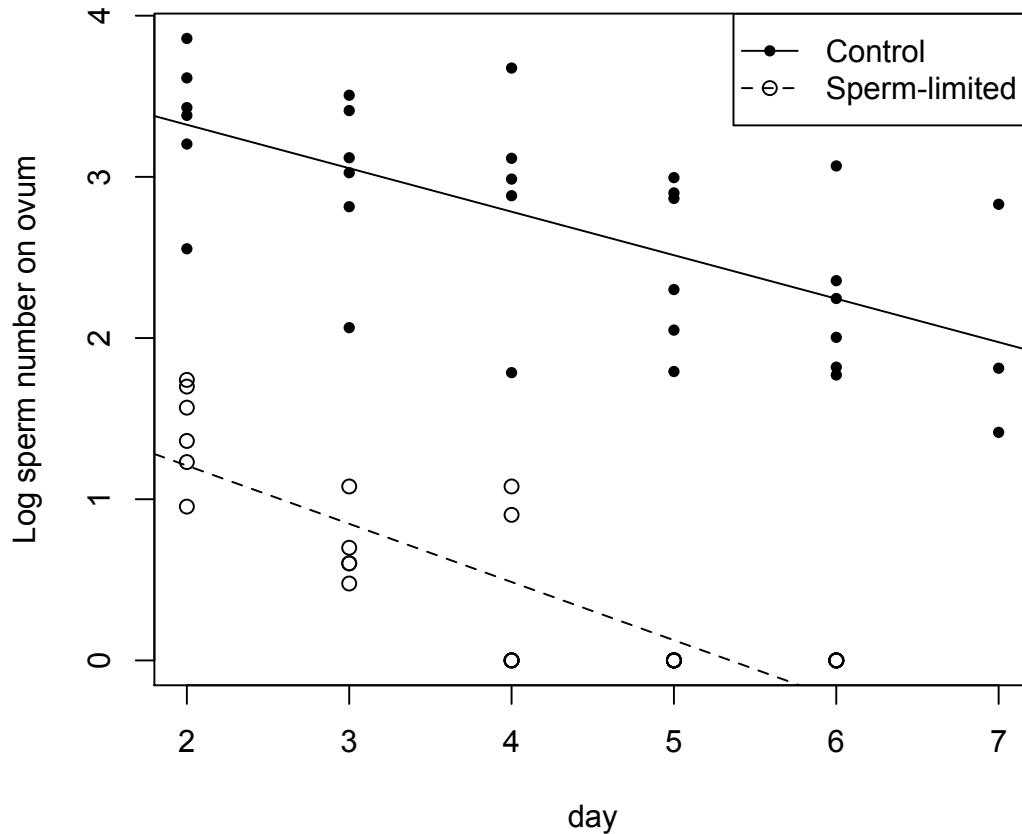


1 **ESM 1 - Figure S1**



2

3 Figure S1. The decline in sperm numbers found on the perivitelline layer of ova

4 following control ( $y = -0.27 + 0.06x$ ;  $R^2 = 0.42$ ;  $F_{(1,30)} = 23.56$ ;  $p < 0.0001$ ) and

5 sperm-limited ( $y = -0.36 + 0.04x$ ;  $R^2 = 0.72$ ;  $F_{(1,29)} = 76.39$ ;  $p < 0.0001$ )

6 inseminations, over the course of the laying sequence in domestic fowl. There

7 was no significant interaction between the effects of insemination mode (control

8 versus sperm-limited) and time ( $F_{(1,59)} = 1.603$ ;  $p = 0.21$ ) on sperm numbers,

9 indicating that the rate of sperm loss did not differ significantly between control

10 and sperm-limited trials.

