

SUPPLEMENTARY DATA

Supplementary Table 1: primers used for qPCR

Primer name	Sequence	Origin (<i>C. lusitaniae</i> , Broad Institute) ^{a)}	Reference
MFS7-R	GGAATGTAAGGCATGGCAGT	supercont_1.2 (1458215-1459123)	(1)
MFS7-F	ATGGTTTTCTCGCCAATGTC	supercont_1.2 (1458215-1459123)	(1)
ABC12-R	GTGCTTCAACCACCACCTTT	supercont_1.3 (1264183-1268415)	(1)
ABC12-F	TGATTGAGTCTGCGGACAAG	supercont_1.3 (1264183-1268415)	(1)
ABC15-F	GTGCTAAGGCGTTTTTCGAG	supercont_1.3 (1682175-1688644)	This study
ABC15-R	TGAGATGCACGCCAGTAGTC	supercont_1.3 (1682175-1688644)	This study
ABC9-F	TCCATTGCAGAGGCTTCTTT	supercont_1.2 (1950802-1957241)	This study
ABC9-R	GTACGCATCCTGGGAACACT	supercont_1.2 (1950802-1957241)	This study
ACT1-F	GTATCGCTGAGCGTATGCAA	supercont_1.4 (88657-89739)	This study
ACT1-R	GATGGATGGTCCAGACTCGT	supercont_1.4 (88657-89739)	This study
ERG11-F	AACACCAGCTACGTGGTTCC	supercont_1.6 (218130-219707)	This study
ERG11-R	AGGCAAACCTGCTCTCCGATA	supercont_1.6 (218130-219707)	This study

^{a)} :Numbering gives the coordinates of the gene of interest according to supercontig data

Supplementary Table 2: primers used for CRISP-Cas9 system

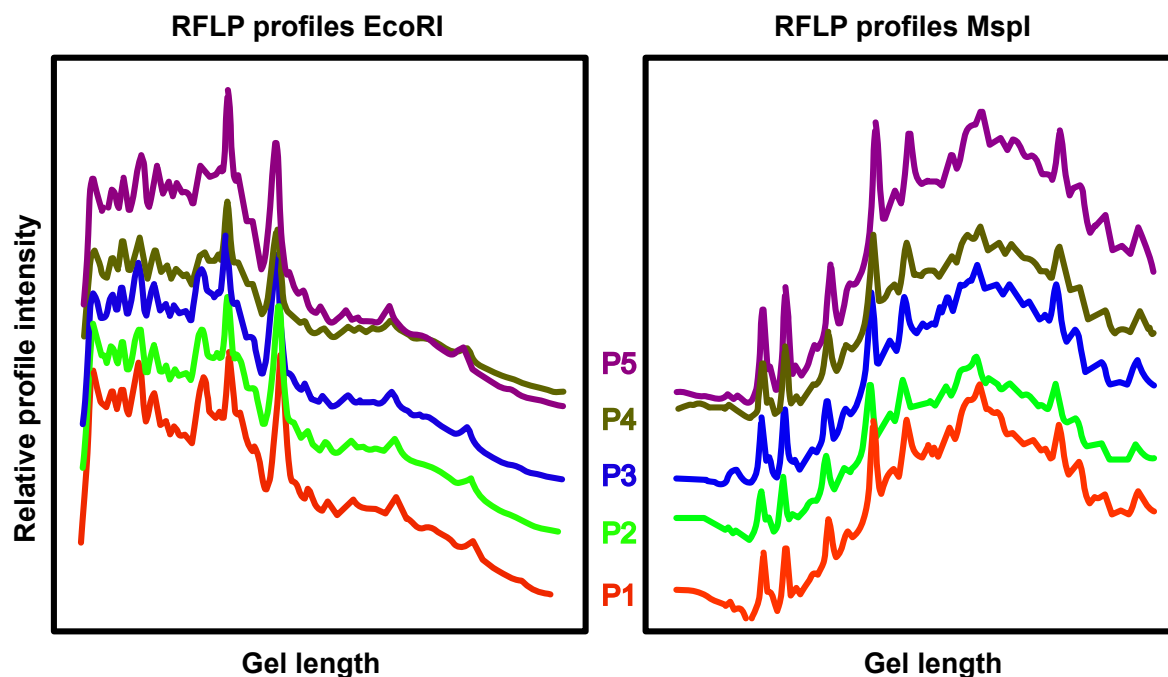
Primer Name	Sequence	Purpose
FKS_crisp_R ^{a)} FKS_crisp_F	GTTGATAACGGACTAGCCTTATTTAACTTG CTATTTCTAGCTCTAAAACA <u>AATATTCAGAAT</u> <u>CGTACTACGATCATT</u> TATCTTTCACTGCGGA GAAGTTTCGAACGCCGAAACATGCGCA TGCGCATGTTTCGGCGTTCGAAACTTCTCCG CAGTGAAAGATAAATGATCGTAGTACGATT <u>CTGAATATTGTTTTAGAGCTAGAAATAGCA</u> AGTTAAAATAAGGCTAGTCCGTTATCAAC	Generation of guide-pMEL10 sequence by complementary assembly of primers
FKS left arm_S636Y ^{b)} FKS right arm_S636Y	GATAGATGGATGTCCTATTTAGTTTGGGTTA CTGTTTTTGCTGCCAAATATTCAGAAT ATTAC <i>TACTTTTTAG</i> CATTGCAGTGGTGGACAAAATTCTAATTGG ATCTCTCAAAGATAAA ACTAAAAAGTAGTAAT <i>ATTCT</i>	Generation of repair fragment for S636Y by overlapping PCR
FKS left arm FKS right arm_S643Y	GATAGATGGATGTCCTATTTAGTTTGGGTTA CTGTTTTTGCTGCCAAATATTCAGAAT CGTAC <i>TACTTTTTAG</i> CATTGCAGTGGTGGACAAAATTCTAATTGG ATCTCTCAAATATAAA ACTAAAAAGTAGTAC <i>GATTCT</i>	Generation of repair fragment for S643Y by overlapping PCR
FKS right arm S643P	CATTGCAGTGGTGGACAAAATTCTAATTGG ATCTCTCAAAG GTA AA ACTAAAAAGTAGTA CGATTCT	Generation of repair fragment for S643P by overlapping PCR using primer FKS left arm
FKS1verif left FKS1verif right	TCTACAAGGCGTTATGTTGCAT CAATCTTGGGTTGCACTTTACA	<i>FKS1</i> verification primers
p426 CRISPR rv p426 CRISPR fw	GATCATTTATCTTTCACTGCGGAGAAG GTTTTAGAGCTAGAAATAGCAAGTTAAAAT AAGGCTAGTC	Inverse PCR with pMEL10

