Supplementary information - "High-throughput sequencing enhanced phage display enables the identification of patient-specific epitope motifs in serum"

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Supplementary Table S1. Primer and barcode DNA sequences used for PCR. The barcode element has been italicised. The reverse primer in the bottom row was used for all reactions.

Supplementary Table S2. Number of sequences in each step of the processing. The provided numbers are the number of sequences that remains at the particular stage. The percentage denotes the percentage of remaining sequences compared to the initial assignment based on barcodes. In brief the steps were (i) associating sequences with a sample based on barcode, (ii) calling 7-mer peptides and (iii) filtering peptides based NNK codon pattern and (iv) quality filtering. The steps are covered in greater detail in the Materials and Methods section.

Supplementary Figure S1. (A) Derivative sequences due to high-throughput sequencing errors. The relationship between the prevalence of f a high-frequent peptide ("High Sequence Counts") and the prevalence of similar peptides where the DNA sequence varies at a single position ("Low Sequence Counts") has been plotted. Note that the axes have different scales. The horizontal rows of observations that are visible at high prevalences corresponds to the various derivative sequences that are observed from a single high-frequent peptide. **(B)** As (A) but plotting the prevalence of derivative sequences where there are two deviations in the DNA sequences compared to the high-frequent peptide. **(C)** As (A) but depicting the sum of all derivative sequences on the "Low Sequence Counts" axis for each high-frequent peptide. A linear relationship between the prevalence of the high-frequent peptide and the sum of the derivative sequences is observed, where peptides with higher prevalences give rise to more derivative sequences. Such a relation is to be expected if the derivative sequences are due to sequencing errors.

Threshold details: The applied thresholds of 500 for single-base deviations (shown in A) and 10,000 for double-base deviations (shown in B) where empically determined. Taking the 500-fold threshold as an example, sequences were removed if they were at least 500 times less prevalent than a similar sequence (differing by a single base pair) in the same sample. Translating this to a per-base error rate was done by the formula P^{20} × ((1-P) / 3) = 1/500 × P^21, where P is the per-base accuracy and 1-P is the per-base error rate. (1-P) /3 assumes that the change to any of the three other bases are equally distributed. Solving for P yields P = 0.994, thus the 500-fold threshold corresponds to a per-base sequencing error rate of 0.6%.

Supplementary Figure S2. Derivative sequences due to high-throughput sequencing errors give rise to "spill-over" between samples. The relationship between the prevalence of a high-frequent peptide ("Count High") and the prevalence of the same peptides in another subject ("Count Low"), analysed on the same sequencing chip has been plotted. Note that the axes have different scales. A maximum of 50,000 and 1,000 is plotted for "Count High" and "Count Low", respectively, has been plotted. The summed prevalence for each peptide across the three selection rounds is depicted. A Pearson correlation coefficient has been calculated and included.

Supplementary Table S3. Number of sequences filtered due to high-throughput sequencing errors. "Derivative" filtering comprise the removal of the derivative sequences plotted in Supplementary Figure S1, i.e. peptides in a sample that are very similar to a high-frequent peptide in the same sample. "Spill-over" filtering encompasses the removal of the derivative sequences plotted in Supplementary Figure S2, i.e. peptides in a sample that are very similar to a high-frequent peptide in a different subject, analysed on the same sequencing chip. The provided numbers are the number of peptide sequences that remains at the particular stage. The percentage denotes the percentage of remaining peptides after the specific filtering step.

Supplementary Table S4. Number of unique peptide sequences filtered due to high-throughput sequencing errors. The table is similar to Supplementary Table S2, however, the number of unique peptides is given instead of total peptide sequence numbers. "Derivative" filtering comprise the removal of the derivative sequences plotted in Supplementary Figure S1, i.e. peptides in a sample that are very similar to a highfrequent peptide in the same sample. "Spill-over" filtering encompasses the removal of the derivative sequences plotted in Supplementary Figure S2, i.e. peptides in a sample that are very similar to a highfrequent peptide in a different subject, analysed on the same sequencing chip. The percentage denotes the percentage of remaining unique peptides after the specific filtering step.

Supplementary Figure S3. The selection process in control samples. **(A)** A plot of the number of unique peptides identified in each control sample for selection round 1-3. **(B)** A stacked bar chart showing the combined frequency of certain rank intervals. Specifically, the frequency of the most frequent peptide (Top1) along with the combined frequencies of the peptides ranked 2-5, 6-20, 21-50, 51-200 and below 200, based on their frequency in a sample. The average of the combined frequencies for all control samples is shown.

Supplementary Table S5. The ten most frequent peptides and their frequencies for each sample. The peptides are separated according selection round (Round 1-3). The peptides with a grey overlay were also observed in a control sample with a prevalence > 1.

Supplementary Table S6. Dominating TUP candidates. Peptides that have been identified with a peptide prevalence >1 in a minimum of 3 different samples, including at least 1 control sample, are listed. The number of samples were a given peptide was observed is listed in the "# of samples" column.

Supplementary Table S7. List of peptides in the significant peptide clusters separated according to selection round.

Supplementary Table S8. The number of peptides in the significant clusters in selection round 2 and 3 separated according to the patient sample they were derived from.

Supplementary Figure S4. Intensity based on control IgE reactivity to Ara h 1 as measured by peptide micro-arrays. The right-bound rolling median (window size 12) of the mean intensity of the triplicate 12 mer peptides overlapping each residue is shown for every control sample. The start position of the epitope identified in the patients, at position 136, has been specifically marked.

Supplementary Figure S5. Frequency of peptides that were included in the significant cluster. The logarithmic peptide frequencies are separated according to selection round (1-3). The horizontal line represents the suggested detection limit of traditional phage display approaches, assuming that 1000 phage colonies are assayed thereby detecting peptides with a frequency above 0.001. Peptides with a rank score of 0 have not been plotted.