11

## 12 **ESM 2: Artificial Insemination in Zebra Finches: Methods**

13

14 In preliminary trials of artificial insemination in the zebra finch (carried out  
15 under license from the UK Home Office and approved by local ethical review), we  
16 noted that successful inseminations consistently resulted in extremely low  
17 sperm numbers reaching and penetrating the ovum (as shown by the number of  
18 sperm and sperm penetration holes in the perivitelline layer; Wishart 1987). We  
19 do not know why these inseminations resulted in such low numbers of sperm  
20 reaching the ovum, but the most likely explanation is that sperm were deposited  
21 into the female at a location more external than they would be under natural  
22 insemination, limiting their ability to enter the reproductive tract. This method  
23 of “artificial insemination” has proved a useful tool for limiting the number of  
24 sperm reaching the ova (see article).

25 To encourage females under the ‘low’ treatment to lay a ‘typical’ clutch (i.e. 5-6  
26 eggs, laid on consecutive days approximately one week after pair formation),  
27 male-female pairs were maintained in paired cages (single cage dimensions 0.6 x  
28 0.5 x 0.4m) separated by a wire divider extending into a double nest-box. This  
29 allowed the pair to interact visually and acoustically but prevented copulations.  
30 Female zebra finches will build a nest and lay eggs normally under these  
31 conditions (NH, pers. obs.). In the control treatment, male-female pairs were  
32 kept in the same cages as used in the sperm-limited treatment, but without a  
33 wire divider. All birds were provided with food and water *ad libitum*,  
34 supplemented with boiled egg and lettuce and maintained at 20°C on a 12h  
35 photoperiod.

36 All females included in the experiment (under both control and sperm-limited  
37 treatments) were habituated to the “artificial insemination” procedure via daily  
38 handling and mock inseminations carried out under license. A mock  
39 insemination involved placing 5  $\mu$ l of sterile water onto the female’s cloaca. For  
40 the ‘low’ treatment, habituated females were artificially inseminated (under  
41 license) between 1400-1600h on the day their first egg was laid, with 5 $\mu$ l semen  
42 taken directly from the seminal glomerus of dissected males. Based on sperm  
43 concentration data from a previous study on the same zebra finch population  
44 (Bennison *et al.* 2015), we estimated the approximate number of sperm in 5 $\mu$ l to  
45 be  $0.675 \times 10^6$ , which is slightly but not remarkably fewer than the number in a  
46 natural ejaculate ( $5.29 \times 10^6 \pm 1.56 \times 10^6$  for rested males, decreasing to  $0.166 \times$   
47  $10^6 \pm 0.08 \times 10^6$  in unrested males; Birkhead *et al.* 1995). Artificial insemination  
48 was identical in procedure to the mock inseminations, except that fresh semen  
49 instead of sterile water was placed on the cloaca. The female was held until the  
50 semen had disappeared into the cloaca (usually within 30s) and then returned to  
51 the cage. Females under the control treatment received a mock insemination at  
52 the same time.

53

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62

63 **ESM 3: Verification of *glmer* models using Bayesian generalised linear**  
64 **models**

65 Bayesian generalised linear models with Markov chain Monte Carlo (MCMC)  
66 estimation in *MCMCglmm*, R Version 3.1.2 (Hadfield 2010) were used to verify  
67 the *glmer* results concerning the effect of the sperm limitation treatment on  
68 fertility and embryo survival (reported in the main text). Due to a lack of  
69 variation in the data, the effect of sperm limitation on fertility in the zebra finch,  
70 and embryo survival in the domestic fowl, was not reanalysed. Results of the  
71 other *glmer* analyses were supported by the results of the *MCMCglmm* analyses:  
72 sperm limitation negatively influenced (a) embryo survival in the zebra finch  
73 (PDM =  $-0.38 \times 10^3$ , L-95% CI =  $-0.73 \times 10^3$ , U-95% CI =  $-0.11 \times 10^3$ , pMCMC <  
74 0.001), and (b) fertility in the domestic fowl (PDM =  $-0.20 \times 10^3$ , L-95% CI =  $-0.38$   
75  $\times 10^3$ , U-95% CI =  $0.03 \times 10^3$ , pMCMC = 0.008).

76

77 Reference

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