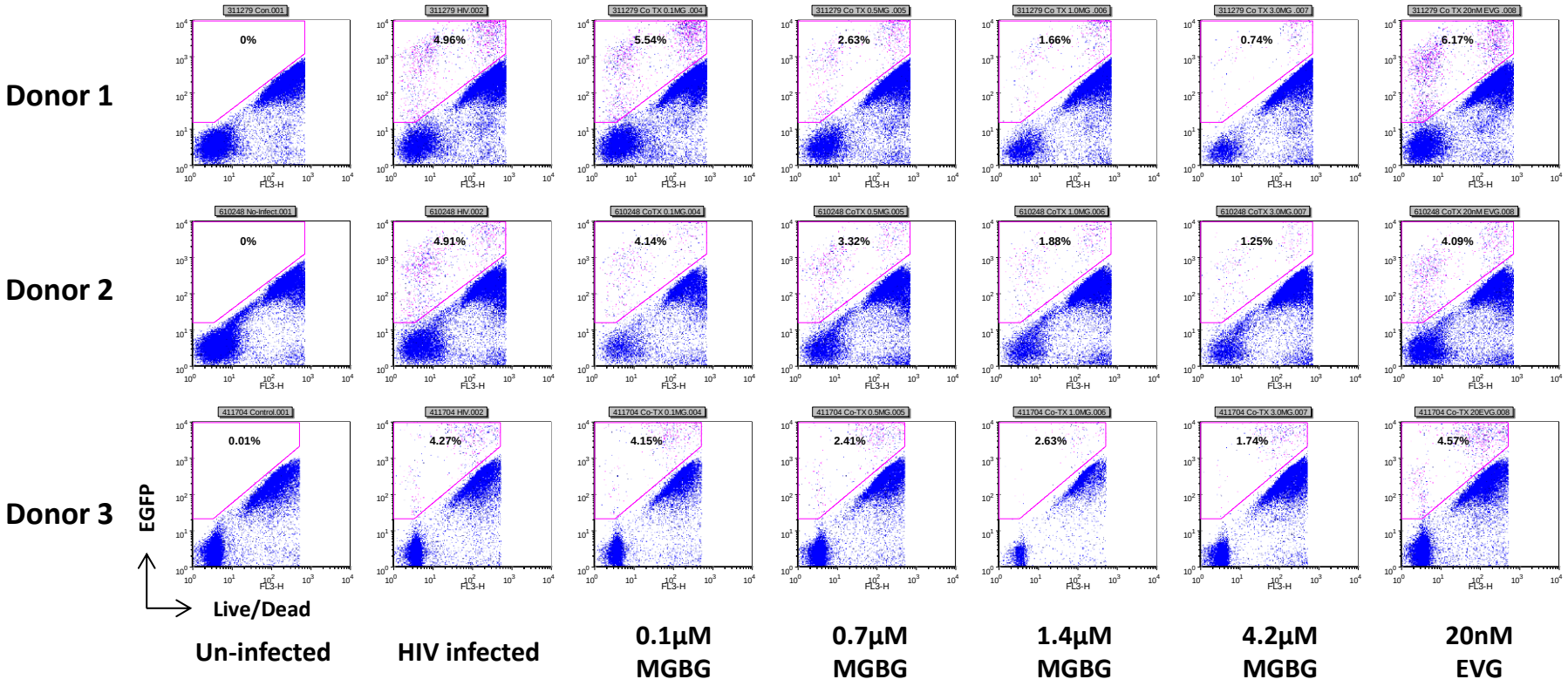

Fig. S5 FACS data for EGFP expression shown in Figure 6B.

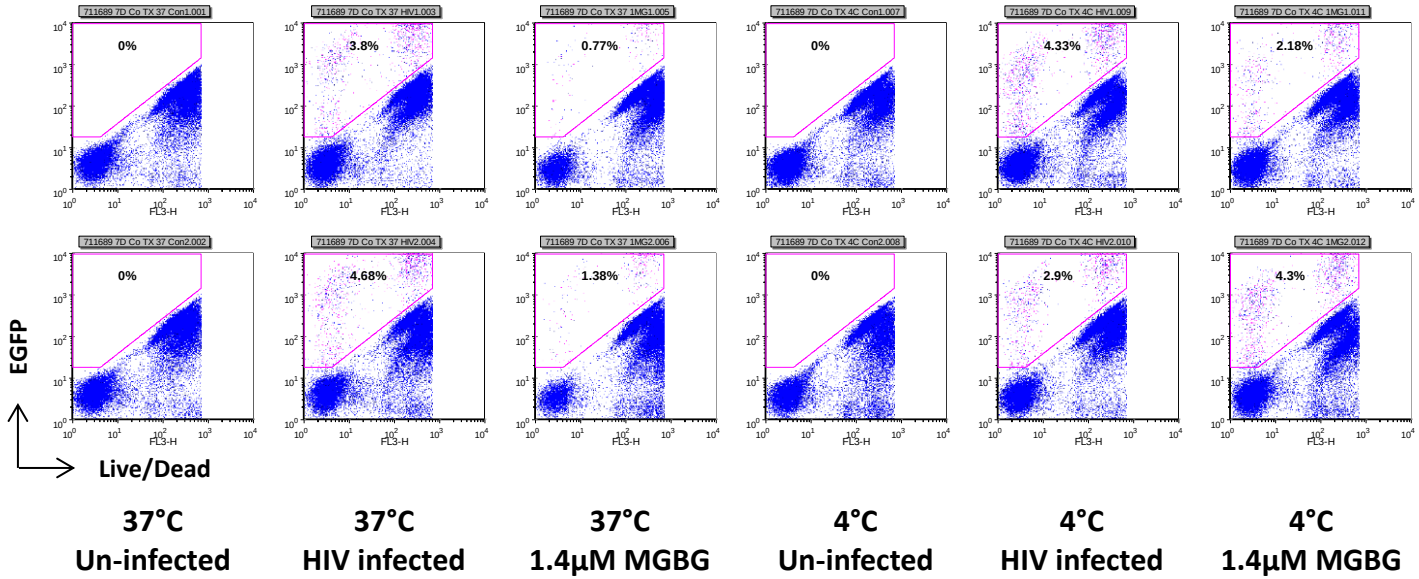

Fig. S6 FACS data for EGFP expression shown in Figure 9. Note that the experiment was done in duplicates with cells from a single donor.

Fig. S2-S6. For flow cytometric analyses, Fully differentiated primary human macrophages were infected with EGFP-tagged HIV and/or treated with the drug as indicated. At the time of harvest, the cells were first stained with the LIVE/DEAD® Fixable Red Dead Cell Stain before flow cytometric analysis. The dot plots showed EGFP expressing population after gating out the debris and dead cells. The regions for the positive cells were further gated based on the autofluorescence pattern of the uninfected cells.