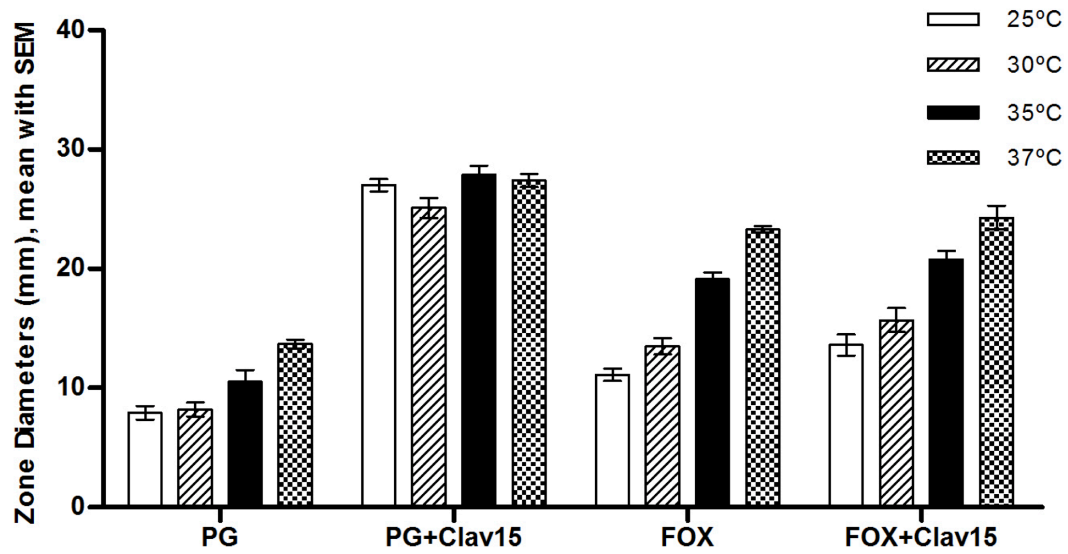
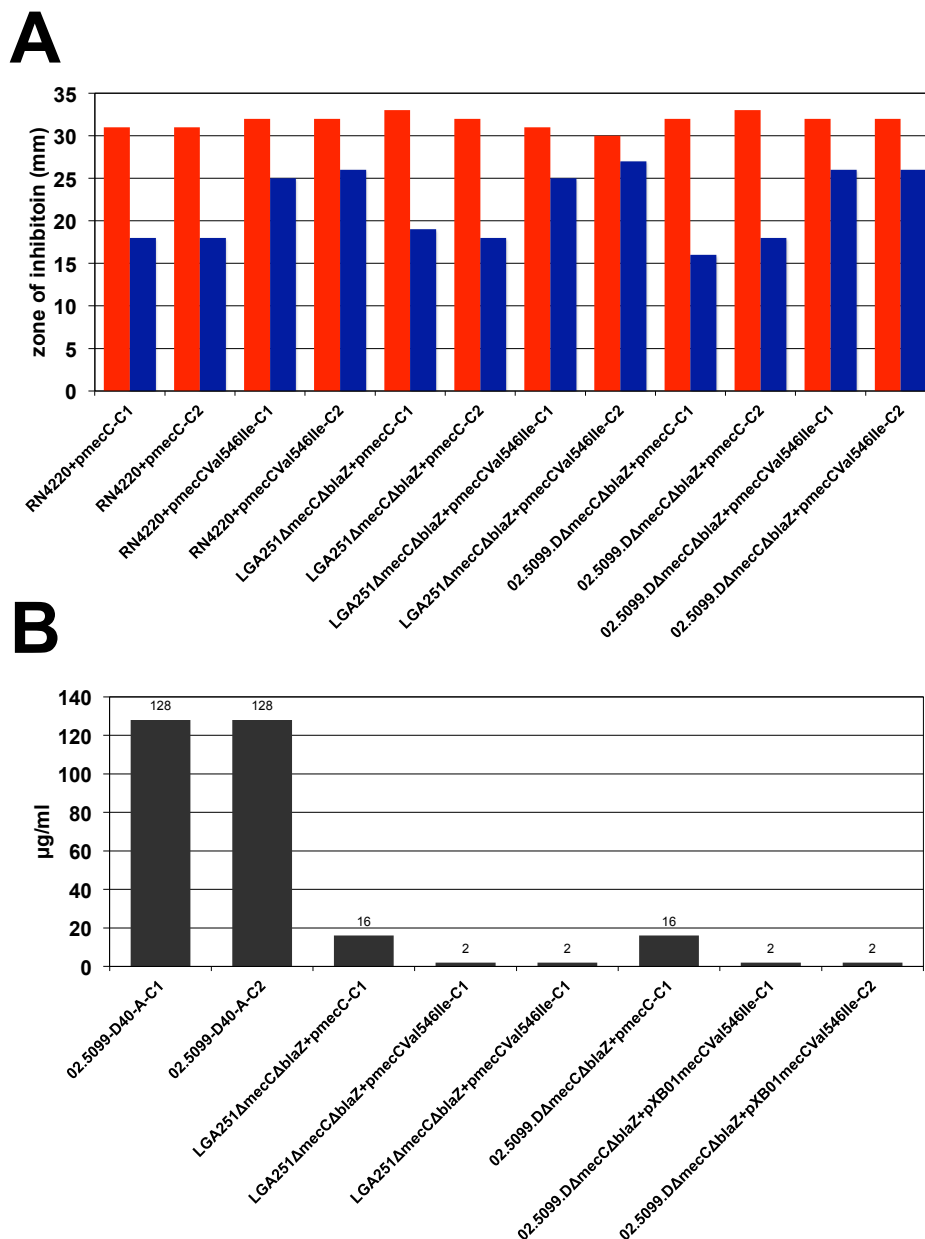


