Supplementary Data

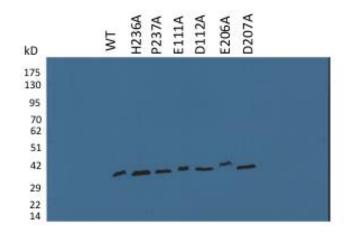
Identification and biochemical characterization of a novel α2,3-sialyltransferase WbwA from pathogenic *Escherichia coli* serotype O104 Diana Czuchry, Paul Desormeaux, Melissa Stuart, Donald L. Jarvis, Khushi L. Matta, Walter A. Szarek, and Inka Brockhausen

Supplementary Figure 1. Sequence comparisons of WbwA^{O104} and sites of mutations. Alignments of sialyltransferases were done using ClustalW2 (35) and were adjusted slightly so that the HP motifs were aligned. The alignments were edited using GeneDoc (36, http://www.psc.edu/biomed/genedoc). WbwA was from *E. coli* O104, NST from *Neisseria meningitidis*, WaaH from *Salmonella enterica*, PM0188 from *Pasteurella multocida* and ST3Gal-I was human. HP mutation sites for WbwA are shown in light blue, whereas the HP motifs in other sequences are shown in dark blue. The ED mutations for WbwA are shown in coral red, whereas the E(D)E(D)G motifs in other sequences are shown in pink. ST3Gal1 has an HP sequence but does not have an E(D)E(D)G motif.

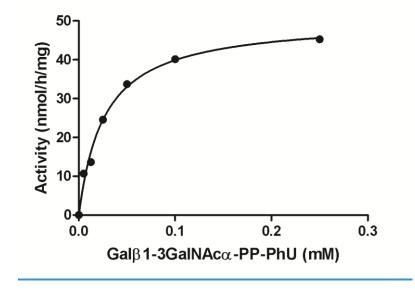
WbwA : NST : WaaH : PM0188 : ST3GalI :				ITLYLDPASLI	/NQGERNAV: PALNQLMDF1	SLLKDKLFN <mark>EI</mark> QNNEDKTHPI	MFLFICM RIFGLSRFKIP	TILQMKVAER: TNLQLLIARS: DNIITQYQNII	IMAQ <mark>HP</mark> GERI IIEKEQLKS\ HFVELKDNRI	YVVLMSENRI /DILFIGDVHI ?TEALFTILD(100 SLRDINFPG WEKYDYYFNQIKD WVKNQYYLKKIQP QYPGNIELDIHLN QWVLELSENLKRL	: 109 : 49 : 109
NST : WaaH : PM0188 :	KAERAYI LCRHSS IAHSVQ	FFHLPYGLNKS IVSQVSKFAAE LIRPILAYRFF	SFNFIPTMAEL KTIHRTRYAK KHLDRVSIQRL	KVKAMLLPKVI KIMESYAKEYI NLY <mark>DDG</mark> SMEYY	KRIYLASLEH HTVFFANFHN /DLEKEENKI	VSIAAFLST PLIHHILSC DISAEIKQAEI	YPDAEIKTFDD ISFSEINTFDD KQLSHYLLTGK	GTGN- GTNN- IKFDNPTIAR	YVWQSAF PVH	-LIQSSSYLG INQKSII (YHFLSTDYF)	* IGGHTFGRSTKVN DEFSVNGTIKRNF MYENKNISSTSKL EKAEFLQPLKEYL NLRESSYGPEIDS	: 202 : 139 : 218
NST : WaaH : PM0188 :	ARMMIG IRKLMG AENYQKI	DWSIAKTRNAS RKYHKDEILKI MDWTAYQQLTE	SDEHYTIFKGL JDAKDYTLFPN PEQQAFYLTLV	KNIM <mark>DDG</mark> RRKI RTNIIEKTEG: GFNDEVKQSLI	MTYLPLFDA: IILVHHNGLI SVQQAKFIF1	SELKAGDETG PDTNNGFKKVI SGTTTWEGNTI	STVRILLGSPD LLGTVYTDALK DVREYYAQQQLI	KEMKEISEKA. NKEDECVFLQI NLLNHFTQAG(AKNFNIQYVA HLQRFIKKKA GDLFIGDHYA	AP <mark>HP</mark> AVDIYIPHPR XIYFKG- <mark>HP</mark> R(320 ELTDYSLYFKDVK RQTYGLSG YDSHQFNGVLNVN GGEINDYILNNAK FIKYVFDNWLQGH	: 300 : 248 : 326
NST : WaaH :	VTTLNS SEMIAE NITNIP	PYVIEDYILRE DIILEYLEQG- ANISFEVLMMI	IKKNPHTRYE -MSLEIYGFN GLLPDKVGGV	IYTFFSGAAL STVQYNLNNI ASSLYFSLPKI	CMKDFPNVHV STIKNYKITS SKISHIIFTS	/YALKPASLPI SP-FLKDSFNI SNKQVKSKED/	* I SNKFEVKGKK SDYWLK PVYAL IGLGFDFNQVS ALNN PYVKVMR FE SNVTATLAS	FTQSGIPILT) V RLGIIDESQV:	FDDKD IFWDSLKQL-	- : 313 - : 412		