a) The 20 nt sequence of the *FKSI* guide is underlined

b) Sequences in italics are overlapping sequences; Bold characters indicate mutated bases

Supplementary Table 3 : *S. cerevisiae* strain descriptions

Strain number	Parent	Genotype	Reference
IMX581	CEN.PK113-5D	<i>MATa ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 FKS1</i>	(2)
DSY4762	IMX581	<i>MATa ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 FKS1^{S636Y}</i>	This study
DSY4763	IMX581	<i>MATa ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 FKS1^{S643P}</i>	This study
DSY4764	IMX581	<i>MATa ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 FKS1^{S643Y}</i>	This study



Supplementary Figure 1 : Profile plots of RFLP analysis. Gel electrophoresis pictures of Fig.2. were analysed with the ImageJ (version 1.49P) and profile plots were generated. The X-axis represents relative densities while Y-axis represents length of profiles.

References

- (1) **Reboutier D, Piednoel M, Boisnard S, Conti A, Chevalier V, Florent M, Gibot-Leclerc S, Da Silva B, Chastin C, Fallague K, Favel A, Noel T, Ruprich-Robert G, Chapeland-Leclerc F, Papon N.** 2009. Combination of different molecular mechanisms leading to fluconazole resistance in a *Candida lusitanae* clinical isolate. *Diagn Microbiol Infect Dis* **63**:188-193.
- (2) **Mans R, van Rossum HM, Wijsman M, Backx A, Kuijpers NG, van den Broek M, Daran-Lapujade P, Pronk JT, van Maris AJ, Daran JM.** 2015. CRISPR/Cas9: a molecular Swiss army knife for simultaneous introduction of multiple genetic modifications in *Saccharomyces cerevisiae*. *FEMS Yeast Res* **15**:fov004-fov004.