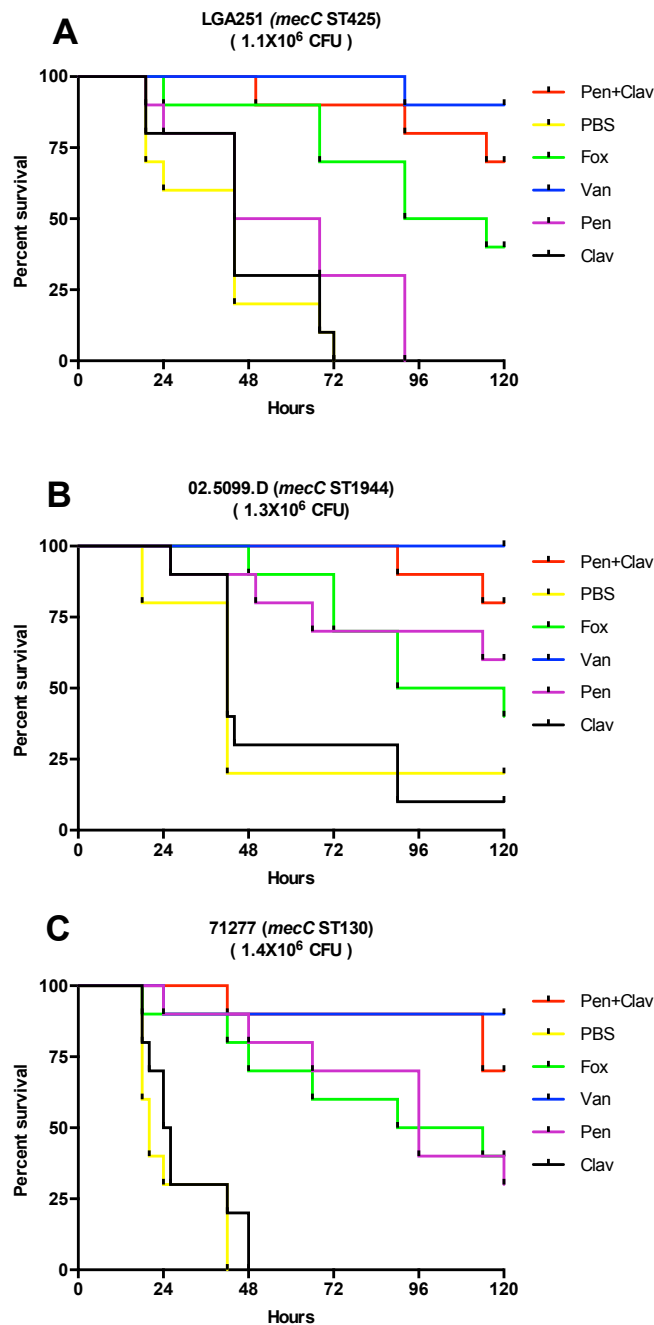
## Supplementary Material



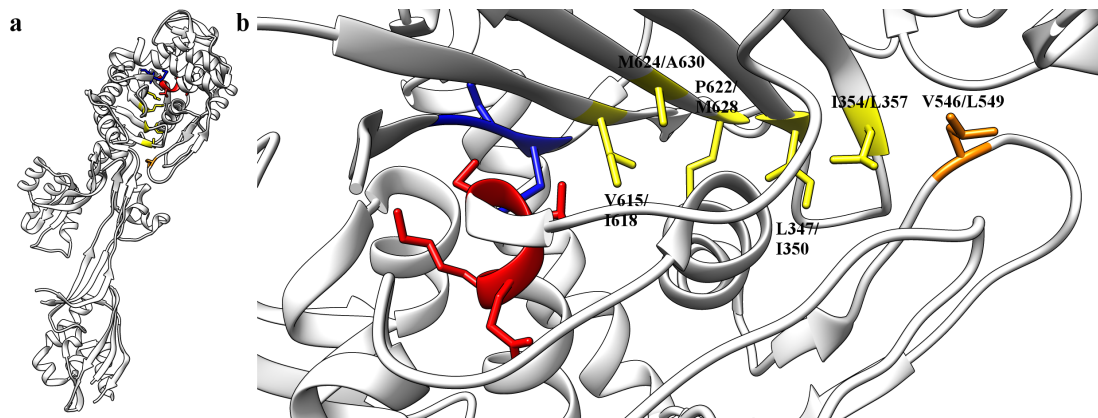
**Figure S1:** The effect of different incubation temperatures (°C) on antimicrobial susceptibility testing for *mecC*-MRSA strains (n=10). Penicillin (PG) and cefoxitin (FOX) disc diffusion was performed on Iso-Sensitest agar (ISA) with or without 15 µg/ml clavulanic acid.



**Figure S2:** Effect of PBP2c Val546Ile substitution. (A) Comparison of disc diffusion results for penicillin (PG) (red bars) and cefoxitin (FOX) (blue bars) of strains RN4220, LGA251Δ*mecC*Δ*blaZ* and 02.5099.DΔ*mecC*Δ*blaZ* carrying plasmid borne copies of wildtype *mecC* and *mecC*<sup>Val546Ile</sup>. C1 and C2 denote clone 1 and clone 2, respectively. (B) Comparison of minimum inhibitory concentrations for cefoxitin (FOX) of wildtype strain and strains RN4220, LGA251Δ*mecC*Δ*blaZ* and 02.5099.DΔ*mecC*Δ*blaZ* carrying plasmid borne copies of wildtype *mecC* and *mecC*<sup>Val546Ile</sup>. C1 and C2 denote clone 1 and clone 2, respectively.



**Figure S3:** Biological repeats of experimental treatment of *Galleria mellonella* infected by *mecC*-MRSA strains: (A) LGA251 (B) 02.5099.D (C) 71277. Ten larvae in each group were experimentally infected and then treated at 2, 24 and 48 hours with Vancomycin ( $50 \text{ mg/kg} / 6.73 \times 10^{-9} \text{ mol}$ ), Penicillin ( $20 \text{ mg/kg} / 1.12 \times 10^{-8} \text{ mol}$ ), Clavulanic acid ( $20 \text{ mg/kg} / 1.69 \times 10^{-8} \text{ mol}$ ), Penicillin / Clavulanic acid, Cefoxitin ( $40 \text{ mg/kg} / 1.78 \times 10^{-8} \text{ mol}$ ), and PBS alone. Fig. shows a representative image of a single experiment.



**Figure S4:** Structural view of PBP2a (A) in context of fold (B) close-up. Residues numbered first PBP2c and PBP2a second. SxxK motif in red and KT/SG motif in blue. Produced using UCSF Chimera (<http://www.cgl.ucsf.edu/chimera/>); from PDB code 1VQQ (PBP2a).

**Table S3:** Comparison of minimum inhibitory concentration of penicillin (PG) and cefoxitin (FOX) after *in vitro* selection for penicillin-resistance in strains lacking the SCCmec type XI encoded *blaZ*. C1, C2, C3 indicate multiple individual colonies tested.

