Supplementary Information

Brightening Gold Nanoparticles: New Sensing Approach Based on Plasmon Resonance Energy Transfer

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Figure S1. a) Normalized absorption spectra of RdBS and RdBS+F. Absorption spectra of RdBS $(1.0 \times 10^{-5}$ M) before (red line) and after (black line) the addition of NaF (10.0 equiv.) in water. The inset is the photograph of (a) RdBS solution, (b) RdBS +NaF. b) Absorption spectrum of compound 2 in water.

Synthesis and characterization of 60 nm gold nanoparticles^[1]

Seed gold nanoparticles were prepared by the citrate-mediated reduction of HAuCl₄. 50 mL of 0.01 wt% HAuCl4 was heated to reflux with vigorous stirring and then 5 mL sodium citrate (38.8 mM) was added quickly to the solution. The mixed solution was continued to heat for 15 min, stopped heating and kept stirring for an additional 15 min. The resulting solution of colloidal particles was filtered and characterized by an absorption maximum at 521 nm using an Ocean optical USB 2000+ UV-Vis spectrometer. These particles were then used as seed particles for the synthesis of 60 nm gold particles. To 25 mL of water, 1 mL of preformed seed gold particles and 100 μL of 0.2 M NH2OH·HCl was mixed. The mixture was stirred vigorously at room temperature and 3.0 mL of 0.1 wt% HAuCl₄·3H₂O was added drop-wise. Diameters were characterized by UV-Vis and SEM.

Figure S2. Characterization of gold nanoseeds and 60 nm GNPs by UV-Vis and SEM. a) UV-Vis spectrum of Au seeds and 60 nm gold nanoparticles; b) TEM image of 60 nm gold nanoparticles.

Figure S3. Dark-field images and scattering spectra of GNPs before and after modification of RdBS. a) The dark-field image of 60 nm gold nanoparticles. b) The scattering spectrum of a typical gold nanoparticle in (a); c) The dark-field image of RdBS modified 60 nm gold nanoparticles; d) The scattering spectrum of a single gold nanoparticle in (c).

Figure S4. Detailed experimental configuration of a dark-field microscopy system. The collected scattering light is imaged by a true color charge-coupled device (CCD) camera and analyzed by a spectrometer.

Figure S5. TEM images of RdBS modified GNPs (a) and after (b) the addition of fluoride ions.

Figure S6. TLC identification test before and after the reaction between RdBS and F.

Figure S7. MS spectrum of the product after the reaction of RdBS with F. Synthesis of compound RdBS

Glycol (3.0g, 48.5 mmol), DCC (1.03 g, 5 mmol) and DMAP (0.6 g, 5 mmol) were dissolved in 20 mL dry DCM, a solution of lipoic acid (1.0 g, 48.5 mmol) in 10 mL dry DCM was added dropwise at 0 ºC. Then the mixture was stirred at room temperature for 5 h before filtered. The filtrate was washed with brine, NaHCO₃ and concentrated in vacuum. The residue was purified by silica gel column chromatography to obtain the pure product as a yellow oil 1 (2.1 g) . ¹H NMR (400 m) MHz, CDCl₃) δ 4.24 – 4.20 (m, 2H), 3.83 (dd, J = 9.3, 5.6 Hz, 2H), 3.58 (dq, J = 12.7, 6.4 Hz, 1H), 3.24 – 3.05 (m, 2H), 2.53 – 2.42 (m, 1H), 2.42 – 2.33 (m, 2H), 2.06 (t, J $= 5.7$ Hz, 1H), 1.92 (dq, J = 13.8, 6.9 Hz, 1H), 1.78 – 1.60 (m, 3H), 1.57 – 1.38 (m, 2H).

The synthetic procedure of 2 was the same as that of 1. ¹H NMR (400 MHz, CDCl₃) δ

8.48 – 8.39 (m, 1H), 7.80 – 7.69 (m, 2H), 7.26 (dd, *J* = 6.9, 1.5 Hz, 1H), 7.11 (d, *J* = 9.5 Hz, 2H), 6.88 (dd, *J* = 9.5, 2.2 Hz, 2H), 6.82 (d, *J* = 2.1 Hz, 2H), 4.18 – 4.10 (m, 2H), 3.63 (dd, *J* = 14.4, 7.1 Hz, 10H), 2.33 – 2.18 (m, 4H), 1.33 (t, *J* = 7.1 Hz, 12H).