Supplementary Figure 2

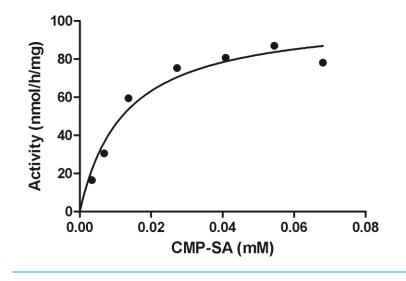
Western blot of wild type and mutant WbwA. Lysates of bacteria containing wild type WbwA (WT) and six WbwA mutants wer run on SDS-PAGE and visualized on a Western blot after treatment with mouse anti-His antibody, goat anti-mouse antibody linked to horseradish peroxidase, and Lightning Plus ECL Chemiluminescence Substrate. All proteins show only one band at about 38 kDa in mass.



Supplementary Figure 3A. Kinetics of WbwA^{O104} using acceptor Gal β 1-3GalNAc α -PP-PhU and bacterial lysate. Assays for SiaT WbwA were carried out as described in the Methods section. The CMP-Sia concentration in the assays was 0.8 mM. The apparent K_M for the acceptor Gal β 1-3GalNAc α -PP-PhU was 0.02 mM and the apparent V_{max} was 50 nmol/h/mg.

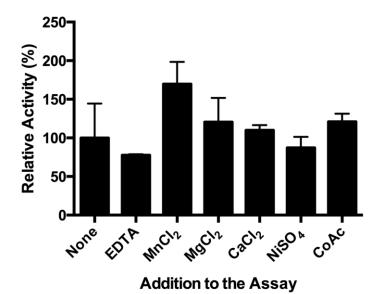


Supplementary Figure 3B. Kinetics for CMP-sialic acid of WbwA^{O104} using acceptor Gal β 1-3GalNAc α -PP-PhU and bacterial lysate. Assays for SiaT WbwA were carried out as described in the Methods section. The acceptor concentration in the assays was 0.1 mM. The apparent K_M value for CMP-sialic acid (CMP-SA) was 0.013 mM and the apparent V_{max} was 90 nmol/h/mg.