Strains	PG $\mu\text{g/ml}$	FOX $\mu\text{g/ml}$
LGA251 $\Delta$ <i>blaZ</i>	0.015	8
LGA251 $\Delta$ <i>blaZ</i> -D9-C1	8	128
LGA251 $\Delta$ <i>blaZ</i> -D9-C2	8	128
LGA251 $\Delta$ <i>blaZ</i> -D9-C3	8	128
02.5099.D $\Delta$ <i>blaZ</i>	0.0625	8
02.5099.D $\Delta$ <i>blaZ</i> -D13-C1	16	128
02.5099.D $\Delta$ <i>blaZ</i> -D13-C2	16	>128
02.5099.D $\Delta$ <i>blaZ</i> -D13-C3	16	>128

Note for the table: 20 hours, at 35°C. MIC breakpoint for PG is 0.12  $\mu\text{g/ml}$ ; for FOX is 4  $\mu\text{g/ml}$ . PG concentration range is 0.03125-128  $\mu\text{g/ml}$ . FOX concentration range is 1-128  $\mu\text{g/ml}$ . D9 = day 9 and D13 = day 13.

**Table S5:** Bacterial strains, plasmids and primers used in this study.

Bacterial strains, plasmids or primers	Description	Reference
<b><i>E. coli</i> strains</b>		
DC10B	$\Delta dcm$ in the DH10B background; Dam methylation only	(1)
<b><i>S. aureus</i> strains</b>		
RN4220	Derived from NCTC8325-4, selected for transformability with DNA from <i>E. coli</i>	(2)
RN4220(pXB01mecC)	RN4220 with wildtype <i>mecC</i>	This study
RN4220(pXB01mecCVal546Ile)	RN4220 with mutated <i>mecC</i>	This study
LGA251	ST425, <i>mecC</i> reference strain	(3)
LGA251 $\Delta$ <i>mecC</i>	LGA251 with <i>mecC</i> deletion	This study
LGA251 $\Delta$ <i>blaZ</i>	LGA251 with <i>blaZ</i> deletion	This study
LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i>	LGA251 with <i>mecC</i> and <i>blaZ</i> deletion	This study
LGA251 $\Delta$ <i>mecC</i> (pXB01)	LGA251 $\Delta$ <i>mecC</i> with empty plasmid pXB01	This study
LGA251 $\Delta$ <i>blaZ</i> (pXB01)	LGA251 $\Delta$ <i>blaZ</i> with empty plasmid pXB01	This study
LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01)	LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> with empty plasmid pXB01	This study
LGA251 $\Delta$ <i>mecC</i> (pXB01- <i>mecC</i> )	LGA251 $\Delta$ <i>mecC</i> complemented with pXB01- <i>mecC</i>	This study
LGA251 $\Delta$ <i>blaZ</i> (pXB01- <i>blaZ</i> )	LGA251 $\Delta$ <i>blaZ</i> complemented with pXB01- <i>blaZ</i>	This study
LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01- <i>mecC</i> )	LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> complemented with pXB01- <i>mecC</i>	This study
LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01- <i>blaZ</i> )	LGA251 $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> complemented with pXB01- <i>blaZ</i>	This study
02.5099.D	CC130, <i>mecC</i> clinical isolate	(3)
02.5099.D $\Delta$ <i>mecC</i> (pXB01)	02.5099.D with <i>mecC</i> deletion	This study
02.5099.D $\Delta$ <i>blaZ</i> (pXB01)	02.5099.D with <i>blaZ</i> deletion	This study