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Diphenyldichlorosilane (2.28 mL, 1.1 mmol), Et3N (2.09 mL) was dissolved in50 mL dry DCM, then a solution of 1 (2.75 g, 1.1mmol) in 20 mL dry DCM was added dropwise at 0 ºC under the protection of argon. The solution was stirred at room temperature for 6 h, then refluxed for 36 h. The solvent was concentrated in vacuum and redissolved in 1:1 ether-hexane 10 mL. The solution was suction filtered and reconcentrated. The crude product 3 was used without further purification for the next step.

Compound 2 (0.5 g, 1.0 mmol) and Et3N (2 mL) were dissolved in 5 mL dry DCM, a solution of crude compound 3 (0.5 g) in 5 mL dry DCM was added dropwise at 0 °C. The mixture was stirred at room temperature for 3 h, and then quenched with saturated NaHCO₃. The solution was wash with H₂O, brine. The residue was purified by silica gel column chromatography to obtain the pure product RdBS 0.3g. ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.17 (d, J = 7.9 Hz, 1H), 7.84 (t, J = 7.5 Hz, 1H), 7.68 (t, J = 7.7 Hz, 1H), $7.47 - 7.35$ (m, 5H), $7.34 - 7.28$ (m, 5H), $7.23 - 7.16$ (m, 1H), 6.96 (t, J = 8.5 Hz, 2H), 6.85 – 6.69 (m, 4H), 4.12 – 4.02 (m, 2H), 3.87 – 3.78 (m, 2H), 3.74 (s, 2H), 3.60 (m, 8H), 3.12 (m, 3H), 2.45 – 2.34 (m, 1H), 2.19 – 2.00 (m, 4H), 1.86 (m, 6H), 1.58 (m, 6H), 1.36 – 1.17 (m, 12H). 13C NMR (101 MHz, CDCl3) δ 173.87, 173.39, 157.74, 155.50, 134.88, 134.73, 133.31 – 133.20, 132.03, 131.70131.29, 130.73, 130.43, 129.61 – 129.41, 127.92, 113.85, 96.35, 66.08, 65.96 – 65.34, 65.14, 61.22, 56.35, 51.03, 46.12, 40.24, 38.50, 34.57, 34.23 – 33.72, 31.94, 29.71, 28.75, 24.60, 22.70, 14.14, 12.66. HRMS m/zcalcd for (M+) C₅₂H₆₁N₂O₇S₂Si, 917.3684; found 917.3688

Figure S9. IR spectrum of compound 1.

Figure S11. IR spectrum of compound 2.

Figure S13.¹³C NMR of RdBS.

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Figure S14. HRMS of RdBS.

Figure S15. IR spectrum of RdBS.

Figure S16. IR spectrum of the product after the reaction of RdBS with F.

The Matlab program for calculating the plasmon resonance scattering intensity for one nanoparticle.

```
file = 'sample.jpg';image = imread(file);imgGray = mat2gray(image, [0 65536]);bins = 100;
```
% findSpots is a function for searching % the scattering spots in the image $spots = findSpots(imgGray);$

```
% estimate the intensity of each spot 
% by averaging the gray value of pixels in the spot
count = length(spots);intensity = zeros(count, 1);for i = 1: count
     positions = spots\{i\};
     l = length(positions);pixelIntensity = zeros(1,1);for j = 1:1pixelIntensity(j) = imgGray(positions(j,1), positions(j,2)); end
```

```
intensity(i) = mean(pixelIntensity);
```
end

% use histogram to distribute the intensity of spots $[h,x]$ = hist(intensity,bins); xlabel('intensity'); ylabel('Counts');

Reference

[1] L. Shi, C. Jing, W. Ma, D.-W. Li, J. E. Halls, F. Marken, Y.-T. Long, *Angew. Chem. Int. Ed.* 2013, *52*, 6011-6014.