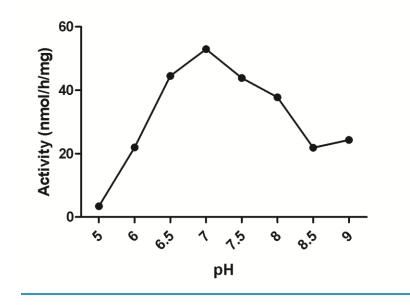


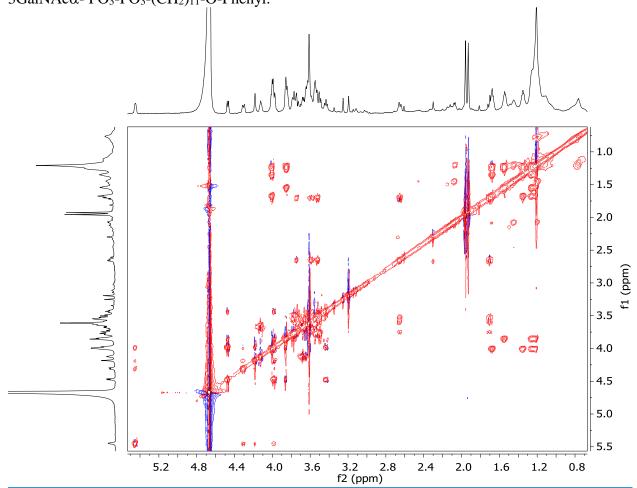
Supplementary Figure 4. Metal ion dependence of WbwA^{O104} activities.

Sialyltransferase assays were carried out using CMP-sialic acid donor and Gal β 1-3GalNAc-PP-PhU acceptor as described in the Methods section. Metal ions were added at 5 mM concentration to the assays. Assays with EDTA at 5 mM and no addition (None) served as negative controls. CoAc, Cobalt acetate. No metal ions were required but Mn²⁺ showed a slight stimulation of activity.

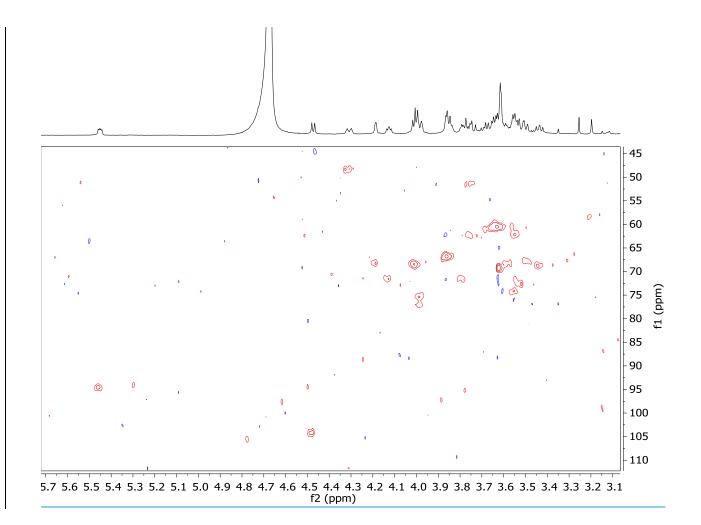


Supplementary Figure 5. WbwA^{O104} activity at different buffer pH values. Sialyltransferase activity of WbwA^{O104} was assayed as described in the Methods section at different buffer pH values.





Supplementary Figure 6A. 600 MHz TOCSY NMR spectrum of WbwA^{O104} product Sialyl α 2-3Gal β 1-3GalNAc α - PO₃-PO₃-(CH₂)₁₁-O-Phenyl.



Supplementary Figure 6B. 600 MHz HSQC NMR spectrum of WbwA⁰¹⁰⁴ product Sialyla2-3Gal β 1-3GalNAca-PO₃-PO₃-(CH₂)₁₁-O-Phenyl.