02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01)	02.5099.D with <i>mecC</i> and <i>blaZ</i> deletion	This study
02.5099.D $\Delta$ <i>mecC</i>	02.5099.D $\Delta$ <i>mecC</i> with empty plasmid pXB01	This study
02.5099.D $\Delta$ <i>blaZ</i>	02.5099.D $\Delta$ <i>blaZ</i> with empty plasmid pXB01	This study
02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i>	02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> with empty plasmid pXB01	This study
02.5099.D $\Delta$ <i>mecC</i> (pXB01- <i>mecC</i> )	02.5099.D $\Delta$ <i>mecC</i> complemented with pXB01- <i>mecC</i>	This study
02.5099.D $\Delta$ <i>blaZ</i> (pXB01- <i>blaZ</i> )	02.5099.D $\Delta$ <i>blaZ</i> complemented with pXB01- <i>blaZ</i>	This study
02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01- <i>mecC</i> )	02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> complemented with pXB01- <i>mecC</i>	This study
02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> (pXB01- <i>blaZ</i> )	02.5099.D $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i> complemented with pXB01- <i>blaZ</i>	This study
<b>Plasmids</b>		
pRMC2	<i>E. coli</i> - <i>S. aureus</i> shuttle vector, Amp <sup>r</sup> , Cm <sup>r</sup>	(4)
pXB01	<i>E. coli</i> - <i>S. aureus</i> shuttle vector pRMC2 with <i>bla</i> deletion; Cm <sup>r</sup>	This study
pXB01- <i>mecC</i>	pXB01 with <i>mecC</i> gene from LGA251	This study
pXB01- <i>mecC</i> <sup>Val546Ile</sup>	pXB01 with <i>mecC</i> gene having a G to A mutation at position 1636 producing a Val546Ile substitution	This study
pXB01- <i>blaZ</i>	pXB01 with <i>blaZ</i> gene from LGA251	This study
pIMAY	pIMC5 with tetracycline inducible <i>secY</i> antisense from pKOR1; Cm <sup>r</sup>	(1)
pIMAY $\Delta$ <i>mecC</i>	Modified pIMAY for deleting <i>mecC</i> gene, encompassing the entire gene, insert amplified from LGA251	This study
pIMAY $\Delta$ <i>blaZ</i>	Modified pIMAY for deleting <i>blaZ</i> gene, encompassing the entire gene, insert amplified from LGA251	This study
pIMAY $\Delta$ <i>mecC</i> $\Delta$ <i>blaZ</i>	Modified pIMAY for deleting <i>blaZ</i> gene, encompassing the entire gene, insert amplified from LGA251 $\Delta$ <i>mecC</i>	This study
<b>Primers</b>		
	<b>Sequence (5'-3') restriction site underlined</b>	<b>Restriction site</b>
de- <i>bla</i> -F	ATATATATGCGGCCGCCTGTCAGACCAAGTTTACTC	NotI
de- <i>bla</i> -R	ATGCGCATGCGGCCGCACTCTTCTTTTCAATATTA TT	NotI
$\Delta$ <i>mecC</i> _A	ATCGGGTACCTAATATGAATGAGTTCATGGC	KpnI
$\Delta$ <i>mecC</i> _B	CTCTTCTCCTTTTGTACTAC	
$\Delta$ <i>mecC</i> _C	TAGTACAAAAGGAGGAAGAGTTCGAATACTTCTTTTG ACTTGG	
$\Delta$ <i>mecC</i> _D	ATCGGAGCTCGTATCGCCACTTTTATTTTGG	SacI
$\Delta$ <i>mecC</i> _outF	GCTAAATGTAATGCTGGATTGAACC	
$\Delta$ <i>mecC</i> _outR	TCTTGTCCATCTGGATAAACAACG	

$\Delta blaZ$ _A	ATATGGTACCAAATATCCCGAGTGATTATCC	KpnI
$\Delta blaZ$ _B	CCTACGTTAGCACCATATTTTTTTTCG	
$\Delta blaZ$ _C	AAATATGGTGCTAACGTAGGGTAAGTGAGGTTGCTG AAATTGTA	
$\Delta blaZ$ _D	ATATGAGCTCCATCATAGAAGTTCCAGACG	SacI
$\Delta blaZ$ _outF	TGCCCGCATTGCATTAGCATTAGGA	
$\Delta blaZ$ _outR	AAATATTGGAAGCAAGCCATAGCAG	
$\Delta blaZ\Delta mecC$ _A	ATATGGTACCGGAGATCTAATAGACAAGTC	KpnI
$\Delta blaZ\Delta mecC$ _outF	GACCATTGGATTCTCTACTTCATCT	
catup	GGATTTTTCGCTACGCTCAA	
catdown	AAAGTACAGTCGGCATTATCTCA	
IM151	TACATGTCAAGAATAAACTGCCAAAGC	(1)
IM152	AATACCTGTGACGGAAGATCACTTCG	(1)
<i>blaZ</i> -F- KpnI	CGCGGGTACCTATATAGGAGGAATAAAATTG	KpnI
<i>blaZ</i> -R-SacI	GCGCGAGCTCTTACTCATTAAATTTTTTAG	SacI
<i>mecC</i> -F-KpnI	CGCGGGTACCCAAAAGGAGGAAGAGATG	KpnI
<i>mecC</i> -R-SacI	GCGCGAGCTCTTCGAATTACTGATCTAT	SacI
<i>mecC</i> f	GTTCACACCTCACTTCTTAAC	
<i>mecC</i> r	GTAATCTTTGTGGAAACGC	
<i>mecC</i> m	GCCGTGTTTATCCATTGAAC	

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