Enzyme	GT family	UniProt #	%Identity
β3-Gal-transferase, WbwC (E. coli O104)	2	Q93NP5	11.8
α4-Gal-transferase, WbwB, putative (E. coli O104)	4	Q93NP7	14.7
α 3-SiaT, ST3Gal1, human	29	Q11201	13.6
α3-SiaT, CstI (Campylobacter jejuni)	42	D2KQ02	13.1
α3/8-SiaT, bifunctional, CstII (<i>Campylobacter jejuni</i>)	42	Q9L9Q5	12.7
α3-SiaT, WbwA (<i>E. coli</i> O104)	52	Q93NP9	100
α3/6-SiaT, NST bifunctional (Neisseria meningitidis)	52	C6S6J1	12.4
α3-SiaT, (Neisseria gonorrhea)	52	P72074	14.2
α3-SiaT (Neisseria meningitidis)	52	P72097	13.0
α 3-SiaT, Lst, PMST2 (<i>Pasteurella multocida</i>)	52	Q9CNC4	14.3
α2-Glc-transferase, WaaH (Salmonella enterica)	52	Q7B6E0	14.6
α 3-SiaT, PdST, multifunctional (<i>Pasteurella dagmatis</i>)	80	C9PR45	12.9
α 3-SiaT (Photobacterium phosphoreum)	80	A5LHX0	13.2
α6-SiaT, PM0188 (Pasteurella multocida)	80	Q9CP67	14.8
<u>α6-SiaT (Photobacterium sp)</u>	80	A8QYL1	11.7

Supplementary TABLE I. Glycosyltransferases potentially related to WbwA

Glycosyltransferases are listed that are encoded by the *E. coli* O104 gene cluster (WbwC, WbwA, WbwB), as well as other bacterial mono- and multi-functional sialyltransferases, possibly related to WbwA, from CAZy GT families 42, 52 and 80, as well as a Glc-transferase from the GT52 family. Human sialyl-T synthase (ST3Gal1) is also included. Uniprot alignments were used to determine the percentage of sequence identity with WbwA from *E. coli* O104. The UniProt ID numbers are shown. The identities of these enzymes with WbwA are low. With the exception of putative Gal-transferase WbwB, all of the bacterial enzymes have an HP sequence, and with the exception of Gal-transferase WbwC, all of the bacterial enzymes have at least one (E(D)-E(D)-G motif.

Compound	Number of carbons	Inhibition (%)		
		WbwA	ST3Gal1	
QT147, 1,4-bis(3-methyl-1H-imidazolium-1-				
yl)butane dichloride	4	10	<1	
QT141, 1,8-bis(3-methyl-1H-imidazolium-1-				
yl)octane dichloride	8	12	<1	
QT142, 1,14-bis(3-methyl-1H-imidazolium-1	-			
yl)tetradecane dichloride	14	19	<1	
QT140, 1,15-bis(3-methyl-1H-imidazolium-1	-			
yl)pentadecane dichloride	15	48	<1	
QT138, 1,16-bis(3-methyl-1H-imidazolium-1	-			
yl)hexadecane dichloride	16	66	>90	
QT136, 1,18-bis(3-methyl-1H-imidazolium-1	-			
yl)octadecane dichloride	18	64	>90	
QT135, 1,20-bis(3-methyl-1H-imidazolium-1	-			
yl)eicosane dichloride	20	90	94	
QT148, 1,20-bis(3-methyl-1H-imidazolium-1	-			
yl)eicosane dimesylate	20	96	>99	
QT149, 1,22-bis(3-methyl-1H-imidazolium-1	-			
yl)docosane dimesylate	22	88	>99	

Supplementary TABLE II. Inhibition of ST3Gal1 and WbwA activities by bis-imidazolium salts

Sialyltransferase assays using purified ST3Gal1 and WbwA in bacterial lysates were performed as described in Material and Methods. Inhibitor concentration in the assays was 0.5 mM. All assays contained 10% MeOH. The concentration of ST3Gal1 acceptor Gal β 1-3GalNAc α -Bn was 0.5 mM. For WbwA assays, the concentration of the acceptor Gal β 1-3GalNAc α -PO₃-PO₃-(CH₂)₁₁-O-phenyl was 0.1 mM. The percentage of inhibition compared to control activity without inhibitor is shown. Number of carbons, the length of the aliphatic carbon chain linking the two imidazolium groups in the chloride or mesylate salts.